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The impacts of the transgenes on the modified crops, non-target soil and terrestrial organisms

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Agriculture forms a basis for survival and advancement of civilization of mankind and economic growth. The production and quality of food depends on nature of cultivars that have hitherto been obtained by mutation and hybridization of selected plants. Since the productivity of many crops has reached a plateau the gene transfer technology is used to break the yield barriers to meet the demand of the growing population of mankind. The gene transfer although yielded intended effects including reduction of emission of greenhouse gases, caused several changes in the modified crop plants, interacted with soil food webs, affected the biota associated with agro system, inflicted damage to mammals and birds, and contaminated both the related and unrelated plant species through horizontal gene transfer. The gene transfer also caused the phenomena of resistance and ecological succession of biota in different habitats

Key words: Gene transfer, transgene, agriculture, modified crops.

INTRODUCTION

Agriculture has been playing a key role in the development of human civilization from Pleistocene era (Peter et al., 2001). The biota that were present at the end of this era were similar to modern ones, brought in selective domestication and cultivation of plants and ensured the food and nutrient security to human population. As this selective method failed to meet the demand of growing population of mankind, cross pollination was resorted to by following the principles of plant genetics established by Gregor Mendel (Druery and Bateson, 1901) in the 19th century and inducted the desired traits into plants. However, the available genetic variability in the gene pool of wild species and the other cultivars had not met the goal for increasing the productivity of the crops. The mutation breeding with the chemicals or radiation has been employed for infusing the desired traits in the crop plants (Muller, 1930). Since these methods had not supported the human population the sexual hybridization was resorted to get the desired traits from different plants into one plant through cross-pollination and improved the productivity. The breeding techniques have reached its peak with the introduction of the dwarf gene into wheat (Borlaug, 1968) and rice (Qin et al., 2008). The dwarf genes and hybrids besides making them highly fertilizers and pesticides responsive, removed poly crop system and laid the foundation for green revolution to prevent the starvation death of billions of people around the world. The uses of monoculture, pesticides and fertilizers have resulted in the degradation of environment, fertility of soils, nutrient status of the crops and health of living beings (Byrnee, 1990; Menzie, 1972; Mozafa, 1993; Sarkar, 1990).

The discovery of DNA's double helix structure by Crick and Watson (1953) held the key to cracking the genetic code which determined how all living things work and this has formed the basis for the modern plant breeding techniques. The modern plant breeding is targeted to identify gene fragment of DNA for a particular trait in any species of plant, animal or microorganism, then isolated, copied, and introduced into an existing crop variety with the aid of Agrobacterium (Joos et al., 1983) or gene gun (Gan et al., 2000) or electroporation (Joshi and Schoenbach, 2000). This modern gene splicing method provided a more precise approach to crop improvement than sexual hybridization and was first developed in the 1983 (Estrella et al., 1983). A large number of crop species were genetically modified with the addition of any one of the traits for high productivity, nutrients, (Babili

et al., 2002), abiotic and biotic stress resistance (Vaeck et al., 1987), to reduce the impacts of global warming by capturing the carbon from the atmosphere(Jansson et al.2010) pharmacological products for molecular farming (Chong and Langridge, 2000), for removal of pollutants from the environment by phytoremediation (Ma, 2010, Pascal et al., 2010,) and also to enhance the production of biofuels in micro algae (Beer et al., 2009) Many millions of hectares of commercially produced transgenic crops have been grown annually in a number of countries and has the greatest impact on global agriculture by way of increasing the farm economy to the extent of \$27 billion, reducing the use of pesticides by 172 million kg, reducing the release of greenhouse gas emissions from agriculture (Brookes and Barfoot, 2000) and water inputs on pesticide application (FAO, 2004) But the genetically manipulated plants do have some unintended effects as other methods (Davis et al., 2004) on the environment, non-target organ-isms, invite the phenomenon of ecological succession, causing biochemical lesions and interact with the health of living beings despite careful selection of crops.

EFFECTS ON THE SOIL ORGANISMS

It has been well established that the genetically manipulated plants affected the soil ecosystem. The genetically manipulated crop species by shedding the plant parts and sending the assimilates through roots as exudates deleteriously affected the useful soil microbes that are involved in the decomposition of the organic matter (Whipps, 1990). Most of the GM crops carry the Cauliflower Mosaic Virus (CaMV) 35S promoter which remains stable for long periods and caused several environmental hazards (Ho et al., 1999). The GM potato lines producing lectins GNA and ConA had altered the composition of starch and affected the microbial activity in soil (Griffiths et al., 2000; Milling et al., 2004). The modified alfalfa to produce lignin peroxidase significantly altered the composition and activity of rhizospheric bacterial communities (Donegan et al., 1999). The Bacillus thuringiensis (Bt) plant residues and root exudates lead to an accumulation of the transgenic gene (Dunfield et al 2009) and protein in the soil (Saxena et al., 1999), persisted long (Tapp and 1998) as that of an insecticide Stotzky, (dichlorodiphenyltrichloroethane) (Edwards, 1976) and lowered the microbial activity representing a potential risk for both target and non-target soil organisms. The Bt soybean and rice did not have any effect on the soil microorganisms (Ferreira et al., 2003; Wu et al., 2006). But the soil amended with Bt rice straw did lower the dehydrogenase activity, methanogenesis, production of hydrogen and anaerobic respiration (Wu et al., 2004). These changes might be due to the increased content of the saturated lipids with a considerable loss of unsaturated lipids from the soils (Dinel et al., 2003). However, a

dramatic change in the micro flora of the root surface was discerned in rice (Liu et al., 2004) that was modified for production of chitinases and glucanases against pathogenic fungi suggesting that the differential effects of genes employed. Some studies had also showed no apparent deleterious effect of the Cryl Ab toxin released by Bt corn plants through root exudates or directly incorporated toxin into soil on earthworms, nematodes, protozoa, culturable bacteria, saprophytic fungi, Collembola and Acaridida (Saxena and Stoztky, 2001) .The Gamasida, Oribatida.and mycorrhizal fungi in microcosms were not affected by growing Bt-maize (deVaufleury et al., 2007). The glyphosate- and glufosinate-resistant Brassica napus and corn did not induce permanent alterations in the composition and diversity of soil and rhizospheric microbial communities (King et al., 2001; Dunfield and Germida, 2003). The abundance of bacteria and fungi was reduced by 23% in soils grown with potato expressing the cysteine proteinase as potato-cyst nematode (Cowgill et al., 2002). The incidence of pathogenic Fusarium fungi was increased by two-fold in glyphosate-resistant soybeans (Larson RL et al. 2006) due to varying soil moisture conditions and altered the root biomass, root nodules and the total nitrogen when sprayed with glyphosate (Means and Kremer, 2007), whereas the glyphosate- and glufosinate-resistant Brassica napus and corn did not induce permanent alterations in the compo-sition and diversity of soil and rhizospheric microbial communities (Siciliano Germida, 1999; Giovannetti 2005). The genetically modified (GM) brinjal expressing the defensin Dm-AMP1had inhibited the growth of both phytopathogenic fungi and by releasing high levels of the antimicrobial protein in root exudates (Turrini et al., 2004).

The plant residues of the GM crops incorporated into soil was decomposed at a slower rate in the cases of Bt corn, canola, cotton, potato, rice and tobacco as compared to their non-Bt lines (Flores et al., 2005) due to the presence of a higher lignin content in the tissues of the plants (Saxena et al., 1999). The delayed decomposition might also be due to differential enrichment of bacterial population which was able to synthesize enzymes for degradation of complex carbohydrates in the Bt crop residues ploughed in situ. The lower rate of degradation of litter might be beneficial for the maintenance of soil structure. But longer persistence of Bt residues might enhance the risks related to toxin accumulation. The germinability and growth of presymbiotic hyphae of Arbuscular mycorrhizal fungus Gomus mosseae that converts the litter into humus, enhanced disease resistance, improved water relations, and increased soil aggregation in soils incorporated with Bt crop residues (Castaldini et al., 2005) and these changes perhaps due to differential rate of release of gene products from the modified crop litter. But the modified tobacco plants for the expression resistance had delayed root colonization by Gomus mosseae fungi (Vierheilig et



Figure 1. Heavy infestation of the Mealy bugs *Phenacoccus solenopsis* (Tinsley) in the cotton system due to ecological succession of insect pests in India.

al., 1993). The activity of soil microflora, dehydrogenase enzymes and respiration of soil and mineralization of nitrogen were reduced while the level of phosphorous was enhanced in soil grown with the Bt cotton (Sarkar et al., 2009). The root nodule bacteria with Bt transgene. Heavy infestation of the Mealy bugs *Phenacoccus solenopsis* (Tinsley) in the cotton system (Figure 1) due to ecological succession of insect pests in India.displaced non-manipulated bacteria in legume root nodules (Skøt et al., 1994) and this may pose serious problems related to the fertility of soils. But the nitrogenfixing Diazotroph bacterial community did not suffer as evidenced in the Bt white spruce plantation (Lamarche and Hamelinthe, 2007).

EFFECTS ON THE MODIFIED PLANTS

The transgenes by interacting with the native ones caused several biochemical changes in the plants and altered morphological traits of the modified plants. The alien genes CP4 EPSP (Gertz et al., 1999), lectin gene (Bernal et al., 2003), phytoene synthase gene (Fray et al., 1995) and Bt gene (Shu et al., 2002) had induced dwarfism in soybean, sugarcane, tomatoes and rice. The GM soybean plants resistant to herbicide were with less chlorophyll and increased susceptibility to stem-splitting at high temperatures (Gertz et al., 1999). The Bt rice was inferior to non-transformed one in terms of plant height,

number of productive tillers per plant, number of grains per panicle and seed setting rate and produced smaller or malformed grains (Shu et al., 2002). The gene MdPG1 in apples had changed the composition of cell walls containing less pectin with malformed stomata which remained open during night (Atkinson et al., 2002) to cause higher transpiration resulting in utilization of more water than the normal trees. The Dandelions manipulated to have compound leaves had irregular leaf form and did not flower (Müller et al., 2006). The GM Arabidopsis plants producing biodegradable polyester were smaller. showed severe changes in metabolism and never produced seeds (Bohmert et al., 2000). The transgenic Lc alfalfa stem and leaves were bright green, synthesized more anthocyanins, lignin and proanthocyanidins with less flavone content and changed the quality of forage (Ray et al., 2003; Shadle et al., 2007). The genes psy and beta-lcy were able to synthesize both B-carotene and xanthophylls in rice (Beyer et al., 2002). The altered biochemical make-up was evidenced in the transgenic pine apple and tobacco (Yabor et al., 2006; Misawa et al., 1994). The DHS gene in Arabidopsis delayed bolting and leaf senescence with bigger rosette leaves, root biomass. catalyzed the conversion of lysine to deoxyhypusine and enhanced yield (Wang et al., 2003). The phytoene synthase gene delayed the germination and increased levels of chlorophyll and carotenoids (Lindgren et al., 2003). The heat-stable enzyme beta-glucanase gene had lowered the grain weight and yield of barely (Horvath et

