Review

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Biocontrol genes from Trichoderma species: A review

Pratibha Sharma*, Vignesh Kumar P, Ramesh R, Saravanan K, Deep S, Sharma M,
Mahesh S and Dinesh S

Division of Plant Pathology, Indian Agricultural Research Institute, New Delhi-110012, India.

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In the world, the traditional agricultural practices are getting affected by various problems such as disease, pest, drought, decreased soil fertility due to use of hazardous chemical pesticides, pollution and global warming. As a result, there is a need for some eco-friendly biocontrol agents that help in resolving the previous mentioned problems. The various types of biological control agents such as bacteria and fungi are involved in biocontrol activity. Among them, fungal genus *Trichoderma* plays a major role in controlling the plant diseases. The species of *Trichoderma* are known to produce different kinds of enzymes which have a significant role in biocontrol activity like cell wall degradation, biotic and abiotic stress tolerance, hyphal growth, antagonistic activity against plant pathogens. By the advance techniques laid in the molecular biology, we can easily isolate, characterize, clone, sequence and express the functions of these genes and can study their functions and role in the biocontrol mechanism. This review article explains about the role, and functions of some major biocontrol genes present in the *Trichoderma* species viz., *Trichoderma harzianum*, *Trichoderma viride*, *Trichoderma atroviride*, *Trichoderma hamatum* and *Trichoderma longibrachiatum*.

Key words: *Trichoderma* spp., genes, cloning, transformation, expression.

INTRODUCTION

The species of *Trichoderma* are well known for their biocontrol activity against many plant pathogens that cause major problems in the current agricultural scenario. Trichoderma species are known for their production of cell wall degrading enzymes which can be used for commercial productions. All living organisms are made up of genes that code for a protein which performs the particular functions. Genes play a major role in the biocontrol process by regulating some signals and lead to the secretion of some enzymes or proteins that help in the degradation of the pathogens and hence they are known as biocontrol genes. Increased expression of the genes helps in enhanced biocontrol activity which helps in promoting the plant growth and prevents the plant from pathogen attack. So the biocontrol genes can be cloned and produced in large amounts for commercial applications (Massart and Jijakli, 2007).

Some genes of *Trichoderma* species can be used to provide resistance to the biotic and abiotic stresses such

as salt, heat and drought (Kuc, 2001). Before using as a commercial product, the biocontrol mechanism of a particular species should be well known (Grondona et al., 1997). The major biocontrol process involves antibiosis, providing plant nutrition and mycoparasitism (Janisiewicz and Korsten, 2002).

Trichoderma species are used widely as biocontrol agents because they have more benefits on plant growth such as promoting plant growth, increasing the nutrient uptake from the soil, and decreasing the activity of the soil borne pathogens that ultimately affect the growth of the plant (Harman et al., 2004). Among the various species of the *Trichoderma*, *Trichoderma harzianum* is considered to be the most effective biocontrol agent (Gao et al., 2002).

BIOCONTROL GENES AND THEIR FUNCTIONS FROM TRICHODERMA SPECIES

The genus *Trichoderma* acts as a biocontrol agent due to its feasible character in fighting against the pathogens. Some major kinds of biocontrol genes that can be easily

^{*}Corresponding author. E-mail: pratibha@iari.res.in.

isolated, cloned and characterized are protease, chitinase, glucanase, tubulins, proteinase, xylanase, monooxygenase, galacturonase, cell adhesion proteins and stress tolerant genes. These genes have their unique functions in the biocontrol mechanism such as cell wall degradation, hyphal growth, stress tolerance, and parasitic activity. Tubulins are structural proteins made of microtubules and they help in studying the cell wall composition of the pathogens (Li et al., 2010). Chitinase helps in the breakdown of the glycosidic bonds. Glucose oxidase catalyses D-glucose to D-glucono-1,5-lactone and hydrogen peroxide are known to have antifungal effect (Ciliento et al., 2004). Xylanase helps in breaking hemicellulose a major component of plant cell walls (Figure 1).

FUNCTIONS OF BIOCONTROL GENES

Cell wall degradation

From *Trichoderma virens*, a gene named *tvsp1* encoding for serine protease was cloned successfully and its function was analyzed. Serine protease has an important role in pathogenesis or biocontrol activity against *Rhizoctonia solani* which affects the cotton seedlings. The gene *tvsp1* was expressed in *Escherichia coli* and cloned using pET-30 vector. Thus, serine protease helps in degrading the fungal cell wall (Pozo et al., 2004).

In *T. harzianum* trichodiene synthase gene *tri*5 was isolated and characterized. This *tri*5 gene was responsible for the synthesis of the enzyme trichothecene which inhibits the protein and DNA synthesis in the cells of the pathogens and inhibits their growth. The trichothecene shows phytotoxic activity against *Fusarium* species. The gene *tri*5 was isolated and by designing of specific primers. The sequence was inserted into pGEM-T vector, cloned and expressed.

