

Full Length Research Paper

Effects of ginger (*Zingiber officinale*) on cadmium toxicity

Egwurugwu, J. N.^{1*}, Ufearo, C. S.², Abanobi, O. C.³, Nwokocha, C. R.⁴, Duruibe, J. O.⁵, Adeleye, G. S.⁶, Ebulomo, A. O.⁶, Odetola, A. O.⁶ and Onwufuji, O.⁶

¹Department of Physiology, College of Medicine and Health Sciences, Imo State University, P.M.B. 2000, Owerri Imo State, Nigeria.

²Department of Physiology, Faculty of Medicine, Nnamdi Azikiwe University, P.M.B. 5001 Nnewi Campus, Nigeria.

³Department of Public Health Technology, School of Health Technology, Federal University of Technology, P.M.B. 1536 Owerri, Imo State, Nigeria.

⁴Department of Physiology, Delta State University, Abraka, Delta State, Nigeria.

⁵Department of Chemistry/Biochemistry, School of Industrial and Applied Sciences, Federal Polytechnic Nekede, P.M.B. 1036 Owerri, Imo State, Nigeria.

⁶Department of Physiology, Faculty of Medicine, Madonna University Elele Campus, P.M.B. 5 Elele, Rivers State, Nigeria.

Accepted 27 July, 2007

Thirty six Wistar rats were divided into six equal groups and investigated for induced cadmium toxicity, and the detoxicating action of ginger on liver-accumulated cadmium. Group 1, the control, were fed with normal rat chow and water for six weeks. Group 2 were fed with normal rat chow and cadmium water (200 ppm Cd in water). Group 3 were fed with rat chow-ginger concentrate (95:5, w/w ratio) and water, while Group 4 were fed with rat chow-ginger concentrate and cadmium water, all for six weeks. Group 5 were fed with normal rat chow and cadmium water initially for one week, followed by rat chow-ginger concentrate and water for five weeks; while Group 6 were fed with rat chow-ginger concentrate for one week, followed by normal rat chow and cadmium water for five weeks. Cadmium accumulated highly in rat livers without ginger administration, and raised serum glutamate oxaloacetate transaminase (GOT) and glutamate pyruvate transaminase (GPT), while ginger lowered these parameters. Ginger had better therapeutic than prophylactic detoxication effects on liver cadmium accumulation, especially as further cadmium intake was stopped. It was concluded that cadmium detoxication by ginger was more effective therapeutically, than prophylactically, as further cadmium intake was avoided.

Key words: Bioaccumulation, cadmium, detoxication, ginger, GOT, GPT, Wistar rats.

INTRODUCTION

Cadmium (Cd) is a biotoxic environmental pollutant, which accumulates in the body tissues, such as the lungs, liver, kidneys, bones, reproductive organs and the immune system. Animals are generally tolerant to low doses of Cd exposure, but respond considerably to high lethal doses. Mortality in animals due to Cd toxicity does

not occur due to cardiotoxicity or nephrotoxicity, but rather by liver injury, because the liver accumulates substantial amounts of Cd after both acute and chronic exposures (Klaassen and Liu, 1998), and Cd pre-treatment does not alter its organ distribution.

Cd is released into the environment from both natural and anthropogenic sources, including agricultural activities (Hutton and Symon, 1986; European Union, 2002; USDOL, 2004; Ogwuegbu and Duruibe, 2005; Duruibe et al., 2007). Activities that cause the release of Cd into the soil, causing soil pollution, result in subse-

*Correspondent author. E-mail: feziechi@yahoo.com. Tel: +234 803 711 7341.

