

# Potential anti-proliferative effects of chemical constituents and hemisynthetic derivatives from *Scadoxus pseudocaulus* (Amaryllidaceae)

Annie Laure Ngankeu Pagning<sup>1,3</sup>, Jean-de-Dieu Tamokou<sup>2</sup>, Bushra Taj Muhammad<sup>3</sup>, David Ngnokam<sup>1</sup>, Leon Azefack Tapondjou<sup>1</sup>, Mohammad Shaiq Ali<sup>3</sup>, Muhammad Waqar Hameed<sup>3</sup>

1. Research Unit of Environmental and Applied Chemistry, Department of Chemistry, Faculty of Science, University of Dschang, P O Box 183, Dschang, Cameroon.
2. Research Unit of Microbiology and antimicrobial Substances, Department of Biochemistry, Faculty of Science, University of Dschang, PO Box 067 Dschang, Republic of Cameroon.
3. International Center for Chemical and Biological Sciences, University of Karachi, Karachi-75270, Pakistan.

## Author e-mails:

Annie Laure Ngankeu Pagning: apagning2000@yahoo.fr, Jean-de-Dieu Tamokou: jtamokou@yahoo.fr, Bushra Taj Muhammad: bushraistaj@gmail.com, David Ngnokam: dngnokam@yahoo.fr, Leon Azefack Tapondjou: tapondjou2001@yahoo.fr, Mohammad Shaiq Ali: shaiq303@hotmail.com, Muhammad Waqar Hameed: mwaqar13@gmail.com

## Abstract

**Background:** Biological significance of *Amaryllidaceae* is well advocated from the literature. In Cameroon, plants from this family are routinely used for the cure of liver, cancer and cardiovascular diseases. To date, no scientific investigation corresponding to the anti-cancer activity of extracts and isolated compounds of *Scadoxus pseudocaulus* is available.

**Objective:** Current study is focused to elaborate the anti-proliferative effects of natural isolates (compounds 1-6, 9) and hemi-synthetic analogs (compounds 7-8) extracted from *S. pseudocaulus*.

**Methods:** Column chromatography of the ethyl acetate extract followed by purification of different fractions led to the isolation of seven compounds (1 – 6, 9). Esterification reaction of compound 6 was carried out using butyryl chlorides and triethylamin to produce two derivatives (7 – 8). The cytotoxic activity was performed after staining of treated cells with fluorescent dye propidium iodide. Dead cells were detected using cytometer FL2 or FL3 channels/filters.

**Results:** Trans-derivative of narciclasine (a natural isolate from *S. pseudocaulus*), was found to be most potent among all tested compounds. Its effects were more significant on low malignant follicular lymphoma (DoHH2 cells) as compared to highly malignant (EBV infected) Burkitts lymphoma (Raji cells).

**Conclusion:** From our results, narciclasine appears to hold the potential of a lead molecule that can be used to bridge the therapeutic gaps in cancer research.

**Keywords:** *Scadoxus pseudocaulus*, *Amaryllidaceae*, 7-deoxy-trans-dihydronarciclasin, farrerol, derivatization, cytotoxic activity.

**DOI:** <https://dx.doi.org/10.4314/ahs.v20i1.53>

**Cite as:** Pagning ALN, Tamokou J-D, Muhammad BT, Ngnokam D, Tapondjou LA, Ali MS, et al. Potential anti-proliferative effects of chemical constituents and hemisynthetic derivatives from *Scadoxus pseudocaulus* (Amaryllidaceae). *Afri Health Sci.* 2020;20(1):469-75. <https://dx.doi.org/10.4314/ahs.v20i1.53>

## Corresponding author:

Jean-de-Dieu Tamokou;  
Research Unit of Microbiology and Antimicrobial Substances, Department of Biochemistry, Faculty of Science, University of Dschang, P. O. Box 067 Dschang, Republic of Cameroon;  
Email: jtamokou@yahoo.fr / jean.tamokou@univ-dschang.org