The presence of *tri*5 gene was confirmed by screening with other *Trichoderma* isolates (Gallo et al., 2004). The expression of gene *tag83* which encodes cell wall degrading enzyme exo-β-1,3-glucanase was isolated from *Trichoderma asperellum* and characterized. The expression analysis of this gene was studied using real time and reverse transcription-polymerase chain reaction (RT-PCR). The enzyme activity of glucanase was studied by comparing with various types of carbon sources like starch, cellulose, chitin, chitosan and cell walls of *R. solani*. The expression of *tag83* gene with *R. solani* showed that glucanase enzyme exhibits parasitic activity against pathogens (Marcello et al., 2010).

T. virens transformants expressed two different kinds of β -1,3 and β -1,6 glucanase genes viz., TvBgn2 and TvBgn3. These genes secrete cell wall degrading enzyme that helps in the biocontrol activity. T. virens GV29.8 wild type and double over expression (DOE) transformant strains were used to detect the enzyme activity against

pathogens like *R. solani*, *Pythium ultimum* and *Rhizopus oryzae* (Djonovic et al., 2007).

A gene, *gluc78* which codes for an antifungal glucan 1,3-β-glucosidase was isolated, cloned and sequenced from Trichoderma atroviride. This gene has its significance in the cell wall degradation of the pathogens. The gene *gluc78* was cloned in pGEM-T vector and the expression analysis was done against pathogens such as R. solani and P. ultimum (Donzelli et al., 2001). From T. harzianum, a glucose repressor gene crel was isolated and characterized. This gene causes the repression of cellulase and xylanase encoding genes. Cellulase and xylanase are the major type of enzymes that involve in the cell wall degradation of the pathogens. The gene was cloned using pTZ57R/T plasmid vector and transformed into E. coli DH 10B and the role of crel gene in cellulase and xylanase expression was studied (Saadia et al., 2008).

 β -Tubulins are structural components of most cells and they interact with benzimidazole fungicides, and play a major role in biocontrol process. This β -tubulin gene was isolated and characterized from T. harzianum. The β -tubulin gene was amplified by PCR, the coding regions and the flanking sequences were identified using inverse and nested PCR. The sequences were analyzed for the presence of motifs for the expression of the gene. The three dimensional model of β -tubulin gene was done by Swiss-model automated comparative protein modeling server (Li and Yang, 2007). From T. virens, a gene, Sm1 a cysteine-rich protein was isolated and expressed. It shows defense activity against diseases in dicot and monocot plants (Buensanteai et al., 2010).

Serine proteases play a key role in the fungal biology and involves in biocontrol activity. From *T. harzianum* a novel serine protease gene named SL41 has been cloned and expressed successfully in *Saccharomyces cerevisiae*. The cDNA of SL41 gene was sequenced and it was cloned in pMD18-T vector and the yields were inserted into *E. coli* DH5- α . Thus, serine proteases were cloned and characterized (Liu et al., 2009).

The gene, *ThPG1* which encodes for endopoly-galacturonase was isolated from *T. harzianum* and characterized. This enzyme involves in the cell wall degradation of the pathogens like *R. solani* and *P. ultimum* and helps in the plant beneficial interactions. The expression study of this gene was studied by comparing the wild and mutant type strains.

The full length cDNA clone of *ThPG1* gene was obtained by polymerase chain reaction and was cloned in pSIL-pG1 vector. The phylogenetic relationship was obtained by Neighbor-joining (NJ) tree method (MoranDiez et al., 2009). A gene, Tv6Gal which codes for endo- β -(1 \rightarrow 6)-galactanase gene was isolated from *T. viride*, cloned and expressed in *E. coli*. Galactanase enzymes belong to the family of arabinogalactan proteins that involve in cell-cell adhesion, cell expansion and cell death. The cDNA clone of the gene Tv6Gal was done by

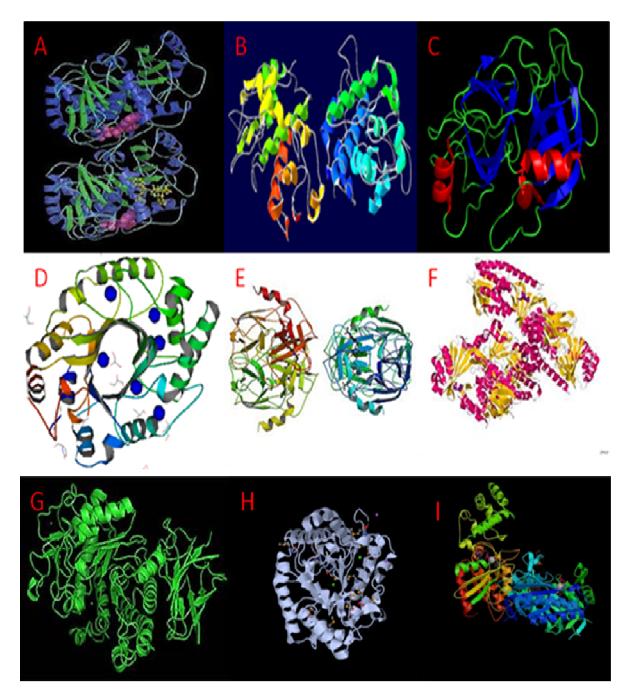


Figure 1. Structure of biocontrol genes: **A)** tubulin, **B)** chitin, **C)** protease, **D)** xylanase, **E)** proteinase, **F)** monooxygenase, **G)** β -endogalactanase, **H)** β -endoglucanase, and **I)** adenylate-Cyclase. (www.google.co.in/images). From *T.harzianum* gene encoding for endochitinase and β -tubulin has been cloned and characterized successfully using pGEM-T vector ongoing work (senior author).

