**Full Length Research Paper**

**Chemical composition and some functional properties of soluble fibro-protein extracts from Tunisian date palm seeds**

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This study is a contribution to give value addition to date palm seeds by extracting an enriched fibro-protein fraction (DSFPE) and to examine the effect of pH on some of its functional properties. For this purpose, DSFPE was prepared from water soluble extracts of defatted Deglet Nour and Allig seeds and obtained by precipitation at pH 4.5. Then, DSFPE was examined for their proximate chemical composition. Significant differences were observed between Deglet Nour and Allig DSFPE: Carbohydrate was 64 against 58%, protein was 33 against 38%, and ash was 2 against 3%, respectively. Glutamic acid presented the largest amount, varying from 17.14% for Deglet-Nour DSFPE to 14.71% for the Allig DSFPE. The essential amino acids (lysine, leucine, threonine, methionine, valine, isoleucine and phenylalanine) were present in the DSFPE of the two studied varieties. Effect of pH on colour and some functional properties were analysed. Colour profiles of Deglet Nour and Allig water soluble extracts were affected by pH (2 to 10). Minimum protein solubility was obtained at pH 3.5 to 4.5 and the maximum at pH 10. Water holding capacity (WHC) and oil holding capacity (OHC) of Deglet Nour and Allig DSFPE were 3.50 to 4.50 g H₂O g⁻¹ of DSFPE and 5.50 to 6.10 g oil g⁻¹ DSFPE respectively. Emulsion and foam properties were analysed at pH 7 and 10. DSFPE presented a slightly higher foam capacity (11 to 14 cm) and lower foam stability at pH 7 or 10. Emulsion capacity of DSFPE was significantly higher at pH 10 (2800 to 3000 ml oil g⁻¹ of protein) than those at pH 7 (2000 to 2400 ml of oil g⁻¹ of protein). Emulsion stability was improved with increase in pH from 7 to 10. These results suggested that the DSFPE have a good potential in food industry and can be used to improve the techno-functional quality for neutral and alkaline food applications with a high commercial value.

Key words: Phoenix dactylifera L, date palm seed, fibre, protein, functional properties.

**INTRODUCTION**

The date palm (Phoenix dactylifera L.) is one of the most cultivated palms around the world. It is commonly found in the Afro-Asiatic dry-band, which stretches from North Africa to the Middle East (Barreveld, 1993). It has a good tolerance to cold and dry-hot climates. The fruit is composed of a fleshy pericarp and seed which constitutes between 10 and 15% of date fruit weight (Hussein et al., 1998; Almana and Mahmoud, 1994; Besbes et al., 2004; Al Farsi et al., 2007, Al Farsi and Lee, 2008; Elleuch et al., 2008).

Chemical composition of date pits showed high amount of fibre (75 to 80%), fat (10 to 13%), proteins (5 to 6%) and ash (El-Shurafa et al., 1982; Devshony et al., 1992; Al-Hooti et al., 1998; Hamada et al., 2002; Besbes et al., 2004a, 2005b; Al Farsi et al., 2007, Al Farsi and Lee, 2008). Presently however, very little use is made of these...
The production of technofunctional ingredient with high quality using a simple extraction by water could give a high value addition to date palm seeds. In this work, we are interested in the optimised extraction process of fraction with high fibre and protein contents from two important cultivars grown in Tunisia: Allig and Deglet Nour dates palm seeds. We then evaluated the chemical composition of DSFPE. Furthermore, we studied the effects of pH on some functional properties of DSFPE and predicted its compatibility in different food systems.

MATERIALS AND METHODS

Sample

Date palm fruits were obtained from the National Institute of Arid Zone (Degach, Tunisia). The seeds of the two cultivars under investigation (Deglet Nour and Allig) were directly isolated from 50 kg of date fruit having the same origin; collected at the “Tamr stage” (full ripeness) and kept at 10°C for a week.

