SHORT COMMUNICATION

CHEMICAL CONSTITUENTS OF THE ETHYL ACETATE EXTRACTS OF THE STEM BARK AND FRUITS OF DICHROSTACHYS CINEREA AND THE ROOTS OF PARKIA BICOLOR

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ABSTRACT. The antibacterial activities of ethyl acetate, methanol and aqueous extracts of the stem bark of *Dichrostachys cinerea* and the roots of *Parkia bicolor* have been evaluated. Ethyl acetate extracts have been investigated, studies that led to a series of known compounds, amongst which many are reported here for the very first time from both the species.

KEY WORDS: Dichrostachys cinerea, Parkia bicolor, Chemical constituents, Antibacterial activity

INTRODUCTION

D. cinerea and P. bicolor both belong to the Leguminosae family (Mimosaceae). Taking previously as two species of the same genus [1, 2], D. cinerea (Linn) and D. glomerata (Forsk) had appeared to be the same species [3]. The other scientific names are Mimosa cinerea Linn, Mimosa glomerata Forsk, Dichrostachys nutans Benth, Mimosa nutans Benth [3]. Stem barks are used in local medicine in decoction against bad coughs in children, treatment of wounds and gynaecological troubles in adults [1-3]. The roots of P. bicolor are used against children measles, woman sterility and sexually transmitted diseases [4].

The antibacterial activities of fruits and leaves of *D. cinerea* have been described previously [5]. The antibacterial activities of ethyl acetate, methanol and aqueous extracts of the stem bark of *D. cinerea* and the roots of *P. bicolor* are investigated in this study.

Previously isolated constituents of *D. cinerea* are triterpenes [1, 2], sterols [2, 6], tannins [7], flavonoids [6, 7] and of *P. bicolor* are fatty acids [8]. This report describes the isolation of new constituents 7,4'-dihydroxyflavon, 7,3',4'-trihydroxyflavon, apigenin, luteonin, monoglycerides of tetracosanoic and 26-hydroxyhexacosanoic acids, 3-α-L-*O*-rhamnopyranosyl-(2*S*,3*R*)-5,7,4'-trihydroxyflavanon and 3-α-L-*O*-rhamnopyranosyl-(2*S*,3*S*)-5,7,4'-trihydroxyflavanon from *D. cinerea* and lupeol, 1'-monoglyceride of octacosanoic acid, lichexanthone, gallic acid and methyl gallate from *P. bicolor*.

EXPERIMENTAL

General. The stem barks and fruits of *D. cinerea* and the roots of *P. bicolor* were collected at Nkolbisson (Yaounde suburban area) in January 2000. Both were identified by M. Kofani, botanist at the National Herbarium in Yaounde, where voucher specimens have been deposited.

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Antibacterial activities

Inhibition zones were recorded by measuring the diameter of the area showing complete inhibition. The growth inhibition was calculated with reference to positive control, using standard microorganisms from Centre Pasteur of Yaounde. Tests were performed using the cupplate agar diffusion protocol [9]. The results are reported in Table 1.

Table 1. Antibacterial ad	ctivities of crude extra	acts from stem bark of h	D. cinerea and the root	s of <i>P. bicolor</i> .
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	Extract	Inhibition zone (mm)						
		Вс	Cd	Kp	Pm	Pa	St	Sp
Dichrostachys cinerea	Aqueous	9.5	10.0	9.0	10.5	12.0	12.0	9.5
(stem bark)	MeOH	10.5	11.0	10.5	10.0	10.0	10.0	11.5
	AcOEt	8.0	-	9.5	7.0	8.0	-	10.0
Parkia bicolor	Aqueous	10.5	10.0	8.0	13.5	12.0	11.0	12.0
(roots)	MeOH	12.0	11.0	10.5	11.0	13.5	10.5	12.5
	AcOEt	8.5	9.5	10.0	-	8.5	7.0	11.0
Amoxicillin.H ₂ O	(20 μg/mL)	15.0	16.0	11.0	13.0	12.0	15.0	13.0
Ampicillin.H ₂ O	$(20~\mu g/mL)$	15.0	18.0	10.0	15.0	15.0	15.0	10.0

Key. Bc: Bacillus cereus; Cd: Corynebacterium diptheriae; Kp: Klebsiella pneumoniae; Pm: Proteus mirabilis; Pa: Pseudomonas aeroginosa; St: Salmonella typhi; Sp: Streptococcus pyogenes.
Test concentration of extracts: 30 mg/mL; 0.1 mL/cup.