RT-PCR, cloned in pGEM-T vector and expressed in *E. coli* (Kotake et al., 2004).

Xylanase producing *Trichoderma* strain SY was isolated from the soil. The gene coding for xylanase, *Xyl* was cloned by RT-PCR. *Xyl* was highly expressed when it was grown in cellulose as an only source of carbon. The full length cDNA of *Xyl* was amplified by PCR and

cloned in pGEM-T vector. The cloned gene was expressed in *E. coli* and the proteins were analyzed using sodium dodecyl sulphate poly acrylamide gel electrophoresis (SDS-PAGE) (Min et al., 2002). From *T. virens*, the g-protein α subunits genes, *TgaA* and *TgaB* were cloned and characterized. This gene exhibits antagonist activity against *R. solani* and *Sclerotium rolfsii*

(Mukherjee et al., 2004) (Table 1).

Biotic and abiotic stress tolerances

Species of *Trichoderma* helps the plant to survive in the abiotic stress conditions. From *T. harzianum* T34 isolate, *hsp*70 gene was cloned and characterized. This gene helps in increasing the fungal resistance to heat and other stresses such as salt tolerance, osmotic and oxidative tolerances. The protein sequences were analyzed using DNAstar package and aligned using CLUSTAL X algorithm. The *hsp*70 gene was cloned in pGEM-T vector and expressed in different isolates of *T. harzianum* (ManteroBarrientos et al., 2008).

From the fungus *T. harzianum*, a gene named *Thkel1* was isolated and characterized. This gene codes for putative kelch-repeat protein which helps in regulating the glucosidase activity and enhances tolerance to salt and osmotic stresses in Arabidopsis thaliana plants. The vector used for cloning was p*SIL-KEL* and was transformed to *T. harzianum*. The expression of this gene was studied by growing the fungal at various biotic and abiotic stress conditions (Hermosa et al., 2011). From *T. virens* glutathione transferase gene *TvGST* was cloned. When transgenic plant expresses this gene against different concentrations of cadmium, it shows tolerance to cadmium accumulation in plants. Thus it acts as cadmium tolerant gene (Dixit et al., 2011).

Mycoparasitism

From 31 isolates of *T. harzianum*, five have been selected namely (T 30, 31, 32, 57 and 78) and from them genes encoding for N-acetyl-B-D-glucosaminidase (exc1 and exc2), chitinase (chit42 and chit33), protease (prb1) and β-glucanase (bgn 13.1) were cloned and expressed. These genes play a major role in the mycoparasitic activity against the pathogens especially Fusarium oxysporum. The expressions of these genes that codes for these enzymes were determined by RT-PCR and their effects against the pathogens were tested by dual plate assay (LopezMondejar et al., 2011). In T. virens, an adenalyte-cyclase encoding gene named tac1 gene was isolated and cloned. This gene has its role in mycoparasitic activity against R. solani and P. ultimum (Mukherjee et al., 2007). ThPTR2 a di or tri peptide transporter gene isolated from T. harzianum CECT 2413 has a significant role in the mycoparasitic activity against Botrytis cinerea.

The cDNA of *ThPTR2* gene was obtained through reverse transcript polymerase chain reaction, transferred into plBRC43 plasmid, cloned and expressed. The sequences were aligned using CLUSTAL-W algorithm and protein binding motifs were discovered. The mycoparasitic expression of the *ThPTR2* gene was analyzed

by dual culture assay (Vizcaino et al., 2006). The gene, qid74 isolated from *T. harzianum* CECT 2413 was found to play a significant role in cell protection and provide adherence to hydrophobic surfaces that helps the fungus in mycoparasitic activity against *R. solani* pathogen. The function of this gene was studied by comparing the expression of genes in wild type transformants and disruptants. The results showed that qid74 gene was responsible for adhesion to the hydrophobic surfaces of the pathogenic fungi and helps in the antagonistic activity (Rosado et al., 2007).

A gene named, *Taabc2* was cloned from *T. atroviride* and characterized. This gene has a significant role in ATP binding cassette (ABC) transporter in cell membrane pump that helps in the mycoparasitic activity. The expression of this gene was found to be more when they uptake the nutrients from the pathogenic fungi. The gene was cloned using pGEM-T vector, expression of the genes were analyzed using RT-PCR.

The antagonist activity against pathogens such as *R. solani*, *B. cinerea*, and *P. ultimum* was done by dual culture plate assay with *T. atroviride* wild and mutant type strains (Ruocco et al., 2009). From *T. harzianum* genes encoding for proteinase *prb1* and endochitinase *ech42* were isolated and characterized. These genes involved in the mycoparasitic activity against *R. solani* and *S. rolfsii*. For the production of these enzymes, the genes were induced by lectin-carbohydrate interaction a diffusible factor. This factor regulates the production of proteinase and endochitinase which helps in the mycoparasitism (Cortes et al., 1998).