al, 2001). The barley with zinc transport protein gene had negative effect on the uptake of zinc in zinc-deficient soil (Ramesh et al., 2004). In some cases, the transgenes yielded non-viable off springs as in barley (Bregitzer et al. 2007). The transgenic birch trees expressing an antifungal enzyme chitinases showed increased susceptibility to leaf spot disease in the field (Pappinen et al., 2002). The delayed germination, reduced the level of chlorophyll with changed structure of plastids and compo-sition of fatty acids, and decreased the level of gamma tocopherol were evidenced in GM Canola (Shewmaker et al., 1999). The tissue that had low chlorophyll content did not express fully Cry1A in maize and cotton(Abel and Adamczyk, 2004). The silencing of gene occurred in Bt cotton at 37°C during bolling stage and altered the levels of toxin, glutamic-pyruvic transaminase, soluble protein, increased levels of protease and free amino acids (Chen et al., 2005). Glyphosate-resistant cotton had produced 40% less loose pollen per stigma with a variety of cellular abnormalities and increased the distance between the anthers and the stigma (Pline et al., 2002). The stems of Bt maize plants had significantly lower levels of alkenes. But they were richer in n-alkanes, n-fatty and unsaturated fatty acids than non-Bt shoots to affect the digestibility in cattle or pigs (Poerschmann et al., 2008). The silking and maturity were delayed with the lower yield, and higher moisture content of grain in Bt maize than in the conventional varieties suggesting that the modified plants drained much of water from the system and yielded less (Ma et al., 2005). The chitinase gene in pineapple had decreased the levels of aldehydes, chlorophyll a, b, phenolics and proteins and thereby reduced ability to perform photosynthesis and withstand stress tolerance (Yabor et al., 2006). The alien gene in sugarcane had modified the sugar metabolism (Vickers et al., 2005a). The sense or antisense constructs engineered had significantly lowered the cane yields with darker juice and higher levels of polyphenol oxidase derived from a sugarcane cDNA library (Vickers et al., 2005b). The glucokinase gene in potato had significantly decreased the amount of starch in the tubers by accumulating the hexose phosphates, organic acids and amino acids and lowered the levels of both alpha-chaconine and solanine glycoalkaloids in the leaves and also caused changes in 88 metabolites including alkaloid content of the plant as evidenced in potatoes (Trethewey et al., 1998; Birch et al., 2002; Roessner et al., 2001a). The blast resistance gene (Coca et al., 2006) in rice had activated the oxidativestress response for enhancing the ceropin A protein synthesis and folding for subsequent secretion (Campo et al., 2008). The genes for the HIV proteins p24 had slowed down the growth of tobacco plants due to yellow pigmentation and induced rearrangement of DNA (Zhang et al., 2002). It was reported that the modified barley for full-length collagen protein of human was found at low and detectable levels in its seeds. But the next generation had fertility problems and produced seeds with no

detectable human collagen (Eskelin et al., 2009). The flavonol-enriched tomatoes with LC and C1 genes had altered levels of at least fifteen substances including citric acid, sucrose, phenylalanine and trigonelline (Le Gall et al., 2003). The antisense DHS gene in tomatoes by increasing the content of chlorophyll enhanced photosynthesis which enabled the plant to have increased starch. But the pollens in these plants were deformed and necessitated cross-pollination for formation of fruits(Wang et al., 2005). The transformed wheat with a novel "hybrid" glutenin subunit (HMW-GS) gene showed an irregular expression of glutenin and changed its expression levels over subsequent generations (Blechl and Anderson, 1996) while the low-molecular-weight glutenin subunit gene had increased the expression of glutenin and decreased the gliadin synthesis in wheat (Masci et al., 2003). The wheat with scab-resistance genes caused localized death of leaf tissue due to formation of necrotic spots on the leaves during the flower formation stage of the crop (Anand et al., 2003). Interestingly, the single-site integration of foreign DNA into Arabidopsis had undergone rearrangements and deleted both plant DNA as well as foreign DNA (Forsbach et al., 2003). The anti-sense shikimate hydroxycinnamoyl transferase gene with PAL2 promoter in alfalfa caused stunting with less biomass and delayed flowering and reduced the enzyme activity and lignin content to improve the digestibility of forage (Shadle et al., 2007). The flax seeds expressing the bacterial phytoene synthase gene crtB, showed elevated level of carotenoid and changed its components, i.e., ct-carotene, 13-carotene, phytoene and lutein (Fuiisawa et al., 2008). The carotenoids are considered as a strong protective agent against lung cancer (Nishino et al., 1995) and lutein is an essential functional component in the human eye (Berendschot et al., 2000). In canola or Arabidopsis or tomato, the phytoene synthase genescr113 or FSY respectively, resulted in delayed germination (Lindgron et al., 2003) produced darker seeds, increased the level of a-carotene, lutein, violaxanthin, zeaxanthin, lycopene and chlorophyll (Shewmaker et al., 1999). There was over accumulation of starch in leaves of potato, inhibition of growth, photosynthesis, induced rapid senescence, decreased fruit setting, sucrose unloading capacity and delayed softening of fruit in tomato that were transformed with the antisense genes(Veramendi et al., 1999; Dai et al.,1999). Also, in tobacco, the phytoene synthase gene resulted in severe phenotypic changes, altered the morphology and pigmentation of leaf (Busch et al., 2002). The Bar transgenic oats exhibited greater growth and increased tolerance to salt stress conditions as against no GM crop (Oraby et al., 2005) in which both the transgenic construct and host DNA got modified (Wojciede et al., 1998) and this phenomenon may have negative consequences on the biodiversity. However the organ specific gene promoter like LP2 was claimed to limit the potential unintended impacts on plants and environment (Thilmony et al., 2009) and such promoter will go in a long

way to help in generating the new crop varieties that may not differ from that of their isogenic lines. The plant volatiles have profound effects on the composition and visitation of insects community to the cropped area. The transgenic scab resistant apple differed quantitatively in four terpenes and an aromatic compound and these modified odour emissions proved to be crucial in the composition of non target insects (Ute et al., 2010).

EFFECTS ON THE NON-TARGET ORGANISMS

The pollen is a potential source for exposure to nontarget insects and consumption of deposited pollens on plants affected the non-target insects as well as pests (Felke et al., 2002). The survival of monarch butterfly Danaus plexippus larvae to adulthood was reduced when they were exposed to Bt maize pollens (Dively et al., 2004). But simultaneous exposure to both Bt anthers and pollens caused an additive effect and resulted in a lower survival of monarch larvae (Anderson et al., 2005) A similar effect was also noticed in Propylea japonica fed with the rice pollen (Bai et al., 2005). The predators and parasites of insect pests of crops were also succumbed to the genetically modified crops (Ahmad et al., 2006). The longevity of the predators Orius tristicolor and Geocoris punctipes that were raised on the prey fed on Bt cotton was significantly decreased by 27 - 28% whereas Nabis sp. and Zelus renardii remained unaffected (Ponsard et al., 2002) and differential susceptibility might either be due differential quantity of food consumed or elimination of the toxin from the system. The hymenopteran parasitoids often showed adverse effects while parasitizing host fed on Bt plants due to the poor quality of the host (Lövei et al., 2005). The Cry1Ac in cotton retarded the development of hymenopteran parasites that were raised on their hosts possibly due to sublethal effects on the host (Baur and Boethel, 2003). Similiarly, an important endoparasitoid of the cotton bollworm Microplitis mediator in China suffered with reduced survival and growth inhibition when its prey Helicoverpa armigera raised on Bt cotton leaf powder (Cry1Ac) (Liu et al., 2005). The survival, developmental period and weight of cocoon were negatively affected in Cotesia marginiventris, Aphidius (Vojtech et al., 2005; Ashouri et al., 2001), Parallorhogas pyralophagus (Tomov et al., 2003) on hosts that were fed on the hosts fed with the modified crops with Cry1Ab, Cry3A and Cry9C. The abundance of some hymenopteran parasitoids was lowered in fields cropped with Bt maize and Bt cotton as compared to Bt corn (Cry1Ab), where the abundance was increased (Dively, 2005; Pilcher et al., 2005). Bt rice did not either resurged the populations of plant hopper or negatively affected its predator Cyrtorhinus lividipennis (Chen et al., 2007)

The predator larvae of *Chrysoperla carnea* had shown a preference for prey fed on the non-transgenic corn as

compared to prey fed on Bt corn (Meier and Hilbeck 2001) which would potentially reduce the exposure of this predator. The tritrophic interaction study revealed that the pupal body mass of lacewing Chrysopa pallen fed with Bt cotton GK12-originated aphid prey was significantly higher than that of the control and more number of females were emerged. These females were laid significantly more number of eggs than the females emerging from larvae maintained on another Bt cotton variety. NuCOTN 99B suggesting that the differing status of the hosts(Guo et al., 2008) might be responsible for this effects. The risks for predators, Lacewings Chrysoperla carnea when feeding on thrips, Frankliniella tenuicornis in or next to Bt maize fields were found to be negligible (Obrist et al., 2005) and a similar effect was noticed in predatory ladybeetle, Propylaea japonica on cotton aphids(Zhu et al., 2006).

The insect predatory pirate bug, lady bird beetles, and carabids did not suffer under Bt corn (Dalv and Buntin. 2005). A five-year field study showed that the Bt cotton had not altered the activity of the natural enemy community on three key pests but a field study made over three years had clearly shown that the population of ladybird beetle was lowered in Bt cotton fields due to availability of reduced number of prey (Naranjo, 2005b). However, the abundance was lower in pure Bt cotton plots as compared to a row mixture of Bt and non-Bt plants in another field study (Sisterson et al., 2004). The overall arthropod diversity and the diversity of pest sub-communities were increased, but the diversity of natural enemy subcommunities were decreased in Bt cotton field (Men et al., 2004) and the density shifts of natural enemies were often ascribed to prev dynamics or plant-mediated indirect causes. The egg plant expressing the Cry3Bb toxin were preferred by the herbivorous spider mites. Tetranychus urticae, but its predator, Phytoseiulus persimilis consumed significantly less number of Bt-fed spider mites (Zemkov et al., 2005). However, the jumping spiders and Lacewings, Chrysoperla carnea were found to be less abundant in Cry1Ab x Vip3A cotton (Dively 2005). But spiders as a whole were more numerous in Bt maize (Ahmad et al., 2006), cotton (Head et al., 2005) and potato (Reed et al., 2001). The Bt potatoes expressing Cry 3Aa had no effect on the aphid Myzus persicae and aphidophagous coccinellids C. septempunctata fed on the aphid (Kalushkov and Nedved, 2005). The recent study elucidated that the ecological function of natural predation on the eggs of Leptinotarsa decemlineata in GM potato plots was not impaired (Arpaia et al., 2009). It was evident that the direct environmental benefits of reduced insecticide applications in Bt-cotton resulted in fewer non-target effects (Whitehouse et al., 2005).

Although the pollen containing the Bt toxin is harmless to bees, their visitation to the fields cropped with modified Canola (van Engelsdorp et al., 2010; Morandin and Winston, 2005) resistant to herbicide was reduced and this phenomenon is considered as a threat to humanity

as predicted by Albert Einstein. The weed seed resources for birds were reduced in glyphosate-resistant beet production areas and this phenomenon poses a great threat to the bird community (Gibbons et al.2006). Interestingly the woodlouse, *Porcellio scaber*, performed better when fed with Cry1Ab maize as compared to the non-transgenic isoline, which was attributed to a better nutritional quality of the Bt corn (Riudavets et al., 2006). The Bt rice grains had affected variously the stored product insects, *Plodia interpunctella*, *S. oryzae* adult and the psocid *Liposcelis bostrichophila* (Escher et al., 2000) suggesting that the Bt toxin is capable of getting into the food chain.