## Introduction

*Scadoxus pseudocaulus* (I. Bjornstad and Friis), a member of the *Amaryllidaceae*, is used in the West Region of Cameroon, for the treatment of liver, cancer and cardiovascular diseases. Previous study demonstrated the antimicrobial, antioxidant and anti-butyrylcholinesterase activities of compounds and extracts of *S. pseudocaulus*.<sup>1</sup> Apart from these biological activities, no conclusive scientific study is reported pertaining to its anti-cancer activity. It is known

that alkaloids of *Amaryllidaceae* family (like the non-basic hydroxylated phenanthridones) possess high cytostatic activity.<sup>2-6</sup> Some other alkaloids like galanthamine, lycorine and narciclasine can only be synthesized in plants of *Amaryllidaceae* family.<sup>7</sup> Alkaloids from this family bear a number of biological activities including anti-microbial and anti-cancer.<sup>6</sup> 7-deoxy-trans-dihydronarciclasin or trans-dihydrolycoricidin (5) is isocarbostryl alkaloid reported from *S. pseudocaulus* like narciclasine (also known as lycoricidinol) who is known for its effects on protein biosynthesis.<sup>8</sup> Narciclasine oil is effective in the treatment for uterine tumors. It also acts as a plant growth modulator.<sup>9</sup> Narciclasine's first bioactivity was observed in 1967, where it was shown to have strong mitosis blocking activity.<sup>10</sup> The reported biological effects of narciclasine includes: anti-proliferative, antitumor/cytotoxic, acetylcholinesterase inhibitory, analgesic, hypotensive, antibacterial and antifungal.<sup>11</sup> Data from studies on HeLa cell line showed that narciclasine and other alkaloids like lycorine, dihydrolycorine, haemanthamine, pretazettine and pseudolycorine can inhibit growth and protein synthesis.<sup>12</sup> Not only narciclasine but also other alkaloid derivatives of *Amaryllidaceae* were reported to have anti-cancer effects.<sup>13,14</sup>

However, narciclasine's group is found to be most potent and effective against cell growth and protein synthesis.<sup>6,15,16</sup> Narciclasine is also found active on murine (p388 lymphoma) and human cancer cell lines (e.g. A549, NS-CLC, PC3 and prostate). Narciclasine was also proposed as potential tool to cure apoptosis resistant metastasizing cancer cells.<sup>17</sup> A study conducted on a panel of 60 human cancer cell lines showed narciclasine's potential cytotoxic effects.<sup>15</sup> Another study reported anti-cancer effects of narciclasine on a variety of cancer cell lines, where fibroblasts were reported to be comparably resistant.<sup>18</sup> Sensitivity of cancer cells to narciclasine was also reported using HUVECs (endothelial) cells.<sup>19</sup> In a series of mechanistic studies, narciclasine was found to induce apoptosis driven cell death in cancer cells either mediated by the death receptors or mitochondria.<sup>18</sup> It was further confirmed using human promyelocytic (HL-60) cells and human oral cavity squamous carcinoma (HSC-2 cells) that it can induce apoptosis even at nano-molar concentrations.<sup>20</sup> The above literature highlighted the significant biological effects of *Amaryllidaceae* family's natural isolates and hemi-synthetic derivatives and advocates further investigations to dis-

cover their anti-cancer tendencies. Therefore, the current study aimed at evaluating the antiproliferative activities of isolated compounds and hemisynthetic derivatives from the whole plant of *S. pseudocaulus*.

## Materials and methods

### Plant material

Whole plant of *Scadoxus pseudocaulus* was collected at Dschang, Menoua Division, West Region of Cameroon, in May 2013. The plant material was identified by Mr. Victor Nana, a botanist at the National Herbarium, Yaoundé, where a voucher specimen (N° 34986/SRF/CAM) was deposited.

### Extraction, isolation and hemi-synthesis of compounds

The extraction and isolation of compounds were done as previously described.<sup>1</sup> Briefly, *S. pseudocaulus* was air-dried and powdered. The powder was macerated at room temperature with MeOH to afford the MeOH extract. The CHCl<sub>3</sub> and EtOAc fractions from the MeOH extract were collected by column chromatography. Purification of the EtOAc fraction yielded seven known compounds (Figure 1). Esterification reaction of compound 6 was carried out using butyryl chlorides and triethylamine to produce two derivatives. The structures of isolated compounds and derivatives were determined through NMR and MS. The data thus obtained was compared with those from the literature.