In *Trichoderma hamatum* the expression of mycoparasitism genes, namely chitinase *chit42* and proteinase *prb1* were analyzed. The expressions of these genes were analyzed by confrontation assay against the plant pathogen *Sclerotinia sclerotiorum*. During sequence analysis the presence of motifs was discovered and that helps in the regular expression of the genes that enhances the parasitic activity against pathogens (Steyaert et al., 2004).

T. longibrachiatum transformants showed over expression of β-1,4-endoglucanase gene egl1. This gene showed biocontrol activity against P. ultimum in damping-off of cucumber. The egl1 gene, coding for endoglucanase was isolated from T. longibrachiatum, cloned and expressed in Saccharomyces cerevisiae. The expression of the gene was compared with the wild type and transformed strains. The results showed that the over expression of egl1 gene showed good biocontrol activity (Migheli et al., 1998). TmkA, mitogen activated protein kinase from T. Virens is known to cause mycoparasitic activity against R. solani and S. rolsfii (Mukherjee et al., 2003).

Antifungal activity

A transcription factor gene named Thctf1 was isolated

Table 1. List of biocontrol genes isolated from different *Trichoderma* species and their functions.

S/N	Name of the isolate and biocontrol gene	Function	Strain identification by DNA sequence analysis with accession numbers	Effect of the gene in biocontrol	Author	Year
1	T.harzianum strain IMI206040 (proteinase prb1 and endochitinase (ech42 genes)	Parasitic activity against Sclerotium rolfsii and Rhizoctonia solani.	Accession numbers not available	Expression of this gene helps in regulation of hydrolytic enzymes.	Cortes et al.	1998
2	T.longibrachiatum wild type strain CECT2606 (β-1,4-endoglucanase gene, egl1)	Biocontrol activity against <i>Pythium ultimum</i> on cucumber.	Accession numbers not available	Shows enhanced biocontrol activity.	Migheli et al.	1998
3	T.harzianum strain P1 74058 (ech42 gene)	Biocontrol activity against Botrytis cinerea and R. solani	Accession numbers not available	Disruption of this gene affects the biocontrol activity.	Woo et al.	1999
4	<i>T.atroviride</i> strain P1 (ATCC 74058) (1,3-β-glucosidase gene, <i>gluc78</i>)	Cell wall degradation of pathogens <i>Pythium</i> and <i>Phytophthora</i> .	GenBank AF253421	Exhibits moderate biocontrol activity.	Donzelli et al.	2001
5	Trichoderma strain SY (Xylanase gene Xyl)	Helps in breakdown of hemicellulose.	GenBank AY156910	Only gene isolation	Min et al.	2002
6	T.virens strain IMI 304061 (TmkA Mitogen Activated Protein kinase gene)	Biocontrol activity against pathogens like S. rolfsii and R. solani.	GenBank AY141978	This gene represses the conidial formation of <i>R. solani</i> .	Mukherjee et al.	2003
7	T.harzianum strain ATCC 90237 (trichodiene synthase <i>tri</i> 5 gene)	A toxic secondary metabolite which is responsible for inhibiting DNA or protein synthesis, and enhances virulence against Fusarium spp.	DDBJ/EMBL/ Gen bank accession number is AJ 784992.	Increases the virulence against <i>Fusarium</i> spp.	Gallo et al.	2004
8	T.virens strain IMI 304061 (TgaA, TgaB genes)	Antagonism against S. rolfsii and R. solani.	GenBank AY186729 (tgaA) and AY168002 (tgaB).	Increases virulence in the plant pathogenic interactions.	Mukherjee et al.	2004
9	<i>T.virens</i> wildtype strain Gv29-8 and an arginine auxotrophic strain, Tv10.4 (<i>tvsp</i> 1 serine protease encoding gene)	Involved in pathogenesis or biocontrol process of <i>R. solani</i> .	GenBank AY242844	Exhibits moderate activity against <i>R. solani</i> .	Pozo et al.	2004
10	T.hamatum strain LU593 (chitinase chit42 and proteinase prb1 gene)	Mycoparasitic activity against Sclerotinia sclerotiorum.	GenBank ITS1-AY241456, chit42-AY258898, prb1-AY258899, xbg1.3-110-AY269826	Exhibits moderate biocontrol activity.	Steyaert et al.	2004

Table 1. Continue.

