Review

Recent advances in pretreatment of lignocellulosic wastes and production of value added products

Godliving Y. S. Mtui

Department of Molecular Biology and Biotechnology, University of Dar es Salaam, P. O. Box 35179, Dar es Salaam, Tanzania. E-mail: gmtui@amu.udsm.ac.tz.

Accepted 13 February, 2009

This study highlights the recent advances in the treatment and value addition of lignocellulosic wastes (LCW) with main focus on domestic and agro-industrial residues. Mechanical, physical and biological treatment systems are brought into perspective. The main value-added products from lignocellulosic wastes are summarized in a manner that pinpoints the most recent trends and the future directions. Physicochemical and biological treatment systems seem to be the most favored options while biofuels, biodegradable composites and biosorbents production paints a bright picture of the current and future bio-based products. Engineered microbes seem to tackle the problem of bioconversion of substrates that are otherwise non convertible by conventional wild strains. Although the main challenge facing LCW utilization is the high costs involved in treatment and production processes, some recent affordable processes with promising results have been proposed. Future trends are being directed to nanobiotechnology and genetic engineering for improved processes and products. The paper presents state of the art review of the dual advantage of handling LCW for cleaner environment and production of renewable bio-products.

Key words: Lignocellulosic wastes, pretreatment systems, value-added products.

INTRODUCTION

Lignocelluloses wastes (LCW) refer to plant biomass wastes that are composed of cellulose, hemicellulose, and lignin. They may be grouped into different categories such as wood residues (including sawdust and paper mill discards), grasses, waste paper, agricultural residues (including straw, stover, peelings, cobs, stalks, nutshells, non food seeds, bagasse, domestic wastes (lignocellulose garbage and sewage), food industry residues, municipal solid wastes and the like (Qi et al., 2005; Roig et al., 2006; Rodríguez et al., 2008). Currently, the second generation bio-products such as bioethanol, biodiesel, biohydrogen and methane from lignocellulose biomass are increasingly been produced from wastes rather than from energy crops (jatropha, switchgrass, hybrid poplar and willow) because the latter competes for land and water with food crops that are already in high demand. The use of food crops such as corn and sugarcane to produce biofuels is increasingly being discouraged due to the current worldwide rise in food prices. In order to minimize food-feed-fuel conflicts, it is necessary to integrate all kinds of biowaste into a biomass economy (Mahro and Timm, 2007). Furthermore, the use of LCW offers a possibility of geographically distributed and greenhouse-gas-favourable sources of products (Rubin, 2008).

The lignocellulosic biomass, which represent the largest renewable reservoir of potentially fermentable carbohydrates on earth (Mtui and Nakamura, 2005), is mostly wasted in the form of pre-harvest and post-harvest agricultural losses and wastes of food processing industries. Due to their abundance and renewability, there has been a great deal of interest in utilizing LCW for the production and recovery of many value-added products (Pandey et al., 2000; Das and Singh, 2004; Foyle et al., 2007). Among the main recovery products include enzymes, reducing sugars, furfural, ethanol, protein and amino acids, carbohydrates, lipids, organic acids, phenols, activated carbon, degradable plastic composites, cosmetics, biosorbent, resins, medicines, foods and feeds, methane, biopesticides, biopromoters, secondary metabolites, surfactants, fertilizer and other miscellaneous products (Tengerdy and Szakacs, 2003; Mtui, 2007; Ubalua, 2007; Galbe and Zacchi, 2007; Demirbas, 2008). Alongside producing these products, the processes also remove wastes from the environment.

The barrier to the production and recovery of valuable materials from LCW is the structure of lignocellulose which has evolved to resist degradation due to crosslinking between the polysaccharides (cellulose and hemicellulose) and the lignin via ester and ether linkages (Yan and Shuya, 2006; Xiao et al., 2007). Cellulose, hemicellulose and lignin form structures called microfibrils, which are organized into microfibrils that mediate structural stability in the plant cell (Rubin, 2008). The main goal of any pretreatment, therefore, is to alter or remove structural and compositional impediments to hydrolysis and subse-quent degradation processes in order to enhance digestibility, improve the rate of enzyme hydrolysis and increase yields of intended products (Mosier et al., 2005; Hendriks and Zeeman, 2009). These methods cause mechanical, physical chemical or biological changes in the plant biomass in order to achieve the desired products.

Technology of LCW bioconversion has long been considered to be rather expensive. However, recent increases in grain prices mean that the switch to second generation bio-products such as biofuels from LCW will reduce competition with grain for food and feed, and allow the utilization of materials like straw which would otherwise go to waste. Technologies that will allow costeffective conversion of biomass into fuels and chemicals consider economy of scale, low-cost pretreatment systems and highly effective and efficient biocatalysts (Schneider and McCar, 2003; Gray et al., 2006).

This work reviews the recent developments in LCW pretreatment, value addition and techno-economic considerations.

PRETREATMENT TECHNOLOGIES FOR LIGNOCELLULOSIC WASTES

Mechanical pretreatment

Mechanically based pretreatment technologies are aimed at reducing the size of LCW to facilitate subsequent treatments. Reduction of biomass size below #20 sieves shows the best mechanical performance (de Sousa et al., 2004). Mechanical pretreatment technologies increase the digestibility of cellulose and hemicellulose in the lignocellulosic biomass. The use of mechanical chopping (de Sousa et al., 2004); hammer milling (Iñiguez-Covarrubias et al., 2001; Mani et al., 2004); grind milling (Mtui and Nakamura, 2005); roll milling (Qi et al., 2005); vibratory milling (Guerra et al., 2006) and ball milling (Inoue et al., 2008) have proved success as a low cost pretreatment strategy. The pulverized materials with increased surface area have been found to facilitate the subsequent physicochemical and biochemical pretreatments of corn stover, barley straw sugar cane baggase, wheat straw, wood waste and municipal solid waste. They result to improved digestibility of cellulose and hemicellulose to glucan and xylan, respectively; they further enhance enzymatic digestibility with lower enzyme loads. Mechanical pretreatment also result to substantial lignin depolymerization via the cleavage of uncondensed-aryl ether linkages (Inoeu et al., 2008). Solubility and fermentation efficiency of the natural lignocellulosic residues is also substantially increased by mechanophysicochemical pretreatment, leading to value-added utilization of these residues (Qi et al., 2005).