Preparation of defatted date palm seeds

The seeds were soaked in water, washed to get rid of any adhering date flesh, and then air-dried. Their relative percentage weight compared with the weight of the fresh fruits was about 11.32% for the Deglet Nour variety and about 10.7% for the Allig variety. Then, they were further dried at about 50°C. Date pits, of each variety, were separately milled in a heavy-duty grinder to pass 1 to 2 mm screens and then preserved at -20°C until analyses.

Lipid extraction was carried out as described by Besbes et al. (2004a) with a SER 148 solvent extractor (Velp Scientifica, Italy) equipped with six Soxhlet posts. The extraction was carried out for 30 min, with thimbles immersed in boiling petroleum ether, and 60 min of reflux washing. After removing solvent, using a rotavapor apparatus, the obtained defatted date seeds were used for preparation of fibro-protein extract.

Extraction procedure of DSFPE

The fibro-protein extract from defatted date seed was prepared according to the Tsaliki et al. (2002) method. The defatted date seed flower was mixed with distilled water (1:10 w/v), adjusted to pH 10 with NaOH and after stirring for at least 40 min, was centrifuged at 6500 g for 20 min at 4°C (Beckman J2-21, USA). Then, the residue was mixed with distilled water (1:5 w/v), readjusted to pH 10 and centrifuged following the same process. The supernatants of both centrifugations were blended and used as mother solution for DSFPE production. This mother solution was adjusted at pH 4.5 with 0.1 HCl, centrifuged, freeze and lyophilized to obtain the DSFPE (Figure 1).

Chemical analysis of powdered seeds

All analytical determinations were performed at least in triplicate. Values of different parameters were expressed as the mean ± standard deviation (±S.D.).

Dry matter was determined according to the Association of Official Analytical Chemists (AOAC, 1995). Nitrogen content of defatted samples was determined by Kjeldahl method, following the method of the AOAC (1995). Protein content of each sample was calculated by multiplying the total nitrogen content by a factor of 6.25 (Besbes et al., 2004a). Protein yield was calculated after the determination of protein content in the powdered seed and in the lyophilised supernatant.

Carbohydrate content was estimated by difference of mean values, that is, 100 - (sum of percentages of moisture, ash, protein and lipids). Ash content was determined after incineration at 550°C, during 8 h, using a muffle furnace (NABER, Germany). It was expressed as percent of dry weight (AOAC, 1995). Amino acids of DSFPE (150 mg) were analysed by a BioChrom 20 plus amino acid analyser according to the method of Bouaziz et al. (2008) after hydrolyses with HCl (6N) at 110°C for 24 h. Amino acids were analysed by chromatographic ionic exchange and detected colorimetrically using Ninhydrin reagent. All amino acids were detected at 570 nm except proline and hydroxyproline, which were detected at 440 nm. The amino acid concentrations were calculated from the standard curves.

Measurements of colour

The CIE Lab parameters (L*, a*, b*) were directly read with a spectrophotocolorimeter MS/Y-2500 (Hunterlab, In., Reston, VA, USA), calibrated with a white tile. In this coordinate system, the L* value is a measure of lightness, ranging from 0 (black) to 100 (white); the a* value ranges from -100 (green) to +100 (red) and the b* value ranges from -100 (blue) to +100 (yellow).

Protein solubility

Nitrogen solubility of proteins at 1% (w/v) from defatted Deglet Nour and Allig seeds were determined by the Bio-Rad’s protein assay kit following the procedure of Bradford (1976). Serial dilutions of bovine serum albumin (BSA) were used for the construction of a standard curve from 0.2 to 1.5 mg/ml.

Protein solutions were adjusted to pH values from 2 to 10. The dispersions were stirred then centrifuged at 6500 g for 20 min (JOUAN CR 42, USA). Protein content of the supernatants was determined. A Shimadzu UV-160 A spectrophotometer was used for absorbance measurements at 595 nm.

