SRY and NR5A1 gene mutation in Algerian children and adolescents with DSD and testicular dysgenesis

Naouel Kherouatou-Chaoui¹, Djalila Chellat-Rezgoune¹, Mohamed Larbi Rezgoune¹, Ken Mc Elreavey², Laaldja Souhem Touabti³, Noreddine Abadi⁴, Dalila Satta¹

- 1. Laboratory of Cellular and Molecular Biology. Frères Mentouri University-Constantine 1, Constantine, Algeria.
- 2. Human Genetic Developmental Unit, Pasteur Institute, Paris, France.

3. Pediatric Urology Unit, C.H.U Sétif, Sétif, Algeria.

4. Laboratory of Biology and Molecular Genetic, University Constantine 3, Constantine, Algeria.

Co-authors Emails:

Djalila Chellat-Rezgoune: rchedjalila@gmail.com; Mohamed Larbi Rezgoune: rezgoune.genetique25@gmail. com; Ken Mc Elreavey: kenneth.mcelreavey@pasteur.fr; Laaldja Souhem Touabti: souhemtouabti@univ-setif.dz; Noreddine Abadi: nourabadi@yahoo.fr; Dalila Satta: dsatta741@gmail.com

Abstract

Background: In humans, sex determination and differentiation is genetically controlled. Disorders of sex development (DSD) result in anomalies of the development of the external and internal genitalia. Variants in transcription factors such as SRY, NR5A1 and SOX9, can cause changes in gonadal development often associated with ambiguity of the external genitalia. **Objectives:** This study has been conducted to determine the frequency, types and associated genetic alterations in patients with DSD in the Algerian population.

Methods: Thirty patients were included. Based on their clinical presentation, thirteen patients presented with ambiguous external genitalia, thirteen patients presented with hypospadias and four patients presented with bilateral undescended testes. Karyotype analysis was performed on peripheral blood lymphocytes using standard R-banding. DNA was isolated from blood leukocytes for PCR reaction and mutational analysis of SRY and NR5A1 was done by direct sequencing.

Results: Most patients with ambiguous genitalia had a 46,XY karyotype. One patient had a deletion of SRY, otherwise no point mutations in SRY or NR5A1 genes were identified. However, a single NR5A1 polymorphism (p.Gly146Ala) in patient with 46,XX DSD has been detected.

Conclusions: The absence of mutations in these genes suggests that there are others genes playing an important role in sex development and differentiation.

Keywords: DSD; consanguinity; karyotyping; SRY; NR5A1; sequencing.

DOI: https://dx.doi.org/10.4314/ahs.v21i3.61

Cite as: Kheronaton-Chaoni N, Chellat-Rezgoune D, Rezgoune ML, Mc Elreavey K, Tonabti LS, Abadi N, et al. SRY and NR5A1 gene mutation in Algerian children and adolescents with DSD and testicular dysgenesis. Afri Health Sci. 2021;21(3). 1491-1497. https://dx.doi. org/10.4314/ahs.v21i3.61

Introduction

Disorders of sex development DSD comprise a series of genetic diseases in which chromosomal, gonadal, or anatomical sex is atypical¹. Ambiguous genitalia are

Corresponding author:

Naouel Kherouatou-Chaoui, Laboratory of Cellular and Molecular Biology, Frères Mentouri University-Constantine 1, Route Ain el Bey, 25000 Constantine, Algeria. Tel/Fax: +213 31811184 Email: naouel.kerou@gmail.com present in about 1 in 2,000 live newborns and reflects a situation where the external genitalia cannot be established as being either male or female. Four steps must occur during sexual development and differentiation: establishment of chromosomal sex at fertilization, formation of undifferentiated gonads, gonadal differentiation into testes or ovaries, and development of the internal and external genitalia².

Error in any of these stages may lead to DSD, which can be classified into the following groups: 46,XX DSD (overvirilisation or masculinisation of a 46,XX female), 46,XY DSD (undervirilisation or undermasculinisation of an XY male), Ovotesticular DSD, 46,XX Testicular

African Health Sciences © 2021 Kherouatou-Chaoui N et al. Licensee African Health Sciences. This is an Open Access article distributed under the terms of the Creative commons Attribution License (https://creativecommons.org/licenses/BY/4.0), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

DSD (XX male), and 46,XY Complete Gonadal Dysgenesis (CGD)^{1,3}.

