Review

Regulation function of nitric oxide (NO) in leaves of plant under environmental stress

Yue-fei Xu¹, Jing-wei Jin², Tie-yuan Liu¹, He Zhou³, Tian-ming Hu¹, Quan-zhen Wang¹ and Ming-xiu Long¹*

¹College of Animal Science and Technology, Northwest A&F University, Yangling, Shaanxi Province 712100, China. ²Institute of Soil and Water Conservation, Chinese Academy of Sciences and Ministry of Water Resources, Yangling, Shaanxi 712100, China.

³College of Animal Science and Technology, China Agricultural University, Beijing 100193, China.

Accepted 29 July, 2011

Nitric oxide is an important signaling molecule involved in many physiological processes. This paper expounds the source and biological features of nitric oxide as well as the cognizance of the role of nitric oxide in biotic and abiotic stresses. Finally, the prospect of development in this field is presented.

Key words: Abiotic stress, biotic stress, nitric oxide, oxidative damage.

INTRODUCTION

Nitric oxide (NO), a fat- and water-soluble small molecule of gas is considered to be a key signaling molecule commonly existing in plant leaves in recent years. In plant, NO is mainly formed by four ways: NO synthase pathway which is similar to that of animal (Delledonne et al., 1998; Durner et al., 1998; Neill et al., 2003), nitrate reductase pathway (Morot-Gaudry-Talarmain et al., 2002), other enzymatic pathways and non-enzymatic pathways (Cooney et al., 1994).

PHYSICO-CHEMICAL PROPERTIES OF NO

NO is more extensive on the growth and development of plant. Early studies have shown that NO may regulate the growth of plant leaves (Leshem et al., 1998), affect the growth of plant roots, be involved in plant photomorphogenesis, stimulate seed germination and play the role of de-etiolation (Beligni and Lamattin, 2000). Treatment with exogenous nitric oxide in horticultural plants can increase NO content in tissues and inhibit ethylene formation, thus, delaying the fruit maturation and senescence (Leshem and Haramaty, 1996). The influence of NO depends on the level of concentration, a low concentration level of NO directly act on cell wall components to make cell wall relax and promote cell expansion, thus promoting the growth of plants and help plants resist stress and delay aging. A high concentration level of NO and superoxide iron reaction forming a toxic peroxynitrite compound which causes membrane lipid over oxidation, lead to membrane leakage and may even spreads into the cytoplasm to attack key enzymes, thereby causing destructive impact (Pyor, 1994).

Iron is a necessary element for synthesis and development of chloroplast. Iron need to go through several layers to reach the chloroplast, and the operating mechanism is unclear at present. NO plays an important role in the distribution of iron in the chloroplast in plant leaves (Sun et al., 2007). Thylakoids and starch grains decreased obviously and exist in iron-deficient corn without NO treatment. Vascular bundle of Zea mays chloroplast, had no obvious difference between irondeficient with NO treatment and without NO treatment regards to morphological characteristics of with chloroplasts, thylakoids and starch grains. There was a complex interaction existing between NO and hormone. NO and peroxynitrite can inhibit the biosynthesis of ethylene by regulating the 1-aminocyclopropane-1-carboxylic acid synthase activity regulators and 1aminocyclopropane-1-carboxylic acid synthase cofactor, so that they can regulate the maturation and senescence of plant tissues. There are more reports about the function of NO and ABA in regulating stoma. Some

^{*}Corresponding author. E-mail: longmingxiu@nwsuaf.edu.cn. Tel/Fax: +86-29-87091953.

studies reported that there is a positive cooperativity in the regulation of stomatal movement. In Arabidopsis thaliana and Pisum sativum, NO may participate in the ABA-induced stomatal closure, which may be the transduction signal of stomatal movement under the ABA regulation. NO and NOS are involved in the synthesis of ABA when the Triticum aestivum root tip is under drought stress. In corn's response to drought stress, NADPH was increased because of ABA-induction, which make the level of oxygen radical elevated, and the oxygen radical promotes the production of NO, proving that the relationship of ABA and superoxide radical and NO are close (Jiang and Zhang, 2002). Cytokinin can induce NO production (Scherer and Holk, 2002), and NO may mediate the occurrence of adventitious roots of ABAinduced cucumber (Samuel et al., 2000). NO can also induce acid synthesis, and salicylic acid can activate NO synthesis pathway of plants (Guo et al., 2003). NO can not only participate in joint action of plant hormones regulating plant growth, but can also be used as the signal substance to induce hormones synthesis. NO is also induced by hormones.

