Biokemistri

An International Journal of the Nigerian Society for Experimental Biology



Original Article

The effects of exposure to multiple stressors of Lead (Pb) and Cypermethrin on biochemical profiles of African catfish, *Clarias gariepinus*

Joseph A. Adeyemi^{1*} and Omowumi O. Adewale²

¹Department of Biological Sciences, College of Science, Engineering and Technology, Osun State University, Osogbo, Osun State, Nigeria. ²Department of Chemical Sciences, College of Science, Engineering and Technology, Osun State University, Osogbo, Osun State, Nigeria.

*Corresponding Author: Joseph A. Adeyemi. E-mail: joseph.adeyemi@gmail.com; Tel.: +234 8075195318

Received: 29 May 2013; Revised: 16 June 2013; Accepted: 17 Jun 2013

ABSTRACT: The African catfish (*Clarias gariepinus*) is a widely cultured fish species in many African countries because of its rich nutritional quality. In this study, the impacts of exposure to common environmental contaminants; a heavy metal (Lead) and a pyrethroid insecticide (Cypermethrin) on the biochemical contents of *C. gariepinus* was investigated. Juveniles of *C. gariepinus* were exposed to borehole water (control), 2 mg/L Pb, 0.5 μ g/L cypermethrin or 2 mg/L Pb + 0.5 μ g/L cypermethrin for 96 h after which the total protein levels, glycogen contents and total cholesterol in the liver and muscle were determined. Fish exposure to cypermethrin and a mixture of cypermethrin and Pb resulted in a significant decrease in glycogen and protein levels but a significant increase in the cholesterol levels in liver and muscle (p < 0.05). There was no significant difference between the control and the group exposed to 2 mg/L Pb. The glycogen and total cholesterol levels were significantly higher in the liver in groups exposed to cypermethrin and the mixture of cypermethrin and Pb (p < 0.05) while the total protein levels were significantly higher in the liver in groups exposed to cypermethrin and the mixture of cypermethrin and Pb (p < 0.05) while the total protein levels were higher in the muscle, although this difference was not statistically significant. The results of this study suggest that environmental pollution of aquatic environments have adverse effects on the health of resident fish as well as other aquatic life.

KEYWORDS: Environmental contamination; African catfish, *Clarias gariepinus*; glycogen content; total protein; cholesterol level.

BKM.2013.017 © 2013 Nigerian Society for Experimental Biology; All rights reserved. Printed in Nigeria This article is downloadable online in PDF format at http://www.bioline.org.br/bk

INTRODUCTION

The aquatic ecosystem is constantly being threatened by the release of various environmental contaminants into the environments (Islam and Tanaka, 2004; Zhou *et al.*, 2008). Various human activities such as agriculture and mining have contributed to the release of different forms of pollutants into the environments. For example, agricultural pesticides and insecticides are routinely applied to agricultural crops and planting fields in order to control pests and unwanted weeds (Anyim, 2004). There are also reports of anthropogenic release of various heavy metals into the environments (Samecka-Cymerman and Kempers, 2001). Lead is one of such heavy metals that are very common in various ecosystems; land, water and air. Lead is released into the

environment through mining activities and wastes from different industrial and commercial usage e.g. in lead batteries, lead pipes, electrical cables, tennis rackets, glazing bar for stained glass etc. While most of these chemicals are most of the times not directly applied into the aquatic environments, however natural phenomenon such as leaching and erosion gets them into aquatic environments; aquatic environments serve as sinks and they can then be accumulated to levels where they interfere with the physiology of resident organisms. Fish being one the most important occupants of the aquatic environments provides human with rich nutritive diet. While various indices such as oxidative stress biomarkers, changes in plasma ions, changes in hormonal levels, changes in biochemical enzymes etc. have been employed to show the impacts of environmental contamination on the physiology and biochemistry of fish inhabiting polluted sites, only few of these indices links the physiological status of stressed fish to the nutritional values that could be derived when such contaminated fish are consumed. However, change in biochemical profiles such as glycogen, protein and lipid levels serves these purposes.