Mutations in several genes are known to cause DSD. Most often the underlying cause of DSD is a variant in a gene or genes regulating gonadal/genital or steroidogenic pathways. The commonest known genetic condition that leads to 46,XX DSD is congenital adrenal hyperplasia (CAH) due to 21α -hydroxylase deficiency. However Among the 46,XY DSD, the most common cause is androgen insensitivity syndrome (AIS) with mutations in the androgen receptor gene¹.

The Sox (SRY-type HMG box) genes encode a group of proteins characterized by the presence of an HMGbox, a 79 amino acid motif which can bind and bend DNA, which is the only part of the testis-determining gene SRY that is conserved between species⁴. Mutations in SRY are associated with 46,XY complete gonadal dysgenesis and are found in approximately 15% of these cases⁵. Most of the mutations detected are located in the HMG domain of the protein. The second most common monogenic cause of 46,XY DSD are mutations involving the NR5A1 gene. NR5A1 regulates multiple genes involved in adrenal and gonadal development steroidogenesis, reproduction and other metabolic functions^{6,7}. Mutations in the human NR5A1 gene has a phenotypic spectrum that ranges from complete testicular dysgenesis with Müllerian structures, through individuals with mild clitoromegaly or genital ambiguity, to severe penoscrotal hypospadias or even anorchia⁸⁻¹⁴. Although not essential for ovarian determination¹⁵, heterozygote mutations in NR5A1 have recently been linked to ovarian insufficiency, thus confirming the role of this factor in ovarian follicologenesis^{16,17}. A link between NR5A1 mutations and the occurrence of male infertility was also found¹⁸.

Recently, a recurrent NR5A1 p.Arg92Trp mutation was also identified in several patients with 46,XX testicular/ ovotesticular DSD, highlighting the role of NR5A1 in the development of both testes and ovaries¹⁹.

In view of the importance of the SRY and NR5A1 genes in normal gonadal development, this first study in Algeria was undertaken with an aim to screen for mutations in these genes in DSD Algerian cases and to describe their clinical and cytogenetics featuresing.

Materials and Methods Patients

Informed consent was obtained from all subjects participating in the study. Thirteen patients with ambiguous external genitalia, thirteen patients with hypospadias and four patients with bilateral testicular ectopy were included. Twenty-eight were children aged 22 days-13 years and two were young adult aged 18-20 years. All patients were of Algerian origin and from different geographic locations. These patients came to our laboratory for karyotype analyses. The controls were normal adult males with 46,XY and females with 46,XX chosen from volunteers. A blood sample was obtained from each subject for karyotype analysis, hormonal analysis and DNA extraction.

Cytogenetic analysis

Peripheral blood lymphocyte cultures were set up using TC199 medium for human lymphocyte culture for 72 hours. Dividing cells were arrested at metaphase stage with Colcemide and fixed using Carnoy acetic solution after treatment with hypotonic solution according to standard procedures. The harvested cells were dropped on clean slides and the chromosomes were studied by RHG method (R-bands after heat denaturation and Giemsa). A total of 30 metaphases were analyzed for each sample and the karyotypic descriptions were according to the ISCN recommendations to look at any numerical or structural chromosomal aberrations²⁰.

PCR and sequencing the coding region of candidate genes

Previously we have analyzed the presence of the SRY gene in leucocytes in all cases by PCR. The complete open reading frames of SRY and each exon of NR5A1 were amplified by polymerase-chain-reaction (PCR) according to a previous report^{16,21}. After amplification, PCR products were electrophoresed in a 2% agarose gel and afterwards the PCR products were purified using exonuclease I and phosphatase alkaline. These products were then sequenced (10-15ng DNA template reaction) on an Automated DNA sequencer using the big dye V1.1 terminator cycle sequencing. Cycling conditions were as follows: initial denaturation at 96°C for 1min and 25 cycles at 96°C for 10 sec, 50°C for 5 s, and 60 °C for 4 min. DNA sequencing of both sense and antisense strands were carried out for all exons of the candidate genes.

Ethics

Ethics approval was obtained from the Ethics Committee of the university hospital center Benbadis of Constantine.