Physical pretreatment

Elevated temperatures and irradiation are the most successful physical treatments in the processing of LCW. Thermogravimetric treatment of wood waste under both inert and oxidant atmospheres from room temperature up to 1100 K leads to moisture loss; hemicellulose, cellulose and lignin decomposition (Lapuerta et al., 2004). On the other hand, pyrolysis of nutshells, straws, sawdust and municipal solid wastes at temperatures of 600 - 1200 K result to yields of char, liquid and gaseous products of up to 55% of the original LSW (Puértolas et al., 2001; Demirbas, 2002; Bonelli, 2003; Chen et al., 2003; Álvarez et al., 2005; Phan et al., 2008; Zabaniotou et al., 2008).

Irradiation can cause significant breakdown of the structure of LSW. Microwave irradiation at a power of up to 700 W at various exposure times resulted to weight loss due to degradation of cellulose, hemicellulose and lignin, and the degradation rates are significantly enhanced by the presence of alkali (Zhu et al., 2005a, 2005b, 2006). In addition, gamma radiation has been shown by Yang et al. (2008) to cause significant breakdown of the structure of powder of 140 mesh wheat straw, leading to weight loss and glucose yield of 13.40% at 500 kGy.

Physicochemical pretreatment

Combined chemical and physical treatment systems are of importance in dissolving hemicellulose and alteration of lignin structure, providing an improved accessibility of the cellulose for hydrolytic enzymes (Hendriks and Zeeman, 2009). The most successful physicochemical preatments include thermochemical treatments such as steam explosion or (steam disruption), liquid hot water (LHW), ammonia fiber explosion (AFEX) and CO₂ explosion (Sun and Cheng, 2002). In these processes, chipped biomass is treated with high-pressure saturated steam, liquid ammonia or CO₂ and then the pressure is swiftly reduced, making the materials to undergo an explosive decompression.

Steam explosion is typically initiated at a temperature of $160 - 260 \,^{\circ}\text{C}$ (corresponding pressure of 0.69 - 4.83MPa) for several seconds to a few minutes before the material is exposed to atmospheric pressure. The processes cause hemicellulose degradation and lignin transformation due to high temperature, thus increasing the potential of cellulose hydrolysis. Addition of H₂SO₄ (or SO_2) or CO_2 in steam explosion of LCW can effectively improve enzymatic hydrolysis, decrease the production of inhibitory compounds, and lead to more complete liquefaction of hemicellulose, glucan, xylan, mannan, galactan, and arabinan (Jeoh and Agblevor, 2001; Sun and Cheng, 2002). Such pretreatments also lead to higher digestion efficiencies during production of monosaccharides, oligosaccharides, lactic acid, antibacterial violet pigments and methane gas (Liu et al., 2002; Kim et al., 2003; Asada et al., 2005; Wang and Chen, 2007; Öhgren et al., 2007). Wet oxidation pretreatment at 200 - 210°C in the presence of alkali or Na₂CO₃ leads to LCW solubilization and better enzymatic convertibility to value-added products (Fox and Noike, 2004; Lissens et al., 2004; Martín et al., 2008).

Liquid hot water (LHW) pretreatment utilizes pressurized hot water at pressure less than 5 Mpa and temperature range of 170 - 230 °C for several minutes followed by decompression up to atmospheric pressure. Bagasse, corn stalk and straws of wheat, rice and barley pretreated by LHW have been reported to effect 80 - 100% hemicellulose hydrolysis, resulting to 45 - 65% xylose (Sun and Cheng, 2002; Sánchez and Cardona, 2008).

On the other hand, in AFEX treatment, the dosage of liquid ammonia ranging from 1 - 2 kg ammonia/kg dry biomass, temperature 90 °C, and residence time of 30 min can significantly improve the saccharification rates (Chundawat et al., 2007; Thomsen and Belinda, 2007). On CO₂ explosion, 75% of the theoretical glucose released during 24 h of the enzymatic hydrolysis has been reported (Sun and Cheng, 2002). Ethanol yield of up to 83% of the theoretical value has been achieved for LCW subjected to physicochemical treatment (Jeoh and Agblevor, 2001).

Chemical pretreatment

Chemicals ranging from oxidizing agents, alkali, acids and salts can be used to degrade lignin, hemicellulose and cellulose from LCW. Poweful oxidizing agents such as ozone and H₂O₂ effectively remove lignin; does not produce toxic residues for the downstream processes; and the reactions are carried out at room temperature and pressure (Sun and Cheng, 2002). Alkali (NaOH, Ca(OH)₂, NaOH-urea, Na₂CO₃) hydrolyses of rice straw (Carrillo et al., 2005); spruce wood waste (Zhao et al., 2007); sugarcane, cassava and peanuts wastes (Thomsen and Belinda, 2007); corn cob (Torre et al., 2008); organic fraction of municipal solid waste (Torres and Lloréns, 2008) have been investigated. When these pretreatments are performed by using 0.5 - 2 M alkali at 120 - 200°C, they substantially facilitate saccharification and improve enzymic hydrolysis of LCW.

Dilute and concentrated acids at high temperature are suited for hydrolysis of LCW. Studies by del Campo et al. (2006) and Karimi et al. (2006) have established that 0.5% H₂SO₄ is optimal for treatment of wastes from vege-

tables and rice straw, respectively. More concentrated H_2SO_4 (up to 2.5 M) has been shown to be able not only to hydrolyse cellulose and hemicellulose, but also in separating lignin and other organic components from LCW (Iranmahboo et al., 2002; Alma and Acemioglu, 2004; Okafoagu and Nzelibe, 2006; Miller et al., 2007; Rahmanet al., 2007). SO₂ and fly ash in flare gas; HNO₃, HCI and polyhydric alcohol in the presence of sulfuric acid are also useful in LCW pretreatment (Fan, 2003; Herrera et al., 2004; Kobayashi et al., 2004; Rodríguez-Chonga et al., 2004; Hassan and Shukry, 2008): Recent studies have shown that when acids are combined with alkali, they play a more effective role in LCW pretreatment than acids and alkalis alone (Damisa et al., 2008).