EFFECTS ON THE MAMMALS AND POULTRY

The genetically modified (GM) crop plants have allergens and caused different kinds of allergies both in the test animals and human beings. A new molecule (SSP 6711) expressed by GM maize was akin to 50 kDa gamma zein is a well known allergenic protein and also carried several synthetic RNA fragments to interact with the mammalian immune system to cause unexpected turbulences (Nordlee, 1996; Zolla et al., 2008; Rosati et al., 2008). A recent study in GM maize had identified the recurrent changes in the immune system involving changes in the number of a special type of lymphocytes and is important in the human (Rachmilewitz et al., 2004) as well as mice immune system (Finamore et al., 2008). The GM peas expressing non-native proteins in plants resulted in the structural variants possessing altered immunogenicity in mice (Prescott et al., 2005). The modified rice seeds with genes from Japanese cedar pollen allergens were shown to function as an effective edible vaccine for the control of pollen allergen-induced responses (Takagi et al., 2006). Interestingly the accumulation of allergen Gly m Bd 30 K (P34) in the seeds of Soya bean was prevented by modifying the plants with a silencing gene (Herman et al., 2003).

The GMO foods were affected the normal physiology and biochemical metabolisms in the mammals. The rats fed with GM maize had significant variations in growth in both the sexes and female suffered from hepatorenal toxicity with increased triglycerides, while the males excreted diminished amount of phosphorus and sodium in the urine (Séralini et al., 2007) and significant disturbances in the function of kidney, liver and some effects on heart, adrenal, spleen and blood cells were also frequently seen between male and female rats fed with GM maize (deVendômois, 2009). The haemorrhage, sickness, and lesions in liver with the altered enzyme production, enlarged pancreas, stomach with mysteriously heavier intestine and proliferation of the gastric mucosa on different parts of the gastrointestinal tract were noticed in rats fed with GM foods (Pusztai et al., 2003) rats fed with the potatoes engineered to produce a

lectin showed proliferative cell growth in both the stomach and intestinal walls (Stanley et al., 1999), partially atrophied livers and slower growth in the brain (Pusztai, 2002). The Cell proliferation was reported to be a precursor to cancer (Pusztai, 2005). The FlavrSavr GM tomato fed rats died within two weeks (Pusztai, 2002) and GM Soya food increased the infant mortality of rat by 55.3% (Ermakova, 2006). Rabbits fed with GM soybean showed higher metabolic activity with altered enzyme production in their livers (Tudisco et al., 2006). The cells in the pancreas of mice fed with the Roundup Ready soy had produced significantly less digestive enzymes (Malatesta et al., 2003). Embryos of GM Soy-fed mice also showed temporary changes in their DNA function (Vecchio et al., 2004). The microscopic analysis of the livers of mice fed with GM soybeans revealed altered gene expression and structural and functional changes. Enzyme production in the hearts of mice was altered by GM Sova (Tudisco et al., 2006). A high pup mortality was characteristic of every litter from mothers fed with the GM soy flour of R21 (Ermakova, 2006). Mice fed with the Bt potatoes showed an abnormal and damaged cells, and proliferative cell growth in ileum (Nagui et al., 1998). The ultrastructural analyses of the liver showed that the nuclei of cell of the GM Soya fed mice were more irregular in shape with a higher percentage of dense fibrillar components with lower proportions of fibrillar centres and granular component (Malatesta et al., 2002). The larger zymogen granule in pancreas with significantly lower αamylase antibody labeling density and pancreatic αamylase were noticed in animals fed with the GM soybean (Caporaloni et al., 2002). In both mice and rats fed with the Roundup Ready soybeans, dramatic changes in the testicles were observed; they were dark blue in colour instead of pink with the altered young sperm cells (Cisterna et al., 2006). The testis had higher number of perichromatin granules with dilated vesicles in cells and smooth endoplasmic reticulum of Sertoli cells. A temporary reduction in antibodies was evident and the female gave birth to fewer smaller babies than the normal one (Velimirov et al., 2008). Both the male and female animals displayed troubling reproductive problems when fed with the GM Soybean (Tudisco et al., 2006). The other study showed that the genetically modified crops had no adverse effects on the multigeneration reproductivedevelopmental ability(Rhee et al., 2005) The GM fruits of the brinial modified with CrylAb and CrylAc sequences were known to have resistance to kanamycin, provided less calories and affected the intake of food as well as feed conversion rate in the test animals besides changing the picture of the blood cells and its chemistry in respect of the hepatic markers alanine albumin, lactosedehydrogenase and aspartate aminotransferases in animals. Interestingly, the milk production and its composition were changed by 10 to 14% in cows fed with the GM fruits (Seralini, 2009). The ileal starch, amino acids digestibility and apparent metabolisable energy (AME) were

significantly reduced in the birds fed with the transgenic up peas (Li et al., 2006). The Round up Soya carrying the stop signal had four harmful RNA variants which are considered as harmful and (Rang et al., 2005) reduced clinically important phytoestrogens levels by 12 to 14% (Lappé et al., 1999). Interestingly the hamsters fed with GM Soya showed the growth of hairs inside the pouches of the mouth and the number of hairy mouthed hamsters was much higher in the third generation of GM soy fed animals than in others (Baranov et al., 2010). The study with human showed that the antibiotic resistance marker gene survived in the gut of the ileostomy patients given one meal of GM Soya and detected the measurable amounts of the full length of the transgene construct (Susana et al., 2002). The full length transgene construct is likely to encode the functional gene provided if it is not methylated (Orend et al., 1995) and penetrate into the nucleus by crossing through the nuclear membrane (Gorlick et al., 1996). The large scale introduction of GM crops in USA had increased the incidence of three or more chronic disease of people from 7 to 13% (Paez et al., 2009).

THE HORIZONTAL GENE TRANSFER

The persistence and movement of transgenic DNA in agricultural and natural systems poses a threat of horizontal gene transfer and possible proliferation of genetically modified DNA into the general environment. Although the natural horizontal gene transfer (Ochiai et al., 1959) is limited by species barriers engineered artificial genetic constructs crossed all species barriers and invaded all genomes. There are many studies confirming that the pollens are contributing much towards the horizontal gene transfer in the plant kingdom (Jarosz et al., 2005). The pollens of GM maize are capable of moving up to 1000m and pollinate rape (Scheffler et al., 1993), sugarbeet (Alibert et al., 2005) and maize (Loos et al., 2003). The GM Arabidopsis thaliana with the Csrl-1gene (Bergelson et al., 1998) and canola with bargene (Snow et al., 1999) were more freely pollinated the natural wild populations of B. rapa than the non GM plant. The F1 hybrids formed due to such trespass of genes competed with other weeds in the commercial agricultural fields (Warwick et al., 2003). The genes flow from GM rice (Chen et al., 2004) crossed into the population of creeping bent grass (Agrostis stolonifera) and Zoysia grass through pollens (Watrud et al., 2004). The low genes flow was also reported from GM rice, red rice and conventional rice (Messeguer et al., 2004), from transgenic rice to red rice (Zhang et al., 2003) and insect resistant rice (Bashir et al., 2004). The non-Bt plants in the refuges known to carry one to two copies of the Bt gene (Heuberger et al., 2008) and such contamination is detrimental to the system. The vector-mediated horizontal gene transfer and recombination could occur to create new pathogenic organisms

(Gebhard and Smalla, 1998) in the biosphere. A rare event of transference of transgenic gene was occurred from the plant to terrestrial bacteria (Nielsen et al., 1998) .The persistence and movement of genes from GM crops through DNA pose a threat for possible proliferation in the general environment. The plant litters on decomposition released DNA spread into the soil rhizosphere, persisted for a long time upto two to several years and retained its transforming activity (Ceccherini et al.2003, Gebhard and Smalla, 1999) in the soils towards naturally competent Bacillus (Blum et al., 1997), Pseudomonas (Meier and Wackernagel, 2003) and Acinetobacter (deVries et al., 2001) depending on homologous sequences in the recipient cells. Such DNA were protected by the clay minerals and other soil particles against degradation (Lorenz and Wackernagel, 1992). Although the DNA in decaying plant cells was rapidly degraded DNA of the appropriate length survived in some soils (Hay et al., 2002) and is prone for uptake. The plasmidic DNA detected in compost and manure was incorporated into soil (Smalla et al., 2001; Loos, 2003). It was known that the transgenic DNA concentration in bulk soil micro arthropods, nematodes, macro arthropods and earthworms was significantly higher than that of background soi1 (Miranda et al., 2009). The gene transfer between GM plants and fungi was detected both in co-culture systems and in planta (Bryngelsson et al., 1988) and could represent a potential risk in the case of fungal pathogens and symbionts living inside the host plant cells. Horizontal gene transfer was occurred from a GM potato line to a bacterial pathogen (Schluter et al., 1995). and the in situ transfer of gene occurred from GM plants to terrestrial bacteria (Kay et al.2002) and also from GM zoysiagrass to the indigenous microorganisms in the cultivated field (Bae et al., 2008). The agro infection of transgenic plants had resulted in causing viable cauliflower mosaic virus disease through the processes of intermolecular recombination (Gal et al., 1992).

All though there were conflicting reports on the fate of GM DNA in the biological system it was observed that DNA could pass through the gut wall into the blood stream and taken up by cells in the blood, liver, spleen and passed through the placenta to the cells of the feoetus and the newborn one (Doerfler and Schubert, 1998). The presence of recombinant DNA fragments and CrylAb protein was detected in the gastrointestinal contents of pigs (Chowdhury et al., 2003), mice (Schubbert et al., 1997), oral cavity and rumen of sheep and the digestive tract (Duggan et al., 2004) that were fed with GM food. The gene from GM Soya crop was transferred into the intestinal bacteria and continued to function in the human system (Netherwood et al., 2004). The fragments of synthetic transgenes were also traced in the blood of piglets fed with GM maize food (Mazza et al., 2005). The transformation activity of DNA of decaying GM maize in ovine saliva, rumen fluid and silage effluent and found the way into aquatic environments was also occurred (Paul et al.,

1989; Duggan et al., 2000). The gills, digestive glands, and gonads of the fresh water Mussels were significantly contaminated by cry1 and cry1Ab genes (Douville et al., 2009). Interestingly the DNA and proteins of transgenic pollen were remained intact in honey for more than seven weeks (Eady et al., 1995). Interestingly the deleted gene npll was able to to take up and integrate npll and other genetic material from transgenic sugar beet through homologous recombination (Gebhard and Smalla, 1998).

