

Prevalence and etiology of mastitis in traditionally managed camels (*Camelus dromedarius*) in selected pastoral areas in eastern Ethiopia

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Abstract

Prevalence and causes of mastitis in traditionally managed camels in selected pastoral areas in eastern Ethiopia were assessed. The prevalence of camel mastitis was determined by the California mastitis test (CMT) and by clinical examinations of the udder and milk samples. A total of 642 udder quarters from 161 camels were examined from three locations for the determination of clinical cases and CMT. Among the CMT positive milk samples and milk samples collected from clinical quarters, 174 randomly selected samples were further examined for identification of the etiological agents of camel mastitis. The overall prevalence of mastitis observed in camel herds examined as assessed by the CMT and clinical examinations of the udder or milk (76.0%) was very high. The high prevalence of mastitis in the camel herds examined could be attributed to the unhygienic milking procedures followed by the camel owners and the poor hygienic condition of the milking area. The bacterial species isolated from the camel milk samples include coagulase negative staphylococci (39.6%), *Streptococcus dysgalactiae* (22.2%), *Corynebacteria* spp. (9%), *Bacillus* spp. (7.6%), *Streptococcus uberis* (7.6%), *Escherichia coli* (6.3%), *Staphylococcus aureus* (4.2%) and *Streptococcus agalactiae* (3.5%). These results suggest that measures need to be taken to improve the health of camels and the quality of camel milk in the study areas.

Keywords: Camel; Causative agent; Mastitis; Prevalence; Udder health.

Introduction

The one-humped camel (*Camelus dromedarius*) plays an important role as a primary source of subsistence in the lowlands of Ethiopia. Camels live in arid and semi-arid areas which are not suitable for crop production and where other livestock species hardly thrive. Because of their outstanding performance in the arid and semiarid environments of eastern Ethiopia where browse and water are limited, pastoralists rely mainly on camels for their livelihood. In these areas, camel is mainly kept for milk production and produce milk for a lon-

ger period of time even during the dry season when milk from cattle is scarce (Bekele Tafesse et al., 2002). Ethiopia possesses over 1 million dromedary camels (FAO, 2002) and the majority of these camels are found in eastern part of the country. The annual camel milk production in Ethiopia is estimated to be 75,000 tones (Getachew Felleke, 2003).

Mastitis is a complex disease occurring worldwide among dairy animals with heavy economic losses. Mastitis results in milk compositional changes such as increase in leukocyte counts, leakage of plasma proteins into the milk, cell damage resulting in leakage of intracellular constituents into milk, change in ion composition and decrease in milk production (Korhonen and Kaartinen, 1995).

Bacterial infections are considered the primary cause of mastitis in domestic animals. The causative agents of bovine mastitis are well defined but as far as camels are concerned, there is paucity of information about the etiological agents associated with camel mastitis. Few available literatures indicate that *Staphylococcus aureus*, *Streptococcus* spp. (Barbour et al., 1985; Abdurahman et al., 1995; Al-Ani and Al-Shareefi, 1997; Younan et al., 2001), *Micrococcus* spp. (Barbour et al., 1985; Al-Ani and Al-Shareefi, 1997), *Streptococcus agalactiae* (Abdurahman et al., 1995; Younan et al., 2001), coagulase negative staphylococci (Abdurahman et al., 1995), *Staphylococcus epidermidis*, *Pasteurella haemolytica* (Al-Ani and Al-Shareefi, 1997), *Escherichia coli* (Abdurahman et al., 1995; Al-Ani and Al-Shareefi, 1997) and *Corynebacterium* spp. (Barbour et al., 1985) have been implicated as causes of mastitis in camels.

There is extensive literature on bovine mastitis and to a lesser extent on ovine and caprine mastitis; however, little is known about mastitis in camels. There is limited information on the prevalence and causative agents of camel mastitis in Ethiopia. The prevalence and causes of mastitis differ markedly due to geographical area and individual herd management (Guidry, 1985). To establish an efficient mastitis control program in a dairy herd, baseline information on the nature of mastitis and economic impact of the problem need to be known (Honkanen-Buzalski and Pyörälä, 1995). The principal steps in mastitis control program are to undertake a preliminary mastitis screening survey and to evaluate the udder health status in the herd (Honkanen-Buzalski and Pyörälä, 1995). This study was, therefore, conducted with the objectives to determine the prevalence of mastitis in traditionally managed camels in Jijiga, Shinile and Errer valley and identify the major etiological agents of camel mastitis at the study areas.