Organic acids such as oxalic, acetylsalicylic and salicylic acid can be used as catalysts in the *organosolv* process whereby an organic or aqueous organic solvent mixture with inorganic acids (HCl or H_2SO_4) are used to break the internal lignin and hemicellulose bonds. The organic solvents used in the process include methanol, ethanol, acetone, ethylene glycol, triethylene glycol and tetrahydrofurfuryl alcohol (Sun and Cheng, 2002). The use of a dicarboxylic acid catalyst, maleic acid, for hemicellulose hydrolysis in corn stover overcomes the technical and economic hurdle of hemicellulose hydrolysis (Lu and Mosier, 2007).

Biological pretreatment

Biological treatment involves the use of whole organisms or enzymes in pretreatment of LCW. Both fungi and bacteria are used for biotreatment of LCW. Commercial preparations of fungal and bacterial hydrolytic and oxidative enzymes are also widely used instead of these microorganisms.

Fungal pretreatment of agricultural residues is a new method for improvement of digestibility (Sinegani et al., 2005). White-, brown- and soft-rot fungi are used to degrade lignin and hemicellulose in waste materials whereby brown rots mainly attack cellulose, while white and soft rots attack both cellulose and lignin. White-rot fungi are the most effective basidiomycetes for biological pretreatment of lignocellulosic materials (Sun and Cheng, 2002). Recent studies have shown that Aspergillus terreus (Emtiazi et al., 2001); Trichoderma spp (Pérez et al., 2002); Cyathus stercoreus (Keller et al., 2003); Lentinus squarrosulus (Shide et al., 2004); Lentinus edodes (Songulashvili et al., 2005; Brienzo et al., 2007); Trametes pubescens (Melamane et al., 2007); Pleurotus spp (Ragunathan and Swaminathan, 2004; Mukherjee and Nandi, 2004; Belewu, 2006; Locci et al., 2008); Penicillium camemberti (Taşeli, 2008), Phanerochaete chrysosporium (Das and Hossain, 2000; Shi et al., 2008) grown at 25 - 35 ℃ for 3 - 22 days resulted to 45 - 75% and 65 - 80% holocellulose and lignin degradation, respectively. The postreatement by anaerobic bioprocesses of LCW effluents that have been pretreated with fungi can lead to higher biogas than the original effluents (Coulibaly et al., 2003). Recombinant strains of Saccharomyces cerevisiae have been genetically engineered to carry out simultaneous saccharification and fermentation (SSF) to produce extracellular endoglucanase and β -glucosidase that are able to ferment cellulose and hemicellulose to 6-carbon and 5-carbon sugars and subsequent fermentation to ethanol (Sedlak and Ho, 2004; van Maris et al., 2006; Haan et al., 2007; Chu and Lee, 2007; Wisselink et al., 2007). In bio-(Ceriporiopsis process, organosolv fungal subvermispora) pretreatment of wood waste for 2 - 8 weeks followed by organic solvent treatment at 140 -200 °C for 2 h has achieved considerable energy efficient delignification and hemicellulose hydrolysis (Itoh et al., 2003; Sánchez and Cardona, 2008).

Bacterial pretreatment of LCW involves both anaerobic and aerobic systems. Anaerobic degradation utilizes mainly mesophillic, rumen derived bacteria (Han and Shin, 2002; Hu and Yu, 2005, 2006; Neves et al., 2006; Hu et al., 2008; Yue et al., 2008). Aerobic-anaerobic systems have an upper hand when it comes to degradation of LCW richer in lignin content (Ammary, 2004; Mshandete et al., 2005, 2008) while in aerobic system alone, actinomycete *Streptomyces griseus* is able to produce high levels of extracellular hydrolytic enzyme that degrade lignocellulose (Arora et al., 2005). *Escherichia coli* and *Klebsiella oxytoca* strains have been genetically engineered to produce microbial biocatalysts that produce bioethanol from lignocellulosic materials (Jarboe et al., 2007; Peterson and Ingram, 2008).

Enzymatic pretreatment of LCW utilize hydrolytic and oxidative enzymes which are mainly derived from fungi and bacteria. Cellulases are usually a mixture of several enzymes. At least three major groups of cellulases are involved in the hydrolysis process: (1) endoglucanase (endo-1,4-glucanohydrolase) which attacks regions of low crystallinity in the cellulose fiber, creating free chainends; (2) exoglucanase or cellobiohydrolase (CBH) (1,4- β -glucan cellobiohydrolase) which degrades the molecule further by removing cellobiose units from the free chainends and (3) β -glucosidase which hydrolyzes cellobiose to produce glucose (Sun and Cheng, 2002). In addition, there are also a number of ancillary enzymes that attack hemicellulose, such as glucuronidase, acetylesterase, feruloylesterase, xylanase, β -xylosidase, galactomannanase and glucomannanase (Nikolov et al., 2000; Draude et al., 2001; Aranda et al., 2004; Mtui and Nakamura, 2005, Roman et al., 2006; Georgieva et al., 2008). During the enzymatic hydrolysis, cellulose is degraded by cellulases to reducing sugars that can be fermented by veasts or bacteria to ethanol.

Ligninolytic enzymes are primarily involved in lignin degradation in oxidative reactions that are mainly free radical driven in the presence (or sometimes absence) of mediators. The main enzymes involved are lignin peroxidase, manganese peroxidase and laccase (Hao et al., 2006; Mtui and Nakamura, 2007, 2008; Mtui and Masalu, 2008). The hydrolytic and oxidative enzymatic reactions are mainly carried out at $30 - 45 \,^{\circ}$ C with low enzyme loading rate at reaction time of 6 - 26 h. All the pretreatment methods discussed above are summarized in Figure 1.

VALUE-ADDED PRODUCTS FROM LIGNOCELLULOSIC WASTES

Advances in industrial biotechnology offer potential opportunities for economic utilization of agro-industrial residues. Biodevelopment of biowastes provide a wide range of affordable renewable value-added products from LCW (Pandey et al., 2000; van Wyk, 2001; Howard et al., 2003).