Results

Karyotype and Molecular Analysis

Table 1 shows the karyotype in the 30 cases with sex anomalies. Thirteen of these patients (43.33%) were re-

ferred as cases with ambiguous genitalia. Nine of them (30%) had 46,XY constitution, among these cases three had uterus. Three of them had 46,XX constitution (10%), one had uterus and ovaries/penis and testicles. The remaining case (3%) with ambiguous genitalia had

peripheral blood karyotype of 47,XYY, ectopic testicles, penis and labia Majora. Thirteen cases (43.33%) had been referred as patients with hypospadias with or without undescended testes with 46,XY constitution. The four remaining cases (13.33%) with undescended testes only had peripheral blood karyotype of 46,XY.

Patient	Age	Phenotype	Clinical data	Karyotype	SRY gene	NR5A1 gene	Consanguinity	Endocrine data
1	13 years	Male Partial gonadal dysgenesis	Hypoplastic Uterus	46,XY	Positive	Normal	+	Ν
2	8 years	dysgenesis Female Partial	ambiguous external	46,XY	Negative	Normal		T(N), FSH↑, LH↑
2	o years	gonadal dysgenesis	genitalia with bilateral inguinal hernia Uterus / ovaries		Regulive	Ttormar		
3	22 days	Male Partial gonadal dysgenesis	Micropenis Enlarged labia majora Hypoplastic uterus	46,XY	Positive	Normal		T↓ at birth
4	5 years	Ambiguous genitalia raised as a boy	Ambiguity of external genitalia Micopenis	46,XY	Positive	Normal		T ↓, FSH (N), LH ↓
5	20 years	Ambiguous genitalia raised as a boy	Prostate Ectopic testicles/ Penis Labia majora	47,XYY	Positive	Normal		T(N), FSH ↑, LH↑
6	18 years	Ambiguous genitalia raised as a boy	Ambiguity of external genitalia	46,XY	Positive	Normal		T(N), FSH ↑, LH↑
7	10 years	Ambiguous genitalia raised as a boy	Ambiguity of external genitalia	46,XY	Positive	Normal		T ↓, FSH (N), LH ↓
8	4 years	Ambiguous genitalia raised as a boy	Left testicle absent Right ectopic testicle Micropenis	46,XY	Positive	Normal		T ↓, FSH (N), LH ↓
9	3 months	Ambiguous genitalia raised as a boy	Ambiguity of external genitalia	46,XY	Positive	Normal		T(N), FSH (N), LH↓
10	23 days	Ambiguous genitalia raised as a boy	Ambiguity of external genitalia	46,XY	Positive	Normal		NA
11	4 years	Male with Hypospadias	Posterior hypospadias	46,XY	Positive	Normal		N
12	13 months	Male with Hypospadias	Vulviform hypospadias	46,XY	Positive	Normal		N
13	5 years	Male with Hypospadias	Posterior hypospadias	46,XY	Positive	Normal		N
14	4 years	Male with Hypospadias	Vulviform hypospadias Renal malformation	46,XY	Positive	Normal	+	N
15	3 years	Male with Hypospadias	Posterior hypospadias	46,XY	Positive	Normal		N
16	4 years	Male with Hypospadias	Posterior hypospadias	46,XY	Positive	Normal	+	N
17	5 years	Male with Hypospadias	Posterior hypospadias	46,XY	Positive	Normal	+	N
18	5 years	Male with Undescended testis	Bilateral undescended testes	46,XY	Positive	Normal		N
19	5 years	Male with Undescended testis	Bilateral undescended testes	46,XY	Positive	Normal		N
20	2 years	Male with Undescended testis	Bilateral undescended testes	46,XY	Positive	Normal		Ν
21	1 year	Male with Undescended testis	Bilateral undescended testes	46,XY	Positive	Normal		N
22	15 months	Male with Hypospadias and undescended testis	Posterior hypospadias with Bilateral undescended testes	46,XY	Positive	Normal	+	N
23	4 years	Male with Hypospadias and undescended testis	Posterior hypospadias with Bilateral undescended testes Micropenis	46,XY	Positive	Normal		Ν
24	3 years	Male with Hypospadias and undescended testis	Vulviform hypospadias with Chordee Bilateral undescended testes	46,XY	Positive	Normal		N
25	3 years	Male with Hypospadias and undescended testis	Vulviform hypospadias with Bilateral undescended testes	46,XY	Positive	Normal		N
26	2 years	Male with Hypospadias and undescended testis	Posterior hypospadias with Bilateral undescended testes	46,XY	Positive	Normal		Ν
27	5 years	Male with Hypospadias and undescended testis	Vulviform hypospadias with Bilateral undescended testes	46,XY	Positive	Normal		N
28	13 years	Male ovotesticular DSD	Uterus Ovaries/Penis /Testicles Hypospadias	46,XX	Negative	Normal	+	T ↑,FSH (N), LH \downarrow
29	13 years	46,XX virilised (gonad status unknown)	Ambiguity of external genitalia	46,XX	Negative	c.437G→C) polymorphism		T ↑,FSH↓ , LH↓
30	6 years	46,XX virilised (gonad	Ambiguity of external genitalia	46,XX	Negative	Normal	1	T ↓,FSH↑ , LH ↑