There are reports of toxic effects of Pb and cypermethrin on aquatic organisms. Cypermethrin results in mortality in the freshwater fish, Poecilia reticulata (Gautam and Gupta, 2008), disruption of glycogen metabolism in the estuarine clam, Marcia opima (Tendulka and Kulkarni, 2012), reduced growth and abnormal changes in gonadal development in Odontesthes bonariensis (Carriquiriborde et al., 2009). Pb is neurotoxic and there are reports of Pb resulting in behavioral impairment in adult zebrafish (Chen et al., 2012), hematological changes in African catfish (Adeyemo, 2009) etc. Despite the vast documentation of these toxicity data, not much has been done to show the likely implications of exposure to mixture of lead and cypermethrin on the alteration of biochemical profiles in fish. This present study is therefore designed to investigate the effects of exposure to either cypermethrin or Pb on the biochemical contents of C. gariepinus, as well as the effects of exposure to the mixture of cypermethrin and Pb. We also showed the tissue-specific patterns in the alteration of biochemical profiles in contaminated C. gariepinus.

MATERIALS AND METHODS

Experimental organisms and contaminant exposure

Juvenile catfish (N=80) of average length 10 \pm 2.5 cm were collected from a commercial fish hatchery located in Abere, Osun State, Nigeria (07°43'55N; 004°31'07E). They were then transported live in plastic containers to the Research Laboratory of the Department of Biological Sciences of the Osun State University, Osogbo, Nigeria. Fish were acclimatized to laboratory conditions for seven days prior to commencement of experiments. During acclimatization, water was exchanged every 24 h and fish were fed with commercial fish pellets twice daily. At the end of the sevenday acclimatization period, fish were divided into four groups of about twenty fish per group; control, 2 mg/L Pb, 0.5 μ g/L cypermethrin and 2 mg/L Pb + 0.5 μ g/L. Experiments were conducted in 7.5 L plastic buckets and contaminant exposure lasted for 96 h. Three separate buckets containing 6-7 fish each were used for each treatment.

Sample preparation for biochemical analyses

After the 96 h period of contaminant exposure, 6 fish were sampled from each treatment group (at least 2 fish from each of the three buckets in the treatment group) and were euthanized with an overdose of benzocaine (200 mg/L), and the liver and muscle were carefully dissected out for biochemical analyses.

Glycogen contents determination

Liver and muscle glycogen content was determined using the anthrone reagent in a method developed by Nicholas et al. (1956) and modified by Reddy et al. (1994). Briefly, liver tissue was homogenized in 10% trichloroacetic acid (TCA). A volume of 0.2 ml of the homogenate was then transferred into a microcentrifuge tube and precipitated by adding 1 ml of absolute ethanol. The solution was then incubated for 3 h at 37 C. After the 3 h incubation, the solution was centrifuged at 3,000 g for 15 min. After this step, 0.2 ml of the supernatant was transferred into another tube for glucose measurement. A 1-ml volume of freshly prepared anthrone reagent was added to the sample and the mixture boiled in a waterbath for 15 min, after which it was allowed to cool to room temperature. The absorbance was then measured at 620 nm using an UV/Vis spectrophotometer (Jenway, Model 6400), and liver and muscle glycogen was expressed as mg glycogen per mg tissue wet weight, using a standard curve obtained with glucose solutions.

Total protein determination

The total protein levels in the liver and muscle were quantitatively determined using the Bradford assay (Bradford, 1976). Bovine serum albumin (BSA) was used as the standard.

Total cholesterol determination

The total cholesterol levels in the liver and muscle samples of C. gariepinus were determined colorimetrically using Liebermann-Burchard method reported in Burke et al. (1974). This reaction is based on reaction between acetic anhydride and cholesterol in chloroform solution to form a characteristic blue colour. Briefly, samples were homogenized in ice-cold phosphate buffer (pH, 7.4). Then, 0.2 ml of the homogenate was transferred into centrifuge tubes that contain about 5 ml of alcohol-acetone solvent solution. The tube was then immersed into a boiling water bath until the solvent began to boil, after which the tubes were removed from the water bath and allowed to cool to room temperature. The content of the tube was then centrifuged lightly, and the supernatant was discarded while the pellet was allowed to evaporate to dryness in a boiling water bath. The residue was then reconstituted by adding 2 ml of chloroform, followed by addition of 2 ml of acetic anhydride/concentrated sulphuric acid mixture. The tubes were left in the dark for about 10 min. after which the absorbance was read at 620 nm. The total cholesterol levels in liver and muscle was expressed as mg total cholesterol per mg tissue wet weight, using a standard curve obtained with cholesterol solutions.