EFFECTS ON RESISTANCE AND ECOLOGICAL SUCCESSION

The ecological succession is a natural phenomenon when one species of competing organism is displaced from the habitat either by nature or anthropological activity (Sundaramurthy, 2002) will lead into the succession of another one as evidenced earlier in the Indian cotton system on the account of gene to gene interaction (Sundaramurthy, 1994). The large scale cultivation of genetically modified crops over several years could increase the selection pressure on pest species, which might result in the development of resistance (Fox, 2003). The resistant population may elicit succession within the species in the given habitat by breeding with the susceptible wild one. Bt cultivars showing large reductions in Cry1Ac concentration over the growing season became more susceptible to insect damage (Olsen et al., 2005). Increased resistance to Bt toxin was noticed in the populations of American bollworm at high temperatures as the high temperature altered the synthesis of the protein metabolism by silencing the gene by way of reducing the glutamic-pyruvic transaminase, soluble protein and increasing protease and free amino acid levels in the leaves (Chen et al., 2004). The insect species diamondback moth (Martinez-Ramirez et al., 1995). European corn borer (Weinzierl et al., 1997) pink bollworm (Wilson et al., 1992) had developed resistance to the transgenic Bt crops and the resistant population would cause succession by breeding with the susceptible one in the ecological niches. The changes in the biochemical make up of the modified crops (Martinez-Ramirez et al., 1995) may accelerate succession of other insect pests of minor importance as noticed earlier with indiscrimate use of synthetic pyrethroids insecticides (Sundaramurthy, 1994) and currently widespread growing the transgenic cotton (Nagrare et al., 2009) in India and China (Lu et al., 2010). The genetically modified crops are also known to cause succession of pathogenic fungi due to availability of certain nutrients. The root exudates containing elevated levels of several amino acids, fruc-tose and maltose in transgenic cotton supported the growth of the fungus, Fusarium oxysporum and resulted in succession (Li et al., 2009). However, GM beet culti-vars showed significantly more injury from the soil fungus Fusarium oxysporum and Rhizoctonia solani when sprayed with glyphosate which could have sequential effect by increasing the inoculum

of soil pathogens and affect other crops in rotation with GM sugar beet (Pasonen et al., 2006). The transgenic birch trees expressing an antifungal enzyme gene had increased the susceptibility to leaf spot disease in the field (Pasonen et al., 2004). The significant biological disadvantage of transgenes is that they may either transform wild weed plants into new or worst weeds or cause ecological succession of exis-ting weed species resistant to weedicides as in Argentina (Vitta et al., 2004).

The techniques involving the use of the restriction enzymes Zinc-finger nucleases for inducing a doublestranded break at their target locus (Shukla et al., 2009) and Targeting Induced Local Lesions in Genomes (TILLING) (Koornneef et al., 1982) that combines chemical mutagenesis with a sensitive mutation detection instrument are showing promise in the field of genetics for improvement of the crop plants (Reynolds et al., 2009) by accelerating the evolutionary process without involving introduction of foreign DNA of any type into a plant. The Cisgenesis (Giddings, 2006) involving a natural gene (Rommens et al., 2004) from the breeder's gene pool that codes for a trait also shows promise in generating the crops with high yield, enhanced nutritive values, resistance to biotic and abiotic stresses. This concept may facilitate second green revolution not only in the Indian agro system (Jacobsen et al., 2008) but also elsewhere for conferring food and nutrition security to mankind without causing any immediate disturbances to the available water resources, health of living beings and biodiversity.

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REFERENCES

Abel C, Adamczyk J (2004). Relative concentration of Cry1A in maize leaves and cotton bolls with diverse chlorophyll content and corresponding larval development of fall armyworm (Lepidoptera: Noctuidae) and southwestern corn borer (Lepidoptera: Crambidae) on maize whorl leaf profiles. J. Econ. Entomol. 97: 1737-1744.

Ahmad A Wilde GE, Whitworth RJ, Zolnerowich G (2006). Effect of corn hybrids expressing the coleopteran-specific cry3Bb1 protein for corn rootworm control on above ground insect predators. J. Econ. Entomol. 99: 1085-1095.

Anderson PL, Hellmich RL, Prasifka JR, Lewis LC (2005). Effects on fitness and behaviour of monarch butterfly larvae exposed to a combination of Cry1Ab expressing corn anthers and pollen. Environ. Entomol. 34: 944-952.

Anand A, Zhou T, Trick HN, Gill BS, Bockus WW, Muthukrishnan S (2003). Greenhouse and field testing of transgenic wheat plants stably expressing genes for thaumatin-like protein, chitinase and glucanase against *Fusarium graminearum*. J. Exptl. Bot. 54: 1101-1111.

Alibert B, Sellierand H, Souvré A (2005). A combined method to study

- gene flow from cultivated sugar beet to ruderal beets in the glasshouse and open field. Eur. J. Agron. 23: 195-208.
- Arpaia S, Schmidt JEU, Di Leo GM, Fiore MC (2009). Oviposition of the Colorado potato beetle (*Leptinotarsa decemlineata*) and natural predation on its egg masses in Bt-expressing fields. Biocont. Sci. Technol. 19: 971- 984.
- Ashouri A, Michaud D, Cloutier C. (2001).Recombinant and classically selected factors of potato plant resistance to the Colorado potato beetle, *Leptinotarsa decemlineata*, variously affecting the potato aphid parasitoid *Aphidius nigripes*. Biol. Control. 46: 401-418.
- Atkinson RG, Schröder R, Hallett I, Cohen D, MacRae EA (2002). Overexpression of Polygalacturonase in Transgenic Apple Trees Leads to a Range of Novel Phenotypes Involving Changes in Cell Adhesion. Plant Physiol. 129: 122-133.
- Bae TW, Vanjildorj E, Song SY, Nishiguchi S, Yang SS, Song IJ, Chandrasekhar T, Kang TW, Kim JI, Koh YJ, Parl SY, Lee J, Lee YE, Ryu KH, Riu KZ, Song PS, Lee HY (2008). Environmental risk assessment of genetically engineered herbicide-tolerant *Zoysia japonica*. J. Environ. Qual. 37: 207-218.
- Babili S, Ye X, Lucca P, Schaub P, Welsch R, Potrykus I (2002).Golden Rice: introducing the beta-carotene biosynthetic pathway into rice endosperm by genetic engineering to defeat vitamin A deficiency. J. Nutr. 132: 506-510.
- Bai YY, Jiang MX, Cheng JA (2005). Effects of transgenic cry1Ab rice pollen on fitness of *Propylea japonica* (Thunberg). J. Pest. Sci. 78: 123-128.
- Bashir K, Husnain T, Fatima T, Latif Z, Mehdi SA, Riazuddin S (2004). Field evaluation and risk assessment of transgenic indica basmati rice. Mol. Breed.13: 301-312.
- Baur ME, Boethel DJ (2003). Effect of Bt-cotton expressing Cry1A(c) on the survival and fecundity of two hymenopteran parasitoids (Braconidae, Encyrtidae) in the laboratory. Biol. Control, 26: 325-332.
- Beer LL, Boyd ES, Peters JW, Posewitz MC (2009). Engineering algae for biohydrogen and biofuel production. Curr. Opinion Biotech. 20: 264-271.
- Bernal JS, Sëtamou M, Fortuitous (2003). Antixenosis in Transgenic Sugarcane: Antibiosis-Expressing cultivar is refractory to ovipositing herbivore pests. Environ. Entomol. 32: 886-894.
- Bergelson J, Purrington CB, Wichmann G (1998) Promiscuity in transgenic plants. Nature. 395: 25. Baranov AS, Chernova OF, Feoktistova N, Yu, Surov AV (2010). A new example of ectopia: Oral hair in some rodent species, Doklady Biol. Sci. 431: 117-120,
- Berendschot TT, Goldbohm RA, Klöpping WA, Van de Kraats J, Van Norel J, Van Norren D (2000). Influence of lutein supplementation on macular pigment, assessed with two objective techniques. Invest. Ophthalmol. Vis. Sci. 41: 3322-3326.
- Beyer P, A1- Birch ANE, Geoghegan IE, Griffiths DW, McNicol JW (2002). The Effect of Genetic Transformations for Pest Resistance on Foliar Solanidine-Based Glycoalkaloids of Potato (Solanum tuberosum), Ann. Appl. Biol. 140: 143-149
- Blechl A, Anderson O (1996). Expression of a Novel High-Molecular-Weight Glutenin Subunit Gene in Transgenic Wheat. Nat. Biotech. 14: 875-879.
- Blum SAE, Lorenz MG, Wackernagel W (1997). Mechanism of retarded DNA degradation and prokaryotic origin of DNAses in nonsterile soil system. Appl. Microbiol. 20: 513-521.
- Borlaug NE (1968). Wheat breeding and its impact on world food supply In Proc.III Intl.Wheat Gen.Symp. Australian Acad. Sci. Canberra. pp. 1-36.
- Bohmert K, Balbo I, Kopka J, Mittendorf V, Nawrath C, Poirier Y, Tischendorf G, Trethewey RN,Willmitzer L(2000). Transgenic Arabidopsis plants can accumulate polyhydroxybutyrate to up to 4% of their fresh weight. Planta, 211: 841-845.
- Bregitzer P, Cooper LD, Hayes PM, Lemaux PG, Singh J, Sturbaum A (2007). Viability and bar Expression are Negatively Correlated in Oregon Wolfe Barley Dominant Hybrids. Plant Biotech. J. 5: 381-388.
- Brookes G, Barfoot P (2000). GM Crops: The global economic and environmental impact-the first nine years 1996-2004. AgBio. Forum, 58: 187-196.
- Bryngelsson T, Gustafsson M, Green B, Lind C (1988). Uptake of host DNA by the parasitic fungus *Plasmodiophora brassicae*. Physiol. Mol.

