#### Review

# Roles of plant soluble sugars and their responses to plant cold stress

Ma Yuanyuan<sup>1</sup>, Zhang Yali<sup>1</sup>, Lu Jiang<sup>1\*</sup>, Shao Hongbo<sup>2,3,4</sup>

<sup>1</sup>The College of Food Science and Nutritional Engineering, China Agricultural University, Beijing 100083, China. <sup>2</sup>State Key Laboratory of Soil Erosion and Dryland Farming on the Loess Plateau, Institute of Soil and Water Conservation, Chinese Academy of Sciences, Yangling 712100, China.

<sup>3</sup>Shandong Key Laboratory of Eco-environmental Science for the Yellow River Delta, Binzhou University, Binzhou256603, China.

Accepted 17 March, 2009

Plants can enhance freezing tolerance by gradual exposure to low temperature known as cold acclimation. Soluble sugars have been confirmed to play an important role during the process. This is based on the fact they are most commonly detected in various species of terrestrial plants that have undergone seasonal cold acclimation. Soluble sugars exert their positive effects to protect plant cells from damage caused by cold stress through many ways including serving as osmoprotectants, nutrients as well as interacting with the lipid bilayer. In addition to these functions, more focus is on their important hormone-like functions as primary messengers in signal transduction nowadays. On the other hand, higher sugar concentrations can trigger leaf senescence, which indicates that the accumulation of soluble sugars during cold acclimation might impact on plants negatively. Further study can be done to investigate the mechanisms by which soluble sugars are involved in the plants' response to the cold signal by advanced molecular biology methodology to examine sugar-regulated gene expression under cold condition.

**Key words:** Plants, sugars, cold acclimation, signal transduction, biotechnology.

#### THE IMPACT OF SOLUBLE SUGARS ON THE COLD ACCLIMATION

Cold stress causes several dysfunctions at cellular level; these include damage to membranes, generation of reactive oxygen species, protein denaturation and accumulation of toxic products etc (Bowers, 1994; Shao et al., 2008). In nature, plants may exhibit cold tolerance due to gradual exposure to low non-freezing temperatures, and the process is known as cold acclimation (Thomashow, 1999).

The advent of molecular biology has propelled research in this area over the past two decades; today some of the events of cold signal perception, transduction and cold acclimation are defined at the molecular level (Shinozaki et al., 2000; Shinozak et al., 2001; Zhu, 2001; Thomashow, 2001; Shao et al., 2006, 2007). Cold acclimation is associated with multiple mechanisms that

include changes in (a) gene expression (b) membrane composi-tion and cryobehaviour (Orvar et al., 2000; Shao et al., 2008) (c) elevation of ABA (Lang et al., 1994) (d) antioxidants (Kang and Saltveit, 2002; Shao et al., 2008) (e) accumulation of substantial amounts of compatible solutes, such as soluble sugars (Ristic et al., 2003; Wanner et al., 1999), amino acids (Lalk et al., 1985; Wanner et al., 1999; Koster, 1991; Shao et al., 2006) and glycine betaine (Kishitani et al., 1994), which are thought to play a role in protection of cells from freezing injury (Figure 1). Although so many changes occur during coldacclimation, soluble sugars have been confirmed to play an important role based on the fact they are most commonly detected in various species of land plants that have undergone seasonal cold acclimation, including angio-sperms, gymnosperms, and lower land plants such as bryophytes (Saka and Larcher, 1987; Rutten and Santarius, 1992). It has been reported that under natural conditions, soluble sugars increase during the onset of winter when plants are subjected to low temperatures;

<sup>&</sup>lt;sup>4</sup>Institute for Life Sciences, Qingdao University of Science & Technology (QUST), Qingdao266042, China.

<sup>\*</sup>Correspondence author. E-mail: shaohongbo@qust.edu.cn, jiang.lu.cau@gmail.com. Tel: 86 532 84023984.

## Cold acclimation induces changes of many genes in various cellular processes

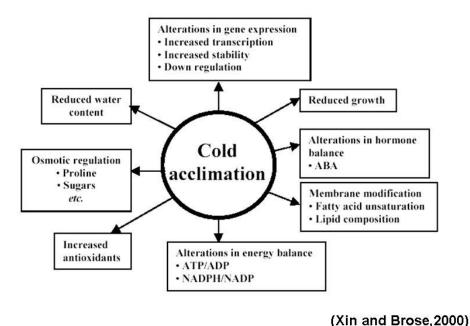


Figure 1. The well known changes during the process of cold acclimation.

conversely, soluble sugars decrease in spring when plants are deacclimating (Sakai, 1962; Siminovitch, 1981). Sugar content also increases during cold acclimation under artificial conditions (Koster and Lynch, 1992; Sasaki et al., 1996; Shao et al., 2006-2008).

