

# Isolation, characterization, and phylogenetic analysis of *copia*-like retrotransposons in the Egyptian cotton *Gossypium barbadense* and its progenitors

Abdel Ghany A. Abdel Ghany<sup>1</sup> and Essam A. Zaki<sup>2\*</sup>

<sup>1</sup>Institute of Efficient Productivity, Zagazig University, Zagazig

<sup>2</sup>Genetic Engineering & Biotechnology Research Institute, GEBRI, Research Area, Borg El Arab, Post Code 21934, Alexandria, Egypt.

Accepted 16 May 2003

We have used the polymerase chain reaction to analyze *copia*-like retrotransposons in the Egyptian cotton and its progenitors. All three cotton species studied contain reverse transcriptase fragments from *copia*-like retrotransposons. Sequence analysis of these reverse transcriptase fragments reveals that each is different from the others, with predicted amino acid diversities between 9 and 94%. The detection of stop codons and insertions/deletions in the derived amino acid sequences of the *Gossypium* RT clones, suggests that these clones represent defective retrotransposons. The presence of these sequences in *G. barbadense* progenitors, however, suggests the presence of active retrotransposons capable of producing new functional copies at an appropriate rate to compensate for the mutational loss of old ones. Phylogenetic analysis provided strong bootstrap support for a monophyletic origin of plant *copia*-like retrotransposons, yet showed high diversity within all species. Our results suggest that both vertical transmission of *copia*-like retrotransposons within *G. barbadense* lineages, and horizontal transmission between *G. barbadense* and its progenitors have played major roles in the evolution of *copia*-like retrotransposons in *Gossypium*.

**Key words:** Genome structure, *Gossypium*, repetitive DNA, polyploidy, sequence diversity, retrotransposons.

## INTRODUCTION

*Copia*-like group retrotransposons are one of the best characterized groups of plant retrotransposons (for review see Kumar and Bennetzen, 1999). They have been reported in a wide range of plant taxa, including angiosperms, gymnosperms, ferns, lycopods, and bryophytes (Konieczny et al., 1991; Flavell et al., 1992; Voytas et al., 1992; Friesen et al., 2001). Their ubiquity in the plant kingdom suggests that they are of very ancient origin (Bennetzen, 2000). In addition, their abundance has played a major role in plant genome structure and evolution (Bennetzen, 2002).

*Gossypium* L. (Malvaceae) has become a useful system for studying plant retrotransposons evolution

(Abdel Ghany and Zaki, 2002; Zaki and Abdel Ghany, 2003). The phylogenetic relationships of the approximately 50 diploid and 5 allotetraploid species of *Gossypium* are well characterized (reviewed in Wendel and Cronn, 2002). The five allotetraploid *Gossypium* species (designated AF-genome) diverged from a single recent allopolyploidization event, and their parental diploids (Wendel, 1989). In this regard, *copia*-like retrotransposons were previously identified in *G. hirsutum* (Vanderwiel et al. 1993). In addition, fluorescent *in-situ* hybridization was used to study their chromosomal distributions (Hanson et al., 1999). In the current study, we isolated, cloned, and sequenced part of the reverse transcriptase (RT) domain of *copia*-like retrotransposons in the Egyptian allotetraploid cotton, *G. barbadense* cotton, and its progenitors. Our results revealed that all three cotton species studied here contain RT fragments from *copia*-like retrotransposons, suggesting that *copia*-like retrotransposons is a standard component of the *Gossypium* genome, and supporting the fact that *copia*-like retrotransposons represents a major component of the plant genome.

\*Corresponding author; Current Address: Department of Biological Sciences, 1392 Lilly Hall of Life Sciences, West Lafayette, IN 47907-1392, Phone (765) 494-9837 Fax (765) 496-1496, E-mail: ezaki@purdue.edu.

**Abbreviations;** PCR: polymerase chain reaction, RT: reverse transcriptase gene.

## MATERIALS AND METHODS

### Plant materials and genomic DNA extraction

*Gossypium* species and cultivars, listed in Table 1, were kindly provided by Dr. Percival. Total DNA was extracted using Qiagen DNeasy kit (Qiagen, Germany).

**Table 1.** *Gossypium* species used in this study: isolated clones and their GenBank accession numbers.

Species	Clone	Accession number
<i>G. arboretum</i>	Arb	U75244
<i>G. barbadense</i>	Bah163	U75221
	Bah185	U75222
	Ashoumni	U75246
<i>G. darwinii</i>	Dar	U75245

### PCR

Total DNA was subject to PCR with specific primers to amplify an approximately 280 bp region of the *copia*-like reverse transcriptase (5'-GGAATTCGAYGTNAARACNGCNTTYT-3') and (5'-GGGATC CAYRTRCTCNACRTANARNA'), where N= A+C+G+T, R= A+G, and Y= T+C (Voytas et al. 1992). DNA amplifications were carried in an ABI GeneAmp PCR system 9700 cyler with a denaturing step at 95°C for 5 min and the step cycle program set for 45 cycles (with a cycle consisting of denaturing 94°C for 30s, annealing at 47°C for 1 min and extension step at 72°C for 2 min), followed by a final extension step at 72°C for 10 min.