quent water pollution (Nriagu and Pacyna, 1988; OECD, 1994; Peplow, 1999). The presence of Cd in agricultural soils from phosphate fertilizers will also result in its increased uptake by plants, accumulating in plant tissues, more especially corns and vegetables (European Union, 2002; André et al., 2005). Human acute and chronic Cd exposures occur through food, air, water, industrial products; and by occupational exposure (Heyer, 1985; Habashi, 1992; Horsfall and Spiff, 1999; Smolters, 2001; USDOL, 2004; Duruibe et al., 2007), and toxicity dysfunctions resulting from Cd ingestion include bone defects, increased blood pressure, myocardic dysfunctions, proteinuria and pulmonary edema. Death of animals may also subsequently occur (Klaassen and Liu, 1998; Telisman et al., 2001; Jarup, 2003; Young, 2005). In an investigation, Cd exposure was shown to be linked to a wide range of mammalian reproductive dysfunctions. Depending on the steroidogenic tissue involved and dosage administered, it enhances or inhibits the synthesis of progesterone; and antenatal exposure results in reduced birth weight and premature birth (Henson and Chedrese, 2004). Another investigation reveals that the bio-effect of Cd on mice depended on dose administered, absorption and distribution in metallothionein-1 transgenic mice (Liu and Klaassen, 1996). Malgorzata (1998) reported that Cd exposure to fishes resulted in 40% mortality 96 h after the end of the exposure due to disturbances in physiological functions in the fishes. Defects due to acute Cd exposure via food are very unusual, while chronic defects are more frequent (Satarug et al., 2004).

Ginger (*Zingiber officinale*) is commonly used as food spice in many Asian and African countries, including Nigeria. It contains a host of compounds, which include acid resins, vitamin C compounds (folic acid, inositol, choline and panthothenic acid), gingerol, sesquiterpene, vitamins B₃ and B₆, volatile oils, and bio-trace elements (Ca, Mg, P and K). The pungency of ginger is due to gingerol, while its aroma is due to volatile oils, which are bisabolone, zingiberene and zingiberol.

The medicinal values of ginger have been intensively reported. Ginger contains Mg, Ca and P, which play important roles in bone formation, and curbing muscle spasm, depression hypertension, convulsion, nausea, gastrointestinal disorders, paralysis, kidney damage, and a host of other biodysfunctions (Lee and Ahn, 1985; Kikuzaki and Nakatani, 1993; Kikuzaki et al., 1994; Meyer et al., 1995). Ginger extracts exhibit anticholinergic and antihistaminic effects (Qian and Liu, 1992), antihypercholesterolemic effect (Janabai et al., 1984; Tanabe et al., 1993), antihyperlipidaemic effect (Bhandari et al., 1998), antiinflammatory effect (Al-Yahya et al., 1989), antiemetic effect (Philips et al., 1993) and lowers the serum glutamate oxaloacetate transaminase (GOT) and glutamate pyruvate transaminase (GPT) levels (Bhandari et al., 2003). Ginger is very useful in the treatment of migraine, motion sickness (Mowrey and Clayson, 1982;

Holtman et al., 1989), and rheumatic disorders (Srivastava and Mustafa, 1989, 1992). It is also an anti-tumor, anticarcinogenic and antitoxic agent (Mascolo et al., 1989; Katiyar et al., 1996; Surh, 1999; Vimala et al., 1999).

Our purpose for carrying out this research was to investigate on the effects of ginger on Cd toxicity, using rats as test specimens. This was done by creating an induced Cd toxicity in the rats, by feeding them with Cd water containing 200 ppm Cd concentration, followed by rat chow mixed with 5% (w/w) ginger. After the test period, the extent of accumulation of Cd in the liver and the antidote effects of ginger on Cd poisoning was evaluated. Literature survey shows different treatment methods for Cd toxicity such as increased intake of Zn, Se, Cu and Ge (Pizent et al., 2001; Paolo et al., 2005), which also act as metallic antioxidant (Yinn and Lin, 1998), and the use of dihydroxyethylthiocarbamate (DHDC), diethylthiocarbamate (DEDC), and dicarboxymethylthiocarbamate (DCDC) in mobilizing metallothionein-bound Cd from some organs and tissues of mice, and promoting its excretion (Gale et al., 1983a,b). However, the use of ginger will be a good viable option since it is a natural plant, which serves as spice for food, hence can be used as food additive, while also detoxicating the body tissues of Cd.