- Plant Pathol. 33: 163-171.
- Busch M, Scutcr A, Ham R (2002). Functional analysis of the early steps of carotenoid biosynthesis in tobacco. Plant Physiol. 128: 439-453
- Byrnee BH (1990). Environmental effects of N fertilizer use An overview Fertil. Res. 26: 209-215.
- Campo S, Manrique S, Garcia-Martinez J, San Segundo B (2008). Production of cecropin A in transgenic rice plants has an impact on host gene expression. Plant Biotech. J. 6: 585-608.
- Caporaloni C, Rossi L, Battistelli S, Rocchi MBL, Tonucci F, Gazzanelli G (2002). Brief communication: Ultrastructural analysis of pancreatic acinar cells from mice fed on genetically modified soybean. J. Anat. 201: 409-415.
- Castaldini M, Turrini A, Sbrana C, Benedetti A, Marchionni M, Mocali S, Fahiani A, Landi S, Santomassimo F, Pietrangeli B, Nuti MP, Miclaus N, Ceccherini MT, Poté J, Kay E, Tran Van V, Maréchal J, Pietramellara G, Nannipieri P, Vogel TM, Simonet P (2003). Degradation and transformability of DNA from transgenic leaves. Appl. Environ. Micriobiol. 69: 673-678.
- Chen D, Ye G, Yang C, Chen Y, Wu Y (2005). The effect of high temperature on the insecticidal properties of Bt cotton. Environ. Exptl. Bot. 53: 333-342.
- Chen M, Liu ZC, Ye G, Y, Shen ZC, Hu C, Peng YF, Altosaar I, Shelton AM (2007). Impacts of transgenic *cry1Ab* rice on non-target plant hoppers and their main predator *Cyrtorhinus lividipennis* (Hemiptera: Miridae) A case study of the compatibility of Bt rice with biological control. Biol. Control. 42: 242-250
- Chen LJ, Lee DS, Song ZP, Suh HS, Lu BR (2004). Gene flow from cultivated rice (*Oryza sativa*) to its weedy and wild relatives. Ann. Bot. 93: 67-73.
- Chen DH,Ye GY, Yang CQ, Chen Y, Wu YK (2004). Effect after introducing *Bacillus thuringiensis* gene on nitrogen metabolism in cotton. Field Crops Res. 87: 235-244.
- Chong DK, Langridge WH (2000). Expression of full-length bioactive antimicrobial human lactoferrin in potato plants. Transgenic Res. 9: 71-78.
- Chowdhury EH, Kuribara H, Hino A, Sultana P, Mikami O, Shimada N, Guruge KS, Saito M, Nakajima Y (2003). Detection of corn intrinsic and recombinant DNA fragments and Cry I Ab protein in the gastro-intestinal contents of pigs fed genetically modified corn Bt11. J. Anim. Sci. 81: 2546-2551.
- Cisterna B, Malatesta M, Martin TE, Biggiogera M (2004).Ultrastructural analysis of testes from mice fed on genetically modified soybean. Euro. J. Histochem. 48: 449-454.
- Coca M, Peñas G., Gómez J, Campo S, Bortolotti C, Messeguer J, Segundo BS (2006). Enhanced Resistance to the Rice Blast Fungus *Magnaporthe grisea* Conferred by Expression of a cecropin A Gene in Transgenic Rice. Planta. 223: 392-406
- Cowgill SE, Bardgett RD, Kiezebrink DT, Atkinson HJ (2002). The effect of transgenic nematode resistance on non-target organisms in the potato rhizosphere. J. Appl. Ecol. 39: 915-932.
- Dai N, Schaffer A, Petreikov M, Shahak Y, GillerY, RatnerK, Levine A, Granot D (1999). Over expression of Arabidopsis hexokinase in tomato plants inhibits growth, reduces photosynthesis, and induces rapid senescence. Plant Cell. 11: 1253-1266.
- Daly T, Buntin GD (2005). Effect of Bacillus thuringiensis transgenic corn for Lepidopteran control on non-target arthropods. Environ. Entomol. 34: 1292-1301.
- Davis DR, Epp MD, Riordan HD (2004). Changes in USDA food composition data for 43 garden crops, 1950 to 1999. J. Am. Coll. Nutr. 23: 669-682.
- De Vaufleury A, Kramarz PE, Binet P, Cortet J, Caul S, Andersen MN, Emmanuelle, Coeurdassier PM, Krogh PH (2007). Exposure and effects assessments of Bt-maize on non-target organisms (gastropods, micro arthropods, mycorrhizal fungi) in microcosms. *Pedobiologia*. 51: 185-194
- Devries J, Meier P, Wackernagel W (2001). The natural transformation of the soil bacteria Pseudomonas stutzeri and *Acinetobacter* sp. by transgenic plant DNA strictly depends on homologous sequences in the recipient cells. *FEMS* Microbiol. Lett. 195: 211-215.
- Devendômois JS, Roullier F, Cellier D, Séralini GE (2009). A comparison of the effects of three GM corn varieties on mammalian health. Int. J.

- Biol. Sci. 5: 706-726.
- Dinel H, Schnitzer M, Saharinen M, Meloche F, Paré T, Dumontet S, Lemee L Ambles A (2003). Extractable soil lipids and microbial activity as affected by Bt and Non -Bt Maize grown on a silty clay loam soil. J. Environ. Sci. Health. 38: 211-219.
- Dively GP (2005). Impact of transgenic VIP3A × Cry1Ab lepidopteranresistant field corn on the non-target arthropod community. Environ. Entomol. 34: 1267-1291.
- Dively GP, Rose R, Sears MK, Hellmich RL, Stanley-Horn DE, Calvin DD, Russo JM, Anderson PL (2004). Effects on monarch butterfly larvae (Lepidoptera: Danaidae) after continuous exposure to Cry1Ab-expressing corn during anthesis. Environ. Entomol. 33: 1116-1125.
- Doerfler W, Schubert R (1998). Uptake of foreign DNA from the environment: the gastro-intestinal tract and the placenta as portals of entry. *Wien Klin Wochenschr.* 110: 40-44.
- Donegan KK, Seidler RJ, Doyle JD, Porteous LA, Digiovanni G, Widmer F, Watrud LS (1999). A field study with genetically engineered alfalfa inoculated with recombinant *Sinorhizobium meliloti*: Effects on the soil ecosystem. Appl. Soil Ecol. 36: 920-936.
- Douville M, Gagné F, André C, Blaise C (2009). Occurrence of the transgenic corn cry1Ab gene in freshwater mussels (*Elliptio complanata*) near corn fields: Evidence of exposure by bacterial ingestion. Ecotoxicol. Environ. Safety, 72:17-25.
- Duggan PS, Chambers PA, Heritage JF, Michael J (2003). Fate of genetically modified maize DNA in the oral cavity and rumen of sheep. British J. Nutr. 89: 159-161.
- Duggan PS, Chambers PA, Heritage J, Forbes JM (2000). Survival of free DNA encoding antibiotic resistance from transgenic maize and the transformation activity of DNA in ovine saliva rumen fluid and silage effluent. FEMS Microbiol. Lett. 191: 71-77.
- Dunfield KE, Germida JJ (2003). Seasonal changes in the rhizosphere microbial communities associated with field-grown genetically modified canola (*Brassica napus*). Appl. Environ. Microbiol. 69: 7310-7318.
- Dunfield KE, Pauls KP, Swanton CJ, Klironomos JN,Trevors JT (2009).Detection of transgenic cp4 epsps genes in the soil food web. Agron.Sustain. Dev. 29:497-501.
- Druery CT Bateson W (1901). Experiments in plant hybridization. J. Royal Hort. Soc. 26: 1-32.
- Edwards CA (1976). Persistent Pesticides in the Environment, CRC Press, Cleveland, Ohio, p. 170.
- Eady C, Twell D, Lindsey K (1995). Pollen viability and transgene expression following storage in honey. Transgenic Res. 4: 226-231.
- Ermakova IV (2006). GMO: Life itself intervened into the experiments. Lett. EcosInf. 2: 3-4.
- Ermakova IV (2006). Genetically modified organisms and biological risks. *Proc.* Intl. Disaster Reduction Conf. (IDRC) Davos, Switzerland August 27th September 1st, pp. 168-172.
- Escher NB, Käch W, Nentwig (2000). Decomposition of transgenic Bacillus thuringiensis maize by microorganisms and woodlice Porcellio scaber (Crustacea: Isopoda). Basic Appl. Ecol. 1: 161-169.
- Eskelin K, Ritala A, Suntio T, Blumer S, Holkeri H, Wahlström EH, Baez J, Mäkinen K, Maria NA (2009). Production of a recombinant full-length collagen type I alpha-1 and of a 45-kDa collagen type I alpha-1 fragment in barley seeds. Plant Biotech J. 7: 657-672.
- Estrella LH, Depicker A, Montagu MV, Schell J (1983). Expression of chimaeric genes transferred into plant cells using a Ti-plasmid-derived vector. Nature. 303: 209-213.
- FAO (2004). The state of food and agriculture2003-2004-agricultural biotechnology, meeting the needs of the poor? Food and Agriculture Organization of the United Nations, Rome. p. 208.
- Felke M, Lorenz N, Langenbruch GA (2002). Laboratory studies on the effects of pollen from Bt maize on larvae of some butterfly species., J. Appl. Entomol.126: 320-325.
- Ferreira LHPL, Molina JC, Brasil C Andrade G (2003). Evaluation of *Bacillus thuringiensis* bioinsecticidal protein effects on soil microorganisms. Fl. Soil. 256: 161-168
- Flores S, Šaxena D, Stotzky G (2005). Transgenic Bt plants decompose less in soil than non-Bt-plants. Soil Biol. Biochem. 37: 1073-1082.
- Finamore A, Roselli M, Britti S, Monastra G, Ambra R, Turrini A, Mengheri E (2008). Intestinal and peripheral immune response to
- MON810 maize ingestion in weaning and old mice. J. Agric. Food

- Chem. 56: 1533-1539.
- Forsbach A, Schubert D, Lechtenberg B, Gils M, Schmidt, RA (2003).Comprehensive characterization of single-copy T-DNA insertions in the Arabidopsis thaliana genome. Plant Mol. Biol. 52: 161-176.
- Fox J (2003). Resistance to Bt toxin surprisingly absent from pests. Nat. Biotechnol. 21: 958-959
- Fray RG, Wallace A, Fraser PD, Valero D, Heddon P, Bramley P Grierson D (1995).Constitutive expression of a fruit phytoene synthase gene in transgenic tomatoes causes dwarfism by redirecting metabolites from the gibberellin pathway. Plant J. 8: 693-701
- Fujisawa M, Watanabe M, Choi SK, Teramoto M, Ohyama K Misawa N (2008). Enrichment of carotenoids in flaxseed (*Linum usitatissimum*) by metabolic engineering with introduction of bacterial phytoene synthase gene crtB. J. Biosci. Bioeng. 105: 636-641.
- Gan W, Grutzendler J, Wong WT, Wong ROL, Lichtman JW (2000). Multicolor "DiOlistic" Labeling of the nervous system using lipophilic dye combinations. Neuron, 27: 219-225.
- Gal S, Pisan B, Hohn T, Grimsley N, Hohn B (1992). Agroinfection of transgenic plants leads to viable cauliflower mosaic virus by intramolecular recombination. Virology, 187: 525-533.
- Gebhard F, Smalla K (1988). Transformation of *Acinetobacter* sp. strain BD 413 by transgenic sugar beet DNA. Appl. Environ. Microbiol. 64: 1550- 1554.
- Gebhard F, Smalla K (1999). Monitoring field releases of genetically modified sugar beets for persistence of transgenic plant DNA and horizontal gene transfer. FEMS Microbiol. Ecol. 28: 261-272.
- Gertz JM, Vencill WK (1999). Hill N.S., Tolerance of transgenic soybean (*Glycine max*) to heat stress. The 1999 Brighton Confer. Weeds. Farnham, Surrey, UK, pp. 835-840.
- Gibbons DW, Bohan DA, Rothery P, Stuart RC, Haughton AJ, Scott RJ, Wilson JD, Perry JN, Clark SJ, Dawson RJG, Firbank LG (2006). Weed seed resources for birds in fields with contrasting conventional and genetically modified herbicide-tolerant crops. Proc. Biol. Sci. 273: 1921-1928.
- Giddings LV (2006). 'Cisgenic' as a product designation. Nat. Biotech. 24: 1329.
- Gorlick D, Mattaj IW (1996). Nucleocytoplasmic transport. Science, 271: 1513-1518.
- Giovannetti M (2005). Impact of Bt Corn on rhizospheric and soil euhacterial communities and on beneficial symbiosis in experimental microcosms. Appl. Environ. Microbiol. 71: 6719-6729.
- Griffiths BS, Geoghegan IE, Robertson WM (2000). Testing genetically engineered potato, producing the lectins GNA and ConA, on non target soil organisms and processes. J. Appl. Ecol. 37: 159-170.
- Guo JY, Wan FH, Dong L, Lovei GL, Han ZJ (2008). Tri-trophic interactions between Bt cotton, the herbivore *Aphisgossypii* Glover (Homoptera: Aphididae), and the predator *Chrysopa pallens* (Rambur) (Neuroptera: Chrysopidae). Environ. Entomol. 37: 263-270.
- Hay I, Morency MJ, Séguin A (2002). Assessing the persistence of DNA in decomposing leaves of genetically modified poplar trees. Can. J. For. Res. 32: 977-982.
- Head G, Moar M, Eubanks M, Freeman B, Ruberson J, Hagerty A, Turnipseed S (2005). A multiyear, large-scale comparison of arthropod populations on commercially managed Bt- and non-Btcotton fields. Environ. Entomol. 34: 1257-1266.
- Herman EM, Helm RM, Jung R, Kinney AJ (2003). Genetic modification removes an immunodominant allergen from Soybean. Plant Physiol. 132: 36-43.
- Ho MW, Ryan A, Cummins J (1999). Cauliflower mosaic viral promoter a recipe for disaster? Microb. Ecol. Health. 11: 194-197.
- Heuberger S, Yafuso C, Hoffman GD, Tabashnik BE (2008). Out crossed cottonseed and adventitious Bt plants in Arizona refuges. Environ. Biosafety, Res. 7: 87-96.
- Horvath H, Jensen LG, Wong OT, Kohl E, Ullrich SE, Cochran J, Kannangara CG, Von Wettstein D (2001). Stability of transgene expression, field performance and recombination breeding of transformed barley lines. Theor. Appl. Gene. 102:1-11.
- Jacobsen E, Nataraja KN (2008). Cisgenics Facilitating the second green revolution in India by improved traditional plant breeding. Cur Sci. 94: 1365-1366.