Table 1. Summary of all characteristics of included patients (n=30).

*T: Testosterone, N: T, FSH, LH Normal, NA: not available

Age- and sex-dependent hormone levels in the normal population are taken as given previously²².

Table 1 showed that six cases (20%) of the 30 had positive consanguinity. Cases 16 and 17 were brothers. The presence or absence of SRY gene for all cases was reported above in Table 1. One case of 46,XY gonadal dysgenesis carried a deletion of SRY. This patient has ambiguous external genitalia with bilateral inguinal hernia and female's internal reproductive organs (uterus and ovaries).

SRY and NR5A1 Genes Direct Sequencing

The SRY gene had only one exon. One pair of primer was designed for this exon. The whole coding sequence of SRY gene was sequenced but no mutation or deletion was found in all patients whose SRY gene was positive.

In the 30 cases with DSD we did not detect a mutation in the whole coding sequence of NR5A1 gene. We identified a single NR5A1 polymorphism. This patient presented with 46,XX DSD. The polymorphism consisted of a G to C transversion at nucleotide position 437 in exon 3 that is predicted to result in a p.Gly146Ala amino acid change.

Discussion

DSD ranging from minor genital abnormalities to complete sex-reversal are the most common birth defects with an incidence rate of almost 3%²³. Genetic studies of human patients presenting with DSD have revealed an increasing number of genes important of sex determination and differentiation. Our genetic study was divided into two main categories: karyotyping and molecular study. Karyotyping of the 30 patients revealed a normal female karyotype 46,XX for 3 cases and a normal male karyotype 46,XY for 26 cases and one karyotype with anomalies in the sex chromosomes 47,XYY. This is in contradiction with the finding of Abou El Ella et al.²⁴ and Anhalt et al.²⁵, who reported that the most common presentation of DSD was females (77% and 70% of cases, respectively). Karyotyping is essential for provisional diagnosis and classification of the DSD cases²⁶. Consanguinity was found in 6 cases of our 30 cases (20%). Two cases with hypospadias and ectopic testis were brothers. It is important to note that the most of the published data on the incidence of DSD available from Western countries are with low rates of consanguinity, and it may not be a true reflection of the worldwide prevalence. There is, however, some evidence for a higher rate of DSD in societies

with a higher rate of consanguinity²⁷. In this context, Abou El Ella et al.²⁴ and Shawky et al.²⁸ found a consanguinity rate of 61.54% and 35.3%, respectively. A retrospective study in Sudan, of the 122 cases of DSD, found that 69 cases were 46,XX DSD and 45 cases were 46,XY DSD. The most common cause of 46,XX DSD was CAH 21-hydroxylase deficiency, whereas androgen insensitivity was the most frequent cause of DSD in 46,XY individuals. 70% of all 122 cases were born to first cousin marriages²⁹. A study of 26 cases of DSD from Sudan indicated that parental consanguinity was observed in 70% of cases³⁰. Consanguinity has been proposed as the cause of a high frequency of 46,XX DSD (CAH, 65.4% of total DSD cases) in one referral center in Saudi Arabia over a 20 year period³¹. In our study most of the cases of ambiguous genitalia had a 46,XY karyotype and only three cases were 46,XX.