### DIFFERENT SUGARS INVOLVED IN THE COLD ACCLIMATION

It has been proved that many kinds of soluble sugars are included in the course of cold acclimation. For instance, European habitats which possess substantial degrees of freezing tolerance have exhibited higher degrees of freezing tolerance in winter than in summer. Increase in freezing tolerance in these mosses appeared to be associated with accumulation of soluble sugars, especially that of sucrose (Rutten and Santarius, 1992). Also, chilling of muskmelon plants led to the accumulation in the source leaves of large amounts of starch, sucrose, and hexoses, as has been reported previously for many plant species (Paul, 1991; Paul, 1990). In addition, it has been demonstrated that the levels of a few oligosaccharides increase during cold acclimation, typically in cereals and woody plants, both of which develop high degrees of freezing tolerance in winter. Galactosylsucrose oligosaccharides such as raffinose and stachvose are commonly found to accumulate during seasonal cold acclimation in woody plants, along with development of freezing tolerance to levels so high that some plants can even tolerate to freezing in liquid nitrogen without cryoprotectants (Sauter et al., 1996).

### VARIOUS STUDIES INVOLVED IN THE EFFECT OF SOLUBLE SUGARS

Aside from the accumulation in endogenous sugars, it has been revealed that exogenous application of soluble sugars resulted in an increase in freezing tolerance in various higher plant species. There has been report that pre-incubation with glucose increased survival after cryopreservation of cabbage leaf cells (Jitsuyama et al., 1997), and pre-incubation with sucrose enhanced survival Jerusalem artichoke cells after cryopreservation (Harris et al., 2004). Moreover, improved freezing tolerance in *Arabidopsis thaliana* protoplasts occurred after the exogenous application of sucrose (Uemura et al., 2003; Li et al., 2006).

Up to now, researchers have isolated several mutant lines of *Arabidopsis* that are impaired in their freezing tolerance after cold acclimation, such as the sfr (sensitivity to freezing) mutants. These mutants have provided researchers a good insight into the relationship between the sugars and freezing resistance in the plants. The freezing tolerance in these plants before cold acclimation is unaffected which allows the freezing sensitivity of the sfr mutants to be specifically attributed to defects in the

acclimation process, rather than generalized debilitation. Using electrolyte leakage assay, the quantitative examination of freezing tolerance of all the sfr mutants was achieved. The results showed all the sfr mutants actually increased their freezing tolerance during cold acclimation, but to a less extent than the wild type. The deficit was greatest in the sfr4 mutant, according to its high degree of sensitivity in survival tests on intact plants (Warren et al., 1996). Subsequently, it was detected that sfr4, alone among the mutants, showed levels of freezing tolerance similar to those of wild-type (WT) plants when grown at ambient temperatures but, unlike WT plants, underwent a reduction in its Suc and Glc contents, in contrast to the increase shown by wild type during cold acclimation (McKown et al., 1996; Uemura and Steponkus, 2003).

In addition to these mutants, there have existed many studies indicating that transgenic plants over-accumulating raffinose have higher degrees of freezing resistance than do wild-type plants (Taji et al., 2002; Pennycooke et al., 2003). Perennial ryegrass plants which were transgenic to express wheat fructosyltransferase genes accumulate increased amounts of fructan and acquire enhanced resistance on a cellular level to freeze (Hisano et al., 2004). It has also been reported that the *Arabidopsis* ecotype C24, which accumulates more raffinose than does the Columbia ecotype, shows greater freezing tolerance than that of the Columbia ecotype (Klotke et al., 2004).

### THE FUNCTIONAL ROLES OF SOLUBLE SUGARS DURING COLD ACCLIMATION

All of these facts suggest critical roles for these sugars as compatible solutes in freezing tolerance. The mechanisms by which sugars act to influence freezing resistance are just beginning to be deciphered. Sucrose glucose, fructose and oligosaccharides such as raffinose as well as stachyose are commonly found in soluble sugars that accumulate along with development of freezing tolerance in higher plants. It has been suggested that these sugars not only serve as osmoprotectants but also play a role in protecting cellular membranes from damage caused by dehydration and freezing through interacting with the lipid bilayer (Anchordoguy et al., 1987; Shalaev and Shalaev, 2001; Shao et al., 2006). It is also proposed that the role of some oligosaccharides is to prevent crystallization of sucrose, thus facilitating glass formation within the cell, leading to protection of membrane phospholipids (Anchordoguy and Leopold, 1989; Crowe et al., 1988). Generally speaking, the accumulation of soluble sugars contributes to the increase in the cryostability of cellular membranes. Increased membrane cryostability is a prerequisite for freezing tolerance because freeze-induced destabilization of cellular membranes is the primary cause of injury in plants (Steponkus et al., 1993; Shao et al., 2008).

