Changes in platelet functional parameters and CD62 P expression in liver cirrhosis


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Abstract

Background: Hepatic impairment, portal hypertension, and multi-systemic damage could occur during liver cirrhosis’s late stage. Bleeding is a complication of hepatic cirrhosis along with several changes including blood platelet count (BPC), mean platelet volume (MPV), platelet crit (PCT) and expression of platelet CD62P. Blood platelet count (BPC), mean platelet volume (MPV), platelet distribution width, and other indices are indirect reflections of CD62P parameters.

Objective: To investigate the changes in platelet functional parameters and CD62 P expression in liver cirrhosis as a possible guide in clinical treatments and prognoses of liver cirrhosis.

Methods: CD62P was tested by flow cytometry in liver cirrhosis. BPC, MPV, and PCT in peripheral blood were tested using an auto blood cell analyzer. Data were analyzed using SPSS11.0.

Results: The values of CD62P and MPV in patients was significantly higher than those of healthy donors (P<0.01), while the values of BPC and PCT were significantly lower than those of the control group (P<0.01)

Conclusions: CD62P, BPC, MPV, and platelet crit (PCT) show several changes in liver cirrhosis. It is useful to understand the relationship between hepatic cirrhosis severity and CD62P, BPC, MPV, PCT, timely monitoring of CD62P for treatment of hepatic cirrhosis in clinical treatment and prognosis.

Keywords: Liver cirrhosis, granula protein, platelet parameters, expression

Introduction

Cirrhosis is the final stage attained by various chronic hepatic diseases after years or decades of slow progression. It can thus be prevented by appropriate screening for chronic liver diseases so that they can be treated in time. Chronic liver diseases do not usually cause any symptoms. However, hepatic impairment, portal hypertension, and multi-systemic damage could occur during the late stage of liver cirrhosis. Moreover, hypersplenism, thrombocytopenia, and consumption of blood coagulation factors and antibodies directed against platelets could lead to bleeding tendency. Granula protein on the platelet surface (CD62P) has very high sensitivity and specificity to platelet activation; thus, it is regarded as a special index of platelet activation. Platelets are formed by fragments of macrophage cytoplasm, and interact with mono-macrophages in the spleen. The mean platelet volume (MPV) reflects the active compounds and granula in the platelet.

The larger the size of the platelet, the more glycogen, adenine, nucleotide, and normal phosphates it contains and the higher its active functions become. Smaller-sized platelets have lower functions. Blood platelet count (BPC), MPV, and other indices are indirect reflections of its parameters. Reports show that BPC, MPV, and platelet crit (PCT) exhibit some changes in liver cirrhosis. Decreased MPV may be related to the presence of endotoxins, as well as to increased immunoglobulin, both having the ability to induce sustained platelet activation, thereby leading to the release of active compounds that cause granula exhaustion, platelet shrinkage, and decreased reservation.

In this study, CD62P was tested by flow cytometry in liver cirrhosis. BPC, MPV, and PCT in peripheral blood were tested using an auto blood cell analyzer. Understanding the relationship between hepatic cirrhosis severity and CD62P, BPC, MPV, PCT, timely monitoring of CD62P, and other platelet parameters is highly essential in preventing hemorrhage and coagulation disorders during the early stage of hepatic cirrhosis and may serve as a guide in clinical treatments and prognoses.

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Methods

Patients
Up to 32 cases of hepatic cirrhosis were studied, among whom were 25 male and 7 female. Their age range was 35-68 years, and the average age was 51±15. The diagnostic criteria were in accordance with the 5th National Infectious Diseases and Parasite Conference, and created from combined clinical manifestations, liver function, ultrasound, and CT scan findings. The patients were classified according to Child–Pugh (REF) as follows: Grade A, 12 cases; Grade B, 10 cases; and Grade C, 10 cases, which was used for assessing the prognosis of cirrhosis. The higher score, the worse of liver reserve function (Grade A, 5-6, the risk of operation is low; Grade B, 7-9, the risk of operation is higher than Grade A; Grade C, e”10, the risk of operation is the highest and the prognosis is the worst). All the patients had no heart, brain, kidney, or diabetic problems; they were also not suffering from acute infections. Six cases were confirmed to have upper gastrointestinal tract bleeding through gastroscope examination and a positive occult blood (OB) test.

The control group comprising 20 cases of normal patients underwent a health checkup. Among the 20 cases, 10 were male and 10 were female, aged 21 to 50 years, with an average age of 38±12. All cases in the patient group and in the control group did not take any anticoagulant one week before the test. This study was conducted in accordance with the declaration of Helsinki. This study was conducted with approval from the Ethics Committee of Linyi People’s Hospital. Written informed consent was obtained from all participants.