Nitrogen solubility index (NSI) was calculated as percent of nitrogen content of the sample to the nitrogen content in the solution (Besbes et al., 2002).
Water and oil holding capacity

The method of Moure et al. (2001) was used with a slight modification. 1 g of protein samples was stirred in 10 ml of distilled water or corn oil and then centrifuged at 6000 rpm for 20 min (JOUAN CR4 22, USA). The volume of the supernatant was measured. The water-holding capacity was expressed as the number of gram of water held by 1.0 g of protein sample. The oil-holding capacity was expressed as the number of gram of oil held by 1.0 g of protein sample.

Surface tension determination

The automated drop volume tensiometer TVT 1 (Lauda, Germany) was employed to perform dynamic measurements. The tensiometer
Table 1. Chemical composition (dry basis) of principal component of DSFPE and protein yields.

<table>
<thead>
<tr>
<th>Component (%)</th>
<th>DSFPE</th>
<th>Allig</th>
<th>Deglet Nour</th>
</tr>
</thead>
<tbody>
<tr>
<td>Dry matter</td>
<td>98.10 ±0.37</td>
<td>98.40±0.48</td>
<td></td>
</tr>
<tr>
<td>Protein</td>
<td>38.68 ±0.13</td>
<td>33.53±0.42</td>
<td></td>
</tr>
<tr>
<td>Carbohydrate</td>
<td>58.77±1.33</td>
<td>63.37±1.70</td>
<td></td>
</tr>
<tr>
<td>Ash</td>
<td>2.55±0.20</td>
<td>3.10±0.28</td>
<td></td>
</tr>
<tr>
<td>Protein yields</td>
<td>23.40±0.37</td>
<td>24.10±0.46</td>
<td></td>
</tr>
</tbody>
</table>

DSFPE: Date seed fibro-protein extract; values in lines with different letters are significantly different (p<0.05).

was connected to a computer.

Drops of solution were formed with a growing formation speed. The life time of the drops was measured as a function of their volume, which made it possible to calculate the surface tension. All measurements were performed at 25 ± 0.5°C. Each measurement was repeated twice. For high concentrations of surfactant, equilibrium surface tension (σ_e) was taken as the mean of the values obtained with the last four drops. For low concentrations, σ_e was deduced by extrapolating the surface tension to time t →∞ in the σ = t⁻¹/² (Blecker et al., 2002).

Foam proprieties measurements

Foam capacity and stability at pH 7 and 10 for solution 1% protein of DSFPE were determined according to Blecker et al. (1997) method. Dispersion (3 ml) was deposited on a porous glass plate (pore size: 40 to 100 µm) in a 30 x 300 mm graduated column. Foam capacity (FC) was defined as the height of foam after injection of air at a constant rate (120 ml min⁻¹) for one min. Foam stability was estimated by monitoring the height of foam vs. time. These measurements took place in triplicate and the values given are the mean values of three measurements.

Emulsion proprieties measurements

The emulsion capacity was determined by a model system described by Blecker et al. (1997); 50 ml of protein solutions (0.1% w/v) adjusted to pH 7 or 10 with 0.1 N NaOH. Then, sunflower oil was added and emulsified using an Ultraturrax T25 (Ika, Staufen, Germany) at 15000 rpm. During emulsification, temperature was maintained at 0°C by immersing the reaction vessel in ice bath. The sudden increase in electrical resistance showed the phase inversion point; the oil phase becomes continuous, which can be determined by electrical conductivity measurements. Emulsion capacity is expressed in g oil g⁻¹ protein.

Emulsion stability was determined using a Turbiscan MA 2000 (Formulaction, Ramonville St Agne, France). Creaming was monitored. Programmable in time, the system enabled the determination of light scattering profiles of the sample. Creaming intensities were obtained by integration of the increasing reflected light peaks of the cream layer formation. It was expressed as percentage with respect to whole reflected light intensity of the sample at time 0 (Blecker et al., 1997).