These conflicting results indicate that consanguinity may lead to an increase in the incidence rates of both 46,XY and 46,XX DSD, depending on the population studied.

The main gene implicated in sex determination in mammals is the sex-determining region on the Y chromosome (SRY), which encodes a transcription factor that is a member of the high mobility group (HMG)-box family of DNA binding proteins and in mammals triggers the development of undifferentiated gonads towards a testicular phenotype³². In humans, zygotes bearing mutations in SRY develop into XY females³³, whilst XX individuals with the presence of SRY typically show a normal male phenotype³⁴ but may occasionally show ambiguous genitalia³⁵. Only one of our 26 patients with 46,XY karyotype was SRY-negative. Gonadal dysgenesis in this patient was caused by the absence of the SRY gene. Two of the XX individuals were SRY-negative, but show a male phenotype and ambiguous genitalia. Most SRY-negative XX males have a high incidence of genital ambiguity or hypospadias³⁶. Direct sequencing of the whole SRY coding region revealed no mutation in all patients studied. Mutations in the SRY gene have been described in patients with DSD^{5,37-43}. Mutations involving the SRY gene are almost exclusively associated with complete rather than partial gonadal dysgenesis and this may explain the lack of SRY mutations in this cohort. Furthermore, in male sexual development there are many other genes, which also play a crucial role. NR5A1 is a key regulator of adrenal and reproductive development and function. Approximately, 191 variants NR5A1 have been identified in humans to date44. In the 30 studied cases with DSD, no mutations were detected

in the NR5A1 gene. One subject who had 46, XX DSD was heterozygous for the p.Gly146Ala (c.437G \rightarrow C) polymorphism that has previously been demonstrated to have approximately 80% of the activity of the wild-type protein⁴⁵. The contribution of this variant to the development of DSD is unknown. It have been reported that this variant, was found to be very frequent in Egyptian cohort with 46, XY DSD (34 %)⁴⁶ which is not the case in our cohort.

The limitation of the current study is small sample size and the fact that only two genes involved in the sex determination were screened.

Conclusion

We found in Algeria that the most common form of DSD associated with ambiguous external genitalia in the newborn is 46,XY DSD. The pathology of the majority of these cases is not explained by mutations in the two most common genes associated with DSD namely, SRY and NR5A1 suggesting that other genes are involved. Finally, we anticipate in the future sequencing a panel of genes and whole exome for genetic diagnosis of Algerian patients with DSD.

Acknowledgements

Authors appreciate the kind participation of the patients and controls. We are grateful to all members of Biology and Genetics Laboratory, of Human Genetic developmental Unit, Pasteur Institute and of Pediatric Urology Unit of Setif for their contributions to this work.

Conflict of Interest Disclosure

The authors declare that they have no conflict of interests.

References

1. Lee PA, Nordenström A, Houk C, Ahmed SF, Auchus R, Baratz A, et al. Global Disorders of Sex Development Update since 2006: Perceptions, Approach and Care. *Horm Res Paediatr*. 2016; 85(3):158-180. DOI: 10.1159/000442975.

2. Massanyi EZ, DiCarlo HN, Gearhart JP, Migeon CJ. Review and management of 46,XY Disorders of Sex Development. *J Pediatr Urol.* 2013; 9: 368-379. DOI: 10.1016/j.jpurol.2012.12.002.

3. Govind BC, Dimitri AP, Kamaldine O, Stephen FM, Paul SB, Foao LPS. Imaging of ambiguous genitalia: classification and diagnostic approach. *Radio Graphics.* 2008; 281891-1904. DOI: 10.1148/rg.287085034.

4. Jiang T, Hou CC, She ZY, Yang WX. The SOX gene

family: function and regulation in testis determination and male fertility maintenance. *Mol Biol Rep.* 2013; 40(3):2187-2194. DOI: 10.1007/s11033-012-2279-3.

5. Salehi LB, Scarciolla O, Vanni GF, Nardone AM, Frajese G, Noveli G, et al. Identification of a novel mutation in the SRY gene in a 46, XY female patient. *Eur J Med Genet.* 2006; 49:494-498. DOI : org/10.1016/j. ejmg.2006.03.003.