Reducing sugars

Fermentable sugars comes first in the value chain of processed LCW with glucose, xylose, xylitol, cellobiose, arabinose, pentose and galactose being the main reduced sugars produced (Akmar and Kennedy, 2001; Saha, 2003; Rodríguez-Chonga et al., 2004; Yáñez et al., 2004; Sepúlveda-Huerta et al., 2006; Tabka et al., 2006; Hanchar et al., 2007; Singh et al., 2008; Li et al., 2008; Kim et al., 2008). In these sugar producing processes, hydrolysable sugars yield of up to 83.3% has been achieved at the reaction temperatures of 37 - 50 ℃ for 6 -179 h at pH 5 - 6. The size of substrate added determines the amount of the saccharification products (Baig et al., 2004). In the enzymatic hydrolysis step using celluclast® supplemented with novozym®, a degree of saccharification of 100% has been achieved (Margues et al., 2008). Some transgenic plant residues have been reported to yield nearly twice as much sugar from cell walls compared to wild-types (Chen and Dixon, 2007). Glucose seems to be the major monosaccharide product from LCW. The challenge facing depolymerization of hemicellulose into fermentable sugars is the requirement for a consortium of enzymes to complete the hemicellulose hydrolysis, leading to high enzyme costs. Efforts to overcome the problem include process improvement and the use of modified microorganisms that produce the required hemicellulose enzymes (Lu and Mosier, 2007; Haan et al., 2007).

Enzymes

Lignocellulosic enzymes, mainly from fungi and bacteria, are important commercial products of LCW bioprocessing used in many industrial applications including chemicals, fuel, food, brewery and wine, animal feed, textile and laundry, pulp and paper and agriculture

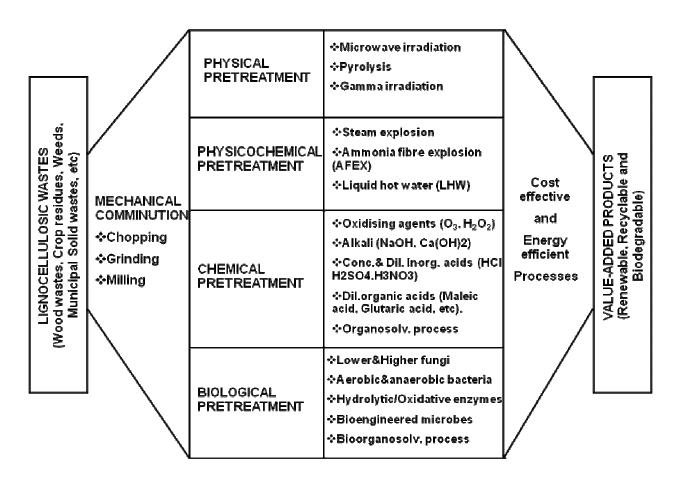


Figure 1. A summary of various methods used in the pretreatment of lignocellulosic wastes.

(Howard et al., 2003). Overall, extracellular enzymes are secondary metabolic products released in the presence of inducers at N-limited media (Mtui and Nakamura, 2007). They include hydrolytic enzymes such as cellulases; hemicellulases and pectinases; degradative enzymes like amylases, proteases; and ligninolytic enzymes like laccases, peroxidases and oxidases. Cellulases production from LCW has been extensively studied (Jecu, 2000; Emtiazi and Nahvi, 2000; El-hawary and Mostafa, 2001; Ögel et al., 2001; Raj and Singh, 2001; Ojumu et al., 2003; Wen et al., 2005; Muthuvelayudham and Viruthagiri, 2006; Pothiraj et al., 2006; Daroit et al., 2007; Gao et al., 2008). Phytases, mannanases and amylases are also produced by microorganisms using LCW as the main feedstock (Bhavsar et al., 2008; Mabrouk et al., 2008).

On the other hand, hemicellulolytic enzymes, mainly xylanases, are produced from a wide range of LCW biomass (Abdel-Sater and El-Said 2001; Rezende et al. 2002; Pandey and Pandey, 2002; Isil and Nilufer, 2005; Haq et al., 2006; Elisashvili et al., 2006; Dobrev et al., 2007; Mohana et al., 2008). Pectinases such as endopolygalacturonase (endo-PG), exo-polygalacturonase (exo-PG) and pectin liase are mainly produced from solid state fermentation processes utilizing agricultural

residues (Silva et al., 2005; Botella et al., 2005, 2007), while protease has been produced by *Penicillium janthinellum* in submerged cultures (Oliveira et al., 2006).

Among the ligninases produced from LCW, laccases are the mostly studied (Nazareth and Sampy, 2003; Moldes et al., 2003, 2004; Couto et al., 2006; Couto and Sanromána, 2006; Mishra and Kumar, 2007; Alcántara et al., 2007; Minussi et al., 2007), followed by Manganese peroxidase and lignin peroxidase (Couto et al., 2001, 2003; Wuyep et al., 2003; Velázquez-Cedeño et al., 2004; Couto and Sanromána, 2005; Alam et al., 2005; Asgher et al., 2006; Songulashvili et al., 2007; Elisashvili et al., 2008).

Very high enzyme activities (31,786 U/L) have been reported when the experiments are carried out under optimal conditions (pH 5.5 - 6: temperature 30 - 45 °C) (Rosales et al., 2007). Recovery of pure enzymes is achieved through 50 - 80% (NH₄)₂SO₄ saturation followed by chromatographical purification techniques (A-el-Gammal et al., 2001; Mtui and Nakamura, 2008). Several efforts have been made to increase the production of enzymes through strain improvement by mutagenesis and recombinant DNA technology. Cloning and sequencing of the various genes of interest could economize the enzymes production processes (Kumar et al., 2008).

Biofuels

Worldwide, there is a growing concern over the fossil oil prices increase, the security of the oil supply and the negative impact of fossil fuels on the environment, particularly greenhouse gas emissions (Hahn-Hägerdal et al., 2006). Conversion of LCW to biofuels provides the best economically feasible and conflict-free second-generation renewable alternatives (Rubin, 2008). Significant advances have been made towards bioconversion of plant biomass wastes into bioethanol, biodiesel, biohydrogen, biogas (methane).