Statistical analysis

Duncan’s est, at the level of P ≤ 0.05 was applied to the data to establish significance of difference between the samples. Statistical analyses were performed on statistical analysis package STATISTICA (Release 5.0 Stat Soft Inc., Tulsa, Oklahoma, USA).

RESULTS AND DISCUSSION

Chemical composition

Table 1 presents the chemical composition of Allig and Deglet Nour DSFPE. Carbohydrate content was about 58.77 to 63.37%, protein content was about 38.68 to 33.53% and ash was about 2.55 to 3.10% for Allig and Deglet Nour DSFPE, respectively. Deglet Nour DSFPE presented higher carbohydrate content compared to Allig DSFPE (P≤0.05). These values indicated the water soluble fibre extracted from defatted date seeds. Significant difference was observed between DSFPE Allig (38%) and DSFPE Deglet Nour (33%) proteins (P≤0.05). These results could be due to the solubility of protein in pH extraction. In fact, precipitation at pH 4.5 improved the DSFPE protein contents in date seeds (5 to 6%) (Besbes et al., 2004a) 14 to 16% in water soluble extracts (Bouaziz et al., 2008) to 33 to 38% in DSFPE. These results show that Allig DSFPE contained more proteins than Deglet Nour DSFPE because of their low protein solubility.

Protein yield was about 23 to 25% of the total date seed proteins (Table 1). Therefore, a large portion of date seed proteins was insoluble. These insoluble fractions are likely to be composed of high-molecular weight polypeptides that are highly aggregated and or cross-linked by disulphide bridges.

DSFPE from Deglet Nour and Allig varieties have a similar amino acid profiles (Table 2). 17 types of amino acids were detected and identified. Glutamic acid (Glu) was the predominant amino acid, followed by arginine (Arg), aspartic acid (Asp), leucine (Leu), lysine (Lys), valine (Val), glycine (Gly), alanine (Ala) and phenylalanine (Phe). DSFPE contained amino acid more than the defatted date seeds reported by Bouaziz et al. (2008). Glutamic acid presented the largest amount, varying from 17.14% for Deglet-Nour DSFPE to 14.71% for the Allig DSFPE. The essential amino acids (lysine, leucine, threonine, methionine, valine, isoleucine and phenylalanine) were present in the DSFPE of the two studied varieties, except tryptophan. The disappearance of tryptophan could be attributed to its destruction during acid hydrolysis and could also account for the loss of cysteine (Salim and Ahmed, 1992).

DSFPE proteins are of a relatively important biological value by reference to the standard egg proteins, considering their wealth in essential amino acids (Fayadh and Al-Showiman, 1990). This result showed the importance of precipitation undergone by the water soluble extract to improve protein contents. These findings could be explained by the low solubility of proteins at pH 4.5.
For these high contents in fibre and protein, DSFPE can be used as ingredient in dietetic food formulations. Incorporation of this fibro-protein fraction (DSFPE) in food formulations like an inexpensive ingredient could be improved by the nutritional and dietetic qualities in the finished product.

### Colour

CIE lab parameters ($L^*$, $a^*$ and $b^*$) for the water soluble extracts from Allig and Deglet Nour seeds are given in Figure 2. The water soluble extracts of Deglet Nour seeds were darker than those of Allig seeds whatever the pH was. $a^*$ increased but $L^*$ and $b^*$ decreased although it is known that Allig seeds are darker. This result can be explained by a better diffusion of pigments during the alkaline extraction.

The redness of date seed water extracts increased with the increase of pH. In addition, brightness and yellowness were reduced with the increase in pH. This variation of colour was reversible if pH decreased. These results could be explained, probably, by the following assumptions: i) the variation of pH modified the protein structures and changed their interactions with pigments. The precipitation of proteins at low pH could be the origin of masked pigments that were imprisoned in the precipitate; ii) the tannin reaction with the acid or the base solution could be the origin of colour variation.