Detection of CD62P
Approximately 2 ml of fasting blood samples from both patient and control groups were collected with 3.84% citric acid sodium. Then the samples were placed in 1% paraformaldehyde at 4 °C for 15 min. The samples underwent centrifugation at 800r/min for 5 min. The platelet-rich supernatant was then separated. Up to 50 µl supernatant was mixed with CD62P monoclonal antibody (Becton-Dickinson) labeled with phycoerythrin, 20 ul aliquot was mixed and used for testing, and another 50 µl platelet-rich plasma was placed in a tube and mixed with phosphate buffer solution (PBS) for use as control. Both tubes were reacted for 20 min away from light, and then mixed with 1 ml of 1% paraformaldehyde at 4 °C. The tubes were poached with PBS and underwent centrifugation at 2000 r/min for 5 min. The supernatant was withdrawn and the procedure was repeated twice, adjusting the platelet concentration to 10×10⁹/L, preserving at 4°C, and testing with flow cytometry assay (Becton-Dickinson) within 24 hours. All data were analyzed using Cell Quest Plot software. Platelet CD62P positive rate was obtained as a result.

Detection of BPC, MPV, and PCT
Up to 2 ml samples with ethylenediaminetetraacetic acid (EDTA) anticoagulant and special anticoagulant (1.5 mg/ml to 2.2 mg/ml EDTA-K₂) were taken from both patient and control groups. Subsequently, they were tested using an M-533 auto blood cell analyzer. The process lasted for approximately 2 hours. Three kinds of whole blood were used as control. When all the results were within the normal range, samples were taken.

Statistical analysis
Data were expressed using mean ± standard deviation (x ± s). T-test was applied to obtain the mean of subgroup comparison in both groups. All data gathered were analyzed with SPSS11.0. P < 0.05 was considered statistically significant.

Results

Comparison of plasma CD62P, BPC, MPV, and PCT
The testing results of CD62P, BPC, MPV, and PCT in the 32 cases of hepatic cirrhosis and in the 20 cases from the control group are shown in Table 1. CD62P and MPV in the cirrhosis group were higher than those in the control group (P < 0.05), whereas BPC and PCT were lower in the control group (P < 0.05). The differences are presented in table 1.

Table 1: Result of CD62P, BPC, MPV, and PCT in cirrhosis group and control group (X±s)

<table>
<thead>
<tr>
<th>Groups</th>
<th>Case</th>
<th>CD62P (%)</th>
<th>BPC (10⁹/L)</th>
<th>MPV (fl)</th>
<th>PCT (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cirrhosis</td>
<td>32</td>
<td>11.84±7.82*</td>
<td>43.96±21.69*</td>
<td>13.26±0.41*</td>
<td>1.06±0.72*</td>
</tr>
<tr>
<td>Control</td>
<td>20</td>
<td>2.74±1.05</td>
<td>95.14±21.75</td>
<td>9.73±0.80</td>
<td>1.98±0.35</td>
</tr>
</tbody>
</table>

Data were expressed using average ± standard deviation (x ± s). T-test was applied to obtain the average of subgroup comparison in both groups. P < 0.05 was considered statistically significant.

*Compared with control group, P < 0.05.
Analysis of CD62P, BPC, MPV, and PCT in liver cirrhosis
The testing value of platelet surface CD62P in Grade A cirrhosis patients was lower than that in Grade B (P < 0.05). BPC, MPV, and PCT showed no difference in both grades. The testing value of platelet surface CD62P in Grade C liver cirrhosis patients was higher than that in Grades A and B (P < 0.05). No marked difference was observed in various grades (P > 0.05). The results of CD62P, BPC, MPV, and PCT in different grades are shown in table 2.

Analysis of CD62P, BPC, MPV, and PCT in bleeding or non-bleeding group
The testing value of platelet surface CD62P (24.79±6.55) was markedly higher than that in the no-bleeding group (8.85±4.21) (P < 0.01), whereas BPC, MPV, and PCT exhibited no significant difference. The testing values of CD62P, BPC, MPV, and PCT in the upper digestive bleeding with hepatic cirrhosis group and in the non-bleeding group are shown in table 3.