11	T.viride IFO31137 (endo- $β$ -1-6-glactanase gene)	A type of arabinogalactan proteins that involves in cell-cell adhesion, expansion and cell death.	Accession number not available	Expression of gene enhances the production of proteins.	Kotake et al.	2004
12	T.atroviride strain P1ATCC 74058 (tga1 gene)	Chitinase formation and production of antifungal metabolites.	GenBank AY190117	Increases the antifungal activity.	Reithner et al.	2005
13	T.virens IMI 304061 (tmkA gene)	Induction of plant systemic resistance and biocontrol activity against $R.\ solani$. (Tested in green house condition)	Accession number not available	Shows increased biocontrol activity.	Viterbo et al.	2005
14	T.harzianum CECT 2413 (ThPTR2 gene)	Di/tri peptide transporter gene, that involves in mycoparasitic activity against <i>B. cinerea</i> .	GenBank AJ868360	Induces peptide transport that enhances mycoparas-istism	Vizcaino et al.	2006
15	T.harzianum CECT 2413 (erg1 gene)	Silencing of the <i>erg1</i> gene enhances resistance to terbinafine that shows antifungal activity.	GenBank AM050097	Shows enhanced biocontrol activity.	Cardoza et al.	2006
16	T.harzianum Rifai CECT 2413 (qid74 gene)	Involved in cell protection and adherence to hydrophobic surfaces that helps in antagonism against <i>R.solani</i> .	Accession number not available	Shows moderate biocontrol activity.	Rosado et al.	2007
17	T.virens Gv29-8 (TvBgn2 and TvBgn3 genes)	These genes help in encoding cell wall degrading enzymes.	Accession number not available	Shows enhanced biocontrol activity.	Dzonovic et al.	2007
18	T.virens IMI 304061 (tac1, adenylate cyclase gene)	Mycoparasitism against <i>R.solani</i> , <i>S.rolfsii</i> , <i>Pythium</i> Spp. and production of secondary metabolism.	Accession number not available	Shows reduced biocontrol activity.	Mukherjee et al.	2007
19	T.harzianum T88 (beta tubulin gene)	Exhibits biocontrol activities like mycoparasitism, and antifungal activity.	GenBank EF027167	Shows moderate biocontrol activity.	Li et al.	2007
20	T.hamatum LU593 (monooxygenase gene)	Antagonist activity against S. sclerotiorum, S. minor and S. cepivorum.	GenBank EU124654	Shows enhanced biocontrol activity.	Carpenter et al.	2008
21	T.harzianum E58 (CRE1 gene)	Production of cellulase and hemicellulase enzymes.	Accession number not available	Shows enhanced biocontrol activity.	Saadia et al.	2008
22	T.harzianum CECT 2413 (T34 hsp70)	Increases fungal resistance to heat and abiotic stresses.	GenBank EU311400	Shows increased biocontrol activity.	MonteroBarrientos et al.	2008

Table 1. Continue.

23	T.harzianum (Th-Chit gene)	Antifungal activity in transgenic tobacco.	GenBank AJ605116	Shows enhanced biocontrol saiprasad et al. 2	2009
24	T.atroviride P1 (ATCC 74058) (Taabc2 gene)	Plays key role in antagonism against R.solani, P.ultimum, and B.cinerea.	Accession number not available	Shows decreased biocontrol Ruocco et al. 2	2009
25	T.reesei QM9414 (ATCC 26921) (TrCCD1 gene)	Helps in hyphal growth, conidiospore development and carotenoid pigment production.	GenBank EF407410	Shows increased biocontrol HuaZhong et al. 2 activity.	2009
26	T.harzianum CECT 2413 (Thctf1transcription factor gene)	Antifungal activity against R.solani, Fussarium oxysporumand B.cinerea and production of 6-pentyl-2H-pyran-2.	GenBank EU551672	Shows enhanced biocontrol Rubio et al. 2 activity.	2009
27	T.harzianum (serine protease gene SL41)	Biocontrol activity against pathogens.	GenBank DQ910533	Shows enhanced biocontrol Liu et al. 2	2009
28	T.harzianum T34 CECT 2413 (endopolygalacturonase ThPG1 gene)	Secretion of plant cell wall degrading enzymes against <i>R. solani</i> and <i>P. ultimum</i> .	GenBank AM421521	Shows enhanced biocontrol MoranDiez et al. 2	2009
29	T.asperellum (Enzymology Group collection,UFG-ICB) (tag 3 gene)	Production of cell wall degrading enzyme glucanase.	Accession number not available	Shows significant biocontrol Marcello et al. 2	2010
30	T.virens strain TvSMOE38 (Sm1 gene, cysteine-rich protein)	A small cysteine-rich protein that induces defense responses in dicot and monocot plants and in protecting crop diseases.	Accession number not available	Shows enhanced biocontrol Buensanteai et al. 2	2010
31	T.virens (strain IMI 304061) (TvGST glutathione transferase gene)	Enhances cadmium tolerance in plants.	GenBank EH628505	Increases the biocontrol activity. Dixit et al. 2	2011
32	T.brevicompactum IBT40841 (tri5 gene)	Production of trichodermin and antifungal activity against <i>C. albicans</i> , <i>C. glabrata</i> and <i>A. fumigatus</i> .	Accession number not available	Shows enhanced biocontrol Tijerino et al., 2	2011
33.	T.harzianum CECT 2413 (Thke11 gene)	Expression of this gene in <i>A.thaliana</i> modulates glucosidase activity, and enhances tolerance to salt and osmotic stresses.	GenBank EU399786	Enhances the biocontrol activity. Hermosa et al. 2	2011
34.	T.harzianum CECT 2413 (genes encoding for NAGases (exc1 and exc2), chitinases (chit42 and chit33), proteases (prb1) and b-glucanases (bgn13.1)	Mycoparasitic activity against F. oxysporum.	Accession number not available	Shows enhanced biocontrol Mondezar et al. 2 activity.	2011

from *T. harzianum*. The gene involves in the production of 6-pentyl-2*H*-pyran-2-one (6-PP) and shows antifungal activity against pathogens such as *R. solani*, *B. cinerea*, and *S. rolfsii*. The sequences were analyzed using Lasergene package and cloned using pGEM-T vector (Rubio et al., 2009).