### Protein solubility

The protein solubility of defatted date seeds from the two studied varieties is shown in Figure 3. The nitrogen solubility profiles presented a maximum at alkaline pH (10) and the minimum at acidic pH (2 to 4.5). Similar effects were reported for protein from *Rosa rubiginosa* (Moure et al., 2001), *Guevina avellana* (Moure et al., 2002) lupin seed (El-Adawy et al., 2001), cotton seed (Tasaliki et al., 2002), sesame seed (Khalid et al., 2003) and defatted *Erythrina variegata* flour (Jyothirmayi et al., 2006). The nitrogen solubility at pH 10 (highest solubility) varied from 84 to 90% and 64 to 79% at pH 7.

Deglet Nour seed proteins were more soluble especially at pH 5 to 10 than Allig seed proteins. For example, at pH 7, protein solubility was 79.1% for Deglet Nour seed proteins against 64.8% for those from Allig seeds.

For the two studied date varieties, the low solubility of date seed proteins was obtained at pH between 4 and 4.5; NSI was minimal (~22 to 26%). For this reason, we chose to carry out precipitation at pH 4.5 to obtain DSFPE. Therefore, this result showed that the isoelectric pH can be ranged between pH 4 and 4.5. However, with further acidification, solubility was inversely increased (at pH 2, NSI = 37 to 39%).

The protein solubility reduction could be due to the decrease of electrostatic repulsions and the hydrophobic interactions induction of protein aggregation at pH 4 to 4.5. El-Adawy et al. (2001), Tasaliki et al. (2002), Khalid et al. (2003) and Jyothirmayi et al. (2006) reported similar

### Table 2. Amino acid composition of DSFPE (mg·g⁻¹ of dry matter).

<table>
<thead>
<tr>
<th>Amino acid (mg/g)</th>
<th>DSFPE Allig</th>
<th>DSFPE Deglet-Nour</th>
</tr>
</thead>
<tbody>
<tr>
<td>Asp</td>
<td>1.96±0.05</td>
<td>1.92±0.06</td>
</tr>
<tr>
<td>Thr</td>
<td>0.74±0.02</td>
<td>0.80±0.09</td>
</tr>
<tr>
<td>Ser</td>
<td>0.92±0.08</td>
<td>0.96±0.05</td>
</tr>
<tr>
<td>Glu</td>
<td>5.69±0.04</td>
<td>5.75±0.01</td>
</tr>
<tr>
<td>Pro</td>
<td>0.82±0.08</td>
<td>0.78±0.07</td>
</tr>
<tr>
<td>Gly</td>
<td>1.28±0.04</td>
<td>1.24±0.06</td>
</tr>
<tr>
<td>Ala</td>
<td>1.05±0.05</td>
<td>1.02±0.04</td>
</tr>
<tr>
<td>Cys-Cys</td>
<td>0.63±0.07</td>
<td>0.40±0.07</td>
</tr>
<tr>
<td>Val</td>
<td>1.34±0.07</td>
<td>1.25±0.09</td>
</tr>
<tr>
<td>Met</td>
<td>0.43±0.04</td>
<td>0.41±0.01</td>
</tr>
<tr>
<td>Ile</td>
<td>0.74±0.02</td>
<td>0.71±0.05</td>
</tr>
<tr>
<td>Leu</td>
<td>1.54±0.02</td>
<td>1.49±0.03</td>
</tr>
<tr>
<td>Tyr</td>
<td>0.46±0.05</td>
<td>0.44±0.02</td>
</tr>
<tr>
<td>Phe</td>
<td>1.03±0.01</td>
<td>0.97±0.03</td>
</tr>
<tr>
<td>His</td>
<td>0.51±0.02</td>
<td>0.50±0.01</td>
</tr>
<tr>
<td>Lys</td>
<td>1.39±0.03</td>
<td>1.33±0.09</td>
</tr>
<tr>
<td>Arg</td>
<td>3.57±0.01</td>
<td>3.48±0.02</td>
</tr>
<tr>
<td>Total</td>
<td>24.10±1.17</td>
<td>23.45±1.14</td>
</tr>
</tbody>
</table>

Values in the same lines with different letters are significantly different (p≤0.05)
Figure 2. Effect of pH on colour of water soluble extract from defatted date seeds. a, Allig; b, Deglet-Nour cultivars (■; L*, ♦; a*, ▲; b*).