- Jansson C, Wullschleger SD, Kalluri UC, Tuskan GA (2010). Phytosequestration: Carbon biosequestration by plants and the prospects of genetic engineering. BioSci. 60: 685-696.
- Joos H, Timmerman B, Montagu MV, Schell J (1983). Genetic analysis of transfer and stabilization of Agro bacterium DNA in plant cells. Embo J. pp. 22151-2160.
- Joshi RP, Schoenbach KH (2000). Electroporation dynamics in biological cells subjected to ultrafast electrical pulses: A numerical simulation study. Phys. Rev. 62: 1025-1033.
- Kalushkov P, Nedved (2005). Genetically modified potatoes expressing Cry 3A protein do not affect aphidophagous coccinellids., J. Appl. Entomol. 129: 401-406
- Kay E, Vogel TM, Bertolla F, Nalin R, Simonet P(2002). In situ transfer of antibiotic resistance genes from transgenic (transplastomic) tobacco plants to bacteria. Appl. Environ. Microbiol. 68: 3345-3351
- King CA, Purcell LC, Vories ED (2001). Plant growth and nitrogenase activity of glyphosate-tolerant soybean in response to foliar glyphosate applications. Agro. J. 93: 179-86.
- Koornneef M, Dellaert LW, Van der Veen JH (1982). EMS- and relationinduced mutation frequencies at individual loci in *Arabidopsis thaliana* (L.) . Heynh. Mutat. Res. 93: 109-123.
- Lamarche J, Hamelinthe RC (2007). Evidence of an impact on the rhizosphere diazotroph community by the expression of *Bacillus thuringiensis* Cry1Ab toxin by Bt White Spruce. Appl. Environ Microbiol. 73: 6577-6583.
- Lappé MA, Bailey EB, Childress C, Setchell KDR (1999). Alterations in clinically important phytoestrogens in genetically modified, herbicidetolerant soybeans. J. Med Food. 1: 241-245.
- Larson RL, Hill AL, Fenwick A, Kniss AR, Hanson LE, Miller SD (2006). Influence of glyphosate on Rhizoctonia and Fusarium root rot in sugar beet. Pest Manage. Sci. 62: 1182-1192.
- Le Gall G, Colquhoun IJ, Davis AL,Collins GJ, Verhoeyen ME(2003).Metabolite profiling of tomato (*Lycopersicon esculentum*) using H NMR spectroscopy as a tool to detect potential unintended effects following a genetic modification. J Agric Food Chem. 51: 2447-2456.
- Li XH, Higgins TJV, Bryden WL (2006). Biological response of broiler chickens fed peas (*Pisum sativum* L.) expressing the bean (*Phaseolus vulgaris* L.) alpha-amylase inhibitor transgene. J. Sci. Food Agric. 86: 1900-1907.
- Li X, Liu BS, Heia D, Liu ZH, Zhou K, Cui J, Luo J, Zheng Y (2009). The effect of root exudates from two transgenic insect-resistant cotton lines on the growth of *Fusarium oxysporum*. Transgenic Res.18: 757-767.
- Lindgrcn LO, Stalbcrg KG, Hoglund AS (2003). Seed-specific over expression of an endogenous Arahidopcic phytoene synthase gene results in delayed germination and increased levels of carotenoids, chloroph II, and abscisic acid. Plant Physiol. 132: 779 785.
- Liu XX, Zhang QW, Zhao JZ, Li JC, Xu BI, Ma XM (2005). Effects of Bt transgenic cotton lines on the cotton bollworm parasitoid *Microplitis mediator* in the laboratory. Biol. Control, 35: 134-141.
- Liu B, Zeng Q, Yan F, Xu H, Xu C (2004). Effects of transgenic plants on soil microorganisms. Plant Soil. 271: 1-13.
- Loos C, Seppelt R, Meier-Bethke S, Schiemann J, Richter O (2003). Spatially explicit modeling of transgenic maize pollen dispersal and cross-pollination. J. Theor. Biol. 225: 241-255.
- Lorenz MG, Wackernagel W (1992). DNA binding to various clay minerals and retarded enzymatic degradation of DNA in a sand/clay microcosm. *in* Gene Transfers and Environment (ed. Gauthier MJ), Springer-Verlag, Germany, pp. 103-113.
- Lövei GL, Arpaia S (2005). The impact of transgenic plants on natural enemies: a critical review of laboratory studies. Entl. Exp. Appl. 114: 1-14.
- Lu Y, Wu K, Jiang Y, Xia B, Li P, Feng H, Wyckhuys KAG, GuoY (2010). Mirid bug outbreaks in multiple crops correlated with widescale adoption of Bt cotton in China. Science, 328: 1151-1154
- Ma JKC (2010).Generation of transgenic plants expressing antibodies to the environmental pollutant microcystin-LR. FASEB J. 24: 882-890.
- Ma BL, Subedi KD (2005). Development, yield, grain moisture, and nitrogen uptake of Bt Corn Hybrids and their conventional nearisolines, Field Crop. Res. 93: 199-211.
- Malatesta M, Caporaloni C, Gavaudan S, Rocchi MBL, Serafini S, Tiberi

- C, Gazzanelli G (2002). Ultrastructural morphometrical and immunocytochemical analyses of hepatocyte nuclei from mice fed on genetically modified soybean. Cell Struct. Funct. 27: 173-180.
- Malatesta M, Biggiogera M, Manuali E, Rocchi MBL, Baldelli B, Gazzanelli G (2003). Fine structural analyses of pancreatic acinar cell nuclei from mice fed on GM Soybean. Eur. J. Histochem. 47: 385-388.
- Martinez-Ramirez, Amparo C, Balthazar Escriche M, Dolores Real, Silva FJ, Ferre J (1995). Inheritance of resistant *Bacillus thuringiensis* toxin in a field population of Diamondback Moth (*Plutella xylostella*). Pesticide Sci. 43: 115-20.
- Mazza R, Soave M, Morlacchini M, Piva G, Marocco A (2005).
 Assessing the transfer of genetically modified DNA from feed to animal tissues. Transgenic Res.14:775-784.
- Means NE, Kremer RJ (2007).Influence of soil moisture on root colonization of glyphosate-treated soybean by Fusarium Species. Soil. Sci, Plant Anal. 38: 1713-1720.
- Men X, Ge F, Edwards CA, Yardim EN (2004). Influence of pesticide applications on pest and predatory associated with transgenic Bt cotton and non-transgenic cotton plants. Phytoparasitica, 32:246-254.
- Meier MS, Hilbeck A (2001). Influence of transgenic *Bacillus* thuringiensis corn-fed prey on prey preference of immature *Chrysoperla carnea* (Neuroptera: Chrysopidae). Basic Appl. Ecol.2:35-44.
- Meier P, Wackernagel W (2003). Monitoring the spread of recombinant DNA from field plots with transgenic sugar beet plants by PCR and natural transformation of *Pseudomonas stutzeri*. Transgenic Res. 12: 293-304.
- Messeguer J, Marfa V, Catala MM, Guiderdoni E, Mele E (2004). A field study of pollen-mediated gene flow from Mediterranean GM rice to conventional rice and the red rice weed. Mol. Breed. 13: 103-112.
- Menzie CM (1972). Fate of pesticides in the environment. Ann. Rev. Ent. 17: 199-222.
- Milling A, Smalla K, Maid FX, Schloter M, Munch JC (2004). Effects of transgenic potatoes with an altered starch composition on the diversity of soil and rhizosphere bacteria and fungi. Plant Soil, 266: 23-39.
- Miranda M, Hart JR, Powell RH, Gulden DJ, Levy-Booth, Misawa N, Masamoto K, Hori T, Ohtani T, Böger P, Sandmann G (1994). Expression of an erwinia phytoene desaturase gene not only confers multiple resistance to herbicides interfering with carotenoid biosynthesis but also alters xanthophyll metabolism in transgenic plants, Plant J. 6: 481-489.
- Morandin LA, Winston ML (2005). Wild bee abundance and seed production in conventional, organic and genetically modified Canola. Eco. Appl.15: 871-881.
- Mozafa A (1993). Nitrogen fertilizers and the amount of vitamins in plants: A review J. Plant Nutri.16: 2479-2506.
- Muller HJ (1930). Types of visible variations induced by X-rays in *Drosophila*; J. Gen. 22: 299-334.
- Müller K, He X, Fischer R, Prüfer D (2006). Constitutive knox1 gene expression in dandelion (*Taraxacum Officinale*, Web.) changes leaf morphology from simple to compound. Planta. 224: 1023-1027.
- Nagui H, Fares AK, El-Sayed (1998). Fine structural changes in the ileum of mice fed on endotoxin treated potatoes and transgenic potatoes. Nat. Toxins,. 6: 219-233.
- Nagrare VS, Kranthi S, Biradar VK, Zade NN, Sangode V, Kakde G, Shukla RM, Shivare D, Khadi BM, Kranthi KR (2009). Widespread infestation of the exotic mealy bug species *Phenacoccus solenopsis* (Tinsley) (Hemiptera: Pseudococcidae) on cotton in India. Bull. Ent. Res. 99: 537-541.
- Naranjo SE (2005b). Long-term assessment of the effects of transgenic Bt Cotton on the function of the natural enemy community. Environ. Entomol. 34: 1211-1223.
- Netherwood T, Martin-Orue SG, Gockling S, Graham J, Mathers JC (2004). Assessing the survival of transgenic plant DNA in the human gastrointestinal tract. Nat. Biotech. 22: 204-209.
- Nielsen KM, Bones AM, Smalla K, Van Elsas JD (1998). Horizontal gene transfer from transgenic plants to terrestrial Bacteria- a rare event? FEMS Microbiol. Rev. 22: 79-103.
- Nishino H (1995). Cancer chemoprevention by natural carotenoids and their related compounds., J Cell. Biochem. Suppl. 22: 231-235.