An additional role of sugars during cold acclimation is

that they may function as nutrients which make plants survive the decreased temperature and dryland conditions as well as recover from freezing-stress (Eagles et al., 1993; Trunova; 1982; Shao et al., 2008).

### MORE FOCUS ON THE ROLES OF SOLUBLE SUGARS AS SIGNALING MOLECULES

Although there have been many functional soluble sugars that act as documented above, nowadays, more and more attention has been put on the important hormone-like functions of soluble sugars as primary messengers in signal transduction. The pivotal role of sugars as signaling molecules is well illustrated by the variety of sugar sensing and signaling mechanisms discovered in free-living microorganisms such as bacteria and yeast (Rolland et al., 2001; Stulke and Hillen, 1999; Wang et al., 2008; Xiao et al., 2009).

Based on the biochemical, molecular, and genetic experiments, it has been concluded that soluble sugars play a central role in the control of plant metabolism, growth, and development. Moreover many researches have revealed interactions that integrate light, abiotic and biotic stress, and hormone signaling (Roitsch, 1999; Sheen, 1999; Smeekens, 2000; Gazzarrini and McCourt, 2001; Finkelstein and Gibson, 2002) and coordinate carbon and nitrogen metabolism (Stitt and Krapp, 1999; Coruzzi and Bush, 2001; Coruzzi and Zhou, 2001; Uddin et al., 2008; Wang et al., 2008; Xiao et al., 2009).

Abiotic and biotic stress stimuli, such as drought, salinity, wounding, and infection by viruses, bacteria, and fungi, can modulate source-sink activities. Because extracellular invertase, a key enzyme for hydrolyzing Suc (Sturm, 1999), is regulated by stress stimuli and hormones, it has been proposed to be a central modulator of assimilate partitioning, integrating sugar, stress, and hormone signals (Roitsch, 1999). In addition, many jasmonate-, ABA-, and stress-inducible genes are coregulated by sugars (Reinbothe et al., 1994; Sadka et al., 1994; Shao et al., 2005-2008; Uddin et al., 2008; Wang et al., 2008).

How plants sense cold is unknown. A fall in temperature can reduce the fluidity of membranes, and this appears to be an effective direct sensor of cold in cyanobacteria. Another possibility is that, in light, photosynthetic cells may sense cold through an effect on photosystem 2 excitation pressure (Gray et al., 1997; Huner et al., 1998). Molecular studies have shown that cold acclimation in higher plants is complex involving multiple regulatory pathways (Fowler and Thomashow, 2002), furthermore while some of the signal transduction pathways interact, others are independent (Xin and Browse, 2000). Thus more than one sensor of cold is possible (Figure 2).

As mentioned above, sugar-sensing and sensing of the environment appear to be part of a complex regulatory web (Gibson, 2000). In relation to cold, high sugar supply

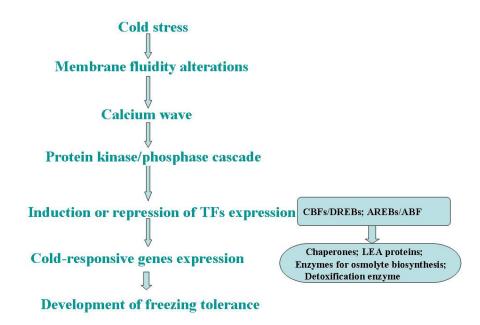


Figure 2. Sugars-involved into the events occurring in plants under cold stress (Shao et al., 2008).

induces fructan accumulation, which is also a usual response of grasses to cold (Winter, 1994; Wang et al., 2008; Xiao et al., 2009).

New insights into the mechanisms by which sugarresponse pathways interact with cold response pathways have been supported by micro-array experiments examining sugar-regulated gene expression under different conditions. Based on the fact, it has been elucidated that many sugar-regulated genes are involved in the coldinduced gene expression.

Cold acclimation is accomplished by the expression of many cold-regulated genes (Thomashow, 2001; Zhu, 2001). In Arabidopsis, these genes are called rd (responsive to dehydration), erd (early responsive to dehydration), Iti (low-temperature induced), kin (cold induced) and cor (cold-regulated). These genes are also induced by dehydration (due to water deficit or high salt) and ABA, and can be collectively called cold-responsive genes. Cold-responsive genes are regulated through Crepeat/dehydration-responsive elements (CRT/DRE) and abscisic acid (ABA)-responsive elements (cis elements), C-repeated binding factors/dehydration-responsive element binding proteins (CBFs/DREBs) and basic leucine zippers (bZIPs) (SGBF1), respectively. It has been reported that expression of key regulators of abiotic stress responses such as CBF3, COR15A, and RD29A was induced by both glucose and ABA. Constitutive expression of CBF3 in transgenic Arabidopsis plants induces expression of target COR (cold-regulated) genes to enhance freezing tolerance in non-acclimated plants. Expression of COR15A and RD29A is regulated by CBF3, suggesting that glucose may contribute to the regulation of cold stress tolerance (Yunhai et al., 2006; Uddin et al., 2008; Wang et al., 2008).