Table 3. CIE Lab parameters (L*, a*, b*) of defatted date seeds and DSFPE from the two studied varieties.

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Allig seed</th>
<th>Deglet Nour seed</th>
<th>DSFPE Allig</th>
<th>DSFPE Deglet Nour</th>
</tr>
</thead>
<tbody>
<tr>
<td>L*</td>
<td>51.28±0.45</td>
<td>56.25±0.58</td>
<td>41.37±0.04</td>
<td>36.97±0.17</td>
</tr>
<tr>
<td>a*</td>
<td>14.58±0.56</td>
<td>13.09±0.23</td>
<td>21.78±0.03</td>
<td>28.55±0.06</td>
</tr>
<tr>
<td>b*</td>
<td>17.42±0.22</td>
<td>20.55±0.15</td>
<td>46.95±0.03</td>
<td>30.33±0.05</td>
</tr>
</tbody>
</table>

Values in the same lines with different letters are significantly different (p≤0.05).

Water and oil-holding capacity (WHC; OHC)

The capacities of water and oil retention of the DSFPE from date seeds are presented in Table 4. WHC of DSFPE varied between 4 and 5 g of water g⁻¹ of the sample. Due to these values, DSFPE from date seeds could be used like ingredient, to improve the sensory properties of the formulated product, by reducing and limiting the phenomenon of syneresis. OHC of DSFPE varied between 5 and 6 g of oil g⁻¹ of the sample. Considering these values of oil retention, the DSFPE from date seeds could be employed like ingredient to stabilize the products rich in oil. These WHC and OHC were a function of size, shape, hydrophilic and hydrophobic interactions and were affected by the presence of carbohydrates, lipids and amino acid residues on the surface, since most non polar amino acid residues and polar groups are not hydrated in the interior (Moure et al., 2001).

These values of WHC and OHC of DSFPE are superior to those of protein concentrate of some seeds, as observed for Rosa Rubiginosa (Moure et al., 2001), sesame (Khalid et al., 2003) and of defatted Erythrina observations.
variegata flour (Jyothirmayi et al., 2006). The OHC of DSFPE showed a lower oil holding capacity than the protein concentrate of Guevina avellana (Moure et al., 2002). Kinsella (1979) explained the mechanism of fat absorption as a physical entrapment of oil and several authors have related the oil absorption capacity to the non polar side chains of the protein as well as to different conformational features of the proteins. Probably, these results cannot only be due to the higher protein contents but also to the higher soluble fibre contents; they are known by their higher capacities of oil and water retention (Macconnell et al., 1974; Fleury and Lahaye, 1991; Elleuch et al., 2008). High oil absorption of DSFPE is essential in the formulation of food systems like sausages, cake batters, mayonnaise and salad dressings. Also, DSFPE have a good water absorption capacity, it can be used in products requiring high water retention.

Surface tension

The surface properties study of DSFPE proteins is necessary to evaluate their capacity to lower the surface tension and thus their aptitude to be acted like surfactant agent. The protein adsorption kinetics on the surface was studied at pH 7 and 10 by using solutions with 1% of proteins prepared by a suitable dilution of the DSFPE from the two studied varieties (Figures 4 and 5).

Differences in behaviours of proteins at pH 7 and 10 were very visible for DSFPE from Allig variety (Figure 4). It seems that the seed proteins of this variety have more active surface at pH 10 probably due to their high solubility at alkaline solutions. Indeed, the surface tension is definitely more reduced. On the other hand, whatever is the pH was (7 or 10), the adsorption kinetic of DSFPE Deglet Nour protein was not affected which shows interesting and comparable surface properties with those of DSFPE Allig at pH 10 (Figure 5). These results could be explained by the higher solubility of Deglet Nour seed proteins than those from Allig seeds (Figure 3).