NO INVOLVED IN ENVIRONMENTAL STRESS

NO involved in abiotic stress

Abiotic stress and biotic stress can induce the production of NO, exogenous and endogenous NO could increase plant stress, but high concentration of NO is toxic to the cells (Durzan and Pedroso, 2002). Abiotic stresses such as drought, salt, heavy metals, extreme temperatures, UV radiation and so on could induce a large number of active oxygen (Uchida et al., 2002; An et al., 2005; Laspina et al., 2005; Song et al., 2006; Chen et al., 2006: Shi et al., 2007; Vital et al., 2008; Zhang et al., 2009; Xu et al., 2010), which can cause oxidative damage and trigger different signaling pathways. NO and active oxygen interaction by different ways, may play an important role as an antioxidant in different stresses. NO can regulate the formation of superoxide and inhibit lipid peroxidation to protect the plant from environmental stress damage, which can explain the potential antioxidation (Neill et al., 2003). In addition, excessive NO may cause nitrosation stress to harm plant, therefore, the balance between NO and active oxygen is very important.

Å study showed that the moisture holding capacity of excised wheat leaves with 150 μ M sodium nitroprusside (a NO donor) pretreatment is 15% higher than with water or NO²⁻/NO³⁻ pretreatment after 3 h (Mata and Lamattina, 2001). It can be observed from wheat seedlings after 7 days drought stress that SNP treatment can help it increase water-retention capacity. The result was consistent with that leaf transpiration rate of detached wheat decreased by 20% which is by SNP treatment. The

studies of Song et al. (2008) have shown that NO may act as signal molecules to induce the antioxidative enzyme activity of *Phragmites communis*, thus inducing the accumulation of active oxygen and the level of membrane lipid peroxidation under the high temperature stress. Leshem, (2001) found that short-term heat stress can increase nitric oxide level in alfalfa. Neill et al. (2003) reported that the role of NO can reflect the antioxidant activity that can reduce the accumulation of cold and the levels of reactive oxygen in heat stress.

Zhao et al. (2001) found that the semiochemical of NO is also involved in plant material salt reaction. NO increased obviously and induced the H⁺-ATPase gene expression of plasmalemma and increase its vitality, finally, ions within the cell achieve re-balanced, thus maintain a relatively high K⁺/Na⁺ ratio and induce the salt tolerance of plant cells. In subsequent experiments, Zhao et al. (2004) used the method of T-DNA insertion in the Arabidopsis AtNOSI genes and found that the NOS activity decreased in a vast scale in mutant, the release of NO reduced, and at the meantime, mutant to salt stress is more sensitive than the wild type, this shows the effect of NO in the regulation of plant response to salt stress.

Under low temperature stress, NO can enhance resistance of plants to stress (Lamattina and Beligni, 2001). The experiments show that low concentrations of NO can reduce chlorophyll breakdown, ion leakage, leaf necrosis and loss and other symptoms in *Solanum tuberosum* leaf induced by herbicide, lipid peroxidation, ribulose 1,5-bisphosphate carboxylase/oxygenase and DI protein degradation, and the rupture of mRNA caused by herbicide can be reduced obviously by NO. NO can also make ETR of isolated chloroplast which are raised by herbicide to returned to normal (Lamattina and Beligni, 2001). All of these suggested that the protective effect of NO on plants might be able to reduce the levels of reactive oxygen in plant tissues.

Using an ozone-sensitive *Arabidopsis* ecotype as materials, research suggests that ozone may promote NOS activity, this effect occurred before the salicylic acid accumulation and cell death (Zhang et al., 2009). Accordingly, they believed that the production of NO increased the sensitivity of *Arabidopsis* ecotype, and may act as a signal molecule induced hypersensitive cell death. In *Nicotiana tabacum*, NO can induce the synthesis of salicylic acid (Durner et al., 1998; Shi et al., 2007; Zheng et al., 2009), it is possible that NO can increase the accumulation of salicylic acid under ozone stress.