Examination of stem bark of D. cinerea

3 kg of air-dried bark powder was extracted at room temperature with a mixture of $CH_2CI_2/MeOH(1:1)$. The evaporation gave 250 g (8.33%) of a residue which was redissolved in 500 mL of 10% aqueous MeOH. The solution was extracted once with ethyl acetate. After evaporation, 96 g (3.2%) of a sticky brown oil was obtained. Chromatography over silica gel, eluted with mixtures of hexane and ethyl acetate in increasing polarities yielded **1** (200 mg, 0.007%), **2** (400 mg, 0.013%), **3** (1.62 g, 0.054%), **4** (30 mg, 0.001%), **5** (26 mg, 0.0009%), **6** (42 mg, 0.0014%), **7** (23 mg, 0.0008%), **8** (25 mg, 0.0008%), **9** (37 mg, 0.0012%), and **10** (28 mg, 0.0009%).

Compounds 1, 2, 3 and 10 were identified as friedelan-3-one, friedelan-3 β -ol, betulinic acid and 3-O- β -D-glucopyranosyl- β -sitosterol, respectively, as revealed by NMR spectroscopy, mass spectrometry, melting point and specific rotations. 1 and 2 have previously been isolated from roots [6] and stem bark [2] of the same plant, while 3 and 10 was reported only from the stem bark [2, 6].

I'-Monoglyceride of tetracosanoic acid 4. m.p. 115-117 °C (lit. 112-114 °C [10]); IR (KBr): $v = 3450, 3560, 1770 \text{ cm}^{-1}$; MS (EI) m/z 308 (13.8%), 134 (64.4); HRMS (EI) $m/z 442.6895 (C_{27}H_{54}O_4)$; cal. 442.6936. The structure was confirmed by a combination of spectroscopic data, 1H NMR, ^{13}C NMR, HMBC and mass.

I'-Monoglyceride of 26-hydroxyhexacosanoic acid 5. m.p. 106-107 °C (lit.103-104 °C [11]); IR (KBr): v = 3625, 3566, 1705 cm⁻¹; MS (EI) m/z 352 (17.8%), 134 (46.8); HRMS (EI) m/z 486.7510 ($C_{29}H_{58}O_5$); cal. 486.7678. The structure was confirmed by 1H NMR, ^{13}C NMR spectra and HMBC.

Apigenin (4,7,4'-trihydroxyflavon) **6**. m.p. 347-348 °C; MS (EI) m/z 270 [M⁺, 100%]; HRMS (EI) m/z 270.1828 ($C_{15}H_{10}O_5$); cal. 270.2369. ¹H NMR and ¹³C NMR data correlate well with those from the literature [12].

7,4'-Dihydroxyflavon 7. m.p. 328-330 °C; MS (EI) m/z 254 [M+, 76.3%]; HRMS (EI) m/z 254.2579 (C₁₅H₁₀O₄); cal. 254.2375. ¹H NMR and the ¹³C NMR spectra were identical to those reported in the literature [13, 14].

Luteonin (5,7,3',4'-tetrahydroxyflavon) 8. m.p. 288-290 °C, MS (EI) m/z 286 [M⁺, 100%]; HRMS (EI) m/z 286.2177 ($C_{15}H_{10}O_6$); cal. 286.2363. ¹H NMR and ¹³C NMR spectra were identical to those reported in the literature [12, 15].

7,3',4'-Trihydroxyflavon 9. m.p. 352-353 °C; MS (EI) m/z 270 [M⁺, 84.5%]; HRMS(EI) m/z 270.2528 (C₁₅H₁₀O₅); cal. 270.2369. ¹H NMR and ¹³C NMR spectra were identical to those reported in the literature [15].

Examination of the fruits of Dichrostachys cinerea

Powdered air-dried fruits (1 kg) were extracted at room temperature with 5 L of a mixture of CH_2Cl_2 -MeOH (1:1). The evaporation of the solvent gave 200 g (20%) of a residue which was redissolved in 400 mL of 10% aqueous MeOH. The solution was extracted once with ethyl acetate. After evaporation, 56 g (5.6%) of a viscous sticky brown oil was obtained. Chromatography over silica gel eluted with mixtures of hexane and ethyl acetate in increasing polarities gave 7 (40 mg, 0.004%) and 9 (29 mg, 0.0029%) previously isolated from the stem barks, 11 (18 mg, 0.0018%), 12 (26 mg, 0.0026%) and 13 (26 mg, 0.0026%).