Statistical analysis

Data were parametrically analyzed since there was no significant deviation from normal distribution. Data were first subjected to a two-way mixed model analysis of variance, factors being the treatment group and experimental buckets (since multiple buckets were used per treatment group). Because the experimental bucket effect was not significant, this factor was eliminated from the model, and the data were subsequently analyzed using a one-way analysis of variance. For each tissue samples, the difference in mean glycogen contents, protein levels and total cholesterol was detected using one-way analysis of variance. This was followed by Tukey's multiple comparison tests whenever there was a significant difference. We also performed a student's t- test in order to show the difference in the tissue glycogen, protein and cholesterol levels between the liver and the muscle. All statistics were performed using JMP version 9.0 software (SAS Inc., 2010). For reporting purposes, data were expressed as mean ± standard error and statistical significance was assumed at $P \le 0.05$.

RESULTS

Liver and muscle glycogen contents

There was a significant difference in the glycogen content in both the liver ($F_{3, 20} = 7.5208$, p = 0.0029) and muscle samples ($F_{3, 20} = 6.3306$, p = 0.0046) among the groups. Fish exposed to cypermethrin and a mixture of cypermethrin and lead had a significantly reduced level of tissue glycogen compared to the control and those exposed to 2 mg/L Pb. The Liver glycogen level is higher in the liver than in the muscle, although this difference was only significant in the groups exposed to cypermethrin and the mixture of cypermethrin and Pb (Figure 1).

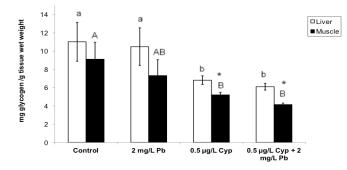


Figure 1: The tissue glycogen contents in *C. gariepinus* exposed to either borehole water (control), 2 mg/L Pb, 0.5μ g/L cypermethrin or 0.5μ g/L cypermethrin + 2 mg/L Pb. Each bar is mean ± standard deviation (n = 6). Bars with different letters are significantly different (upper case = muscle, lower case = liver). The asterisk indicates a significant difference between the liver and muscle levels for a particular treatment

Liver and muscle protein level

There was a significant reduction in the total protein level in both the liver ($F_{3, 20} = 8.7402$, p = 0.0046) and muscle samples ($F_{3, 20} = 9.8306$, p = 0.0032) after contaminant exposure. As in glycogen levels, total protein decreased significantly in fish exposed to either $0.5 \mu g/L$ cypermethrin or those exposed to the mixture of cypermethrin and Pb. However, unlike in glycogen content data where the values were higher in the liver, the total protein levels in the muscle were higher in all the treatment groups but the difference was not significant (Figure 2).

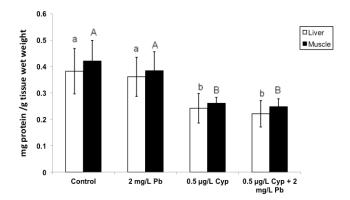


Figure 2: The tissue total protein levels in *C. gariepinus* exposed to either borehole water (control), 2 mg/L Pb, 0.5μ g/L cypermethrin or 0.5μ g/L cypermethrin + 2 mg/L Pb. Each bar is mean ± standard deviation (n = 6). Bars with different letters are significantly different (upper case = muscle, lower case = liver)

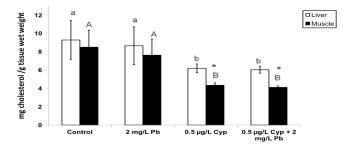


Figure 3: The tissue total cholesterol levels in *C. gariepinus* exposed to either borehole water (control), 2 mg/L Pb, 0.5μ g/L cypermethrin or 0.5μ g/L cypermethrin + 2 mg/L Pb. Each bar is mean ± standard deviation (n = 6). Bars with different letters are significantly different (upper case = muscle, lower case = liver). The asterisk indicates a significant difference between the liver and muscle levels for a particular treatment

Liver and muscle total cholesterol level

The total cholesterol level was significantly higher in the liver ($F_{3, 20} = 5.6404$, p = 0.0146) and in the muscle ($F_{3, 20} = 6.5402$, p = 0.0096) in the groups exposed to 0.5 μ g/L cypermethrin and those exposed to the mixture of cypermethrin and lead compared to the control and the group exposed to just 2 mg/L Pb. The total cholesterol levels in the liver were significantly higher than the levels in the muscle in the groups exposed to cypermethrin and Pb (Figure 3).