### Cytotoxicity assay

To assess the anti-cancer potential of natural isolates and hemi-synthetic analogs, Burkitt's and Follicular lymphoma (B lymphoma) cell lines were selected. These cell lines were a gift from Prof. Dr. Daniel Hoessli, Switzerland and from Dr A. Kluin-Nelemans, Groningen, Netherlands, respectively. Monkey Vero cells (African green monkey kidney cells, normal non-cancer cells, ATCC No. CCL-81), obtained from the American Type Culture Collection (ATCC), was also used in this study. The cell lines were maintained at 37 °C in a humidified 5% CO<sub>2</sub> environment in Roswell Park Memorial Institute 1640 medium (RPMI; Caisson) with 1% L-glutamine, 1% penicillin/streptomycin (Gibco, Invitrogen), supplemented with 10% foetal bovine serum (FBS, PAA laboratories). The cytotoxicity analysis was performed after staining of treated cells with fluorescent dye propidium iodide (PI)

(excitation wavelength = 536 nm; emission wavelength = 617 nm).<sup>21,22</sup> Principally, PI can't cross the intact plasma membrane and therefore, cannot stain live cells. However, after the cells have lost membrane integrity (i.e. dead cells), this dye enters into the cells and intercalate with the cellular DNA. Dead cells thus fluoresce and can be detected using cytometer FL2 or FL3 channels/filters. To generate dose response curve of the set of standard and test compounds, cells were seeded in a 96 well plate. Each well contains 0.13x10<sup>6</sup> cells in final reaction volume of 200  $\mu$ L. Cells treated with bosutinib + RPMI 1640 served as positive control whereas cells left untreated + 0.5% (v/v) DMSO + RPMI 1640 were used as negative control.

### Statistical analysis

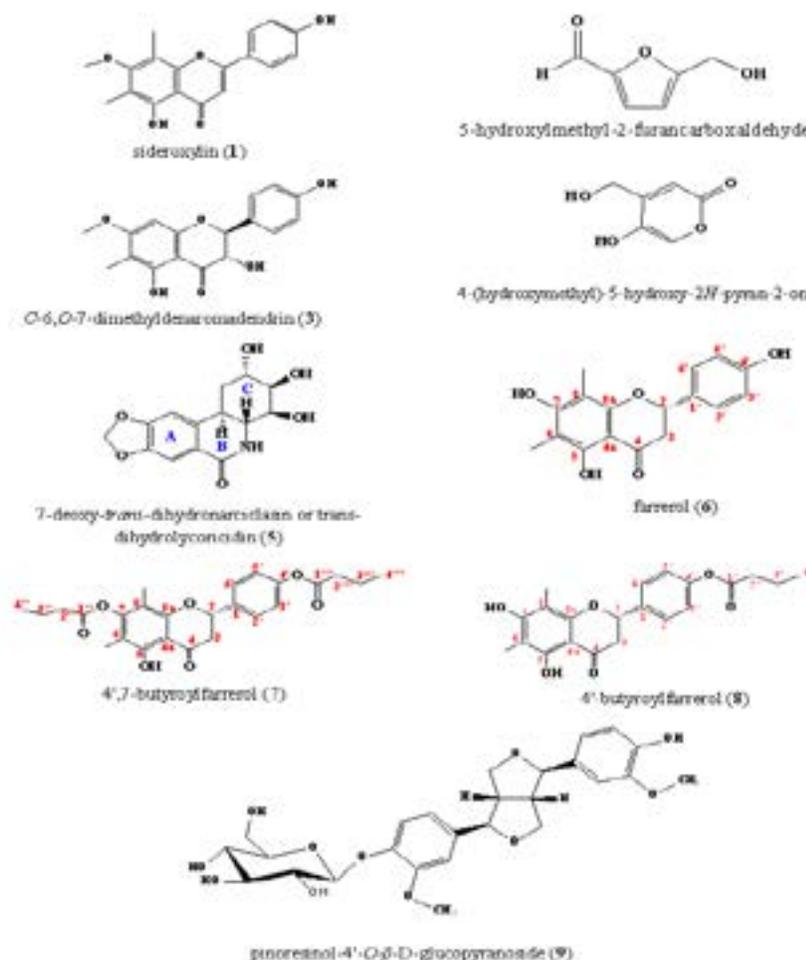
Data was analyzed by one-way analysis of variance followed by Waller-Duncan Post Hoc test and Statistical

Package for Social Sciences software (SPSS, version 12.0). The results were expressed as mean  $\pm$  standard deviation (SD). Differences between groups were considered significant when  $p < 0.05$ .