In recent years, some studies reported that NO was involved in plant response by heavy metals stress (Kopyra and Gwozdz, 2003; Jin et al., 2010). They used lupine bean as the test material to see if exogenous NO can increase superoxide dismutase (SOD), catalase (CAT) and peroxidase (POD) and other active oxygen metabolism enzyme activity, thus reduce the heavy metals on lupine seed germination and seedling growth inhibition. Exogenous NO can reduce oxidative stress obviously induced by cadmium, reduce Cd stress on sunflower growth inhibition and chlorophyll degradation, enhance CAT activity and increase glutathione and ascorbic acid content in cell (Laspina et al., 2005). This showed that exogenous NO can increase antioxidant level in plants, thereby improving the resistance to cadmium stress.

NO in biotic stress

The NO level increases more rapidly in the process of plants and pathogens reaction. NO is closely related to disease resistance plant of allergic reactions (hypersensitive response, HR) programmed cell death (PCD) and systemic acquired resistance (SAR). With a laser scanning confocal microscope, Foissner et al. (2000) proved that the combination of tobacco epidermal cells and pathogens will make NO product, scavenger of NO and NO synthase inhibitors prevent this outbreak. It was found that nitric oxide synthase activity increases rapidly in soybean suspension cells induced by directional cell bacteria, Pseudomonas elicitor. In the action between tobacco and pseudomonas, the nitric oxide donor can cause allergic reaction (Huang and Knopp, 1997).

HR is triggered by rapid and transient ROS large formation. In plants, nitric oxide through two ways: directly or indirectly affect plant disease resistance. On one hand, NO mainly prevent the spread of pathogens from the injection site to participate in allergic mediated disease resistance of plants (Hammond-Kosack and Jones, 1996; Manjunatha et al., 2009). One of the plant cell response to pathogens invasion is oxygen burst, which produce large amount of reactive oxygen, with the occurrence of oxygen burst, and NO is also rapidly generated, which directly kill bacteria. On the other hand, indirect effects of NO can also induce the HR of plant: this is believed to be the main way of signal transduction to induce resistance gene expression. Oxygen burst can release H_2O_2 and O^2 , which can react with NO to produce a more active lethal pathogen, O2- and ONOOwhich can react against pathogenic invasion. At the same time, NO can stimulate the infected cell walls lignification (Ferrer and Barcelo, 1999; Palmieri et al., 2010), involved in the regulation of cell death and defense mechanisms induced during pathogen defense (Morot-Gaudry-Talarmain et al., 2002).

NO can obviously reduce cell death caused by ion leakage and potato leaf infected by pathogen. This process is controlled by cPTIO (carboxy-2-pheny 1-4,4,5,5-tetrmethyliidazoline-1-oxy13-oxide, cPTIO). As an antioxidant, NO can offset the bane action mediated by the reactive oxygen (Beligni and Lamattina, 1999; Palmieri et al., 2010).

PROSPECT OF RESEARCH ON NO

Animal cells synthesize NO primarily by the activity of NOS. However, at present, the mechanism of NO production in plant organs is still a controversial problem (Crawford et al., 2006; Moreau et al., 2008). Previous studies have shown that NO source in plants includes enzymatic sources like nitric oxide synthase-like enzymes and nitrate reductase (NR), and non-enzymatic sources (Neill et al., 2003). NR was shown to be involved in NO production in several physiological situations, such as ABA-induced stomatal closure (Bright et al., 2006) and auxin-induced lateral root development (Kolbert et al., 2008). However, most researches still supported that NO generation was mainly mediated by a putative NOS-like enzyme which catalyses the formation of NO from Larginine, rather than NR (Tian et al., 2007; Corpas et al., 2008). Furthermore, several studies have identified NOSdependent NO generation during plants exposure to stress conditions (Corpas et al., 2008; Chaki et al., 2009).