Production of ethanol from sugars or starch from sugarcane and cereals, respectively, impacts negatively on the economics of the process, thus making ethanol more expensive compared with fossil fuels. Hence, the technology development focus for the production of ethanol has shifted towards the utilization of residual lignocellulosic materials to lower production costs (Howard et al., 2003). Currently, research and development of saccharification and fermentation technologies that convert LCW to reducing sugars and ethanol, respectively, in eco-friendly and profitable manner have picked tempo with breakthrough results being reported (Lin and Tanaka, 2006; Prasad et al., 2007; Patel et al., 2007; Pasha et al., 2007; Tahezaden and Karimi, 2007; Sánchez and Cardona, 2008). Ethanol yield of 6 - 21% has been obtained through fermentation of agricultural and municipal residues (Akin-osanaiye et al., 2005; Mtui and Nakamura, 2005; Sjöde et al., 2007; Li et al., 2007; Cara et al., 2008; Sørensen et al., 2008). While microaeration enhances productivity of bioethanol from LCW using ethanologenic E. coli (Okuda et al., 2007), simultaneous saccharification and fermentation (SSF) using recombinant Saccharomyces cereviasiae result to as high as 62% of the theoretical value (Itoha et al., 2003). The principal benefits of performing the enzymatic hydrolysis together with the fermentation, instead of in a separate step after the hydrolysis, are the cofermentation of both hexoses and pentoses during SSF, reduced end-product inhibition of the enzymatic hydrolysis and the reduced investment costs (Kádár and Réczey, 2004; Olofsson et al., 2008). Life cycle assessment (LCA) shows that bio-ethanol from LCW results to reductions in resource use and global warming (von Blottnitz and Curran, 2007). The long-term benefits of using waste residues as lignocellulosic feedstocks will be to introduce a sustainable solid waste management strategy for a number of lignocellulosic waste materials; contribute to the mitigation in greenhouse gases through sustained carbon and nutrient recycling; reduce the potential for water, air, and soil contamination associated with the land application of organic waste materials; and to broaden the feedstock source of raw materials for the bio-ethanol production industry (Champagne, 2007).

Biodiesel is a renewable fuel conventionally prepared by transesterification of pre-extracted vegetable oils and animal fats of all resources with methanol, catalyzed by strong acids or bases (Liu and Zhao, 2007). They are fatty acid methyl or ethyl esters used as fuel in diesel engines and heating systems (Ito et al., 2005). Production of biodiesel from lignocellulosic residues such as olive oil wastes has been a subject of research towards improving the thermal waste treatment systems and cleaner energy production (Arvanitoyannis et al., 2007a, 2007b). Since the current supplies from LCW based oil crops and animal fats account for only approximately 0.3%, biodiesel from algae is widely regarded as one of the most efficient ways of generating biofuels and also appears to represent the only current renewable source of oil that could meet the global demand for transport fuels (Schenk et al., 2008).

Hydrogen has been considered a potential fuel for the future since it is carbon-free and oxidized to water as a combustion product (Najafpour et al., 2004). While conventional burning or composting seem to be the most cost-effective hydrogen production methods, bacteria such as Enterobacter aerogenes and Clostridium sp isolates can convert saccharified LCW biomass into biohydrogen (Ito et al., 2005). Biohydrogen production from agricultural residues such as olive husk pyrolysis (Ca lar and Demirba, 2002); conversion of wheat straw wastes into biohydrogen gas by cow dung compost (Fan et al., 2006); bagasse fermentation for hydrogen production (Singh et al., 2007) generate up to 70.6% gas vields. System optimization for accessibility of polysaccharides in LCW and the use of genetically efficient bacterial strains for agrowaste-based hydrogen production seem to be the ideal option for clean energy generation. Hydrogen gereration from inexpensive abundant renewable biomass can produce cheaper hydrogen and achieve zero net greenhouse emissions (Zhang et al., 2007).

Biogas production from lignocellulosic materials is a steady anaerobic process where methane rich biogas comes mostly from hemicellulose and cellulose. Anaerobic biomethane production is an effective process for conversion of a broad variety of agricultural residues to methane to substitute natural gas and medium calorific value gases (Demirbas and Ozturk, 2005). Biogas containing 55 - 65% methane can be produced from jute caddis - a lignocellulosic waste of jute mills by anaerobic fermentation, using cattle dung as sole source of inoculum (Banik, 2004). Anaerobic digestion of poultry droppings, cow dung and corn stalk can give up to 137.16 L of biogas from 0.28 m³ digester (Anozie et al., 2005). Mesophilic aerobic pretreatment to delignify sisal pulp waste prior to its anaerobic digestion has been shown to improve methane yields (Mshandete et al., 2005, 2008).

Overall, the success of biofuels production from LCW is dependent on the optimal performance and cost effectiveness of pretreatment and product generation processes.

Organic acids

Organic acids are some of the products of ligninolytic residues fermentations via environmentally friendly integrated processes. Volatile fatty acids including acetic acid, propionic acids and butyric acid are produced from a wide range of LSW such as cereal hulls (Jin et al., 2002, 2004, 2006); bagasse residues (Henrique et al., 2005); food wastes (Lim et al., 2008) and sisal leaf decortications residues (Mshandete et al., 2008). In addition, lactic acid is produced from waste sisal stems (Muruke et al., 2006), sugarcane bagasse (Adsul et al., 2007) and kitchen waste (Ohkouchi and Inoue, 2007) by using Lactobacillus isolates. Furthermore, formic acid, levulinic acid, citric acid, valeric acid, caproic acid and vanillinic acid are obtainable from bioprocessing of LCW (Olson, 2001; Chaudhary and Sharma, 2005; Mshandete et al., 2008; Ibrahim et al., 2008). Overall, organic acids production requires batch or continuous incubation conditions, the average reaction parameters being 35°C, pH 6.0, hydraulic retention time (HRT) of up to 8 days and organic loading rates of 9 g/l d. Product yields of up to 39.5 g/l have been reported (Lim et al., 2008).

Compost

Compost, a nutrient-rich, organic fertilizer and soil conditioner, is a product of humification of organic matter. This process is aided by a combination of living organisms including bacteria, fungi and worms which transform and enhance lignocellulosic waste into humic-like substances (Eyheraguibel et al., 2008). Vermicomposting is the bio-oxidation and stabilization of organic matter involving the joint action of earthworms and microorganisms, thereby turning wastes into a valuable soil amendment called vermicompost (Benitez et al., 2005; Aira et al., 2006, 2007). Substrates suitable for making humus-rich compost include cereal straw and bran (Hart et al., 2003); urban wastes (Taiwo and Oso, 2004); water hyacinth (Chatterjee et al., 2005); lemon tree prunings, cotton waste and brewery waste (García-Gómez et al., 2005); horticultural wastes (Wen-Jing et al., 2004, Lopez et al., 2006); olive, palm and grape wastes (Salètes et al., 2004; Alburguerque et al., 2006; Cayuela et al., 2006; Arvanitoyannis et al., 2007a). While bacteria inoculants such as Bacillus shackletonni, Streptomyces thermovulgaris and Ureibacillus thermosphaericus are used to improve the composting process (Vargas-Garcı et al., 2007), ligno-cellulolytic fungi inocula (e.g. Trichurus spiralis) may also be used in a pretreatment process before composting in order to reduce the resistance of the substrate to biodegradation (Hart et al., 2003; Vargas-García et al., 2007). A new earthworm strain of Perionyx sansibaricus is able to humify a substrate combination of guar gum industrial waste, cow dung and saw dust (Suthar, 2007). Composting can, therefore, be considered as a low-cost technology to convert agroindustrial LCW into value-added biofertilizers.