- Nordlee JA (1996). Identification of Brazil-Nut allergen in transgenic soybeans. New England J. Med. 334: 688-692.
- Obrist LB, Klein H, Dutton A, Bigler F (2005). Effects of Bt maize on Frankliniella tenuicornis and exposure of thrips predators to preymediated Bt toxin. Ent. Expt. Appl. 115: 409-416.
- Ochiai K, Yamanaka T, Kimura K, Sawada O (1959). Inheritance of drug resistance (and its transfer) between Shigella strains and Between Shigella and E. coli strains. *Hihon Iji Shimpor*. 34: 1861.
- Olsen KM, Daly JC, Holt HE, Finnega EJ (2005). Season-long variation in expression of cry1Ac gene and efficacy of *Bacillus thuringiensis* toxin in transgenic cotton against *Helicoverpa armigera* ((Lepidoptera: Noctuidae). J. Econ. Ent. 98: 1007-1017.
- Oraby HF, Ransom CB, Kravchenko AN, Sticklen MB (2005). Barley HVA1 gene confers salt tolerance in R3 transgenic oat. Crop Sci. 45: 2218-2227
- Orend G, Knoblauch M, Kammer C, Tjia ST, Schmitz B, Linkwitz A, Meyer G, Maas J, Doerfler W (1995). The initiation of de novo methylation of foreign DNA integrated into a mammalian genome is not exclusively targeted by nucleotide sequence. J. Virol. 69:1226-1242
- Paez KA, Zhao L, Hwang W (2009). Rising out-of-pocket spending for chronic conditions: A ten-year trend. Health Affairs, 28: 15-25.
- Pappinen A, Degefu Y, Syrjala L, Keinonen K (2002). Vanweissenberg, K., Transgenic silver birch (*Betula pendula*) expressing sugar beet chitinase 4 shows enhanced resistance to *Pyrenopeziza betulicola*, Plant. Cell. Rept. 20: 1046-1051.
- Pascal M, Drake W, Barbi T, Drever MR, Van Dolleweerd CJ, Porter AJR, Paul WJ, Jeffrey WH, David AW, De Flaun MF, Cazares LH(1989).Turnover of extracellular DNA in eutrophic and oligotrophic freshwater environments of Southwest Florida. Appl. Environ. Microbiol. 55: 1823-1828.
- Pasonen HL, Seppänen SK, Degefu Y, Rytkönen A, Von Weissenberg K, Pappinen A (2004). Field performance of chitinase transgenic silver birches (*Betula pendula*): resistance to fungal diseases.Theor. Appl. Gene.109: 562-570.
- Peter RJ, Boyd R, Bettinger RL (2001). Was agriculture impossible during the Pleistocene but mandatory during the Holocene? A climate change hypothesis. Am. Antiq. 66: 387-411.
- Pline WA, Viator R, Wilcut JW, Edmisten KL, Thomas JF, Wells R (2002). Reproductive abnormalities in glyphosate-resistant cotton due to lower CP4–EPSPS levels in male reproductive tissue. Weed Sci. 50: 438-477.
- Pilcher CD, Rice ME, Obrycki JJ (2005). Impact of transgenic *Bacillus thuringiensis* corn and crop phenology on five non-target arthropods. Environ. Entomol. 34: 1302-1316.
- Poerschmann J, Rauschen S, Langer U, Augustin J, Górecki T (2008). Molecular level lignin patterns of genetically modified Bt-Maize MON88017 and three conventional varieties using tetramethylammonium hydroxide (TMAH)-induced thermochemolysis. J. Agric. Food Chem. 56: 906-913
- Ponsard S, Gutierrez AP, Mills NJ (2002). Effect of Bt-toxin (Cry1Ac) in transgenic cotton on the adult longevity of four heteropteran predators. Environ. Entomol. 31: 1197-1205.
- Prescott VE, Campbell PM, Moore A, Mattes J, Rothenberf ME, Foster PS, Higgins TJV, Hogan SP (2005). Transgenic expression of bean alpha-amylase inhibitor in peas results in altered structure and immunogenicity. J. Agric. Food Chem. 53: 9023-9030.
- Pusztai A (2002). Can science give us the tools for recognizing possible health risks of GM food. Nutr. Health, 16: 73-84.
- Pusztai A (2005). Facts behind the GM pea controversy: epigenetic, transgenic plants and risk assessment, Proc. Conf., Dec. 1st 2005 (Frankfurtam Main, Germany: Literaturhaus, 2005).
- Pusztai A, Bardocz S, Ewen ŚWB (2003). Genetically Modified Foods: Potential Human Health Effects. In: Food Safety: Contaminants and Toxins (Ed. By JPF D'Mello). CAB International, Wallingford, Oxon, UK. pp. 347-372.
- Qin R, Qiu Y, Cheng Z, Shan Z, Guo X, Zhai H, Wan J (2008). Genetic analysis of a novel dominant rice dwarf mutant 986083D. *Euphytica*,160: 379-387.
- Rachmilewitz D, Katakura K, Karmeli F, Hayashi T, Rains C, Rudensky B, Akira S, Takeda K, Lee J, Takabayashi K, Raz E (2004). Toll-like receptor 9 signaling mediates the anti-inflammatory effects of

- probiotics in murine experimental colitis. Gastroenterology, 126: 520-528.
- Ramesh S, Choimes AS, Schachtman DP (2004). Over-expression of an *Arabidopsis* zinc transporter in *Hordeum vulgare* increases short-term zinc uptake after zinc deprivation and seed zinc content, Plant Mol. Biol. 54: 373-85.
- Rang A, Linke B, Jansen B (2005). Detection of RNA variants transcribed from the transgene in Roundup Ready soybean. Eur. Food Res. Technol. 220: 438-443.
- Ray H, Yu M, Auser P, Blahut-Beatty L, McKersie B, Bowley S, Westcott N, Cullman B, Lloyd A, Gruber MY (2003). Expression of anthocyanins and proanthocyanidins after transformation of alfalfa with maize Lc, Plant Physiol. 132:1448-1463.
- Reed GL, Jensen AS, Riebe J, Head G Duan JJ (2001). Transgenic potato and conventional insecticides for Colorado potato beetle management: comparative efficacy and non-target impacts. Ent. Exp. Appl. 100: 89-100.
- Reynolds M, Foulkes MJ, Slafer GA, Berry P, Parry MAJ, Snape JW, Angus WJ (2009). Raising yield potential in wheat. J. Exp. Bot. 60: 1899-1918.
- Rhee GS, Cho DH, Won YH, Seok JH, Kim SS, Kwack SJ, Lee RD, Cha SY, Kim JW, Lee BM, Park KL, Choi KS (2005). Multigeneration reproductive and developmental toxicity study of bar gene inserted into genetically modified potato on rats. J. Toxicol. Environ. Health A. 68: 2263-2276.
- Riudavets J, Gabarra R, Pons MJ, Messeguer J (2006). Effect of transgenic Bt rice on the survival of three nontarget stored product insect pests. Environ. Entomol. 35:1432-1438.
- Roessner U, Luedemann A, Brust D, Fiehn O, Linke T, Willmitzer L, Fernie AR (2001a). Metabolic profiling allows comprehensive phenotyping of genetically or environmentally modified plant systems. Plant Cell 13: 11-29.
- Rommens CM (2004). All-native DNA transformation: a new approach to plant genetic engineering, Trends Plant Sci. 9: 457-464.
- Rosati A, Bogani P, Santarlasci A, Buiatti M (2008). Characterisation of 3' transgene insertion site and derived mRNAs in MON810 Yield gaurd maize. Plant Mol. Biol. 67: 271-281.
- Sarkar MC (1990). Long term effect of fertilizers on soil ecosystem. Fertil. News, 36: 81-85.
- Sarkar B, Patra AK, Purakayastha TJ, Mallavarapu M (2009). Assessment of biological and biochemical indicators in soil under transgenic Bt and non-Bt cotton crop in a sub-tropical environment. Environ. Monitor Assess. 156: 595-604.
- Saxena D, Stotzky D (2001). *Bt* corn has a higher lignin content than non-*Bt* corn Am. J. Bot. 88:1704-1706.
- Schluter K, Futterer J, Potrykus I (1995). Horizontal gene-transfer from a transgenic potato line to a bacterial pathogen (*Erwinia-chrysanthem*) occurs, if at all, at an extremely low-frequency. Bio.Tech. 13: 1094-1098.
- Schubbert R, Renz D, Schmitz B, Doerfler W (1997). Foreign (M13) DNA ingested by mice reaches peripheral leukocytes, spleen and liver via the intestinal wall mucosa and can be covalently linked to mouse DNA. Proc. Natl. Acad. Sci. USA, 94: 961-966.
- Shadle G, Chen F, Reddy MSS, Jackson L, Nakashima J, Dixon RA (2007). Down-regulation of hydroxycinnamoyl CoA: shikimate hydroxycinnamoyl transferase in transgenic alfalfa affects lignification, development and forage quality. Phytochem, 68: 1521-1529.
- Shewmaker CK, Sheehy JA, Daley M, Colburn S, Ke DY (1999). Seedspecific over expression of phytoene synthase: increase in carotenoids and other metabolic effects. Plant J. 20: 401-412
- Shu QY, Cui HR, Ye, GY, Wu DX, Xia YW, Gao W, Altosaar I (2002). Agronomic and morphological characterization of Agrobacterium-transformed Bt rice plants. *Euphytica*, 127: 345-352.
- Skøt L, Timms E, Mytton L (1994). The Effect of Toxin-Producing Rhizobium Strains on Larvae of Sitona flavescens Feeding on Legume Roots and Nodules. Plant Soil, 163: 141-150
- Saxena D, Flores S, Stotzky G (1999). Insecticidal Toxin in Root Exudates from Bt Corn., Nature, 402: 480.
- Saxena D, Stoztky G (2001). *Bacillus thuringiensis* (Br) Toxin Released from Root Exudates and Biomass of Bt Corn Has No Apparent Effect on Earthworms, Nematodes, Protozoa, Bacteria, and Fungi in Soil.