The present of NO in plant resistance is mainly through exogenous donor of NO, NO synthase inhibitor, NO removal agent under environmental stress effects on plant physiological indicators, analysis of NO in the application of stress response and gradually to gene transcription, enzyme protein and other molecular mechanism of action. Under almost all environmental stress factors, NO can play its physiological role. However, exogenous nitric oxide through which function intermediary or signal transduction components affect the response of plants to stress factors, by plant hormones, reactive oxygen metabolism, stress proteins or genes associated with stress and functional protein need to be deeply explored further with extensive research in depth. For example, In the high temperature stress, Ca²⁺, abscisic acid, ethylene, salicylic acid and stress-related genes, antioxidant system and HSPS and so on are all related to heat tolerance of plant, but in the process of stress signal transmitting from outside to inside, the order of exogenous or endogenous NO and these function intermediary and whether there is chain among them are worthy to be clarified. In addition, according to Pastori and Foyer (2002), plants adapt to biotic and abiotic stress by starting a series of physiological events, these physiological events begin with the perception of stress stimuli, and end at the expression of target genes. The signaling components mainly contain signaling molecules and their transformation, transcription regulation elements, target genes, and finally the stress response, including plant morphology and physiological and biochemical changes. From an evolutionary point of view, the function of components close to the end of stress response is single, and components near the beginning of stress response is likely to be all the stress response mutual pathway or factor, this "cross resistance" can make plants adapt to a number of other stresses when experiencing a particular stress. Since NO play a role in so many plant stress responses, it is suggested that NO

in the stress signal chain or the signal net should be close to the beginning of transduction. But how exactly, stress stimulation is transferred to NO and how to pass on ulteriorly are not clear. NO itself is likely to be the main component of plant "cross resistance ", which is worthy of further study.

ACKNOWLEDGEMENTS

This work was supported by Key Projects in the National Science and Technology Pillar Program in the Twelfth Five-year Plan Period (No. 2011BAD17B05); Special Research Funds for Public Service of Ministry of Agriculture, China (No. 200903060); Northwest A&F University, Scientific Research Foundation, Dr. (No. 2010BSJJ007); Tang Zhongying Breeding Fund of Northwest A&F University (No. A212020901) and Agricultural research program in Shaanxi Province (No. 2011K01-06).

REFERENCES

- An LZ, Liu YH, Zhang MX, Chen T, Wang XL. Effects of nitric oxide on growth of maize seedling leaves in the presence or absence of ultraviolet-Bradiation. J. Plant Physiol. 162: 317-326.
- Beligni MV, Lamattin AL (2000). Nitric oxide stimulates seed germination and deetiolation, and inhibits hypocotyis elongation, three lightinducible response in pants. Planta, 210: 215-221.
- Beligni MV, Lamattina L (1999). Nitric oxide counteracts cytotoxic processes mediated by reactive oxygen species in plant tissues. Planta, 208: 337-344.
- Bright J, Desikan R, Hancock JT, Weir IS, Neill SJ (2006). ABA-induced NO generation and stomatal closure in Arabidopsis are dependent on H_2O_2 synthesis. Plant J. 45: 113–122.
- Chaki M, Fernández-Ocaña AM, Valderrama R, Carreras A, Esteban FJ, Luque F, Gómez-Rodríguez MV, Begara-Morales JC, Corpas FJ, Barroso JB (2009). Involvement of reactive nitrogen and oxygen species (RNS and ROS) in sunflower-mildew interaction. Plant Cell Physiol. 50: 265–279.
- Chen Y, Zhang M, Chen T, Zhang Y, An L (2006). The relationship between seasonal changes in anti-oxidative system and freezing tolerance in the leaves of evergreen woody plants of Sabina. S. Afr. J. Bot. 72: 272-279.
- Cooney RV, Harwood PJ, Custer LJ, Franke AA (1994). Light mediated conversion of nitrogen dioxide to nitric oxide by carotenoids. Environ. Health Persp. 102: 460-462.
- Corpas FJ, Chaki M, Fernández-Ocaña A, Valderrama R, Palma JM, Carreras A, Begara-Morales JC, Airaki M, del Río LA, Barroso JB (2008). Metabolism of reactive nitrogen species in pea plants under abiotic stress conditions. Plant Cell Physiol. 49: 1711–1722.
- Crawford NM, Galli M, Tischner R, Heimer YM, Okamoto M, Mack A (2006). Response to Zemojtel *et al*: plant nitric oxide synthase: back to square one. Trends Plant Sci. 11: 526–5277.
- Durner J, Wendehenne D, Klessig DF (1998). Defense gene induction in tobacco by nitric oxide. cyclic GMP, and cyclic ADP-ribose. Proc. Natl. Acad. Sci. USA. 95: 10328-10333.
- Durzan DJ, Pedroso MC (2002). Nitric oxide and reactive nitrogen oxide species in plants. Biotechnol. Genet. Eng. Rev. 19: 293-337.
- Ferrer MA, Barcelo AR (1999). Differential effects of nitric oxide on peroxidase and H₂O₂ production by the xylem of Zinnia elegans. Plant Cell Environ. 22: 891-897.
- Foissner I, Wendehenne, Langebartels C, Durner J (2000). *In vivo* imaging of an elicitor induced nitric oxide burst in tobacco. Plant J. 23: 817-824.