EXPERIMENTAL

Ginger was ground and sieved to a particle size of 250 µm. The rat chow – ginger concentrate (5% w/w of ginger in rat chow) was prepared by mixing normal chow and ginger at 95:5 w/w ratio and stored in a dessicator, while Cd – water concentrate (Cd-H₂O) was prepared at 200 ppm Cd concentration in water (1 g CdCl₂ in 5 litre of H₂O).

The specimens (thirty six (36) Winstar rats), weighing about 180 g each, were randomly divided into six equal groups and labeled as Groups 1, 2, 3, 4, 5 and 6, and confined in iron cages. They were allowed a 2-week period to acclimatize with their new environment, after which the experiment proceeded for 6 weeks. Group 1 served as the control, which were fed with normal rat chow and water for 6 weeks. Group 2 were fed with normal rat chow and Cd-H₂O; Group 3 were fed with rat chow – ginger concentrate and water; and Group 4 were fed with rat chow-ginger concentrate and Cd-H₂O, all for 6 weeks. Group 5 were fed with normal rat chow and Cd-H₂O for the first week *ab initio*, then with rat chow – ginger concentrate and water from the second week to the sixth week; whereas Group 6 were fed with rat chow – ginger concentrate for one week, then normal rat chow and Cd-H₂O for the remaining 5 weeks. The grouping and feeding patterns are summarized in Table 1. All administrations were through the oral route.

Enzyme and heavy metal analyses were conducted on the specimens at two-week interval, and two specimens were harvested from each group for each set of analyses. Blood samples were collected from the specimens, from which serum was extracted after coagulation for GPT and GOT analyses. Liver was harvested from dissected specimen and homogenized, and the supernatant solution was extracted for Cd analysis (Gale et al., 1983a,b). Cd was analyzed by atomic absorption spectrophotometer (AAS, UNICAM 919), while GPT and GOT analyses were done according to Reitman-Frankel method (Reitman and Frankel, 1957).

Table 1. Summary of specimen grouping and six weeks feeding pattern.

Week	Groups					
	1*	2	3	4	5	6
1	F + W	F + W _{Cd}	F _g + W	F _g + W _{Cd}	F + W _{Cd}	F _g + W
2	F + W	F + W _{Cd}	F _g + W	F _g + W _{Cd}	F _g + W	F + W _{Cd}
3	F + W	F + W _{Cd}	F _g + W	F _g + W _{Cd}	F _g + W	F + W _{Cd}
4	F + W	F + W _{Cd}	F _g + W	F _g + W _{Cd}	F _g + W	F + W _{Cd}
5	F + W	F + W _{Cd}	F _g + W	F _g + W _{Cd}	F _g + W	F + W _{Cd}
6	F + W	F + W _{Cd}	F _g + W	F _g + W _{Cd}	F _g + W	F + W _{Cd}

* = Control; F = feed (rat chow); W = water; F_g = feed-ginger concentrate; W_{Cd} = Cd-H₂O (200 ppm Cd in water).

Table 2. Effects of Cd and/or ginger on serum GPT.

Week	Serum GPT concentrations (units/l)					
	Group 1*	Group 2	Group 3	Group 4	Group 5	Group 6
2	4.00	2.67	4.67	4.11	2.88	3.92
4	3.96	2.72	3.47	3.55	2.89	2.38
6	3.88	2.88	1.92	2.42	2.88	0.94

* = Control.

Table 3. Effects of Cd and/or ginger on serum GOT.

Week	Serum GOT concentrations (units/l)					
	Group 1*	Group 2	Group 3	Group 4	Group 5	Group 6
2	4.15	3.19	8.56	3.71	2.83	3.77
4	4.02	3.96	6.89	4.00	3.34	3.45
6	3.86	4.91	4.91	4.34	3.91	3.17

* = Control.

Table 4. Results of Cd concentration in the Liver

Week	Liver Cd concentration (ppm or mg/l)					
	Group 1*	Group 2	Group 3	Group 4	Group 5	Group 6
2	ND	8.59	ND	3.24	1.95	3.24
4	ND	8.38	ND	3.24	2.09	3.84
6	ND	12.05	ND	5.41	1.46	7.81

* = Control; ND = not detected.