### THE INVOLVEMENT OF SUGARS IN LEAF SENESCENCE

Soluble sugars help ensure an adequate supply of building materials and energy to carry out specific developmental programs. For example, in-vivo and invitro experiments suggest that in some plant species, increasing sugar levels delay seed germination and stimulate the induction of flowering and senescence. In other words, it may be beneficial for plants to adjust the timing with which nutrient-intensive events occur to supply enough materials and energy for successful completion of those events. Levels of sugars, such as sucrose, have been postulated to affect the timing at which some plant species flower (Bernier et al., 1993). Soluble sugar levels have also been shown to affect other phase changes, such as the onset of senescence (Paul and Pellny, 2003). Endogenous leaf sugar levels tend to increase during senescence. Similarly, application of exogenous sugars stimulates the early stages of senescence.

Leaf senescence is the last step in the life cycle of plants. During the process, materials used to build up leaves during vegetative growth are remobilized and transported into the developing organs (Smart, 1994; Smart et al., 1995; Williams et al., 1997).

Although senescence occurs in an age-dependent manner in many species (Noode`n, 1988), its initiation and progression can be modulated by a variety of environ-

mental factors such as temperature, mineral deficiency, and drought conditions, as well as by internal factors such as plant growth regulators (Grbic and Bleecker, 1995; Buchanan-Wollaston, 1997; Nam, 1997; Weaver et al., 1997; Dai et al., 1999; Uddin et al., 2008; Wang et al., 2008).

Since senescence can be induced by a large number of different conditions, and several signaling pathways are involved in the regulation of senescence (Buchanan-Wollaston, 2003), it is possible that environmental signals are integrated by sugar signaling. For example, experiments on sugar regulated senescence indicate that leaf senescence can be induced by growing Arabidopsis plants in the presence of 2% glucose in combination with low nitrogen supply (Pourtau et al., 2004; Wingler et al., 2004). Affymetrix GeneChip data confirmed that changes in gene expression during glucose-induced senescence are characteristic of developmental senescence. Similarly. Stessman et al. (2002) found that hexoses accumulate in senescing Arabidopsis leaves. However, the question remains as to what causes the strong accumulation of hexoses despite the decline in photosynthetic carbon assimilation in senescing leaves. A possible source of hexoses is the breakdown of starch. In addition, Jongebloed et al. (2004) showed that phloem blockage by callose deposition could lead to an age-dependent sugar accumulation.

#### THE DUAL ROLES OF SUGARS POSSIBLY INVOLVED IN ENCOUNTERING COLD STRESS

During cold acclimation, soluble sugars accumulated which could theoretically have adverse impact on photosynthesis as well as induce senescence. Nevertheless, cold-acclimated leaves do not show the typical repression of photosynthesis by sugars (Strand et al., 1997; Li and Huang, 2008). On the other hand, it has been hypothesized that photosynthetic activity is regulated in a feedback manner by soluble sugars. The photosynthetic machinery might be down-regulated by sugars due to the cold-girdling petioles to prevent sugar export out of the leaf. Also, in these cases, genes related to photosynthesis were repressed, including those for chlorophyll binding protein and Rubisco (Smeekens, 2000).

However more complicated it is, there have been studies showing increased flux through the Suc biosynthetic pathway that reduces inhibition of photosynthesis in the cold (Strand et al., 2003). It was therefore conceivable that cold acclimation could interact with metabolic regulation of senescence by inhibiting the induction of senescence by sugar accumulation (Zhao et al., 2008; Shao et al., 2008; Bressan et al., 2009).

#### **CONCLUSION AND PERSPECTIVES**

Alterations in the levels of soluble sugars, such as glucose and sucrose have been shown to affect the

freezing tolerance in the plants. What roles of sugars play during cold acclimation is complicated by the fact that sugars can act by affecting osmotic potentials as well as by functioning as signaling molecules. On the other hand, sugars affect developmental programs ranging from embryogenesis to senescence, which makes the research more complex. On the basis of the fact that leaves which have developed in the cold do not display the typical down-regulation of photosynthesis in response to sugar accumulation, it is possible that induction of the cold acclimation pathway have resulted in decreased sugar sensitivity and thus delayed senescence.