## Results

### Chemical analysis

The phytochemical investigation of the CHCl<sub>3</sub> and EtOAc fractions from the MeOH extract of *S. pseudo-caulus*, afforded six known compounds namely sideroxylin (1),<sup>23</sup> 5-hydroxymethyl-2-furancarboxaldehyde (2),<sup>24</sup> C-6,O-7-dimethyldemarcandendrin (3),<sup>25</sup> 4-(hydroxymethyl)-5-hydroxy-2H-pyran-2-one (4),<sup>26</sup> 7-deoxy-trans-dihydronarciclasin or trans-dihydrolycoricidin (5),<sup>15</sup> farrerol (6)<sup>25,27</sup> and pinoresinol-4'-O- $\beta$ -D-glucopyranoside (9).<sup>28</sup> Esterification reaction of farrerol (6) was carried out using butyryl chlorides and triethylamin to produce two derivatives namely 4',7-butyrylfarrerol (7) and 4'-butyrylfarrerol (8)<sup>1</sup> (Figure 1).



**Figure 1:** Structures of isolated compounds (1-6, 9) and hemisynthetic derivatives (7-8)

## Cytotoxic activities

The cytotoxic activities of compounds<sup>1-9</sup> were evaluated against two cancer cell lines and normal non-cancer cells (Vero cells) (Table 1, Figures 2 – 3). Both cancer cell lines were found sensitive to all tested compounds ( $0.46 \pm 0.18$  to  $46.15 \pm 3.44$ ), however, the highest activity was observed for compound 5 (-deoxy-trans-dihydronarciclasin or trans-dihydrolycoricidin) ( $46.15 \pm 3.44$  for Raji cells and  $39.62 \pm 1.67$  for DOHH2 cells) (Figures 2 - 3). For Raji cells,  $114.47 \mu\text{M}$  concentration of compound

5 was required for the induction of death in 50% cell population, however,  $134.28 \mu\text{M}$  were found sufficient for DoHH2 cells (Table 1). The test compounds were non-toxic to normal cells (results not shown) whereas the Selectivity Index (SI) values of the compound 5 against the Raji and DOHH2 cells are 13.07 and 11.14 indicating its good selectivity on the test cancer cell lines (Table 1). Bosutinib was used as a standard drug in the study. Its IC<sub>50</sub> dose for Raji cells observed to be  $63.17 \mu\text{M} \pm 3.65$  and for DoHH2 cells  $68.30 \mu\text{M} \pm 1.06$  after 48 hr incubation time (Table 1).

**Table 1:** Cytotoxicity (IC<sub>50</sub>) and Selectivity index (SI) of compound 5 against Raji and DOHH2 cells.

| Compounds        | Cytotoxicity (IC <sub>50</sub> in $\mu\text{M}$ ) |                     |                      | Selectivity index (SI) |                    |
|------------------|---|---------------------|----------------------|------------------------|--------------------|
|                  | Raji cells  | DOHH2 cells         | Vero cells           | Raji cells             | DOHH2 cells        |
| <b>5</b>         | $114.47 \pm 0.53^a$                               | $134.28 \pm 1.34^b$ | $1496.46 \pm 1.84^c$ | $13.07 \pm 1.71^a$     | $11.14 \pm 0.98^a$ |
| <b>Bosutinib</b> | $63.17 \pm 3.65^a$                                | $68.30 \pm 1.06^b$  | $1298.83 \pm 1.09^c$ | $20.56 \pm 1.54^a$     | $19.01 \pm 1.23^a$ |

SI: IC<sub>50</sub> on Vero cells/IC<sub>50</sub> on cancer cells; \*SI obtained from average IC<sub>50</sub>; each IC<sub>50</sub> value represents the Mean  $\pm$  SD (n = 3); on the same line, IC<sub>50</sub>/SI values marked with different superscript letters are significantly different (p < 0.05).