Table 2: Testing result CD62P, BPC, MPV, PCT of platelet surface (X±s)

<table>
<thead>
<tr>
<th>Group</th>
<th>Case</th>
<th>CD62P (%)</th>
<th>BPC (10^9/L)</th>
<th>MPV (fl)</th>
<th>PCT (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Grade A</td>
<td>12</td>
<td>6.68±2.31</td>
<td>52.00±21.67</td>
<td>10.51±0.39</td>
<td>1.32±0.51</td>
</tr>
<tr>
<td>Grade B</td>
<td>10</td>
<td>9.20±4.06*</td>
<td>40.50±22.14</td>
<td>11.76±0.52</td>
<td>1.24±0.76</td>
</tr>
<tr>
<td>Grade C</td>
<td>10</td>
<td>20.67±7.61**</td>
<td>37.50±17.54</td>
<td>13.26±0.41</td>
<td>1.06±0.72</td>
</tr>
</tbody>
</table>

Data were expressed using average ± standard deviation (X±s). T-test was applied to obtain the average of subgroup comparison in both groups. P < 0.05 was considered statistically significant. Grade B compared with Grade A, P < 0.05; Grade C compared with Grade A and B, P < 0.05

Table 3: Result of CD62P, BPC, MPV, and PCT on surface of platelet in upper digestive bleeding and non-bleeding group with hepatic cirrhosis (±s)

<table>
<thead>
<tr>
<th>Group</th>
<th>Case</th>
<th>CD62P (%)</th>
<th>BPC (10^9/L)</th>
<th>MPV (fl)</th>
<th>PCT (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bleeding</td>
<td>6</td>
<td>24.79±6.55*</td>
<td>35.67±27.80</td>
<td>11.20±1.20</td>
<td>1.05±0.71</td>
</tr>
<tr>
<td>Non-bleeding</td>
<td>26</td>
<td>8.85±4.21</td>
<td>46.58±21.90</td>
<td>10.60±0.80</td>
<td>1.31±0.50</td>
</tr>
</tbody>
</table>

Data were expressed using average ± standard deviation (X±s). T-test was applied to obtain the average of subgroup comparison in both groups. P < 0.05 was considered statistically significant.

*Comparison between bleeding group and non-bleeding group, P < 0.05.

Discussion
CD62P is a surface marker for platelet activation, which is present in the granules of platelets. CD62P is a member of selectin family. It is expressed by activated platelets, megakaryocytes, and vascular endothelial cells. It is one of the important membrane proteins, which serve for coagulation. CD62P mediates the interaction between platelets and other cells, platelets aggregation and adherence by improving the interaction of cells. Platelets originate from bone marrow megakaryocytes and the number of megakaryocytes indicates the function of hematogenesis in the bone marrow. The testing value of BPC reflects the generation and decay of platelets. MPV reflects the metabolization of bone marrow megakaryocytes. PCT is a parameter reflecting the condition of granula and active substances in platelets. Thus the detection of BPC, MPV and PCT in peripheral blood is used for treatment of liver cirrhosis clinically since bleeding is a complication of liver cirrhosis.

This study showed that CD62P in liver cirrhosis patients was higher than that in the control group, whereas the number of platelets were lower in the control group. This finding indicates that the increase in CD62P was related to the overdestruction and activation of platelets in vivo. Therefore, CD62P in hepatic cirrhosis patients was higher than that in the control group. CD62P was also related to the Child-Pugh classification, indicating that the platelets were highly activated. To some extent, the destruction of platelets along with the severity of liver cirrhosis increased the value of CD62P and destroyed more
platelets. Meanwhile, CD62P was higher in the group with liver cirrhosis combined with upper GI bleeding than in the non-bleeding group, indicating the relation of platelet activation and liver cirrhosis severity. Therefore, testing the value of CD62P revealed the severity of cirrhosis. The number of platelets in various groups did not exhibit marked difference, indicating that this factor was not related to upper digestive bleeding. However, the number of platelets was related to platelet function. Thus, CD62P is a good index that can be used to determine the activation and function of platelets.

This study found that the number of platelets and PCT was lower in liver cirrhosis patients, but their MPV was increased. Thompson et al. postulated that large-volume platelets contain more body density, higher activity, more rapid metabolism, more powerful adhesive capacity, and more bleeding. They found that MPV and CD62P were higher in liver cirrhosis patients than in the control group, whereas BPC and PCT were lower in hepatic cirrhosis patients than in the control group, indicating the decrease in the number of platelets and the change in quantity. The increase in CD62P and MPV could cause hypersplenism and destroy platelets. The mononuclear macrophage system compensated for the excretion of large-volume platelets, whereas the platelets themselves exhibited morphology changes, such as giant cells and abnormalities, possibly meeting the body's coagulate function requirement.

Conclusion
Dynamic monitoring of the expression of CD62P and the parameters of platelets is helpful in ascertaining the activities and function status of platelets. It is also useful in discovering early bleeding and coagulation disorders caused by hepatic cirrhosis, thus leading to early and accurate treatment and prognosis. Furthermore, results obtained from the dynamic monitoring of the expression of CD62P and the parameters of platelets can serve as guidelines for clinical treatments and prognosis.

References