T. brevicompactum encodes, tri5 a trichodiene synthase gene. The over expression of this gene helps in the production of trichodermin which shows antifungal activity against S. cerevisiae, Kluyveromyces marxianus, Candida albicans, C. glabrata, C. tropicalis and Aspergillus fumigatus. The sequences were analyzed using DNAstar package and aligned using CLUSTAL-X algorithm for analyzing the phylogenetic relationship. The gene tri5 was cloned in pURSPT5 and transformed into T. brevicompactum (Tijerino et al., 2011).

From *T. harzianum*, endochitinase gene named *Th-Chit* was isolated, characterized and that gene confers antifungal activity in transgenic tobacco plant. Chitinase are one of the cell wall degrading proteins that help in the antifungal activity. The gene, *Th-Chit* was cloned using pTZ57R vector, and sequencing of the cloned cDNA was done by ABI prism automated DNA sequencer. From this the full length chitinase gene was isolated and then it is cloned in a binary vector named pIIHR-Th-Chit. The gene was transferred to tobacco plant and their presence was analyzed by polymerase chain reaction amplification from the control and transformed plants. Thus, Th-Chit gene confers antifungal activity against *A. alternata* (Saiprasad et al., 2009).

The *erg1* gene from *T. harzianum* was cloned and characterized. This gene encodes an enzyme named squalene epoxidase, which helps in the synthesis of ergosterol and silencing of this gene provides resistance to terbinafine, an antifungal compound. The antifungal activity was checked with *Saccharomyces cerevisiae*. pSIL-E1 vector was used to clone the gene *erg1*. Sequencing was done by DNAstar package and aligned using CLUSTAL-W algorithm. This is the first terpene biosynthesis gene characterized from *Trichoderma* genus (Cardoza et al., 2006).

Tga1 gene, the G protein α subunit of *T. atroviride* involves in production of chitinase and antifungal metabolites. Chitinase are the proteins that are involved in degrading the cell walls of pathogenic fungus. The sequences were cloned in the pGEM-T vector and characterized. The antifungal activity was determined by dual culture technique by plating wild type and mutant $\Delta tga1$ strain of *T. atroviride* against plant pathogens such as *R. solani*, *B. cinerea*, and *S. sclerotiorum*. The antifungal activity between the wild and mutant type strains were analyzed by altering the tga1 gene (Reithner et al., 2005).

In *T. harzianum*, a gene, viz., *ech42* codes for endochitinase was studied. The gene was cloned in pAN7-1 vector. Disruption of this gene affects the biocontrol activity of the fungus. The antifungal activity was tested

against pathogens like *B. cinerea*, and *R. solani* with the wild type and disruptant strains (Woo et al., 1999). From *T. hamatum* monooxygenase gene was isolated and characterized. This gene helps in the antifungal activity against some pathogens like *S. sclerotiorum*, *Sclerotinia minor*, and *Sclerotium cepivorum*. The expression of monooxygenase gene was influenced by until it had made contact with the two fungal species, and the expression seems to be more particularly at pH 4.

The DNA was isolated from *T. hamatum*, genomic library was constructed and it was sequenced. Agrobacterium mediated gene transformation was done with the help of pG3K02 and the gene was expressed. The promoter region of the monooxygenase gene was analyzed for the presence of motifs which helps in the regular expression of the genes. Thus, *T. hamatum* monooxygenase gene plays it significant role in the antagonist activity (Carpenter et al., 2008).

Hyphal growth

A new gene, *TrCCD1* from *Trichoderma reesei* was isolated and characterized. This gene involves in carotenoid metabolism that helps in the development of conidiospores and hyphal growth in *T. reesei*. The function of the gene was analyzed by comparing two mutant types named *ccdO* and *ccdP* (carotenoid cleavage dioxygenase) with the parental type. The T-DNA insertion of fungal genome was sequenced using specific primer, multiple sequence alignment was done using CLUSTAL-W algorithm and phylogenetic relationship was done by neighbor joining method (Zhong et al., 2009).

CONCLUSION

The various types of *Trichoderma* species were involved in the biocontrol activity and their mechanism of action were well known by the characterization and expression of the genes present in them. The fungal pathogens were known to cause major diseases in the agricultural scenario. So, most of the farmers were using hazardous chemical pesticides which cause major problems in the yield, ultimately affecting the land, soil fertility and remains toxic when consumed by humans and animals.

By using various microbial biocontrol agents, this problem has been reduced. The genes isolated from these biocontrol agents has been found to play an essential role in biocontrol activity therefore, with the help of genetic engineering techniques still more number of beneficial genes should be discovered in developing the agriculture for our future generations.

REFERENCES

Buensanteai N, Mukherjee PM, Horwitz BA, Cheng C (2010).