DISCUSSION

Fish are consumed for their rich nutritive value and affordability. Fish, being one of the most abundant groups in the aquatic ecosystem are sometimes exposed to all sorts of environmental assaults such as heavy metal pollution and agricultural chemicals contamination, which can consequently have a serious effect on the nutritive quality of fish inhabiting the contaminated aquatic environments. In Nigeria, the African catfish is probably the most cultured fish species owing to its numerous desirable characteristics; high tolerance of extreme environmental conditions, ease of cultivation, high guality flesh, distinctive taste and texture, relatively low fat and absence of intramuscular spines (El-Shebly, 2006; Khwuanjai et al., 1997; Luckhoff, 2005).

The results of this study indicate that environmental pollution with heavy metals and run-offs of agricultural chemicals into aquatic environments have adverse effects on the biochemical contents of fish inhabiting contaminated sites. C. gariepinus that were exposed to cypermethrin and lead showed a significant decrease in glycogen contents compared to the control. This result is consistent with the findings of Tendulka and Kulkarni (2012), which reported disruption of glycogen metabolism in the estuarine clam, Marcia opima exposed to cypermethrin. The decreased glycogen levels in the exposed fish could be due to increased glycogenolysis in order to make more glucose available in response to a higher energy demand associated with contaminant exposure (Hargreaves et al., 1995). In addition, the low glycogen levels in stressed fish could be due to contaminants' interference with the mechanisms involved glycogen synthesis since there are reports of heavy metals and organic pesticides directly modulating the processes involved in glycogen biosynthesis (Misra et al., 1991). The higher liver glycogen contents in comparison to the lower level in the muscle could be due to the fact that liver is principally involved in numerous metabolic activities and more importantly, contaminant detoxification occurs in the liver (Sahi et al., 2010). These processes are energetically expensive, so a higher quantity of energy substrate molecules in the liver is guite understandable.

The tissue protein level in *C. gariepinus* decreased significantly after exposure to cypermethrin and a mixture of

cypermethrin and lead. The reduced protein level in the liver and muscle of exposed fish could infer a possible use of protein as additional energy substrate molecule so as to augment the available glycogen. There are reports that in circumstances of low glucose or glycogen levels, other molecules could be used by the cells to produce energy required for essential metabolic processes (Brooks, 1987). It is also possible that the reduction in protein level is actually due to more utilization of protein in the repair of damaged tissue as a result of contaminant exposure or possibly a reduction in the rate of synthesis of protein. (Reichert et al., 1998). The total protein level in the muscle is higher than the level measured in the liver. This may be an indication that the muscle of C. gariepinus is richer in protein than the liver, although one cannot also overrule the possibility that the lower protein level in the liver is due to the fact that more energy demanding metabolic activities occurs in the liver, which could have used some of the available protein as an alternative source of energy. In contrast to the reduced glycogen and total protein levels in the liver and muscle of exposed fish, exposure to cypermethrin and lead resulted in an increase in the levels of total cholesterol in the liver and muscle. This may be an indication that contaminant exposure could disrupt the catabolism of cholesterol thus enhancing the build-up of cholesterol in the muscle and liver (Dutta and Haghighi, 1986). This may have a direct negative impact on the physiology of exposed fish, and in particular to man when such fish with high cholesterol levels are consumed.

In conclusion, we show in this study that the alteration in tissue biochemical profiles is a reliable index of stress in *C. gariepinus*. While there was no direct evidence that exposure to 2 mg/L Pb has a significant adverse effect on the biochemical profile of *C. gariepinus*, however there are some evidence of synergistic effects in this study since alteration in the biochemical profile was more significant in fish exposed to a mixture of cypermethrin and Pb, in comparison to those exposed to just only cypermethrin.

ACKNOWLEDGEMENTS

This work is funded in part through financial support from African-German Network for Excellence in Science (AGNES) grant awarded to Joseph Adeyemi.

REFERENCES

Adeyemo OK (2007) Haematological profile of *Clarias gariepinus* (Burchell, 1822) exposed to Lead. *Turkish Journal of Fisheries and Aquatic Sciences* 7: 163-169.

Anyim A (2004) Effects of insecticidal treatment on the yield and control of major insect pests of soybean (*Glycine max* (L.) Merrill) in South-Eastern Nigeria. *International Journal of Agriculture and Rural Development* 4: 100-109.

Bradford MM (1976) A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the

principle of protein–dye binding. *Analytical Biochemistry* 72: 248–254.

Brooks GA (1987) Amino acid and protein metabolism during exercise and recovery. *Medicine and Science in Sports and Exercise* 19: 150-156.