## Discussion

The current investigation was carried out to evaluate the antiproliferative activities of isolated compounds and hemisynthetic derivatives from the whole plant of *S. pseudocaulis*. The findings of the present study revealed lesser sensitivity of Raji cells to compound 5 that can be due to EBV (Epstein Barr Virus) infection. This virus encodes a number of viral proteins that makes these cells resistant to apoptosis and promote growth.<sup>29-31</sup> One of these proteins is latent membrane protein (LMP2A) that is membrane bound and shares structural resemblance to B cell surface receptor protein (BCR). LMP2A is known to play role in relaying survival signal thus promoting survival of B lymphoma cells.<sup>32-34</sup>

7-deoxy-trans-dihydronarciclasin, (an enantiomer) is a well reported potent anti-neoplastic agent.<sup>35</sup> Trans derivative of narciclassine is found to be more active as compared to cis form.<sup>13</sup> 7-deoxy-trans-dihydronarciclasin has significant anti-cancer effects and the detailed SAR studies elucidated that this molecule has pharmacophore moiety which induces apoptosis.<sup>15,19,36-40</sup> The tri-hydroxyl-

ated ring C of this compound is also considered to be a critical part. In fact, the substitution of tri-hydroxylated ring C with hydrophobic moieties led to decrease in anti-cancer capacity and the same was observed upon loss of three hydroxyl groups.<sup>37,41-44</sup> Thus, it could be seen that 7-deoxy-trans-dihydronarciclasin has significant cytotoxic effects and can induce apoptosis in cancer cell lines making it a lead molecule in cancer research. The antiproliferative effects of sideroxylin against ovarian cancer cells are through the induction of mitochondrial dysfunction and the activation of PI3K and MAPK signal transduction.<sup>45</sup> Selectivity Index (SI) of active compounds was determined in order to investigate, whether the cytotoxic activity was specific to cancer cells. The SI of the samples is defined as the ratio of cytotoxicity (IC<sub>50</sub> values) on normal cells (Vero cells) to cancer cells:  $\text{SI} = \text{IC}_{50} \text{ on Vero cells} / \text{IC}_{50} \text{ on cancer cells}$ . The Selectivity Index (SI) values of the compound 5 against the Raji and DOHH2 cells are 13.07 and 11.14 and could be considered as good when taking in consideration that the ratio for a good therapeutic index for a remedy or drug should be  $\geq 10$ .<sup>46</sup> These results are consistent with the use of compound<sup>5</sup> for treating B lymphoma.

## Conclusion

Present investigation has revealed that isolated compounds and hemisynthetic derivatives from *S. pseudocaulus* are active against the tested cancer cell lines and non-toxic against Vero cells (non-cancer cells). The pattern of response revealed that EBV infected Burkitt lymphoma is less sensitive to 7-deoxy-trans-dihydronarciclasine as compared to Follicular lymphoma. Greater cytotoxic effect on slow growing follicular lymphoma signifies its metabolic stability which can be exploited for slow progressing malignancies. Further investigations are needed to screen them against other cancer types and human cell line of normal tissues, including bone marrow cells to justify the traditional use of *S. pseudocaulus* as an anticancer substance. Detailed mechanistic studies are also needed to find the mode of action of this compound on the molecular pathway(s) potentially leading to cell death.

## Acknowledgments

This research work was supported under the umbrella of TWAS-ICCBS Postgraduate Fellowship Program. We are grateful to the University of Dschang for financing some consumable items used in this work.

## Conflict of interest

None declared.