RESULTS AND DISCUSSION

The results of the various analyses are summarized in Tables 2 - 4, and reported figures are averages of four readings; two readings for each parameter from each of the two rat specimens. While the GPT and GOT values for the control slightly decreased within the period, Group 2 specimens showed increasing GPT and GOT. Groups 3 and 6 showed decreasing GOT and GPT, with a

steeper decreasing trend for Group 3 GOT. While GOT values increased for Groups 4 and 5, GPT values decreased for Group 4, but remained relatively constant throughout the test period for Group 5. Serum GPT and GOT levels increased with Cd administration without ginger (Group 2), and decreased with ginger administration without Cd (Group 3). Hence while Cd raises serum GPT and GOT levels, ginger lowers these parameters. Results for the simultaneous administration of Cd

and ginger to Group 4 specimens showed a dominating effect of ginger on GPT and Cd on GOT. The initial administration of Cd to Group 5 specimens, followed by ginger after the first week for toxicity therapeutic effects, showed no response on GPT, but serum GOT levels increased. Whereas the initial administration of ginger for Cd toxicity prophylaxis on Group 6 followed by Cd administration lowered both serum GPT and GOT.

AAS results showed that Cd concentration in the liver increased tremendously for Group 2 specimens, but remained constant for the first 4 weeks for Group 4, with values still relatively very low after 6 weeks compared with those of Group 2. This is attributable to the detoxication effect of ginger. In Group 5, liver Cd concentration had lowest values; also, this is attributable to the therapeutic detoxication effects of Cd on ginger, thereby showing better effects than the prophylactic effects of ginger for Group 6. The prophylactic effect of ginger on Cd toxicity in Group 6 specimens was effective for the first 4 weeks; after which the values of liver Cd concentration soared high. Thus, it is suggestive that the detoxication of Cd by ginger cannot be effectively achieved by the prophylactic administration of ginger, like medical vaccination.

Conclusion

Overall, the results of these investigations showed that both ginger and Cd altered serum GPT and GOT levels. Ginger had both prophylactic and therapeutic Cd detoxication effects on the specimens, but ginger therapy was more effective as more Cd intake was avoided. Further research can be conducted employing longer durations of induced Cd poisoning and ginger administration and also using lactating rats. Other body organs, such as kidney, heart, lungs, as well as breast milk can be analyzed for Cd accumulation and the prophylactic and therapeutic effects of ginger or another known viable antidote.