Further studies about the actual roles that sugars play in plants' response to cold stimuli can be done by scanning lists of sugar-regulated genes expression under cold condition using various advanced technologies such as microarray experiments. On the basis of these research investigations, we might get new insight into mechanisms by which sugar-response pathways interact with other response pathways in the cold-triggered response. At the same time, we will have a promising perspective by bioengineering plant responses to cold stress by sugar functions.

#### **ACKNOWLEDGEMENTS**

This work was supported by National "948" Program (grant no.2006-G26).

#### **REFERENCES**

Anchordoguy TJ, Rdolph AS, Carpenter JF, Crowe, JH (1987). Modes of interaction of cryoprotectants with membrane phospholipids during freezing. Cryobiol. 24: 324-331.

Bernier G, Havelange A, Houssa C, Petitjean A, Lejeune P (1993). Physiological signals that induce flowering. Plant Cell 5: 1147-1155.

Bowers MC (1994). Environmental effects of cold on plants. In: Wilkinson, R.E. (Ed.), Plant-Environment Interactions. Marcel Dekker, New York, pp. 391-411.

Bressan R, Bohbert H, Zhu JK (2009). Abiotic stress tolerance:from gene discovery in model organisms to crop improvement. Mol. Plant 2:1.2

Browse J, Xin Z (2002). Temperature sensing and cold acclimation. Curr. Opin. Plant Biol. 4: 241- 246.

Buchanan-wollaston V (1997). The molecular biology of leaf senescence. J. Exp. Bot. 48: 181-199.

Buchanan-wollaston V, Earl S, Harrison E, Mathas E, Navabpour S, Page T, Pink D (2003). The molecular analysis of leaf senescence: a genomics approach. Plant Biotech. J. 1: 3-22.

Coruzzi G, Bush DR (2001). Nitrogen and carbon nutrient and metabolite signaling in plants. Plant Physiol. 125: 61-64.

Coruzzi GM, Zhou L (2001). Carbon and nitrogen sensing and signaling in plants: Emerging 'matrix effects.' Curr. Opin. Plant Biol. 4: 247-253.

Crowe JH, Crowe LM, Carpenter JF, Rudolph AS, Wistrom CA, Spargo BJ, Anchordoguy TJ (1988). Interactions of sugars with membranes. Biochem. Biophy. Acta 947: 367-384.

Dai N, Schaffer A, Petreikov M, Shahak Y, Giller Y, Ratner K, Levine K, Granot D (1999). Overexpression of Arabidopsis hexokinase in tomato plants inhibits growth, reduces photosynthesis and induces rapid senescence. Plant Cell 11: 1253-1266.

Eagles CF, Williams J, Louis DV (1993). Recovery after freezing Effect of sugar concentration on cold acclimation in *Avena sativa* L. *Lolium perenne* L. and *L. multiorum* Lam. New Phytol. 123: 477-483.