## References

1. Ngankeu PAL, Tamokou JD, Khan ML, Ali MI, Hameed A, Ngnokam D, Tapondjou LA, Kuate JR, Ali MS. Antimicrobial, antioxidant and butyrylcholinesterase inhibition activities of extracts and isolated compounds from *Scadoxus pseudocaulus* and semi-synthetic farrerol derivatives. *South African Journal of Botany* 2016; 102: 166-174. doi.org/10.1016/j.sajb.2015.06.009
2. Hoshino O. In *The Alkaloids*; Cordell GA, Ed.; Academic: San Diego, CA, 1998; Vol. 51, pp 181–236.
3. Rinner U, Hudlicky T. Synthesis of Amaryllidaceae constituents-an update. *Synlett* 2005; 2005(03): 365-387. doi: 10.1055/s-2005-862382
4. Chapleur Y, Chretien F, Ibn Ahmed S, Khaldi M. Chemistry and synthesis of highly oxygenated alkaloids from Amaryllidaceae: lycoricidine, narciclasine, pancratistatin and analogs. *Current Organic Synthesis* 2006; 3(3): 341-378. doi : 10.2174/157017906777934872
5. Manpadi M, Kornienko A. Total syntheses of pancratistatin. A review. *Organic Preparations and Procedures International* 2008; 40(2): 107-161. doi: 10.1080/00304940809458083
6. Kornienko A, Evidente A. Chemistry, biology, and medicinal potential of narciclasine and its congeners. *Chemical Reviews* 2008; 108(6): 1982-2014. doi: 10.1021/cr078198u
7. Diamond A, Desgagné-Penix I. Metabolic engineering for the production of plant isoquinoline alkaloids. *Plant Biotechnology Journal* 2016; 14(6):1319-1328. doi: 10.1111/pbi.12494.
8. Garreau de Loubresse N, Prokhorova I, Holtkamp W, Rodnina MV, Yusupova G, Yusupov M. Structural basis for the inhibition of the eukaryotic ribosome. *Nature* 2014; 513: 517-522. doi: 10.1038/nature13737.
9. Riddle JM. Ancient and medieval chemotherapy for cancer. *Isis* 1985; 76(3): 319-330. doi.org/10.1086/353876.
10. Ceriotti G. Narciclasine: an antimetabolic substance from *Narcissus* bulbs. *Nature* 1967; 213: 595-596. doi: 10.1038/213595a0
11. He M, Qu C, Gao O, Hu X, Hong X. Biological and pharmacological activities of Amaryllidaceae alkaloids. *Royal Society Chemistry Advances* 2015; 5(21): 16562-16574. doi: 10.1039/c4ra14666B.
12. Jimenez A, Santos A, Alonso G, Vazquez D. Inhibitors of protein synthesis in eukaryotic cells: comparative effects of some Amaryllidaceae alkaloids. *Biochimica and Biophysica Acta (BBA)-Nucleic Acids and Protein Synthesis* 1976; 425(3): 342-348.
13. Evidente A, Kireev AS, Jenkins AR, Romero AE, Steelant WF. Biological evaluation of structurally diverse Amaryllidaceae alkaloids and their synthetic derivatives: discovery of novel leads for anticancer drug design. *Planta Medica* 2009; 75(5): 501-507. doi: 10.1055/s-0029-1185340
14. Evidente A, Kornienko A. Anticancer evaluation of structurally diverse Amaryllidaceae alkaloids and their synthetic derivatives. *Phytochemistry Reviews* 2009; 8(2): 449-459. doi: 10.1055/s-0029-1185340
15. Pettit GR, Pettit III GR, Backhaus RA, Boyd MR, Meerow AW. Antineoplastic agents, 256. Cell growth inhibitory isocarbostryls from *Hymenocallis*. *Journal of Natural Products* 1993; 56(10): 1682-1687. doi: 10.1021/np50100a004.
16. Ingrassia L, Lefranc F, Mathieu V, Darro F, Kiss R. Amaryllidaceae isocarbostryl alkaloids and their derivatives as promising antitumor agents. *Translational Oncology* 2008;1(1): 1-13.
17. Lefranc F, Dubois S, Ingrassia L, Van Quaquebeke E, Darro F, Kiss R. Narciclasine displays potent and selective anti-tumor effects by impairing cancer cell migra-