REFERENCES

- Al-Yahya MA, Rafatulla S, Mossa JS (1989). Gastro-Protective Activity of Ginger on Albino Rats, *Am. J. Chi. Med.* 17: 51-56.
- Andre LO, Paolo RG, Silvana CJ, Josino CM (2005). Dietary Intake and Health Effects of Selected Toxic Elements, *Braz. J. Plant Physiol.* 17(1): 79-93.
- Bhandari U, Sharma JN, Zafar R (1998). The Protective Action of Ethanolic Ginger (*Zingiber officinale*) Extract in Cholesterol-Fed Rabbits, *J. Ethnopharmacol.* 61(2): 167-171.
- Bhandari U, Shamsher AA, Pillai KK, Khan MSY (2003). Antihepatotoxic Activity of Ginger Ethanol Extract in Rats. *Pharm. Biol.* 41(1): 68-71.
- Duruibe JO, Ogwuegbu MOC, Egwurugwu JN (2007). Heavy Metal Pollution and Human Biotoxic Effects, *Int. J. Phys. Sci.* 2(5): 112-118.
- European Union (2002). Heavy Metals in Wastes, European Commission on Environment (http://ec.europa.eu/environment/waste/studies/pdf/heavy_metalsrep_ort.pdf).
- Gale GR, Atkins LM, Walker EM, Smith AB (1983). Effects of Combined Treatment with Diethyldithiocarbamate and Diethylenetriaminepentaacetate on Organ Distribution and Excretion of Cadmium, *Ann. Clin. Lab. Sci.* 13(5): 424-431.
- Gale GR, Atkins LM, Walker EM, Smith AB, Jones MM (1983). Mechanism of Diethyldithiocarbamate Dihydroxyethylthiocarbamate and Dicarboxymethylthiocarbamate Action of Distribution and Excretion of Cadmium, *Ann. Clin. Lab. Sci.* 13(6): 474-481.
- Habashi F (1992). Environmental Issues in the Metallurgical Industry – Progress and Problems, *Environmental Issues and Waste Management in Energy and Mineral Production*, Balkema, Rotterdam: 1143-1153.
- Henson MC, Chedrese PJ (2004). Endocrine Disruption by Cadmium; a Common Environmental Toxicant with Paradoxical Effects on Reproduction, *Exp. Biol. Med.*, 299: 383-392.
- Heyer NJ (1985). Neurological Disorder in Three Aluminium Smelter Workers, *Arch. Int. Med.*, 145(11): 1972-1975.
- Holtman S, Clarke AH, Sherer H, John M (1989). The Anti-Motion Sickness Mechanism of Ginger; a Comparative Study with Placebo, *Acta Otolaryngol.* 108(3-4): 168-174.
- Horsfall MN, Spiff AI (1999). Speciation of Heavy Metals in Intertidal Sediments of the Okirika River System (Nigeria), *Bull. Chem. Soc. Ethiop.*, 13(1): 1-9.
- Hutton M, Symon C (1986). The Quantities of Lead, Cadmium Mercury and Arsenic Entering the UK Environment from Human Activities, *Sci. Total Environ.* 57: 129-150.
- Janabai G, Sakthi DT, Meeraran S (1984). Effect of Ginger on Serum Cholesterol Levels, *Ind. J. Nutr. Dietet.* 21(12): 433-436.
- Jarup L (2003). Hazards of Heavy Metal Contamination, *Brit. Med. Bull.*, 68: 167-182.
- Katiyar SK, Agarwal R, Mukhtar H (1996). Inhibition of Tumor Promotion in SENCAR Mouse Skin by Ethanol Extract of *Zingiber officinale* Rhizome, *Cancer Res.* 56(5): 1023-1030.
- Kikuzaki H, Nakatani N (1993). Antioxidant Effects of Ginger Constituents, *J. Food Sci.* 58(6): 1407-1410.
- Kikuzaki H, Kawasaki Y, Nakatani N (1994). Structure of Antioxidative Compounds of Ginger, *ACS Symp. Ser.* 547: 237-243.
- Klaassen CD, Liu J (1998). Induction of Metallothionein as an Adaptive Mechanism Affecting the Magnitude and Progression of Toxicological Injury, *Environ. Health Perspect.*, 106(Suppl. 1): 297-300.
- Lee IK, Ahn SY (1985). The Antioxidant Activity of Gingerol, *Korean J. Food Sci. Technol.* 17(2): 55-59.
- Liu J, Klaassen CD (1996). Absorption and Distribution of Cadmium in Metallothionein-1 Transgenic Mice, *Fundamental Appl. Toxicol.* 29: 294-300.
- Malgorzata W (1998). Changes in Selected Blood Indices of Common Carp after Exposure to Cadmium, *Acta. Vet. Brno.*, 67: 289-293.
- Mascolo N, Jain R, Jain SC, Capasso F (1989). Ethnopharmacological Investigation of Ginger (*Zingiber officinale*), *J. Ethnopharmacol.* 27(1-2): 129-140.
- Meyer K, Schwartz J, Crater D, Keyes B (1995). *Zingiber officinale* (Ginger) Used to Prevent 8-MOP Associated Nausea, *Dermatol. Nurs.*, 74(4): 242-244.
- Mowrey DB, Clayson DE (1982). Motion Sickness, Ginger and Psychophysics, *Lancet*, 1(8273): 655-657.
- Nriagu JO, Pacyna J (1988). Quantitative Assessment of Worldwide Contamination of Air, Water and Soil by Trace Metals, *Nature*, 333: 134-139.
- Ogwuegbu MOC, Duruibe JO (2005). Environmental Pollution and Biotoxicity of Certain Heavy Metals. In: *International Conference of the Chemical Society of Nigeria; Chemistry and the Nigerian Economy*, Yewa, Maiduguri, Nigeria, pp. 285-288.
- Organization of Economic Co-operation and Development, OECD (1994). Cadmium, Risk Reduction Monograph, Environmental Directorate, Paris, France: 5.
- Paolo B, Luca DG, Niu Q, Marcella R, Castellani ML, Isabella L, Paola T, Kouri M, Nichola V, Volpe RA, Marco C, Roberto P, Mario DG (2005). Inhibitory Effects of Cadmium on Peripheral Blood Mononuclear Cell Proliferation and Cytokine Release are Reversed by Zinc and Selenium Salts, *Ann. Clin. Lab. Sci.* 35: 115-129.
- Peplow D (1999). Environmental Impact of Mining in Eastern Washington, Center for Water and Watershed Studies Fact Sheet, University of Washington, Seattle.
- Philips S, Ruggier R, Hutchinson SE (1993). *Zingiber officinale* (Ginger)