- FInkelstein RR, Gibson SI (2002). ABA and sugar interactions regulating development: Cross-talk or voices in a crowd? Curr. Opin. Plant Biol. 5: 26-32.
- Fowler S, Thomashow MF (2002). Arabidopsis transcriptome proling indicates that multiple regulatory pathways are activated during cold acclimation in addition to the CBF cold response pathway. Plant Cell, 14: 1675-1690.
- Gazzsrrini S, Mccourt P (2001). Genetic interactions between ABA, ethylene and sugar signaling pathways. Curr. Opin. Plant Biol. 4: 387-391.
- Gibson SI (2000). Plant sugar-response pathways. Part of a complex regulatory web. Plant Physiol. 124: 1532-1539.
- Gray GR, Chauvin LP, Sarhan F, Huner NPA (1997). Cold acclimation and freezing tolerance. Plant Physiol. 114: 467-474.
- Grbic V, Bleecker AB (1995). Ethylene regulates the timing of leaf senescence in Arabidopsis. Plant J. 8: 595-602.
- Harris W, Lynch PT, Hargreavs AJ, Bonner PLR (2004). Cryopreservation of Helianthus tuberosus cell suspension cultures: The effect of preculture treatments on cytoskeletal proteins and transglutaminase activity. Cryo-Lett. 25: 213-217.
- Hisano H, Kanazawa A, Kawakami A, Yoshida M, Shimamoto Y, Yamada T (2004). Transgenic perennial ryegrass plants expressing wheat fructosyltransferase genes accumulate increased amounts of fructan and acquire increased tolerance on a cellular level to freezing. Plant Sci.167: 861-868.
- Huner NPA, Oèquist G, Sarhan F (1998). Energy balance and acclimation to light and cold. Trends Plant Sci. 3: 224-230.
- Itsuyama Y, Suzuki T, Harada T, Fujikawa S (1997). Ultrastructural study of mechanism of increased freezing tolerance to extracellular glucose in cabbage leaf cells. Cryo-Lett. 18: 33-44.
- Jongebloed U, Szederke´nyi J, Hartig K, Schobert C, Komor E (2004). Sequence of morphological and physiological events during natural ageing of a castor bean leaf: sieve tube occlusions and carbohydrate back-up precede chlorophyll degradation. Physiol. Plant. 120: 338-346.
- Kang HM, Saltveit ME (2002). Activity of enzymatic antioxidant defence systems in chilled and heat shocked cucumber seedling radicles. Physiol. Plant.113: 548-556.
- Kishitani S, Watanabe K, Yasuda S, Arakawa K, Takabe T (1994). Accumulation of glycinebetaine during cold acclimation and freezing tolerance in leaves of winter and spring barley plants. Plant Cell Environ. 17: 89-95.
- Klokte J, Kopaka J, Gantzke N, Heyer AG (2004). Impact of soluble sugar concentrations on the acquisition of freezing tolerance in accessions of Arabidopsis thaliana with contrasting cold adaptation evidence for a role of raffinose in cold acclimation. Plant Cell Environ. 27: 1395-1404.
- Koster KL (1991). Glass formation and desiccation tolerance in seeds.Plant Physiol. 96: 302-304.
- Koster KL, Lynch DV (1992). Solute accumulation and compartmentation during the cold acclimation of puma rye. Plant Physiol. 98: 108-113.
- Lalk I, Do"rfflung K (1985). Hardening, abscisic acid, proline and freezing resistance in two winter wheat varieties. Physiol. Plant. 63: 287-292.
- Lang V, Mantyl AE, Welin B, Sundberg B, Palva ET (1994). Alterations in water status, endogenous abscisic acid content, and expression of rab18 gene during the development of freezing tolerance in Arabidopsis thaliana. Plant Physiol. 104: 1341-1349.
- Li YH, Lee KK, Walsh S, Smith C, Hadingham S, Sorefan K, Cawley G, Michael W (2006). Establishing glucose- and ABA-regulated transcription networks in Arabidopsis by microarray analysis and promoter classification using a Relevance Vector Machine. Genome Res. 16: 414-427.
- Li YS, Huang MB (2008). Pasture yield and soil water deletion of continuous growing alfalfa in the Loess Plateau of China. Agric. Ecosys. Environ.124: 24-32.
- Mckown R, Kuroki G, Warren G (1996). Cold responses of Arabidopsis mutants impaired in freezing tolerance. J. Exp. Bot. 47: 1919-1925.
- Nam H (1997). The molecular genetic analysis of leaf senescence. Curr. Opin. Biotech. 8: 200-207.
- Noode'n LD (1988). The phenomenon of senescence and aging. In:

- Noode'n LD, Leopold AC, eds. Senescence and aging in plants. San Diego: Academic Press pp. 2-50.
- Orvar BL, Sangwan V, Omann F, Dhindsa RS (2000). Early steps in cold sensing by plant cells: the role of actin cytoskeleton and membrane fluidity. Plant J. 23: 785-794.
- Paul MJ, Lawlor DW, Driscoll SP (1990). The effect of temperature on photosynthesis and carbon fluxes in sunflower and rape. J. Exp. Bot. 41: 547-555.
- Paul MJ, Driscoll SP, Lawlor DW (1991). The effect of cooling on photosynthesis, amounts of carbohydrate and assimilate export in sunflower. J. Exp. Bot. 42: 845-852.
- Paul MJ, Pellny TK (2003). Carbon metabolite feedback regulation of leaf photosynthesis and development. J. Exp. Bot. 54: 539-547.
- Pennycooke JC, Jones ML, Stushnoff C (2003). Down-regulating alphagalactosidase enhances freezing tolerance in transgenic petunia. Plant Physiol. 133: 901-909.
- Pourtau N, Mare's M, Purdy S, Quentinm N, Rue"l A, Wingler A (2004). Interactions of abscisic acid and sugar signalling in the regulation of leaf senescence. Planta 219: 765-772.
- Rolland F, Winderickx J, Thevelein JM (2001). Glucosesensing mechanisms in eukaryotic cells. Trends Biochem. Sci. 26: 310-317.
- Roitsch T (1999). Source-sink regulation by sugar and stress. Curr. Opin. Plant Biol. 2: 198-206.
- Reinbothe S, Mollenhauer B, Reinbothe C (1994). JIPs and RIPs: The regulation of plant gene expression by jasmonates in response to environmental cues and pathogens. Plant Cell, 6: 1197-1209.
- Ristic LA, Ashworth EN (1993). Changes in leaf ultrastructure and carbohydrates in Arabidopsis thaliana L. (Heyn) cv. Columbia during rapid cold acclimation. Protoplasma 172: 111-123.
- Rutten D, Santarius KA (1992). Relationship between frost tolerance and sugar concentration of various bryophytes in summer and winter. Oecologia 91: 260-265.
- Sadka A, Dewald DB, May GD, Park WD, Mullet JE(1994). Phosphate modulates transcription of soybean VspB and other sugar-inducible genes. Plant Cell 6: 737-749.
- Sakai A (1962). Studies on the frost-hardiness of woody plants: I. The causal relation between sugar content and frost-hardiness. Cont. Inst. Low Temp. Sci. B11: 1-40.
- Saka A, Larcher W (1987). Frost Survival of Plants: Responses and Adaptation to Freezing Stress. Springer, Berlin.
- Sasaki H, Ichimura K, Oda M (1996). Changes in sugar content during cold acclimation and deacclimation of cabbage seedlings. Ann. Bot. 78: 365-369.
- Sauter JJ, Wisniewski M, Witt W (1996). Interrelationships between ultrastructure, sugar levels, and frost hardiness of ray parenchyma cells during frost acclimation and deacclimation in poplar (Populus Canadensis Moench robusta) wood. J. Plant Physiol. 149: 451-461.
- Shalaev EY, Steponkus PL (2001). Phase behavior and glass transition of 1,2-dioleoylphosphaditylethanolamine (DOPE) dehydrated in the presence of sucrose. Biochem. Biophy. Acta-Biomembr. 1514: 100-
- Sheen J (1999). C4 gene expression. Ann. Rev. Plant Physiol. Plant Mol. Biol. 50: 187-217.
- Shinozaki K, Yamaguchi-shinozaki K (2000). Molecular responses to dehydration and low temperature: differences and cross-talk between two stress signaling pathways. Curr. Opin. Plant Biol. 3: 217-223.
- Siminovitch D (1981). Common and disparate elements in the processes of adaptation of herbaceous and woody plants to freezing: a perspective. Cryobiol.18: 166-185.
- Smart CM (1994). Gene expression during leaf senescence. New Phytol.126: 419-448.
- Smart CM, Hosken SE, Thomas H, Greaves JA, Blair BG, Schuch W (1995). The timing of maize leaf senescence and characterization of senescence-related cDNAs. Physiol. Plant. 93: 673-682.
- Smeekens S (2000). Sugar-induced signal transduction in plants. Ann.Rev. Plant Physiol. Plant Mol. Biol. 51: 49-81.
- Steponkus PL, Uemura M, Webb MS (1993). A contrast of the cryostability of the plasma membrane of winter rye and spring oat-two species that widely differ in their freezing tolerance and plasma membrane lipid composition. In PL Steponkus, ed, Adv. Low-Temp. Biol. 2: 211-312.
- Stessman D, Miller A, Spalding M, Rodermel S (2002). Regulation of