- tion through a phosphocofilin-mediated increase of actin stress fibers. *Neuro-oncology* 2008;10(6):1142.
18. Dumont P, Ingrassia L, Rouzeau S, Ribaucour F, Thomas S, Roland I, Darro F, Lefranc F, Kiss R. The Amaryllidaceae isocarbostryril narciclasine induces apoptosis by activation of the death receptor and/or mitochondrial pathways in cancer cells but not in normal fibroblasts. *Neoplasia*, 2007; 9(9): 766-776.
  19. Ingrassia L, Lefranc F, Dewelle J, Pottier L, Mathieu V, Spiegl-Kreinecker S, Sauvage S, El Yazidi M, Dehoux M, Berger W, Van Quaquebeke E. Structure-activity relationship analysis of novel derivatives of narciclasine (an Amaryllidaceae isocarbostryril derivative) as potential anticancer agents. *Journal of Medicinal Chemistry* 2009; 52(4): 1100-1114. doi: 10.1021/jm8013585.
  20. Jitsuno M, Yokosuka A, Hashimoto K, Amano O, Sakagami H, Mimaki Y. Chemical constituents of *Lycoris albiflora* and their cytotoxic activities. *Natural Product Communications* 2011; 6(2): 187-192.
  21. Boyd MR, Paull KD. 1995. Some practical considerations and applications of the National Cancer Institute in vitro anticancer drug discovery screen. *Drug Development Research* 1995; 34(2): 91-109. doi.org/10.1002/ddr.430340203
  22. Donnou S, Galand C, Touitou V, Sautès-Fridman C, Fabry Z, Fisson S. Murine models of B-cell lymphomas: promising tools for designing cancer therapies. *Advances in Hematology* 2012; Article ID 701704, 13 pages. doi:10.1155/2012/701704
  23. Hillis WE, Isoi K. Variation in the chemical composition of *Eucalyptus sideroxyylon*. *Phytochemistry* 1965; 4(4): 541-550. doi: 10.1016/s0031-9422(00)86214-7.
  24. Pyo MK, Jin JL, Koo YK, Yun-Choi HS. Phenolic and furan type compounds isolated from *Gastrodia elata* and their anti-platelet effects. *Archives of Pharmacal Research* 2004; 27(4): 381-385.
  25. Arthur HR. A new optically active flavanone from the leaves of *Rhododendron farrerae*, tate. *Journal of the Chemical Society* 1955; 0: 3740-3742. doi:10.1039/JR9550003740.
  26. Lin A, Lu X, Fang Y, Zhu T, Gu Q, Zhu W. Two new 5-hydroxy-2-pyrone derivatives isolated from a marine-derived fungus *Aspergillus flavus*. *The Journal of Antibiotics* 2008 ; 61(4): 245-249. doi: 10.1038/ja.2008.36.
  27. Youssef DT, Ramadan MA, Khalifa AA. Acetophenones, a chalcone, a chromone and flavonoids from *Pancreatium maritimum*. *Phytochemistry* 1998; 49(8): 2579-2583. doi: 10.1016/S0031-9422(98)00429-4
  28. Ouyang M, Wein YS, Zhang ZK, Kuo YH. Inhibitory activity against Tobacco Mosaic Virus (TMV). Replication of pinoresinol and syringaresinol lignans and their glycosides from the root of *Rhus javanica* var. *roxburghiana*. *Journal of Agricultural and Food Chemistry* 2007; 55: 6460-6465. doi: 10.1021/jf0709808
  29. Milner AE, Johnson GD, Gregory CD. Prevention of programmed cell death in burkitt lymphoma cell lines by bcl-2-dependent and-independent mechanisms. *International Journal of Cancer* 1992; 52(4): 636-644. doi: 10.1002/ijc.2910520424.
  30. Imadome KI, Shirakata M, Shimizu N, Nonoyama S, Yamanashi Y. CD40 ligand is a critical effector of Epstein-Barr virus in host cell survival and transformation. *Proceedings of the National Academy of Sciences* 2003;100(13): 7836-7840. doi: 10.1073/pnas.1231363100.
  31. Matusali G, Arena G, De Leo A, Di Renzo L, Mattia E. Inhibition of p38 MAP kinase pathway induces apoptosis and prevents Epstein Barr virus reactivation in Raji cells exposed to lytic cycle inducing compounds. *Molecular Cancer* 2009; 8(1): 18. doi: 10.1186/1476-4598-8-18.
  32. Caldwell RG, Wilson JB, Anderson SJ, Longnecker R. Epstein-Barr virus LMP2A drives B cell development and survival in the absence of normal B cell receptor signals. *Immunity* 1998; 9(3): 405-411. doi.org/10.1016/S1074-7613(00)80623-8.
  33. Longnecker R. Epstein-Barr virus latency: LMP2, a regulator or means for Epstein-Barr virus persistence? *Advances in Cancer Research* 2000; 79: 175-200.
  34. Merchant M, Caldwell RG, Longnecker R. The LMP2A ITAM is essential for providing B cells with development and survival signals in vivo. *Journal of Virology* 2000; 74(19): 9115-9124. doi: 10.1128/JVI.74.19.9115-9124.2000
  35. Szántó G, Hegedűs L, Mattyasovszky L, Simon A, Simon Á, Bitter I, Tóth G, Tőke L, Kádas I. An expedient total synthesis of ent-(-)-7-deoxy-trans-dihydro-narciclasine. *Tetrahedron* 2009; 65(40): 8412-8417. doi: 10.1016/j.tet.2009.07.092
  36. McNulty J, Mo R. Diastereoselective intramolecular nitroaldol entry to lycoricidine alkaloids. *Chemical Communications* 1998; 0(8): 933-934. doi : 10.1039/A800097B.
  37. McNulty J, Mao J, Gibe R, Mo R, Wolf S, Pettit GR, Herald DL, Boyd MR. Studies directed towards the refinement of the pancratistatin cytotoxic pharmacophore. *Bioorganic & Medicinal Chemistry Letters* 2001; 11(2): 169-172. doi: 10.1016/S0960-894X(00)00614-4
  38. Pettit, GR, Eastham SA, Melody N, Orr B, Herald