The variation effects of protein concentration (0.5 and 1%) at pH 10 and 7 on the surface adsorption kinetics for DSFPE Allig and Deglet Nour are presented in Figures 4 and 5, respectively.

At pH 10, similar results were detected between the two studied DSFPE. Thus, it seems that an increase in the protein concentration from 0.5 to 1% improved the surface tension lowering (t = 63 s, protein concentration = 1%, surface tension ~ 56 to 57 mN/m, and protein concentration = 0.5%, surface tension ~ 60 to 62 mN/m).

At pH 7, significant difference was detected between the two studied DSFPE. For DSFPE-Allig protein, surface tension kinetics was not affected by protein concentration and these have a similar reduction at 1 or 0.5%. This result could be probably due to the low solubility of these proteins (Figure 4).

However, for DSFPE Deglet Nour protein, the increase of protein concentration from 0.5 to 1% reduced the surface tension. For example, at time = 60 s and at 0.5% of DSFPE Deglet Nour protein, surface tension was reduced from 62.10 to 57.40 mN/m at 1% (Figure 5).

From all of these results of surface tension measurements, it is clear that the DSFPE proteins had surface tension effects at pH 7 and 10 which were reduced with the increase in protein concentration especially for DSFPE Deglet Nour. These results can be due to the higher solubility of DSFPE Deglet Nour protein at pH 7 to 10. These finding have certainly an impact on foam and emulsion proprieties of DSFPE. Consequently, DSFPE can be used like surface-active agents in some food applications considering their capacity to reduce the surface tension.

Foaming properties

Figures 6 and 7 present the foam capacity and stability of the produced foams from DSFPE at pH 7 and 10, respectively. Generally, high foam capacity (1% of proteins from DSFPE) was found at pH 10 (13.5 to 14 ml). At this pH, foam capacities of DSFPE from Allig and Deglet Nour seed were comparable (P ≥ 0.05). At pH 7, foam capacities were lower than those obtained at pH 10 (11 to 11.5 ml) (Figure 6). This result was comparable to those obtained by Khalid et al. (2003) which studied the influence of pH on sesame seed proteins. Also, similar result was reported by Chandi and Sogi (2007) that the foam capacity of rice bran protein concentrate improved as the pH was increased from acidic (pH 5) to alkaline (pH 9) pH. The protein solubility increased with alkalinity, and foam capacity also increased, resulting in higher overrun of the solutions. These results, are probably due to the fibre - protein and fibre - fiber interactions at pH 7 and 10 and the hydrophobic - hydrophilic balance.

The foam stability of the foam formed from DSFPE was studied at pH 7 and 10. Whatever the pH was, the DSFPE did not present a remarkable stability. Probably, this result can be due to the presence of a high contents and nature of fibre on DSFPE. It may be possible that nature and contents of fibre have a negative effect on the foam stability of the formed network. In addition, Chandi and Sogi (2007), and Sogi et al. (2002) observed that pH setting had a considerable effect on the volume and

Table 4. WHC and OHC of DSFPE from defatted date seeds.

<table>
<thead>
<tr>
<th>DSFPE</th>
<th>Allig</th>
<th>Deglet Nour</th>
</tr>
</thead>
<tbody>
<tr>
<td>WHC (g H2O/g DSFPE)</td>
<td>4.34± 0.11</td>
<td>3.94± 0.70</td>
</tr>
<tr>
<td>OHC (g oil/g DSFPE)</td>
<td>6.09± 0.30</td>
<td>5.63± 0.90</td>
</tr>
</tbody>
</table>

Values in the same lines with different letters are significantly different (p≤0.05).
stability of foams (Figure 7).

**Emulsifying properties**

Emulsions are formed due to the presence of hydrophobic and hydrophilic groups of proteins. Emulsion capacity is the parameter most commonly estimated in the various studies on oil in water.