6. Parker KL, Shimmer BP. Steroidogenic factor 1: a key determinant of endocrine development and function. *Endocr Rev.* 1997; 18: 361-377. DOI: 10.1210/edrv.18.3.0301.

7. Ferraz-de-Souza B, Lin L, Achermann JC. Steroidogenic factor-1 (SF-1, NR5A1) and human disease. *Mol Cell Endocrinol.* 2011; 336(1-2):198-205. DOI: 10.1016/j. mce.2010.11.006.

8. Philibert P, Zenaty D, Lin L, Soskin S, Audran F, Léger J, et al. Mutational analysis of steroidogenetic factor 1 (NR5a1) in 24 boys with bilateral anorchia: a French collaborative study. *Hum Reprod.* 2007; 22: 3255-3261. DOI: 10.1093/humrep/dem278.

9. Lin L, Achermann JC. Steroidogenic factor-1 (SF-1, Ad4BP, NR5A1) and disorders of testis development. *Sexual Dev.* 2008; 2(4–5): 200-209. DOI: 10.1159/000152036.

10. Köhler B, Lin L, Mazen I, Cetindag C, Biebermann H, Akkurt I, et al. The spectrum of phenotypes associated with mutations in steroidogenic factor 1 (SF-1, NR5A1, Ad4BP) includes severe penoscrotal hypospadias in 46,XY males without adrenal insufficiency. *Eur J Endocrinol.* 2009; 161:237-242. DOI: 10.1530/EJE-09-0067.

11. Allali S, Muller JB, Brauner R, Lourenco D, Boudjenah R, Karageorgou V et al. Mutation Analysis of NR5A1 Encoding Steroidogenic Factor 1 in 77 Patients with 46, XY Disorders of Sex Development (DSD) Including Hypospadias. *PloS One*. 2011; 6 :(10) 1-8. DOI: 10.1371/journal.pone.0024117.

12. Brandt T, Blanchard L, Desai K, Nimkarn S, Cohen N, Edelmann L, et al. 46,XY disorder of sex development and developmental delay associated with a novel 9q33.3 micro deletion encompassing NR5A1. *Eur J Med Genet.* 2013; 56: 619-623. DOI: 10.1016/j. ejmg.2013.09.006.

13. Ferlin A, Rocca MS, Vinanzi C, Ghezzi M, Di Nisio A, Foresta C. Mutational screening of NR5A1 gene encoding steroidogenic factor 1 in cryptorchidism and male factor infertility and functional analysis of seven undescribed mutations. *Fertil Steril.* 2015; 104(1):163-9. DOI: 10.1016/j.fertnstert.2015.04.017.

14. Rocca MS, Ortolano R, Menabo S, Baronio F, Cassio

A, Russo G, et al. Mutational and functional studies on NR5A1 gene in 46,XY disorders of sex development: identification of six novel loss of function mutations. *Fertil Steril.* 2018; 109 (6):1105-1113. DOI: 10.1016/j. fertnstert.2018.02.123.

15. Biason-Lauber A, Schoenle EJ. Apparently normal ovarian differentiation in a prepubertal girl with transcriptionally inactive steroidogenic factor 1 (NR5A1/SF-1) and adrenocortical insufficiency. *Am J Hum Genet.* 2000; 67(6): 1563-1568. DOI: 10.1086/316893.

16. Lourenco D, Brauner R, Lin L, De Perdigo A, Weryha G, Muresan M, et al. Mutations in NR5A1 associated with ovarian insufficiency. *N Engl J Med.* 2009; 360: 1200-1210. DOI: 1210, 10.1056/NEJMoa0806228.

17. Warman DM, Costanzo M, Marino R, Berensztein E, Galeano J, Ramirez PC, et al. Three new SF-1 (NR5A1) gene mutations in two unrelated families with multiple affected members: within-family variability in 46, XY subjects and low ovarian reserve in fertile 46,XX subjects. *Horm Res Paediatr.* 2011; 75: 70-77. DOI: 10.1159/000320029.

18. Röpke A, Tewes AC, Gromoll J, Kliesch S, Wieacker P, Tüttelmann F. Comprehensive sequence analysis of the NR5A1 gene encoding steroidogenic factor 1 in a large group of infertile males. *Eur J Hum Genet.* 2013; 21(9):1012-1015. DOI: 10.1038/ejhg.2012.290.