Burke RW, Diamondstone BI, Velapoldi RA and Menis O (1974) Mechanisms of the Liebermann-Burchard and Zak color reactions for cholesterol. *Clinical Chemistry* 20: 794-801.

Carriquiriborde P, Díaz J, López GC, Ronco AE and Somoz GM (2009) Effects of cypermethrin chronic exposure and water temperature on survival, growth, sex differentiation, and gonadal developmental stages of *Odontesthes bonariensis* (Teleostei). *Chemosphere* 76: 374–380.

Chen J, Chen Y, Liu W, Bai C, Liu X, Liu K, Li R, Zhu JH and Huang C (2012) Developmental lead acetate exposure induces embryonic toxicity and memory deficit in adult zebrafish. *Neurotoxicology and Teratology* 34: 581-586.

Dutta HM and Haghighi AZ (1986) Methylmercuric chloride and serum cholesterol level in the bluegill (*Lepomis macrochirus*). *Bulletin of Environmental Contamination and Toxicology* 36: 181-185.

El-Shebly AA (2006) Evaluation of growth performance, production and nutritive value of the African catfish, *Clarias gariepinus* cultured in earthen ponds. *Egyptian Journal of Aquatic Biology and Fisheries* 10: 55-67.

Gautam PP and Gupta AK (2008) Toxicity of cypermethrin to the juveniles of freshwater fish *Poecilia reticulate* (Peters) in relation to selected environmental variables. *Natural Product Radiance* 7: 314-319.

Hargreaves M, McConell G and Proietto J (1995) Influence of muscle glycogen on glycogenolysis and glucose uptake during exercise in humans. *Journal of Applied Physiology* 78: 288-292.

Islam MS and Tanaka M (2004) Impacts of pollution on coastal and marine ecosystems including coastal and marine fisheries and approach for management: a review and synthesis. *Marine Pollution Bull*etin 48: 624–649.

Khwuanjai H, Ward FJ and Pornchai J (1997) The effect of stocking density on yield, growth and mortality of African catfish (*Clarias gariepinus* Burchell, 1822) cultured in cages. *Aquaculture* 152: 67-76.

Luckhoff PD (2005) Application of the condition factor in the production of African sharptooth catfish, *Clarias gariepinus*. Unpublished M.Sc. Thesis, University of Stellenbosch, South Africa. 62pp.

Misra V, Kumar V, Pandey SD and Viswanathan PN (1991) Biochemical alterations in fish fingerlings (*Cyprinus carpio*) exposed to sublethal concentration of linear alkyl benzene sulphonate. Archives of Environmental Contamination and Toxicology 21: 514-517.

Nicholas V, Roe JH, Longley GW and Joseph HR (1956) The determination of glycogen in liver and muscle by use of anthrone reagent. *Journal of Biological Chemistry* 220: 583-593.

Reddy DS, Ghanathay VV, Reddy SLN and Shankariah K (1994) Hepatotoxic effects of hexachlorocyclohexane on carbohydrate metabolism of a freshwater fish *Channa punctatus* (Bloch). *Bulletin of Environmental Contamination and Toxicology* 53: 733-739.

Reichert WL, Myers MS, Peck-Miller K, French B, Anulacion BF, Collier TK, Stein JE and Varanasi U (1998) Molecular epizootiology of genotoxic events in marine fish: Linking contaminant exposure, DNA damage, and tissue-level alterations. *Mutation Research* 411: 215–225.

Sahi J, Grepper S and Smith C (2010) Hepatocytes as a tool in drug metabolism, transport and safety evaluations in drug discovery. *Current Drug Discovery Technologies* 7: 188-198.

Samecka-Cymerman J and Kempers AJ (2001) Concentrations of heavy metals and plant nutrients in water, sediments and aquatic macrophytes of anthropogenic lakes (former open cut brown coal mines) differing in stage of acidification. *Science of The Total Environment* 281: 87-98.

Tendulkar M and Kulkarni A (2012) Cypermethrin-induced toxic effect on glycogen metabolism in estuarine clam, *Marcia opima* (Gmelin, 1791) of Ratnagiri Coast, Maharashtra. *Journal of Toxicology* 2012: 576804.

Zhou Q, Zhang J, Fu J, Shi J and Jiang G (2008) Biomonitoring: An appealing tool for assessment of metal pollution in the aquatic ecosystem. *Analytica Chimica Acta* 606: 135-150.