- DL, McGregor J, Knight JC, Doubek D.L., Pettit GR, Garner LC, Bell JA. Isolation and structural modification of 7-Deoxynarciclasine and 7-Deoxy-trans-Dihydronarciclasine. *Journal of Natural Products* 2006; 69(1): 7-13. doi: 10.1021/np058068l
39. Pettit GR, Melody N, Herald DL, Knight JC, Chapuis JC. Antineoplastic Agents. 550. Synthesis of 10b (S)-Epi-pancratistatin from (+)-Narciclasine. *Journal of Natural Products* 2007; 70(3): 417-422. doi: 10.1021/np068046e.
40. McNulty J, Nair JJ, Griffin C, Pandey S. Synthesis and biological evaluation of fully functionalized seco-pancratistatin analogues. *Journal of Natural Products* 2008; 71(3): 357-363. doi: 10.1021/np0705460.
41. Mondon A, Krohn K. *Chemistry of narciclasine. Chemische Berichte* 1975; 108(2): 445-463.
42. Baez A, Vazquez D. Binding of [3H] narciclasine to eukaryotic ribosomes A study on a structure-activity relationship. *Biochimica et Biophysica Acta (BBA)-Nucleic Acids and Protein Synthesis* 1978; 518(1): 95-103. doi: 10.1016/0005-2787(78)90119-3.
43. Rinner U, Hillebrenner HL, Adams DR, Hudlicky T, Pettit GR. Synthesis and biological activity of some structural modifications of pancratistatin. *Bioorganic & Medicinal Chemistry Letters* 2004; 14(11): 2911-2915. doi: 10.1016/j.bmcl.2004.03.032.
44. McNulty J, Larichev V, Pandey S. A synthesis of 3-deoxydihydrolycoricidine: refinement of a structurally minimum pancratistatin pharmacophore. *Bioorganic & Medicinal Chemistry Letters* 2005; 15(23): 5315-5318. doi: 10.1016/j.bmcl.2005.08.024.
45. Park S, Lim W, Jeong W, Bazer FW, Lee D, Song G. Sideroxylin (*Callistemon lanceolatus*) suppressed cell proliferation and increased apoptosis in ovarian cancer cells accompanied by mitochondrial dysfunction, the generation of reactive oxygen species, and an increase of lipid peroxidation. *Journal of cellular Physiology* 2018; 233(11): 8597-8604. doi: 10.1002/jcp.26540.
46. Caamal-Fuentes E, Torres-Tapia LW, Simá-Polanco P, Peraza-Sánchez SR, Moo-Puc R. Screening of plants used in Mayan traditional medicine to treat cancer-like symptoms. *Journal of Ethnopharmacology* 2011; 135: 719-724. doi: 10.1016/j.jep.2011.04.004.