- Expression and purification of biologically active *Trichoderma virens* proteinaceous elicitor Sm1 in *Pichia pastoris*. Protein Express. Purif. 72: 131-138
- Cardoza RE, Vizcaino JA, Hermosa MR, Gonzalez FJ, Llobell A, Monte M, Gutierrez S (2006). Cloning and characterization of the erg1 gene of Trichoderma harzianum: Effect of the erg1 silencing on ergosterol biosynthesis and resistance to terbinafine. Fungal Genet. Biol. 43: 164-178.
- Carpenter MA, Ridgway HJ, Stringer AM, Hay AJ, Stewart A (2008). Characterization of a *Trichoderma hamatum* monooxygenase gene involved in antagonistic activity against fungal plant pathogens. Curr. Genet. 53: 193-205.
- Ciliento R, Woo SL, DiBenedetto P, Ruocco M, Scala F, Soriente I, Ferraioli S, Brunner K, Zeilinger S, Mach RL, Lorito M (2004). Genetic improvement of *Trichoderma* ability to induce systemic resistance. J. Zhejiang Univ. 30(4): 423.
- Cortes C, Gutierrez A, Olmedo V, Inbar J, Chet I, Herrera Estrella A (1998). The expression of genes involved in parasitism by *Trichoderma harzianum* is triggered by a diffusible factor. Mol. Gen. Genet. 260: 218-225.
- Dixit P, Mukherjee PK, Ramachandran V, Eapen S (2011). Glutathione transferase from *Trichoderma virens* enhances cadmium tolerance without enhancing its accumulation in transgenic *Nicotiana tabacum*. PLoSONE, 6(1): 1-15.
- Djonovic S, Vittone G, MendozaHerrera A, Kenerley CM (2007). Enhanced biocontrol activity of *Trichoderma virens* transformants constitutively coexpressing β-1,3- and β-1,6-glucanase genes. Mol. Plant Pathol. 8(4): 469-480.
- Donzelli BGG, Lorito M, Scala F, Harman GE (2001). Cloning, sequence and structure of a gene encoding an antifungal glucan 1,3-β-glucosidase from *Trichoderma atroviride* (*T. harzianum*). Gene, 277: 199-208.
- Gallo A, Mule G, Favilla M, Altomare C (2004). Isolation and characterization of a trichodiene synthase homologous gene in *Trichoderma harzianum*. Physiol. Mol. Plant Pathol. 65: 11-20.
- Gao KX, Liu XG, Liu YH, Zhu TB, Wang SL (2002). Potential of *Trichoderma harzianum* and *T.atroviride* to control *Botryosphaeria* berengeriana f. sp. piricola, the cause of apple ring rot. J. Phytopathol. 150: 271-276.
- Grondona I, Hermosa MR, Tejada M, Gomis MD, Mateos PF, Bridge P, Monte E, Garcy aAcha I (1997). Physiological and biochemical characterization of *Trichoderma harzianum*, a biocontrol agent against soil borne fungal plant pathogens. Appl. Environ. Microbiol. 63: 3189-3198
- Harman GE, Howell CR, Viterbo A, Chet I, Lorito M (2004). Trichoderma species-Opportunistic, avirulent plant symbionts. Nat. Rev. Microbiol. 1: 43-56.
- Hermosa R, Botella L, Keck M, Jimenez JA, MonteroBarrientos M, Arbona V, GomezCadenas A, Monte E, Nicolas C (2011). The overexpression in *Arabidopsis thaliana* of a *Trichoderma harzianum* gene that modulates glucosidase activity, and enhances tolerance to salt and osmotic stresses. J. Plant Physiol. 168: 1295-1302.
- Janisiewicz W, Korsten L (2002). Biological control of postharvest diseases of fruits. Annu. Rev. Phytopathol. 40: 411-441.
- Kotake T, Kaneko S, Kubomoto A, Haque MA, Kobayashi H, Tsumuraya Y (2004). Molecular cloning and expression in *Escherichia coli* of a *Trichoderma viride* endo-β-(1→6)-galactanase gene. Biochem. J. 377: 749-755.
- Kuc J (2001). Concepts and direction of induced systemic resistance in plants and its
- application. Eur. J. Plant Pathol. 107: 7-12.
- Li M, Yang Q, Song J (2010). Three tubulin genes of *Trichoderma harzianum*: Alpha, Beta, and Gamma. Braz. Arch. Biol. Technol. 53(4): 811-816.
- Li M, Yang Q (2007). Isolation and characterization of a β -tubulin gene from *Trichoderma harzianum*. Biochem. Genet. 45: 529-534.
- Liu Y, Yang Q, Song J (2009). A new serine protease gene from Trichoderma harzianum is expressed in Saccharomyces cerevisiae. Appl. Biochem. Microbiol. 45(1): 22-26.
- LopezMondejar R, Ros M, Pascual JA (2011). Mycoparasitism-related genes expression of *Trichoderma harzianum* isolates to evaluate their efficacy as biological control agent. Biol. Control, 56: 59-66.