19. Knarston IM, Robevska G, Van den Bergen JA, Eggers S, Croft B, Yates J, et al. NR5A1 gene variants repress the ovarian-specific WNT signaling pathway in 46,XX disorders of sex development patients. *Hum mu-tat.* 2019; 40(2): 207-216. DOI: 10.1002/humu.23672.

20. ISCN 2013: An International System for Human Cytogenetic Nomenclature. Shaffer LG, McGowan-Jordan J, Schmid M (Eds). KARGER, 2013.

21. Brauner R, Neve M, Allali S, Löhrs U, Schwarz HP, Kuhnle U. Clinical, Biological and Genetic Analysis of Anorchia in 26 Boys. *PLoS One.* 2011; 6 (8) e23292. DOI: 10.1371 /journal.pone.0023292.

22. Juniarto AZ, Van der Zwan YG, Santosa A, Ariani MD, Eggers S, Hersmus R, et al. Hormonal evaluation in relation to phenotype and genotype in 286 patients with a disorder of sex development from Indonesia. *Clin Endocrinol* (Oxf). 2016; 85(2):247-257. DOI : 10.1111/cen.13051.

23. Tannour-Louet M, Mazen I, Soliman H, Louet JF, Yatsenko S, Meyers L, et al. Identification of de novo copy number variants associated with human disorders of sexual development. *PLoS One*. 2010; 5 (10), e15392. DOI: 10.1371/journal.pone.0015392.

24. Abou El Ella SS, Tawfik MAM, Ellahony DM, Anees NM. Cytogenetic and molecular study in intersex. *Egypt*

J Med Hum Genet. 2012; 13: 281-289. DOI:10.1016/j. ejmhg.2012.06.003.

25. Anhalt H, Neely EK, Hintz R. Ambiguous Genitalia. *Pediatr Rev.* 1996; 17(6):213-220. DOI:10.1542/ pir.17-6-213.

26. Audi L, Ahmed SF, Krone N, Cools M, McElreavey K, Holterhus PM, et al. GENETICS IN ENDOCRI-NOLOGY: Approaches to molecular genetic diagnosis in the management of differences/disorders of sex development (DSD): position paper of EU COST Action BM 1303 'DSDnet'. *Eur J Endocrinol.* 2018; 179(4):197-206. DOI: 10.1530/EJE-18-0256.

27. Bashamboo A, McElreavey K. Consanguinity and Disorders of Sex Development. *Hum Hered.* 2014; 77:108-117. DOI: 10.1159/000360763.

28. Shawky RM, El-Awady MY, Elsayed SM, Hamadan GE. Consanguineous mating among Egyptian population. *Egypt J Med Hum Genet*. 2011; (12):157-163. DOI: 10.1016/j.ejmhg.2011.07.001.

29. Abdullah MA, Saeed U, Abass A, Lubna K, Weam A, Ali AS, et al. Disorders of sex development among Sudanese children: 5-year experience of a pediatric endocrinology clinic. *J Pediatr Endocrinol Metab.* 2012; 25: 1065-1072. DOI: 10.1515/jpem-2011-0467.

30. Ellaithi M, Kamel A, Saber O, Hiort O. Consanguinity and Disorders of Sexual Developments in the Sudan. *Sudan J Med Sci.* 2011; 6:267-270. eISSN: 1858-5051.

31. Al Jurayyan N. Ambiguous genitalia: two decades of experience. *Ann Saudi Med.* 2011; 31:284-288. DOI: 10.4103/0256-4947.81544.

32. Sinclair AH, Berta P, Palmer MS, Hawkins JR, Griffiths BL, Smith MJ, et al. A gene from the human sex determining region encodes a protein with homology to a conserved DNA binding motif. *Nature*. 1990; 346:240-244. DOI: 10.1038/346240a0.

33. Berta P, Hawkins J, Sinclair A, Taylor A, Griffiths BL, Goodfellow PN, et al. Genetic evidence equating SRY and the testis-determining factor. *Nature*. 1990; 348 : 448-450. DOI: 10.1038/348448a0.