- An Antiemetic for Day Case Surgery, *Anesthesia*, 48(8): 715-717.
- Pizent A, Jurasovic A, Telisman S (2001). Blood Pressure in Relation to Dietary Cadmium Intake, Alcohol Consumption, Blood Lead and Blood Cadmium in Female Non- Smokers, *J. Trace Elem. Med. Biol.*, 15: 123-130.
- Qian DS, Liu ZS (1992). Pharmacological Studies of Antimotion Sickness of Ginger, *Chung Kuo Chung Hsi I Chieh Ho Tsa Chih.*, 12(2): 95-98.
- Reitman S, Frankel S (1957). A Colorimetric Method for the Determination of Serum Glutamic Oxalacetic and Glutamic Pyruvic Transaminases, *Am. J. Clin. Pathol.*, 28: 56-63.
- Satarug S, Ujin P, Vanavanitkun Y, Baker JR, Moore JR (2004). Influence of Body Iron-Ore Status and Cigarette Smoking on Cadmium Body Burden of Healthy Thai Women and Men, *Toxicol. Lett.* 148: 177-185.
- Smolders E (2001). Cadmium Uptake by Plants, *Int. J. Occup. Med. Environ. Health*, 14: 177-183.
- Srivastava KC, Mustafa T (1989). Ginger (*Zingiber officinale*) and Rheumatic Disorders, *Med. Hypoth.* 1: 25-28.
- Srivastava KC, Mustafa T (1992). Ginger (*Zingiber officinale*) in Rheumatic and Musculoskeletal Disorders, *Med. Hypoth.*, 4: 342-348.
- Surh Y (1999). Molecular Mechanisms of Chemopreventive Effects of Selected Dietary and Medicinal Phenolic Substances, *Mutat. Res.*, 428(1-2): 305-327.
- Tanabe M, Chen YD, Saito KI, Kano Y (1993). Cholesterol Biosynthesis Inhibitory Component from *Zingiber officinale* Roscoe, *Chem. Pharm. Bull.*, 41(4): 710-713.
- Telisman S, Jurasovic J, Pizent A, Cvitkovic P (2001). Blood Pressure in Relation to Biomarkers of Lead Cadmium, Copper, Zinc and Selenium in Men without Occupational Exposure to Metals, *Environ. Res.* 87: 57-68.
- United States Department of Labor (2004). Occupational Safety and Health Administration (OSHA); Safety and Health Topics: Heavy Metals, USDOL Washington DC (www.osha.gov/SLTC/metalsheavy/index.html).
- Vimala S, Norhanom AW, Yadav M (1999). Anti-Tumor Promoter Activity in Malaysian Ginger Rhizome Used in Traditional Medicine, *J. Cancer* 80(1-2): 110-116.
- Yinn SJ, Lin TH (1998). Effects of Metallic Antioxidants on Cadmium-Catalyzed Peroxidation of Arachidonic Acid, *Ann. Clin. Laborat. Sci.* 28(1): 43-50.
- Young RA (2005). Toxicity Profiles: Toxicity Summary for Cadmium, Risk Assessment Information System, RAIS, University of Tennessee (www.rais.ornl.gov/tox/profiles/cadmium.shtml).