- ManteroBarrientos M, Hermosa R, Nicolas C, Cardoza RE, Gutierrez S, Monte E (2008). Overexpression of a *Trichoderma* HSP70 gene increases fungal resistance to heat and other abiotic stresses. Fungal Genet. Biol. 45: 1506-1513.
- Marcello CM, Steindorff AS, Silva SP, Silva RN, Bataus LAM (2010). Expression analysis of the exo-β-1,3-glucanase from the mycoparasitic fungus *Trichoderma asperellum*. Microbiol. Res. 165: 75-81
- Massart S, Jijakli HM (2007). Use of molecular techniques to elucidate the mechanisms of action of biocontrol agents: A review. J. Microbiol. Methods, 69: 229-241.
- Migheli Q, GonzalezCandelas L, Dealessi L, Camponogara A, RamonVidal D (1998). Transformants of *Trichoderma longibrachiatum* overexpressing the β -1,4-endoglucanase gene *egl*1 show enhanced biocontrol of *Pythium ultimum* on cucumber. Biol. Control, 88(7): 673-677.
- Min YS, Kim BG, Lee C, Ho Gil H, Joong Hoon AHN (2002). Purification, Characterization, and cDNA cloning of Xylanase from fungus *Trichoderma* strain SY. J. Microbiol. Biotechnol. 12(6): 1-5.
- MoranDiez E, Hermosa R, Ambrosino P, Cadoza RE, Gutierrez S (2009). The ThPG1 endopolygalacturonase is Required for the *Trichoderma harzianum*-Plant beneficial interaction. Am. Phytopathol. Soc. 22(8): 1021-1031.
- Mukherjee M, Mukherjee PK, Kale PS (2007). cAMP signalling is involved in growth, germination, mycoparasitism and secondary metabolism in *Trichoderma virens*. Microbiology, 153: 1734-1742.
- Mukherjee PM, Latha J, Hadar R, Horwitz BA (2004). Role of two G-protein alpha subunits, *Tga*A and *Tga*B, in the antagonism of plant pathogens by *Trichoderma virens*. Appl. Environ. Microbiol. 70(1): 542-549.
- Mukherjee PM, Latha J, Hardar R, Horwitz BA (2003). *Tmk*A, Mitogen activated Protein Kinase of *Trichoderma virens* is involved in biocontrol properties and repression of conidiation in the dark. Eukaryot. Cell 2: 446-455.
- Pozo MJ, JongMin B, Garcia JM, Kenerley CM (2004). Functional analysis of *tvsp1*, a serine protease-encoding gene in the biocontrol agent *Trichoderma virens*. Fungal Genet. Biol. 41: 336-348.
- Rosado IV, Rey M, Codon AC, Govantes J, MorenoMateos MA, Benitez T (2007). QID74 Cell wall protein of *Trichoderma harzianum* is involved in cell protection and adherence to hydrophobic surfaces. Fungal Genet. Biol. 44: 950-964.
- Reithner B, Brunner K, Schuhmacher R, Peissl I, Seidl V, Krska R, Zeilinger S (2005). The G protein α subunit Tga1 of *Trichoderma atroviride* is involved in chitinase formation and differential production of antifungal metabolites. Fungal Genet. Biol. 42: 749-760.
- Rubio MB, Hermosa R, Reino JL, Collado IG, Monte E (2009). *Thctf1* transcription factor of *Trichoderma harzianum* is involved in 6-pentyl-2H-pyran-2-one production and antifungal activity. Fungal Genet. Biol. 46: 17-27.
- Ruocco M, Lanzuise S, Vinale F, Marra R, Turra D, LoisWoo S, Lorito M (2009). Identification of a New Biocontrol Gene in *Trichoderma atroviride*: The Role of an ABC Transporter Membrane Pump in the interaction with different plant-pathogenic fungi. Am. Phytopathol. Soc. 22(3): 291-301.
- Saadia M, Ahmed S, Jamil A (2008). Isolation and cloning of *cre1* gene from a filamentous fungus *Trichoderma harzianum*. Pak. J. Bot. 40(1): 421-426.
- Saiprasad GVS, Mythili JB, Anand L, Suneetha C, Rashmi HJ, Naveena C, Ganeshan G (2009). Development of *Trichoderma harzianum* gene construct conferring antifungal activity in transgenic tobacco. Indian J. Biotechnol. 8: 199-206.
- Steyaert JM, Stewart A, Jaspers MV, Carpenter M, Ridgway HJ (2004). Co-expression of two genes, a chitinase (*chit42*) and proteinase (*prb1*), implicated in mycoparasitism by *Trichoderma hamatum*. Mycologia, 96(6): 1245-1252.
- Tijerino A, Cardoza RE, Moraga J, Malmierca MG, Vicente F, Aleu J, Collado IG, Gutierrez S, Monte E, Hermosa R (2011). Overexpression of the trichodiene synthase gene tri5 increases trichodermin production and antimicrobial activity in Trichoderma brevicompactum. Fungal Genet. Biol. 48: 285-296.
- Vizcaino JA, Cardoza RA, Hauser M, Hermosa R, Rey M, Lobell A, Becker JM, Gutierrez S, Monte E (2006). *ThPTR2*, a di/tri-peptide

- transporter gene from *Trichoderma harzianum*. Fungal Genet. Biol. 43: 234-246.
- Woo SL, Donzelli B, Scala F, Mach R, Harman GE, Kubicek CP, Sorbo GD, Lorito M (1999). Disruption of the *ech42* (endochitinase-encoding) gene affects biocontrol activity in *Trichoderma harzianum* P1. Am. Phytopathol. Soc. 12(5): 219-229.
- Zhong YH, Wang TH, Wang XL, Zhang GT, Yu HN (2009). Identification and characterization of a novel gene, *TrCCD1*, and its possible function in hyphal growth and conidiospore development of *Trichoderma reesei*. Fungal Genet. Biol. 46: 255-263.