Materials and methods

Description of the study area

This study was conducted in Shinile, Jijiga and Errer valley in eastern part of Ethiopia. The areas are characterized by unreliable and erratic rainfall with a precipitation ranging from 300 to 600 mm per annum, high ambient temperature ($>30^{\circ}\text{C}$), sparsely distributed vegetation dominated by *Acacia* species, cactus and bushy woodlands (Bekele Tafesse, 2001). These are arid and semi-arid lowlands lying at an altitude of 500-1500 m above sea level and are not suitable for crop production. In these areas, camels are herded by nomadic pastoralists who rely mainly on livestock husbandry for their livelihood.

Survey on prevalence and traditional control methods of mastitis in camels

A single-visit, multiple-subject diagnostic survey (ILCA, 1990) was used to assess the occurrence of mastitis and traditional management practices used to control mastitis in camels. A total of 73 households who own camels and who are familiar with camel husbandry were selected from Jijiga ($n = 31$), Shinile ($n = 32$) and Errer valley ($n = 10$) using purposive sampling technique. Households at each location were selected based on accessibility of the village and willingness of the camel owners to take part in the interview.

The camels were at different stages and numbers of lactation, and they were of various age groups. Information about traditional management, herd size, milking frequency, milking procedure, occurrence of mastitis, and traditional mastitis control methods was obtained from camel owners by means of a semi-structured questionnaire. The camels were fed exclusively on natural browse, watered on the average every 3-4 days, herded during the daytime on communal grazing lands and kept at night in traditional enclosures (Corral) made of thorny bushes and tree branches as protection from predators. The camels were milked on the average three times a day.

Milk sample collection

A total of 642 udder quarters (410 from Jijiga, 72 from Shinile and 160 from Errer valley) were examined and 634 quarter milk samples were aseptically collected from 161 lactating camels and examined for clinical cases and/or subjected to the California mastitis test (CMT). Milk samples were collected from individual quarters during morning milking time. Before milking, all quarters were carefully examined by visual observation and palpation. The teat ends

were thoroughly washed with water and dried with clean towel. Then teats were disinfected using swabs which were dipped into 70% ethanol. The first three to four squirts of milk from each quarter were drawn onto a strip cup and examined for the presence of clots, flakes, blood or pus and change in the colour of milk. Approximately two to three ml of milk from each quarter was drawn onto paddle cups for the CMT test. Then ten ml of milk was collected from each teat directly into clean and sterile plastic tubes and transported to the laboratory in an icebox for bacteriological examination.

Determination of prevalence

Prevalence of clinical mastitis was determined by visual observation and palpation of the udders in addition to milk samples.

California mastitis test

CMT on camel milk samples was conducted using the method described by Schalm and Noorlander (1957) immediately after collecting the milk samples. Scores represented four categories: 0, negative (-) or trace (\pm); 1, positive (+); 2, positive (++) and 3, positive (+++). Negative (-) and trace (\pm) reactions were considered as "negatives" and different intensities of positive reactions (+, ++, +++) were considered as "positives".

Bacteriological examinations

Among the CMT positive milk samples (433) and milk samples collected from clinical quarters (47), 174 samples were randomly selected and used for bacteriological analysis. Bacterial isolation and identification were done at the International Livestock Research Institute laboratory (Debre Zeit, Ethiopia) according to standard procedures. Each milk sample (10 μ l) was streaked onto a plate of blood agar (5% defibrinated sheep blood) (Oxoid, Hampshire, England). Plates were incubated at 37°C for 48 h. The colonies grown were subjected to the following tests as recommended by the National Mastitis Council (NMC, 1987): morphology, haemolysis pattern and Gram stain. Gram-positive cocci were tested for catalase, and catalase-positive isolates further tested with coagulase test. Streptococci were identified by performing CAMP, escilin, hippurate, raffinose, salicin, mannitol, and inulin tests. Gram-negative rods were further differentiated by testing for motility, lactose fermentation (growth on MacConkey agar) and by using oxidase test.

Statistical analysis

Descriptive statistics was used to analyze the data using the Minitab software version 12.21 (Minitab, 1998).

Results

The average herd size per household and milking procedure in study area are indicated in Table 1. Variation in milking procedure was observed between the three locations studied (Table 1). In all the three locations, milk ejection was initiated by letting the calves to suckle their dams before milking. Washing the udder/teats of camels is not practiced in all the three locations prior to milking. The majority of camel owners in Shinile areas (81.3%, n = 32) wash their hands before milking; however, small proportion (30%, n = 10) of the respondents in Error valley reported that they wash their hands before milking. Half of the respondents in Shinile and Error valley reported that they wash and smoke milking utensils before milking camels. In Jijiga, the camel owners neither wash milking utensils nor their hands before milking.