Biocomposites

Biodegradable polymers constitute a loosely defined family of polymers that are designed to degrade through the action of living organisms. Such commercially available biodegradable polymers are polycaprolactone, poly (lactic acid), polyhydroxyalkanoates, poly (ethylene glycol), and aliphatic polyesters like poly (butylene succinate) (PBS) and poly (butylene succinate-co-butylene adipate) (Tserki et al., 2006). Lignocellulosic material-thermoplastic polymer composites are among the emerging products of LCW. In most cases, lignocellulosic biomass flour is used as the reinforcing filler and polypropylene as the thermoplastic matrix polymer to manufacture particle-reinforced composites (Yang et al., 2004). Natural fibres from LCW are considered to be of low-cost by-products, environmentally friendly and practically sustainable raw materials (Georgopoulos et al., 2005). Evaluations of LCW fiber plastic composites utilizing wood fibre wastes (Bhattacharyya and Jayaraman, 2003; Yuan et al., 2004; Schilling et al., 2004; Ashori, 2008); wheat and rice straw (Digabel et al., 2004; Yang et al., 2004a); jute/cotton, sisal/cotton and ramie/cotton hybrid fabrics (Mishra et al., 2004; Alsina et al., 2005; Jacob et al., 2006); non-wood plant fibres (Ndazi et al., 2006); waste newsprint paper (Madani et al., 2004; Baroulaki et al., 2006); flax and hemp (Tserki et al., 2006); oil palm wastes (Shaji et al., 2006; John et al., 2008); cotton gin waste (Bourne et al., 2007); banana fibres (Pothan et al., 2007); cereal husks (Yang et al., 2004b, 2007; López et al., 2007); tissue paper wastes and corn peels (Lertsutthiwong et al., 2008); bagasse (Habibi et al., 2008) and nanofibers from the agricultural residues (Alemdar and Sain, 2008) have shown that such composites are suitable for making products that have improved biodegradability, mechanical strength, thermal stability, electrical conductivity and recyclability.

Treated LCW wastes are also used in the construction industry for manufacturing of light-weight agro-gypsum panels (Basta et al., 2002) and lightweight sand concretes (Reis, 2006; Bederina et al., 2007) with improved structural and thermal properties. Biocomposites are very promising in producing sustainable current and future green materials to achieve durability without using toxic chemicals. The challenge facing the biocomposite industry is to make materials that have better rubber/fiber interface, improved wettability and compatibility.

Food and feed

Bioconversion of lignocellulosic agro-residues through mushroom cultivation and single cell protein (SCP) production offer the potential for converting these residues into protein-rich palatable food and reduction of the

ussis et al., 2007; Zhang et a ts for research on bioactive o

environmental impact of the wastes. Mushrooom cultivation provides an economically acceptable alternative for the production of food of superior taste and quality which does not need isolation and purification (Israilides and Philippoussis, 2003; Philippoussis et al., 2007). Cultivation of edible mushrooms such as Lentinus spp, Lentinula spp, Leonotis spp, Pleurotus spp, Agaricus spp, Agrocybe spp, Volvariella spp, Lentinus spp and Grifola spp is achievable on a wide range of LCW substrates such as wood waste, corncob meal, wheat straw, barley straw, soybean straw, cereal bran, cotton waste, sorghum stalk, banana pseudostem, hazelnut husks, waste tea leaves, dry weed plants, peanut shells, waste paper and olive mill wastewater (Morais et al., 2000; Philippoussis et al., 2001; Yildiz et al., 2002; Oku, 2004; Kalm and Sargin, 2004; Silva et al., 2005; Özçelika and Pekşen, 2007; Peker et al., 2007; Das and Mukherjee, 2007; Akyüz and Yildiz, 2008; Gaitán-Hernández and Salmones, 2008; Rani et al., 2008), Mushrooms with increased number of fruit bodies and high contents of protein and total carbohydrates are obtained when LCW substrates are used in combination.

On the other hand, SCP production from LCW offers a potential substrate for conversion of low-quality biomass into an improved animal feed and human food. SCP is the protein extracted from cultivated microbial biomass. It can be used for protein supplementation of a staple diet by replacing costly conventional sources like soymeal and fishmeal to alleviate the problem of protein scarcity. Moreover, bioconversion of agricultural and industrial wastes to protein-rich food and fodder stocks has an additional benefit of making the final product cheaper (Anupama and Ravindra, 2000). Removal of nucleic acids and toxins from SCP is key to ensure the safety of food and feed. Among the SCP obtained from LCW using agricultural wastes as the main growth media, Saccharomyces cerevisiae, Trichoderma reesei and Kluyveromyces marxianus top the list (Robinson and Nigam, 2003; Chaudhary and Sharma, 2005). SCP yield of 51 and 39.4% efficiency of conversion of beet-pulp into protein has been reported ffrom the above strains. Solid state fermentation of LCW seems to be the most preferred culturing method, while cloning is being considered as a suitable technique for improvement of SCP production (Anupama and Ravindra, 2000).

Medicines

LCW provides a suitable growth environment for mushrooms that comprise a vast source of powerful new pharmaceutical products. In particular, *Lentinula edodes*, *Tremella fuciformis* and *Ganoderma lucidum* contain bioactive compounds such as anti-tumor, anti-inflammatory, anti-virus and anti-bacterial polysaccharides. Moroever, they contain substances with immunomodulating properties, as well as active substances that lower choresterol (Israilides and Philippoussis, 2003; Mtui 1405

Philippoussis et al., 2007; Zhang et al. 2007). Future prospects for research on bioactive compounds from fungi grown on such cheap and ubiquitous substrates look bright and could lead to breakthroughs in the search for antibacterial, antiviral and anticancer chemotherapies.