Isoastilbin (3-O-α-L-rhamnopyranosyl (2S3R)-5,7,4'-trihydroxyflavanon) 11. m.p. 285-286 °C, MS (EI) m/z 288 (25%), 165 (9.6), 153 (100); $[\alpha]_D^{26}$: –199 (c 0.27, EtOH) (lit.: –196°; c 0.27, EtOH [16]); CD (DMSO, c 1 g/L) $[\Theta]_{255}^{26}$: –25 x 10³ $[\Theta]_{295}^{26}$: +50 x 10³. The structure was determined using ¹H-NMR, ¹³C-NMR, HMQC, HMBC, NOESY and mass spectra. The absolute configuration was obtained from the circular dichroism spectrum [17, 18].

Neoastilbin (3-O-a-L-rhamnopyranosyl (2S3S)-5,7,4'-trihydroxyflavon) 12. m.p. 178-180 °C, $[\alpha]_D^{26}$: -72.5° (c 0.56, EtOH) (lit.: -71.1°, c 0.55, EtOH [16]); CD (DMSO, c 1 g/L) $[\Theta]_{255}^{26}$: -25 x 10³, $[\Theta]_{295}^{26}$: +12 x 10³. The structure was determined by the use of ¹H NMR, ¹³C NMR, HMQC, HMBC, NOESY and mass spectra. The absolute configuration was obtained from the circular dichroism spectrum [17, 18].

Catechine 13. m.p. 249-250 °C; MS (EI) m/z 290 [M+, 23%] 170 (100), 86 (47.8); HRMS (EI) m/z 290.2316 ($C_{15}H_{14}O_6$); cal. 290.2680. The ¹H NMR and ¹³C NMR spectra were identical to those reported in the literature [19].

Examination of the roots of Parkia bicolor

Powdered air-dried roots (3 kg) were extracted at room temperature with 15 L of a mixture of CH₂Cl₂-MeOH (1:1). The evaporation of the solvent gave 300 g (10%) of a residue which was redissolved in 500 mL of 10% aqueous MeOH. The solution was extracted once with ethyl acetate. After evaporation, 87 g (2.9%) of a dark viscous and sticky mixture was obtained. Chromatography over silica gel eluted with mixtures of hexane and ethyl acetate in increasing

polarities yielded **14** (18 mg, 0.0006%), **15** (58 mg, 0.002%), **16** (65 mg, 0.002%), **17** (30 mg, 0.001%) and **18** (40 mg, 0.0013%).

Lichexanthone (1-hydroxy-8-methyl-3,6-dimethoxyxanthone) 14. m.p. 189-190 °C (lit. 189-191 °C [20]); MS (EI) m/z 286 [M+, 100%], 257(43); HRMS (EI) m/z 286.2794 ($C_{16}H_{14}O_{5}$); cal. 286.2793. The structure was determined using ¹H NMR, ¹³C NMR, HMQC, HMBC and mass spectra [21].

Lupeol 15. m.p. 215-216 °C; MS (EI) m/z 426. The structure was determined by the use of ¹H NMR and ¹³C NMR data and by comparison with authentic sample.

1'-Monoglyceride of octacosanoic acid **16**. m.p. 107-108 °C (lit. 110-111 °C [22]); MS (MALDI-TOFF) m/z 520 [M⁺+ Na⁺ - H]; MS (EI) m/z 134 (100%), 294 (12.3%), 378 (17.8%) and 498 (3.2%). The structure was determined by the use of ¹H NMR, ¹³C NMR spectra and HMBC data that were similar to those of compound **4**, and in concordance with the mass spectrum.

Methyl gallate 17. m.p. 201-203 °C (lit. 200-202 °C [23]), MS (EI) m/z 184 [M⁺, 100%]. The structure was determined using ¹H NMR, ¹³C NMR and mass spectra.

Gallic acid 18. m.p. 249-250 °C (lit. 251-252 °C [23]); MS (EI) m/z 170 [M⁺, 100%]. The structure was determined by the use of ${}^{1}H$ NMR, ${}^{13}C$ NMR and mass spectra.

CONCLUSION

Extensive NMR studies were performed in this work. The values for all the protons and the carbons were assigned by the use of ¹H NMR, ¹³C NMR, HMBC, HMQC and NOESY spectra. The absolute configurations of stereogenic centres were determined using the circular dichroism spectra.

All tested extracts showed antibacterial activities. The results may provide a justification to the traditional usage of these plants in the treatment of some infectious diseases. At this level of study, we cannot recommend the intensification of their usage. Tests on toxicity of the extracts are in progress as well as antibacterial tests on isolated compounds.

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