### Cloning and sequencing of PCR-amplified fragments

Expected PCR-amplified fragments were excised from the agarose gel and purified using Qiagen Gel Extraction kit (Qiagen, Germany). Purified DNA fragments were then cloned in pCR 4-TOPO vector with TOPO TA cloning kit (Invitrogen, USA) in the competent *E. coli* strain TOPO 10. Plasmid DNA was isolated using QIA Spin mini-prep kit (Qiagen, Germany), and sequenced in both directions using BigDye Sequencing Kit and ABI 377 DNA sequencer (ABI, USA).

### Alignments and phylogenetic analysis

Pairwise and multiple DNA sequence alignment were carried out using CLUSTALW (1.82) (<http://www2.ebi.ac.uk/clustalw>; Thompson et al. 1994). Bootstrap neighbor-joining tree was generated using MEGA 2.1 (Kumar et al., 2001) from CLUSTALW alignments.

### Accession numbers

DNA sequences, reported in the current study, were deposited in the NCBI nucleotide sequence database, GenBank, and are listed in Table 1.

## RESULTS AND DISCUSSION

PCR amplification with degenerate primers for the *copia*-like reverse transcriptase (RT) domain (Voytas et al., 1992) produced 5 putative RT clones: 3 from *G. barbadense*, and 1 clone from *G. arboretum* and *G. darwinii* respectively (Table 1). Blast search confirmed the RT nature of the cloned products. The high amino acid similarities, observed in the Blast search, supports the interpretation that the 5 sequences generated in this study represent portions of *copia*-like retrotransposons RT genes.

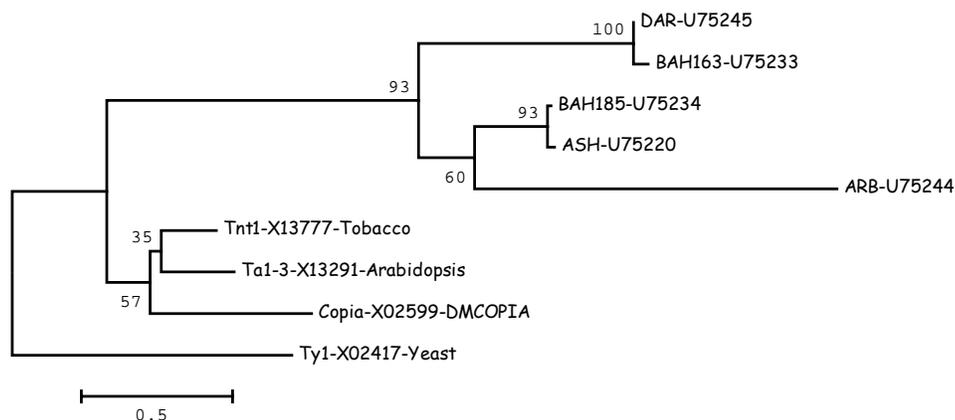
Extreme sequence diversity amongst RT genes of the *copia*-like retrotransposons has been observed both within and between plant species (Flavell et al., 1992). Furthermore, they are more closely related to elements present in other plant species (Voytas et al., 1992). A similar pattern of sequence heterogeneity is observed in *G. barbadense* and its progenitors. Comparative nucleotides and amino acids sequences using CLUSTALW were performed (Figure 1). In addition, pairwise comparisons (Table 2) showed amino acid diversities of 28% (Bah163/Bah185), and 26% (Bah163/Ash) within *G. barbadense* cultivars, and 9% (Arb/Dar), and 10% (Arb/Bah163) between *Gossypium* species.

**Table 2.** Amino acids pairwise comparisons of *copia*-like putative RT sequences in *G. barbadense* and its progenitors.