Biosorbents

Adsorbents obtained from plant wastes are feasible replacements for costly conventional methods of removing pollutants such as heavy metals ions, dyes, ammonia and nitrates from the environment. The use of lignocellulosic agrowastes is a very useful approach because of their high adsorption properties, which results from their ion-exchange capabilities. Agricultural wastes can be made into good sorbents for the removal of many metals, which would add to their value, help reduce the cost of waste disposal, and provide a potentially cheap alternative to existing commercial carbons (Krishnani and Ayyappan, 2006). Chemically modified plant wastes such as rice husks/rice hulls, spent grain, sugarcane bagasse/fly ash, sawdust, wheat bran, corncobs, wheat and soybean straws, corn stalks, weeds, fruit/vegetable wastes, cassava waste fibres, tree barks, azolla (water fern), alfalfa biomass, coirpith carbon, cotton seed hulls, citrus waste and soybean hulls show good adsorption capacities for Cd, Cu, Pb, Zn and Ni (Ahmedna et al., 2004; Basso et al., 2004; Dupont et al., 2005; Harman et al., 2007; Šćiban et al., 2008; Ngah et al., 2008; Zubair et al., 2008). They are usually modified with formaldehyde in acidic medium, NaOH, KOH/K₂CO₃ and CO₂, or acid solution or just washed with warm water (Tsai et al., 2001; Šcibanet al., 2008). Scanning electron micrographs with energy spectra shows that heavy metals are immobilized via two possible routes: adsorption and cation exchange on hypha, and the chelation by fungal metabolite (Huang et al., 2008).

LCW have also been shown to be able to adsorb dyes from aqueous solutions. Adsorption of reactive dyes by sawdust char and activated carbon (Gan et al., 2004); ethylene blue by waste Rosa canina sp. seeds (Gürses et al., 2006); anionic dyes by hexadecyltrimethylammoniummodified coir pith (Namasivayam and Sureshkumar, 2006); and methylene red by acid-hydrolysed beech sawdust (Batzias and Sidiras 2007) have been reported. Ammonia and nitrate removal by using agricultural waste materials as adsorbents or ion exchangers have also been studied (Orlando et al., 2002; Kishore et al., 2006). Prehydrolysis enhances the adsorption properties of the original LCW material due to the removal of the hemicelluloses during sulphuric acid treatment, resulting in the 'opening' of the lignocellulosic matrix's structure, the increasing of the surface area and the activation of the material's surface owing to an increase in the number of dye binding sites (Batzias and Sidiras, 2007). The main value-added products from LCW are generally summarized in Figure 2.

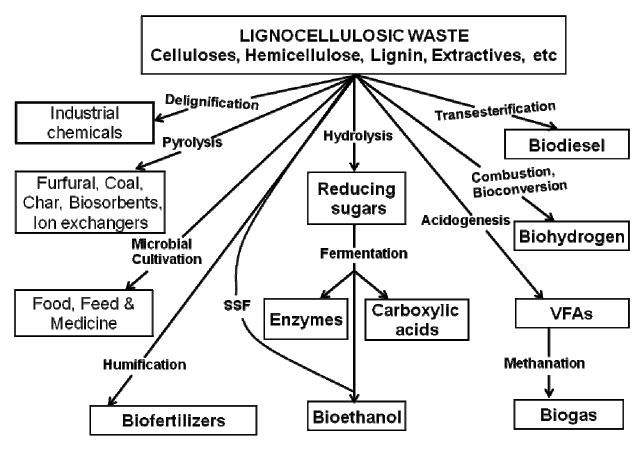


Figure 2. The main value-added products from lignocellulosic wastes (SSF=simultaneous fermentation and saccharification, VFAs = volatile fatty acids).

TECHNO-ECONOMIC EVALUATION

Technologies that are being developed for commercial pretreatment and value addition of LCW face technical and economical impediments. Therefore, cost effective technical innovations that allow cost-effective conversion of biomass into fuels and chemicals are mandatory. These technologies include low-cost thermochemical pretreat-ment, highly effective enzymes and efficient and robust fermentative microorganisms (Gray et al., 2006; Cardona and Sánchez, 2007; Vertès et al., 2008). The choice of appropriate feedstocks and processes ought to consider those which consume less electricity, produce fewer emissions in total, and has less of a human health impact (Kemppainen and Shonnard, 2005).

The high cost of enzymes presents a significant barrier to commercialization of bio-based products. In the simplest terms, the cost is a function of the large amount of enzyme protein required to break down polymeric sugars in cellulose and hemicellulose to fermentable monomers. In recent years, significant effort has been expended to reduce the cost by focusing on improving the efficiency of known enzymes, identification of new, more active enzymes, creating enzyme mixes optimized for selected pretreated substrates, and minimization of enzyme production costs (Merino and Cherry, 2007).

Ethanol's future role as a fuel hinges on several factors including feedstock availability, processing costs and supportive political framework. Improvements in pretreatment and advances in biotechnology, especially through process combinations can bring the ethanol production overall process efficiency to 68%. Also, a combined effect of higher hydrolysis-fermentation efficiency, lower specific capital investments, increase of scale, cheaper biomass feedstock costs and using genetically engineered microorganisms that can convert xylose and/or pentose to ethanol can greatly improve ethanol production efficiency and reduce the cost of the production (Sun and Cheng, 2002; Carlo et al., 2005). Processes that produce only ethanol from lignocellulosics display poor economics. The large market for ethanol makes it possible to achieve economies of scale that reduce sugar costs, and co-producing chemicals promises greater profit margins or lower production costs for a given return on investment. For the large processing plants, the production costs are significantly reduced compared to small plants (Wyman, 2003; Murphy and McCarthy, 2005). Yield improvements in all major steps of LCW processing would enable lower capital requirements, thus improving the economics and lowering

investment risk (Bohlmann, 2006). Bioproducts from LCW will continue to be the keystone of industrial biotechnology-based economy whereby biorefineries leverage common raw materials and unit operations to integrate diverse processes to produce demand-driven product portfolios (Otero et al., 2007). High product yields and less energy-demanding processes could be achieved by increasing the dry matter content resulting in higher products, and thereby improving the overall process economy (Sassner et al., 2008).