Of the 73 households interviewed, the majority of the respondents (96%) reported that they use traditional medicines to treat camel mastitis. The most common traditional practices used to treat camel mastitis were use of various plant species and branding with hot iron (Table 1). The camel owners use extracts from the roots, leaves, seeds and exudates of different plant species to treat camel mastitis (Table 4).

Camel mastitis is known by different names in the study areas (Table 1). Gofla is the predominant type of camel mastitis in the study areas and it causes a significant decline in milk yield as reported by the respondents. It is a clinical type characterized by swelling of the udder. Arar (Carcar) is a mild type and the second prevalent type of camel mastitis in the areas. It causes swelling of the udder and release of pus from the teats. Jid is the third abundant type of mastitis in camels. It is a chronic form and causes blind teats.

Out of the 161 camels examined, 13 animals (8.1%) showed clinical cases characterized by swollen, reddened, hot and hardened udder and alteration in the colour and consistency of the milk. Of the 642 quarters examined, eight teats (1.2%) were blind. The prevalence of sub-clinical mastitis in camel herds as assessed by the CMT was 67.4% (Table 2). Out of the total 161 camels examined, 57 (35.4%) were infested by ticks (Table 1). Among the milk samples subjected

to bacteriological examination, 144 (82.8%) yielded mastitis pathogens (Table 3).

Table 1. Herd size (mean \pm SD), milking procedure, tick infestation and traditional control measures of camel mastitis in Jijiga, Shinile and Errer valley

Parameters	Area			Overall average
	Jijiga (n = 31)	Shinile (n = 32)	Errer valley (n = 10)	
Herd size per household (mean \pm SD)	33.5 \pm 30.6	11 \pm 11.4	13.2 \pm 10.4	21.4 \pm 24.5
Milking procedure (% of total respondents)				
Wash udder/teats before milking.	none	none	none	none
Wash hands before milking.	none		30%	39.7%
Wash/smoke milk utensils before milking.	none	50%	50%	28.8%
Let the calf to suckle before milking.	100%		100%	100%
Traditional control method (% of total respondents)				
Use leaves, roots & exudates of various plant spp.	48.4%	68.8%	80%	61.6%
Branding with hot iron.	25.8%	31.3%	0	24.7%
Local name of camel mastitis				
Gofla	45.2%	68.8%	40%	54.8%
Arar (Carcar)	64.5%	25%	20%	41.1%
Jid	9.7%	0	60%	12.3%
Tick infestation (% of total herd) ^a				35.4%

^aTotal number of camels examined were 161; n = number of households.

Table 2: California mastitis test (CMT) scores in quarter milk samples and clinical examination of udder quarters of camels in Jijiga, Shinile and Errer valley (n = 642)

CMT scores						Visual examination		Mastitis Prevalence
	-ve	Trace	+1	+2	+3	Clinical ^a	Blind	
Jijiga (n = 410)	56 (13.7)	39 (9.5)	165 (40.2)	92 (22.4)	27 (6.6)	28 (6.8)	3 (0.73)	76.8%
Shinile (n = 72)	8 (11.1)	21 (29.2)	21 (29.2)	11 (15.3)	7 (9.7)	2 (2.8)	2 (2.8)	59.7%
Errer (n = 160)	4 (2.5)	26 (16.3)	45 (28.1)	48 (30)	17 (10.6)	17 (10.6)	3 (1.9)	81.3%
Average (n = 642)	68 (10.6)	86 (13.4)	231 (35.9)	151 (23.5)	51 (7.9)	47 (7.3)	8 (1.2)	76.0%

^aClinical case refers to swollen, reddened, hardened and hot udder; Values in the Table indicate the frequency of each variable measured at each location and values in parenthesis represent the proportion (%) of the variables measured; n = number of quarters examined.

Table 3. Bacterial species isolated from quarter milk samples (n = 174) obtained from traditionally managed camels in Jijiga, Shinile and Errer valley

Bacterial species	Number of isolates	% of total isolates
Coagulase negative staphylococci	57	39.6
Streptococcus dysagalactiae	32	22.2
Corynebacteria spp.	13	9.0
Bacillus spp.	11	7.6
Streptococcus uberis	11	7.6
Escherichia coli	9	6.3
Staphylococcus aureus	6	4.2
Streptococcus agalactiae	5	3.5
Total	144	100

Table 4. Major plant species traditionally used to treat camel mastitis in Jijiga, Shinile and Errer valley