34. Fechner PY, Marcantonio SM, Jaswaney VJ, Stetten G, Goodfellow PN, Migeon CJ, et al. The role of the sex determining region Y gene (SRY) in the etiology of 46,XX maleness. *J Clin Endocrinol Metab.* 1993; 76:690-696. DOI: 10.1210/jcem.76.3.8383144.

35. Kusz K, Kotecki M, Wojda A, Szarras-Czapnik M, Latos-Bielenska A, Warenik-Szymankiewicz A, et al. Incomplete masculinisation of XX subjects carrying the SRY gene on an inactive X chromosome. *J Med Gene*. 1999; 36:452-456. PMCID: PMC1734388.

36. Huang B, Wang S, Ning Y, Lamb A, Bartley J. Au-

tosomal XX Sex Reversal Caused by Duplication of SOX9. *Am J Hum Genet.* 1999; 87:349-353. DOI: 10.1002/(sici)1096-8628(19991203)87:4<349::aidajmg13>3.0.co;2-n.

37. Shahid M, Dhillon VS, Hussain Z, Masa JF, Aslam M, Raish M, A et al. Analysis of the SRY gene in two sex-reversed XY sisters identifies two new novel point mutations in the high mobility group box domain. *Fertil Steril.* 2008 ; 90(4) : 1199.e1-8. DOI: 10.1016/j.fertn-stert.2007.11.062.

38. Laino L, Majore S, Preziosi N, Grammatico B, De Bernardo C, Scommegna S, et al. Disorders of sex development: a genetic study of patients in a multidisciplinary clinic. *Endocr Connect.* 2014; 3(4):180-192. DOI :10.1530/EC-14-0085.

39. Andonova S, Robeva R, Sirakov M, Mainhard K, Tomova A, Ledig S, et al., A Novel SRY Gene Mutation p.F109L in a 46,XY Female with Complete Gonadal Dysgenesis. *Sex Dev.* 2015 ; 9(6) :333-337. DOI : 10.1159/000443807.

40. Tajouri A, M'sahli M, Hizem S, Jemaa LB, Maazoul F,M'rad Redha, et al. A Novel Nonsense Mutation p.L9X in the SRY Gene Causes Complete Gonadal Dysgenesis in a 46,XY Female Patient. *J Genet Syndr Gene Ther.* 2016 ; 7:300. DOI : 10.4172/2157-7412.1000300. 41. Fan W, Wang B, He S, Zhang T, Yin C, Chen Y, et al. A Novel Missense Mutation 224G>T (R75M) in SRY Coding Region Interferes with Nuclear Import and Results in 46, XY Complete Gonadal Dysgenesis. *PloS One.* 2016; 11(12), e0168484. DOI: 10.1371/journal.pone.0168484.

42. Wang X, Xue M, Zhao M, He F, Li C, Li X. Identification of a novel mutation (Ala66Thr) of SRY gene causes XY pure gonadal dysgenesis by affecting DNA binding activity and nuclear import. *Gene.* 2018 ; 651:143-151.DOI:10.1016/j.gene.2018.01.076.

43. Raveendran SK, Ramachandran L, Joseph L, Asokan AK, Raj S, George A, et al. A novel SRY gene mutation c.266 A>T (p.E89V) in a 46,XY complete gonadal dysgenesis patient. *Andrologia*. 2019 ; 51(9) :e13377. DOI: 10.1111/and.13377.

44. Fabbri-Scallet H, Sousa L M, Maciel-Guerra AT, Guerra-Júnior G, Mello M P. Mutation update for the NR5A1 gene involved in DSD and infertility. *Hum Mutat.* 2020, 41(1):58-68. DOI: 10.1002/humu.23916.

45. WuQiang F, Yanase T, Wei L, Oba K, Nomura M, Okabe T, et al. Functional characterization of a new human Ad4BP/SF-1 variation, G146A. *Biochem Biophys Res Commun.* 2003; 311:987-994. DOI: 10.1016/j. bbrc.2003.10.096.

46. Tantawy S, Mazen I, Soliman H, Anwar G, Atef A, El-Gammal M, et al. Analysis of the gene coding for steroidogenic factor 1 (SF1, NR5A1) in a cohort of 50 Egyptian patients with 46,XY disorders of sex development. *Eur J Endocrinol.* 2014, 170 (5): 759-767. DOI: 10.1530/EJE-13-0965.