- Guo FQ, Okamoto M, Crawford NM (2003). Indentification of a plant nitric oxide synthase gene involved in hormonal signaling. Science, 302: 100-103.
- Hammond-Kosack KE, Jones JD (1996). Resistande Gene-dependent Plant Defense Responses. Plant Cell, 8: 1773-1791.
- Jiang M, Zhang J (2002). Abscisic acid and antioxidant degense in plant cells. Acta Bot. Sin. 215: 1022-1030
- Jin JW, Xu YF, Huang YF (2010). Protective effect of nitric oxide against arsenic-induced oxidative damage in tall fescue leaves. Afr. J. Biotechnol. 9(11): 1619-1627.
- Kolbert Z, Bartha B, Erdei L (2008). Exogenous auxin-induced NO synthesis is nitrate reductase-associated in *Arabidopsis thaliana* root primordial. J. Plant Physiol. 165: 967–975.
- Kopyra M, Gwozdz EA (2003). Nitric oxide stimulats seed germination and counteracts the inhibitory effect of heavy metal and salinity on root growth of Lupinus luteus. Plant Physiol. Biochem. 41: 1011-1017.
- Lamattina L (1999). Nitric oxide protects against cellular damage produced by methylviologen herbicides in potato plant. Nitric Oxide: Biol. Chem. 3: 199-208.
- Laspina NV, Groppa MD, Tomaro ML, Benavides MP (2005). Nitric oxide protects sunflower leaves against Cd-induced oxidative stress. Plant Sci. 169: 323-330.
- Laspina NV, Groppa MD, Tomaro ML, Benavides MP (2005). Nitric oxide protects sunflower leaves against Cd-induced oxidative stress. Plant Sci. 169(2): 323-330.
- Leshem Y (2001). Nitric oxide in plants. London, UK: Kluwer Academic Publishers, pp. 1-5.
- Leshem YY, Haramaty E (1996). The characterization and contrasting effects of the nitric oxide free radical in vegetative stress and sentence of *Pisum sativum* Linn. Foliage. J. Plant Physiol. 148: 258-263
- Leshem YY, Wilisr BH, Ku VV (1998). Evidence for the function of the free radical gas nitric oxide as an endogenous matu-ration and senescence regulating factor in higher plant. Plant Physiol. Biochem. 36: 825-833.
- Manjunatha G, Deepak S, Geetha PN, Niranjan-Raj S, Kini RKShetty HS (2009). Hypersensitive reaction and P/HRGP accumulation is modulated by nitric oxide through hydrogen peroxide in pearl millet during *Sclerospora graminicola* infection. Physiol. Mol. Plant. 74: 191-198.
- Mata CG, Lamattina L (2001). Nitric oxide induces stomatal closure and enhances the adaptive plant responses against drought stress. Plant Physiol. 126: 1196-1204.
- Moreau M, Lee GI, Wang Y, Crane BR, Klessig DF (2008). AtNOS/A1 is a functional *Arabidopsis thaliana* cGTPase and not a nitric oxide synthase. J. Biol. Chem. 283: 32957–32967.
- Morot-Gaudry-Talarmain Y, Rockel P, Moureaux T, Quilleré I, Leydecker M, Kaiser W, Morot-Gaudry J (2002). Nitrite accumulation and nitric oxide emission in relation to cellular signaling in nitrite reductase antisense tobacco. Planta, 215(5): 708-715.
- Morot-Gaudry-Talarmain Y, Rockel P, Moureaux T, Quilleré I, Leydecker MT, Kaiser WM, Morot-Gaudry JF (2002). Nitrite accumulation and nitric oxide emission in relation to cellular signaling in nitritereductas eantisense tobacco. Planta, 215(5): 708-715.
- Neill SJ, Desikan R, Clarke A. Hancock JT (2002). Nitric oxide is a novel component of abscisic acid signaling in stomatal guard cells. Plant Physiol. 128: 13-16.
- Neill SJ, Desikan R, Hancock JT (2003). Nitric oxide signalling in plants. New Phytol. 159(1): 11-35.
- Palmieri MC, Lindermayr C, Bauwe H, Steinhauser C, Durner J (2010). Regulation of plant glycine decarboxylase by S-nitrosylation and glutathionylation. Plant Physiol. 152: 1514-1528.
- Pastori GM, Foyer CH (2002). Common components Networks, and pathways of cross-tolerance to stress, The central role of "redox" and abscisic acid-mediated controls. Plant Physiol. 129: 460-468.
- Pyor WA (1994). Nitric oxide and peroxynitrite. Soc. Free. Rad. Res. Newslet. 4: p. 12.
- Samuel MA, Miles GP, Ellis BE (2000). Ozone treatment rapidly activates MAPK signaling in plants. Plant J. 22: 367-376
- Scherer GF, Holk A (2000). NO donors mimic and NO inhibitor cytolinin action in betalain accumulation in *Amoranthus candatus*. Plant