Vernacular name (Somali)	Category	Scientific name	Part of plant used
Qalaan	shrub	Cadaba rotundifolia	Leaf
Aday (Caday)	shrub	Salvadora persica	Leaf and root
Dhamajo	tree	Crotolaria albicallis	Exudate
Malmaal	tree	Boswellia spp.	Exudate (Resin)
Shifu	herb	Trigonella foenum	Seed
Irgin	shrub	Euphorbia tirucalli	Exudate
Je'ee (jei, jiic)	shrub	Maerua angolensis	Leaf and root

Discussion

The overall prevalence (76.0%) of mastitis in camel herds as determined by the CMT and clinical examinations of the udder and the milk samples is higher than that reported by Obied et al. (1996) who found an overall mastitis prevalence of 66.8% in Sudanese camel herds. However, the present finding is consistent with the findings of Woubit Salah et al. (2001) who reported high prevalence of mastitis in dromedary camels in Borena areas of southern Ethiopia. The apparently high prevalence of mastitis in the camel herds examined might be attributed to the high tick infestation rate. The infested udders (98.3%) had one or more CMT positive quarters. Tick infestation can predispose camel udders to bacterial infection (Abdurahman, 1996; Obied et al., 1996).

Environmental factors play significant role in the prevalence of sub-clinical (Sandholm, 1995) and clinical (Honkanen-Buzalski and Pyörälä, 1995) mastitis

in dairy animals. The high prevalence of mastitis observed in the camel herds examined in the present study could be associated with the enclosures where the camels were kept overnight. Camels were kept overnight in an open Corral made of thorny bushes and tree branches which might have contributed to udder injuries and predisposed the udder to infection by environmental pathogens causing mastitis. An injury to the udder causing tissue damage may be reflected in a considerable increase in somatic cell counts in milk (Saloniemi, 1995). Moreover, the unhygienic milking procedure (Table 1) and the generally poor management practiced in the study area might also have contributed to the high prevalence of mastitis in the camel herds examined.

The high percentage of mastitis pathogens isolated from camel milk samples examined in the present study is consistent with the findings of Woubit Salah et al. (2001) who reported that 74% of the CMT positive quarter milk samples of camels in Borena area of southern Ethiopia yielded pathogenic bacteria. Gram-positive cocci were the main cause of mastitis in the camels and constituted 93.8% of the total isolates. This finding is in line with that reported by Obied et al. (1996) and Woubit Salah et al. (2001). Among the bacterial isolates, coagulase negative staphylococci (CNS) were identified as the predominant mastitis causing organisms in the camels studied. This agrees with the report of Abdurahman (1996) who found that CNS and *Staphylococcus aureus* represented 61.1% and 38.9%, respectively of the total isolates and considered as the main organisms that cause mastitis in the Bactrian camel. *Streptococcus dysagalactiae* was the second most common cause of mastitis in the camel herds examined in this study. This finding agrees with that reported previously by other researchers (Abdurahman, 1996; Woubit Salah et al., 2001; Guliye et al., 2002). *Streptococcus agalactiae* and *Staphylococcus aureus* were reported to be the most common causes of camel mastitis in eastern Sudan (Obied et al., 1996) and Kenya (Younan et al., 2001).

The bacteria isolated from camel milk samples in the present study are types that cause both contagious and environmental mastitis. Correct and good milking techniques are essential in the prevention of both environmental and contagious mastitis. The teats must be cleaned with individual clothes dipped in hot water. The fact that the pathogens isolated from camel milk samples in the present study are bacteria that cause both environmental and contagious mastitis suggest that proper management and adequate hygienic condition of the environment (enclosures) are required in order to minimize occurrence of mastitis in the study area.

Pastoralists use various traditional (ethno-veterinary) practices to treat sick camels (Bornstein, 1993; Hussein, 1993). The different plant species traditionally used to treat camel mastitis in the study area would be of paramount importance in the future to develop modern drugs which could be used to treat mastitis in camels. Thus, detailed scientific study needs to be conducted on these plant species to determine their potential medicinal properties.

Conclusion

This study revealed high prevalence of mastitis in camel herds in the study areas. The high prevalence of mastitis was attributed to inadequate hygienic condition of the dairy environment and tick infestation. Thus, good management practices with proper sanitation and tick control measures are required to prevent the incidence of mammary infection in camels in the study areas. The predominant etiological agents of camel mastitis in the study area were found to be coagulase negative staphylococci. The isolation of genera of pathogenic bacteria from the camel milk samples suggests the need for strict hygienic measures during the production and handling of camel milk to reduce public health hazards. Education of the camel owners about the importance of hygienic milking practices would minimize the adverse effect of mastitis on the yield and quality of camel milk.

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