Clone	Arb	Bah163	Bah185	Ash
Bah163	10			
Bah185	26	28		
Ash	26	26	94	
Dar	9	93	33	33

The detection of either stop codons and insertions/deletions that have caused frame shifts in the derived amino acid sequences of the *Gossypium* RT clones, suggests that these clones represent defective retrotransposons. In addition, a number of the retrotransposons in the Egyptian cotton and its progenitors obviously are not functional and currently must be evolving as pseudogenes. The long-term survival of these sequences, however, suggests the presence of active retrotransposons capable of producing new functional copies at an appropriate rate to compensate for the mutational loss of old ones. It is noteworthy that the majority of plant *copia*-like retrotransposons are thought to be rarely active (Kumar and Bennetzen, 1999). Relationships among the derived amino acid sequences of the 5 clones with each other and other





**Figure 2.** Phylogenetic tree showing relationship between reverse transcriptase amino acid sequences of *G. Barbadense* and its progenitors and plant, yeast, and *Drosophila copia*-like retrotransposons. The Neighbor-Joining method (Saitou and Nei, 1987) was used to construct the tree. The numbers on the branches represent bootstrap support for 1,000 replicates. Names refer to the accession number of the nucleotide sequences that encode the corresponding reverse transcriptase genes.

required to further clarify their evolutionary relationships. In conclusion, we suggest that both vertical transmission of *copia*-like retrotransposons within *G. barbadense* lineages, and horizontal transmission between *G. barbadense* and its progenitors have played major roles in the evolution of *copia*-like retrotransposons in *Gossypium*.

## ACKNOWLEDGEMENT

This work was supported by a grant from the US-Egypt Science and Technology Foundation to E.A. Zaki.

## REFERENCES

- Abdel Ghany AA, Zaki EA (2002). Cloning and sequencing of an *envelope*-like gene in *Gossypium*. *Planta* 216:351-353.
- Bennetzen JL (2002). Mechanisms and rates of genome expansion and contraction in flowering plants. *Genetica*. 115:29-36.
- Bennetzen JL (2000). Transposable elements contributions to plant gene and genome evolution. *Plant Mol. Biol.* 42:347-353.
- Eickbush TH, Furano AV (2002). Fruit flies and humans respond differently to retrotransposons. *Curr. Opin. Genet. Dev.* 12:669-674.
- Flavell AJ, Dunbar R, Anderson R, Pearce SR, Hartley R, Kumar A (1992). Ty1-*copia* group retrotransposons are ubiquitous and heterogeneous in higher plants. *Nucleic Acids Res.* 20:3639-3644.
- Friesen N, Brandes A, Heslop-Harrison J (2001). Diversity, origin and distribution of retrotransposons in conifers. *Mol. Biol. Evol.* 18:1176-1188.
- Hanson RE, Islam-Faridi N, Crane CF, Zwick MS, Czeschin DG, Wendel JF, McKnight TD, Price HJ, Stelly DM (1999). Ty1-*copia*-retrotransposons behaviour in a polyploid cotton. *Chromosome Res.* 8:73-76.
- Konieczny A, Voytas DF, Cummings MP, Ausubel FM (1991). A superfamily of Arabidopsis thaliana retrotransposons. *Genetics*. 127:801-809.
- Kumar S, Tamura K, Jakobsen IB, Nei M (2001). MEGA2: molecular evolutionary genetics analysis software. *Bioinformatics* 17:1244-1245.
- Kumar A, Bennetzen JL (1999). Plant retrotransposons. *Annu. Rev. Genet.* 33:479-532.
- Peterson-Burch BD, Voytas DF (2002). Genes of the Pseudoviridae (Ty1/copia Retrotransposons). *Mol. Biol. Evol.* 19:1832-1845.
- Saitou N, Nei M (1987) The neighbour-joining method: a new method for reconstructing phylogenetic trees. *Mol. Biol. Evol.* 4:406-425.
- Stuart-Rogers C, Flavell AJ (2001). The evolution of Ty1-*copia* group retrotransposons in gymnosperms. *Mol. Biol. Evol.* 18:155-163.
- Thompson JD, Higgins DG, Gibson TJ (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Res.* 22:4673-6480.
- VanderWiel PS, Voytas DF, Wendel JF (1993). *Copia*-like retrotransposable element evolution in diploid and polyploid cotton (*Gossypium* L.). *J. Mol. Evol.* 36:429-447.
- Voytas DF, Cummings MP, Konieczny A, Ausubel FM, Rodermel S (1992). *Copia*-like retrotransposons are ubiquitous among plants. *Proc. Natl. Acad. Sci. USA.* 89:7124-7128.
- Wendel JF, Cronn R (2002). Polyploidy and the evolutionary history of cotton. *Advances in Agronomy* 78:139-186.
- Wendel JF (1989). New world cottons contain old world cytoplasm. *Proc. Natl. Acad. Sci. USA.* 86:4132-4136.
- Zaki EA, Abdel Ghany AA (2003). Molecular distribution of *gypsy*-like retrotransposons in cotton *Gossypium* Spp. *Afr. J. Biotechnol.* 2:124-128.