Growth Regul. 32: 345-350.

- Shi QH, Ding F, Wang XF, Wei M (2007). Exogenous nitric oxide protect cucumber roots against oxidative stress induced by salt stress. Plant Physiol. Bioch. 45: 542-550.
- Song LL, Ding W, Shen J, Zhang ZG, Bi YR, Zhang LX (2008) Nitric oxide mediates abscisic acid induced thermotolerance in the calluses from two ecotypes of reed under heat stress. Plant Sci. 175: 826-832.
- Song LL, Wei D, Zhao MG, Sun BT, Zhang LX (2006). Nitric oxide protects against oxidative stress under heat stress in the calluses from two ecotypes of reed. Plant Sci. 171: 449-458.
- Sun BT, Jing Y, Chen KM, Song LL, Chen FJ, Zhang LX (2007).Protective effect of nitric oxide on iron deficiency-induced oxidative stress in maize (*Zea mays*). J. Plant Physiol. 164: 536-543.
- Tian QY, Sun DH, Zhao MG, Zhang WH (2007). Inhibition of nitric oxide synthase (NOS) underlies aluminum-induced inhibition of root elongation in *Hibiscus moscheutos*. New Phytol. 174: 322–331.
- Uchida A, Jagendorf AT, Hibino T, Takabe T (2002). Effects of hydrogen peroxide and nitric oxide on both salt and heat stress tolerance in rice. Plant Sci. 163: 515-523.
- Vital SA, Fowler RW, Virgen A, Gossett DR, Banks SW, Rodriguez, J (2008). Opposing roles for superoxide and nitric oxide in the NaCl stress-induced upregulation of antioxidant enzyme activity in cotton callus tissue. Environ. Exp. Bot. 62: 60-68.

- Xu YF, Sun XL, Jin JW, Zhou H (2010). Protective effect of nitric oxide on light-induced oxidative damage in leaves of tall fescue. J. Plant Physiol. 167: 512-518.
- Zhang LG, Zhou S, Xuan Y (2009). Protective effect of nitric oxide against oxidative damage in *Arabidopsis* leaves under ultraviolet-B irradiation. J. Plant Biol. 52: 135-140.
- Zhao LQ, Zhang F, Guo JK Yang LL, Li BB, Zhang LX (2004). Nitric oxide as a signal in salt resistance in the calluses from two ecotypes of reed. Plant Physiol. 134: 849-857.
- Zhao MG, Tian QY, Zhang WH (2007). Nitric oxide synthase-dependent nitric oxide production is associated with salt tolerance in *Arabidopsis*. Plant Physiol. 144: 206-217.
- Zhao Z, Chen G, Zhang C (2001). Interaction between reactive oxygen species and nitric oxide in drought induced abscisic acid synthesis in root tips of wheat seedlings. Aust. J. Plant Physiol. 28: 1055-1061.
- Zheng CF, Jiang D, Liu FL, Liu WC, Jing Q, Cao WX (2009). Exogenous nitric oxide improves seed germination in wheat against mitochondrial oxidative damage induced by high salinity. Environ. Exp. Bot. 67: 222-227.