- photosynthesis during Arabidopsis leaf development in continuous light. Photosynthesis Res. 72: 27-37.
- Stitt M, Krapp A (1999). The interaction between elevated carbon dioxide and nitrogen nutrition: The physiological and molecular background. Plant Cell Environ. 22: 583-621.
- Strand A, Hurry V, Gustafsson P, Gardestro m P (1997). Development of Arabidopsis thaliana leaves at low temperatures releases the suppression of photosynthesis and photosynthetic gene expression despite the accumulation of soluble carbohydrates. Plant J. 12: 605-614
- Strand A, Foyer CH, Gustafsson P, Gardestro"m P, Hurry V (2003). Altering flux through the Suc biosynthesis pathway in transgenic Arabidopsis thaliana modifies photosynthetic acclimation at low temperatures and the development of freezing tolerance. Plant Cell Environ. 26: 523-535.
- Stulke J, Hillen W (1999). Carbon catabolite repression in bacteria. Curr. Opin. Microbiol. 2: 195-201.
- Sturm A (1999). Invertases: Primary structures, functions, and roles in plant development and sucrose partitioning. Plant Physiol.121: 1-8.
- Shao HB, Liang ZS, Shao MA (2006). Osmotic regulation of 10 wheat (*Triticum aestivum L.*) genotypes at soil water deficits. Biointer. 47: 132-139.
- Shao HB, Chen XY, Chu LY, Zhao XN, Wu G, Yuan YB, Zhao CX, Hu ZM (2006). Investigation on the relationship of Proline with wheat anti-drought under soil water deficits. Biointer. 53: 113-119.
- Shao HB, Guo QJ, Chu LY, Zhao XN, Su ZL, Hu YC, Cheng JF (2007). Understanding molecular mechanism of higher plant plasticity under abiotic stress. Biointer. 54: 37-45.
- Shao HB, Jiang SY, Li FM, Chu LY, Zhao CX, Shao MA, Zhao XN, Li F (2007). Some advances in plant stress physiology and their implications in the systems biology era. Biointer. 54: 33-36.
- Shao HB, Chu LY, Shao MA (2008). Calcium as a versatile Plant Signal Transducer under Soil Water Stress. BioEssays 30: 634-641.
- Shao HB, Chu LY, Shao MA, Zhao CX (2008). Advances in functional regulation mechanisms of plant aquaporins: Their diversity, gene expression, localization, structure and roles in plant soil-water relations (Review). Mol. Membr. Biol. 25: 1-12.
- Shao HB, Chu LY, Lu ZH, Kang CM (2008). Primary antioxidant free radical scavenging and redox signaling pathways in higher plant cells. Int. J. Biol. Sci. 4: 8-14.
- Zhao H, Zhang ZB, Shao HB, Xu P, Foulkes MJ (2008). Genetic Correlation and Path Analysis of Transpiration Efficiency for Wheat Flag Leaves. Environ. Exp. Bot. 64:128-134.
- Shao HB, Chu LY, Jaleel CA, Zhao CX (2008). Water-deficit stress-induced anatomical changes in higher plants. CR. Biol. 331: 215-225. Shao HB, Li B, Wang BC, Tang K, Liang YL (2008). A study on differentially expressed gene screening of *Chrysanthemum* plants under sound stress. CR. Biol. 331: 329-333.
- Taji T, Ohsumi C, Iuchi S, Seki M, Kasuga M, Kobayashi M, Yamaguchi-shinozaki K, Shinozaki K (2002). Important roles of drought- and cold-inducible genes for galactinol synthase in stress tolerance in Arabidopsis thaliana. Plant J. 29: 417-426.
- Thomashow MF (1999). Plant cold acclimation: freezing tolerance genes and regulatory mechanisms. Ann. Rev. Plant Physiol. Plant Mol. Biol. 50: 571-599.

- Thomashow MF (2001). So what's new in the field of plant cold acclimation? Lots! Plant Physiol.125: 89-93.
- Trunova TI (1982). Mechanisms of winter wheat hardening at low temperature. In: Li PH, Sakai A ed. Plant cold hardiness and freezing stress New York: Academic Press 2: 41-47.
- Uemura M, Steponkus PL (2003). Modification of the intracellular sugar content alters the incident of freeze-induced membrane lesions of protoplasts isolated from Arabidopsis thaliana leaves. Plant Cell Environ. 26: 1083-1096.
- Uemura M, Warren G, Steponkus PL (2003). Freezing sensitivity in the sfr4 mutant of Arabidopsis is due to low sugar content and is manifested by loss of osmotic responsiveness. Plant Physiol. 131: 1800-1807
- Uddin Mdl, Qi YH, Yamada S, Shibuya I, Deng XP, Kwak SS, Kaminaka H, Tanaka K (2008). Overexpression of a new rice vacuolar antiporter regulating protein OsARP improves salt tolerance in tobacco. Plant Cell Physiol. 49: 880-890.
- Wang ZY, Li FM, Xiong YC, Xu BC (2008). Soil-water threshold range of chemical signals and drought tolerance was mediated by ROS homestasis in winter wheat during progressive soil drying. J. Plant Growth Regul. 27: 309-319.
- Wanner LA, Junttila O (1999). Cold-induced freezing tolerance in Arabidopsis. Plant Physiol. 120: 391-400.
- Warren G, Mckown R, Marin A, Teutonico R (1996). Isolation of mutants affecting the development of freezing tolerance in Arabidopsis thaliana (L.) Heynh. Plant Physiol.111: 1011-1019.
- Weaver L, Himelblau E, Amasino R (1997). Leaf senescence: gene expression and regulation. In: Setlow JK, ed. Genetic engineering. principles and methods. New York: Plenum Press 19: 215-234.
- Williams RJ, Leopold AC (1989). The glassy state in corn embryos. Plant Physiol. 89: 977-981.
- Wingler A, Mare's M, Pourtau N (2004). Spatial patterns and metabolic regulation of photosynthetic parameters during leaf. New Phytol. 161: 781-789.
- Xin Z,Browse J (2000). Cold comfort farm: the acclimation of plants to freezing temperatures. Plant Cell Environ. 23: 893-902.
- Xiao BZ, Chen X, Xiang CB, Tang N, Zhang QF, Xiong LZ (2009). Evaluation of seven function-known candidate genes for heir effects on improving drought resistance of transgenic rice under field conditions. Mol. Plant 2: 73-83.
- Zhu JK (2001). Cell signaling under salt, water and cold stresses. Curr. Opin. Plant Biol. 4: 401-406.