The effects of pH on the emulsion capacity of DSFPE were determined at pH 7 and 10 (Table 5). Similar result was observed between DSFPE Allig and DSFPE Deglet Nour (P ≥ 0.05). The emulsion capacity of DSFPE was important (2000 to 3000 ml oil g\(^{-1}\) of protein) and improved with the increase in pH. Emulsion capacity of DSFPE was significantly higher at pH 10 (2800 to 3000 ml oil g\(^{-1}\) of protein) than at pH 7 (2000 to 2400 ml of oil g\(^{-1}\) of protein). Also, it was probably that a relationship between emulsion properties and the nitrogen solubility (Figure 3) of the studied DSFPE existed. This result suggests that the improvement of emulsification capacity could be due to the presence of soluble proteins and fibres. Moure et al. (2001) and Khalid et al. (2003) reported similar relationships between emulsification

![Figure 4. Adsorption kinetics of 0.5 and 1% DSFPE A protein at pH 7 and 10.](image1)

![Figure 5. Adsorption kinetics of 0.5 and 1% DSFPE DN protein at pH 7 and 10 (DSFPE DN : Date Seed Fibro-Protein Extract from Deglet Nour)](image2)
capacity and pH for soybean, groundnut and guar proteins.

Good emulsification stability of DSFPE from date seeds was observed (Figure 8). The kinetics of creaming shows well, on the one hand, that the emulsion oil / protein solution, prepared starting from the DSFPE of Allig seeds at pH 10, was more stable than that produced at pH 7. In addition, the emulsions oil / protein solution, prepared starting from the Deglet-Nour DSFPE, presented kinetics of creaming almost identical and similar at pH 10 and 7. These results show that DSFPE have good potential to act as a suitable emulsifier under various conditions of pH from food systems.

These results are in accordance with those of the surface tension kinetics and nitrogen solubility. This finding is in agreement with the general correlation between surface tension, foam capacity and stability and emulsification capacity and stability and nitrogen solubility found in previous studies (Moure et al., 2001; Khalid et al., 2003). Thus, it seems that solubility strongly improved the properties of surface and especially the emulsion stability of DSFPE from the two studied varieties.

**Conclusion**

Date palm seeds could be an excellent source of functional foods components considering the protein, fat,
Table 5. Emulsion capacities of DSFPE at pH 7 and 10.

<table>
<thead>
<tr>
<th>Emulsion capacities (ml oil/g of protein)</th>
<th>pH 7</th>
<th>pH 10</th>
</tr>
</thead>
<tbody>
<tr>
<td>DSFPE Allig</td>
<td>2000&lt;sup&gt;ab&lt;/sup&gt;±530</td>
<td>2840&lt;sup&gt;ab&lt;/sup&gt;±380</td>
</tr>
<tr>
<td>DSFPE Deglet Nour</td>
<td>2400&lt;sup&gt;ab&lt;/sup&gt;±310</td>
<td>3000&lt;sup&gt;ab&lt;/sup&gt;±240</td>
</tr>
</tbody>
</table>

Values in the same lines and rows with different letters are significantly different (p≤0.05).

Figure 8. Turbiscan creaming kinetic measurements of emulsion prepared with 0.1% DSFPE protein at pH 7 and 10.

mineral and carbohydrate contents. From the data presented in this work, we can conclude that the Deglet Nour and Allig DSFPE obtained after aqueous extraction at pH 10 and precipitation at pH 4.5 showed a good and comparable nutritional and dietetic qualities (all of essential amino acids, higher fibre content) and functional properties (colour, solubility, WHC, WHC, foam and emulsion proprieties). The DSFPE can be used as a natural dye to change the colour of some food formulations. It was found to be highly soluble from neutral to alkaline pH. Therefore, DSFPE gave an interesting foam capacity and excellent emulsion stability. Extraction and incorporation of DSFPE in neutral and alkaline food could give an important value addition to date seeds.

REFERENCES