- Soil Biol. Biochem. 33: 1225-1230.
- Scheffler JA, Parkinson R, Dale PJ (1993). Frequency and distance of pollen dispersal from transgenic oilseed rape (*Brassica napus*). Transgenic Res. 2: 356-364.
- Seralini ĞE (2009). Effects on health and environment of transgenic (or GM) *Bt* brinjal. University of Caen, France, Sci. Council Comm. Ind. Res. and Info. Gen. Eng. (CRIIGEN), p. 19.
- Séralini GE, Cellier D, De Vendomois J (2007). New analysis of a rat feeding study with genetically modified maize reveals signs of hepatorenal toxicity. Arch. Environ. Conta. Toxicol. 52: 596-602
- Shukla VK, Doyon Y,Miller JC, DeKelver RC,Moehle EA,Worden SE, Mitchell JC, Arnold NL, Sunita G, Meng X, Choi VM, Rock JM, Wu Y, Katibah GE, Zhifang G, McCaskill D, Simpson MA, Blakeslee B, Greenwalt SA, Butler HJ,Hinkley SJ, Zhang L, Rebar EJ, Gregory PD, Urnov D (2009). Precise genome modification in the crop species *Zea mays* using zinc-finger nucleases. Nature, 459: 437-441.
- Siciliano SD, Germida JJ (1999). Taxonomic Diversity of Bacteria Associated with the Roots of Field-Grown Transgenic *Brassica napus* cv. Excel and *B. rapa* cv. Parkland., *FEMS* Microbiol. Ecol, 29: 263-272.
- Sisterson MS, Biggs RW, Olson C, Carrière Y, Dennehy TJ, Tabashnik BE (2004). Arthropod abundance and diversity in Bt and non-Bt cotton fields. Environ. Entomol, 33: 921-929
- Smalla K, Wieland G, Buchner A, Zock A, Parzy J, Kaiser S, Roskot N, Heuer H, Berg G (2001). Bulk and rhizosphere soil bacterial communities studied by denaturing gradient gel electrophoresis: Plant-dependent enrichment and seasonal shifts revealed. Appl. Environ. Microbiol, 67: 4742-4751.
- Snow AA, Andersen B, Jorgensen RB (1999). Costs of transgenic herbicide resistance introgressed from Brassica napus into weedy *B. rapa*. Mol. Ecol. 8: 605-615.
- Stanley W, Ewen B, Pusztai A (1999). Effect of diets containing genetically modified potatoes expressing *Galanthus nivalis* lectin on rat small intestine, *Lancet*. 354:1353-1354.
- Sundaramurthy VT (2002). The integrated insect management system and its effects on the environment and productivity of cotton. Outlook. Agric. 31: 95-106.
- Sundaramurthy VT (1994). Upsurgence of whitefly *Bemisia tabacci* Gen. in the cotton eco system in India. Oulook. Agric. 21: 79-83.
- Susana M, Martín-Orúe SM, Anthony G, O'Donnell AG, Ariño J, Trudy Netherwood T, Harry J, Gilbert HJ, Mathers JC (2002). Degradation of transgenic DNA from genetically modified soya and maize in human intestinal simulations. British J. Nutr. 87: 533-542.
- Takagi H, Hirose S, Yasuda H, Takaiwa F (2006). Biochemical safety evaluation of transgenic rice seeds expressing T Cell epitopes of Japanese cedar pollen allergens. J. Agric. Food Chem, 54: 9901-9905.
- Tapp H, Stotzky G (1998). Persistence of the insecticidal Toxin from Bacillus thuringensis suhsp. Kurstaki in Soil. Soil Biol. Biochem. 30: 471-476.
- ThilmonyR,Guttman M, Thomson JG, Blechl AE (2009). The LP2 leucine-rich repeat receptor kinase gene promoter directs organ-specific, light-responsive expression in transgenic rice. Plant Biotech. J. 7: 867-882.
- Tomov BW, Bernal JS, Vinson SB (2003). Impacts of transgenic sugarcane expressing GNA lectin on parasitism of Mexican rice borer by *Parallorhogas pyralophagus* (Marsh) (Hymenoptera: Braconidae). Environ. Entomol. 32: 866-872
- Trethewey RN, Geigenberger P, Riedel K, Hajirezaei MR, Sonnewald U, Stitt M, Riesmeie RJW, Willmitzer L (1998). Combined expression of glucokinase and invertase in potato tubers leads to a dramatic reduction in starch accumulation and a stimulation of glycolysis. Plant J. 15:109-118
- Tudisco R, Lombardi P, Bovera F, D'Angelo D, Cutrignelli MI, Mastellone V, Terzi V, AvalloeL, Infascelli F (2006). Genetically Modified Soya Bean in Rabbit Feeding: Detection of DNA Fragments and Evaluation of Metabolic Effects by Enzymatic Analysis. Anim. Sci. 82: 193-199.
- Turrini A, Shrana C, Pitto L, Castiglione RM, Giorgetti L, Briganti R, Bracci T, Evangelista M, Nuti MP, Giovannetti M (2004). The Antifungal Dm-AMP1 Protein from *Dahlia merckii* Lehm. Expressed in *Solanum melongena* is released in Root Exudates and Differentially

- Affects Pathogenic Fungi and Mycurrhizal Symbiosis. New Phytu. 163: 393-403.
- Ute V, Anja RS, Cesare G, Silvia D (2010). Comparison between volatile emissions from transgenic apples and from two representative classically bred apple cultivars. Transgenic Res. 19: 77-89.
- Vaeck M, Reynaerts A, Höfte H, Jansens S, De Beuckeleer M, Dean C, Marc ZM, Montagu V, Leemans J (1987). Transgenic plants protected from insect attack. Nature, 328: 33-37
- Van Engelsdorp D, Meixner MD (2010). A historical review of managed honey bee populations in Europe and the United States and the factors that may affect them. J. Invert. Pathol, 103: 80-95.
- Vecchio LB, Cisterna B, Malatesta M, Martin TE, Biggiogera M (2004).

 Ultrastructural Analysis of Testes from Mice Fed on Genetically Modified Soybean. Eur. J. Histochem. 48: 449-454.
- Velimirov A, Binter C, Zentek J (2008). Biological effects of transgenic maize NK603 × MON810 fed in long term reproduction studies in mice. Rept. Forschungsberichte der Sektion IV, Band 3.Institut für Ernährung and Forschungs institut für biologischen Landbau, Vienna, Austria, November,
- Veramendi J, Roessner MU, Renz A, Willmitzer L, Trethewey RN (1999). Antisense repression of hexokinase1 leads to an over accumulation of starch in leaves of transgenic potato plants but not significant changes in tuber metabolism. Plant Physiol. 121: 123-134
- Vickers JE, Grof CPL, Bonnett GD, Jackson PA, Morgan TE (2005a). Effects of tissue culture biolistic transformation and introduction of PPO and SPS gene constructs of performance of sugarcane clones. Aust. J. Agric. Res. 56: 57-68.
- Vickers JE, Grof CPL, Bonnett GD, Jackson PA, Knight DP, Roberts SE, Robinson SP (2005b). Over expression of polyphenol oxidase in transgenic sugarcane results in darker juice and raw sugar. Crop Sci. 45: 354-362.
- Vierheilig H, Alt M, Neuhaus JM, Boile T, Wiemken A (1993). Colonization of Transgenic *Nicotiana sylvestris* Plants, Expressing Different Forms of *Nicotiana tabacum* chitinase, by the Root Pathogen *Rhizoctonia solani* and by the mycorrhizal symbiont *Glomus mosseae*. Mol. Plant-Microbe. Interact. 6: 261-264
- Vitta JI, Tuesca D, Puricelli E (2004). Widespread use of glyphosate tolerant soybean and weed community richness in Argentina. Agric. Ecosyst. Environ. 103: 621-624.
- Vojtech E, Meissle M, Poppy GM (2005). Effects of Bt maize on the herbivore *Spodoptera littoralis* (Lepidoptera: Noctuidae) and the parasitoid *Cotesia marginiventris* (Hymenoptera: Braconidae). Transgenic Res. 14: 133-144
- Wang TW, Lu L, Zhang CG, Taylor C, Thompson JE (2003). Pleiotropic effects of suppressing deoxyhypusine synthase expression in Arabidopsis thaliana. Plant Mol Biol. 52: 1223-1235.
- Wang T, Zhang C, Wu W, Nowack L, Madey E, Thompson JE (2005). Antisense Suppression of Deoxyhypusine Synthase in Tomato Delays Fruit Softening and Alters Growth and Development. Plant Physiol. 138: 1372-1382.
- Warwick SI, Simard MJ, Legere A, Beckie HJ, Braun L, Zhu B, Mason P, Seguin-Swartz G,Stewart CN (2003). Hybridization between transgenic *Brassica napus* L. and its wild relatives: *Brassica rapa* L., Raphanus raphanistrum L., Sinapis arvensis L., and Erucastrum gallicum Willd. OE Schulz. Theor. Appl. Gen. 107: 528-539
- Watrud LS, Lee EH, Fairbrother A, Burdick C, Reichmann JR, Bollman M, Storm M, King G, Van de Water P (2004). Evidence for landscape-level, pollen-mediated gene flow from genetically modified creeping bent grass with CP4 EPSPS as a marker. Proc. Natl. Acad. Sci. 101: 14533-14538
- Crick FHC, Watson JD (1953). Molecular Structure of Nucleic Acids- A Structure for Deoxyribose Nucleic Acid. Nature, 171: 737-738
- Weinzierl R, Pierce C, Steffey K (1997). Preliminary Results of the 1997 Summer Survey for Bt-Resistant European Corn Borers. Pest Manage. Crop Dev. Bull. 22: 183-184.
- Whipps JM (1990). Microbial Interactions and Biocontrol in the Rhizosphere. J. Exp. Bot. 52: 487-511
- Whitehouse MEA, Wilson LJ, Fitt GP (2005). A comparison of arthropod communities in transgenic Bt and conventional cotton in Australia. Environ. Entomol. 34: 1224-1241
- Wilson DF, Flint HM, Deaton RW, Fischhoff DA, Perlak FJ, Armstrong

- TA, Fuchs RL, Berberich SA, Parks NJ, Stap BR (1992). Resistance of cotton lines containing a *Bacillus thuringiensis* toxin to pink bollworm (Lepidoptera: Gelechiidae). J. Econ. Ent. 85: 1516-1521
- Wojciede P, Pawlowski P, Somers DA (1998). Transgenic DNA integrated into the oat genome is frequently interspersed by host DNA. Proc.Natl.Acad.Sci. 95: 12106-12110
- Wu W, Ye Q, Min H (2006). Effect of straws from Bt-transgenic rice on selected biological activities in water-flooded soil Environ. Pollut. 143: 449-455
- Wu W, Ye Q, Min H, Duan X, Jin W (2004). Bt transgenic rice straw affects the culturable microbiota and dehydrogenase and phosphatase activities in a flooded paddy soil. Soil Biol. Biochem. 36: 289-295
- Yabor L, Arzola M, Aragón C, Hernández M, Arencibia A, Lorenzo JC (2006). Biochemical side effects of genetic transformation of pineapple. Plant Cell Tissue Organ Cult. 86: 63-67.
- Zemkov RG, Zemek R, Schmidt JEU, Hilbeck A (2005). Altered host plant preference of *Tetranychus urticae* and prey preference of its predator *Phytoseiulus persimilis*(Acari: Tetranychidae, Phytoseiidae) on transgenic Cry3b egg plants. Biol. Control, 33: 293-300.
- Zhang G, Rodrigues L, Rovinski B, White KA (2002). Production of HIV-1 p24 protein in transgenic tobacco plants. Mol. Biotech. 20: 131-136.
- Zhang NY, Linscombe S, Oard J (2003). Out-crossing frequency and genetic analysis of hybrids between transgenic glufosinate herbicideresistant rice and the weed, red rice. Euphytica, 130: 35-45.

- Zhu S, Su J, Liu X, Du L, Yardim EN, Ge F (2006). Development and reproduction of *Propylaea japonica* (Coleoptera: Coccinellidae) raised on *Aphis gossypii* (Homoptera: Aphididae) fed transgenic cotton. Zool. Stud. 45: 98-103.
- Zolla L, Rinalducci S, Antonioli P, Righetti PG (2008). Proteomics as a complementary tool for identifying unintended side effects occurring in transgenic maize seeds as a result of genetic modifications. J. Proteome Res. 7:1850-1861.