FUTURE TRENDS

Although pretreatment systems and the concomitant release of bio-products from LCW have been greatly improved by new technologies, there are still challenges that need further investigations. These challenges include development of more efficient pretreatment and production technologies, bioprospecting and development of stable genetically engineered microorganisms, improved gene cloning and sequencing technologies and enhancement of productions based on economies of scale for more efficient and cost effective conversions of LCW into value-added products.

So far, lignocellulosic biomass has been the most promising economically viable and renewable source of biohydrogen and biodiesel. However, the second generation microalgal systems seem to be more advantageous in that they: (1) have a higher photon conversion efficiency (as evidenced by increased biomass yields per hectare): (2) can be harvested batchwise nearly all-year-round, providing a reliable and continuous supply of oil: (3) can utilize salt and waste water streams, thereby greatly reducing freshwater use: (4) can couple CO_2 -neutral fuel production with CO_2 sequestration: (5) produce non-toxic and highly biodegradable biofuels (Schenk et al., 2008). Therefore, extensive research is now being directed toward that end. Plant fibers as fillers and reinforcements for polymers are currently the fastest-growing type of polymer additives. Nanobiotechnology seems to take charge as far as the use of LCW nanofibres in plastic composites is concerned (Alemdar and Sain, 2008). It is envisaged that nano materials from renewable biowastes will be the main focus of future research.

Conclusion

This work has attempted to take a broad analysis of most of the research done worldwide in the past nine years regarding the enormous diversity of value-added products from pretreated lignocelluloses wastes (LCW). Various pretreatment and production systems providing technical and economic feasibility to harness the renewable materials while at the same time cleaning up the environment have been highlighted. Physicochemical and biological pretreatment systems appear to be the mostly preferred methods, while the use of treated LCW as bio-based adsorbents of pollutants, and the production of degradable biofuels and biopolymers from LCW have drawn a lot of research interest. Current and future research trends are directed towards the developments and applications of engineered organisms to tackle the challenges encountered from using conventional naturally occurring strains. At the same time, production of nano materials from pretreated LCW provides bright future prospects in the biocomposites industry. Although the main challenge facing LCW pretreatment and valueaddition industry over the next few years will be to reduce processing and production costs, this work looked at studies that suggest the ways out of this problem. The review serves as a valuable reference material for a wide range of scientists and technologists in the relevant fields.

ACKNOWLEDGEMENTS

Sida/SAREC through the International Science Program, is gratefully acknowledged for financial support. The Uppsala University, Sweden, is thanked for logistical support. The University of Dar es Salaam, Tanzania, is appreciated for granting leave of absence.

REFERENCES

- Abdel-Sater MA, El-Said AHM (2001). Xylan-decomposing fungi and xylanolytic activity in agricultural and industrial wastes. Int. Biodeterioration Biodegrad. doi:10.1016/S0964-8305(00)00113-X. 47(1): 15-21.
- Adsul MG, Varma AJ, Gokhale DV (2007). Lactic acid production from waste sugarcane bagasse derived cellulose. Green Chem. doi: 10.1039/b605839f. 9: 58-62.
- A-el-Gammal A, M-Ali A, Kansoh AL (2001). Xylanolytic activities of *Streptomyces* sp. 1--taxonomy, production, partial purification and utilization of agricultural wastes. Acta Microbiol. Immunol. Hung. 48(1): 39-52.
- Ahmedna M, Marshall WE, Husseiny AA, Rao RM, Goktepe I (2004). The use of nutshell carbons in drinking water filters for removal of trace metals. Water Res. doi:10.1016/j.watres.2003.10.047. 38(4): 1062-1068.
- Aira M, Monroy F, Domínguez J (2006). Eisenia fetida (Oligochaeta, Lumbricidae) Activates Fungal Growth, Triggering Cellulose Decomposition During Vermicomposting. J. Microbial. Ecol. doi: 10.1007/s00248-006-9109-x. 52(4): 738-747.
- Aira M, Monroy F, Domínguez J (2007). Microbial Biomass Governs Enzyme Activity Decay during Aging of Worm-Worked Substrates through Vermicomposting. Environ. Qual. doi: 10.2134/jeq2006.0262. 36: 448-452.
- Akin-osanaiye BC, Nzelibe HC, Agbaji AS (2005). Production of ethanol from Carica papaya (pawpaw) agro waste: effect of saccharification and different treatments on ethanol yield. Afric. J. Biotechnol. 4(7): 657-659.
- Akmar PF, Kennedy JF (2001). The potential of oil and sago palm trunk wastes as carbohydrate resources. J. Wood Sci. Technol. 35(5): 467-473. doi: 10.1007/s002260100107.
- Akyüz M, Yildiz A (2008). Evaluation of cellulosic wastes for the cultivation of *Pleurotus eryngii* (DC. ex Fr.) Quel. Afr. J. Biotechnol. 7(10): 1494–1499.
- Alam Z, Mahmat ME, Muhammad N (2005). Solid state bioconversion of oil palm biomass for ligninase enzyme production. Artif. Cells, Blood

Substit. Biotechnol. 33(4): 457-466(10).

- Alburquerque JA, Gonzálvez J, García D, Cegarra J (2006). Effects of bulking agent on the composting of "alperujo", the solid by-product of the two-phase centrifugation method for olive oil extraction. Process Biochem. 41(1): 127-132. doi:10.1016/j.procbio.2005.06.006.
- Alcántara T, Góme J, PazosM, Sanromán Á (2007). Enhanced production of laccase in *Coriolopsis rigida* grown on barley bran in flask or expanded-bed bioreactor. World J. Microbiol. Biotechnol. 23(8): 189-1194. doi 10.1007/s11274-006-9334-y.
- Alemdar A, Sain M (2008). Isolation and characterization of nanofibers from agricultural residues – Wheat straw and soy hulls. Bioresour. Technol. 99(6): 1664-1671. doi:10.1016/j.biortech.2007.04.029.
- Alma MH, Acemioglu B (2004). A kinetic study of sulfuric acid-catalyzed liquefaction of wood into phenol Chem. Eng. Comm. 191(7): 968-980. doi: 10.1080/00986440490276173.
- Alsina OLS, de Carvalho LH, Filho FGR, d'Almeida JRM (2005). Thermal properties of hybrid lignocellulosic fabric-reinforced polyester matrix composites. Polymer Testing 24(1): 81-85. doi:10.1016/j.polymertesting.2004.07.005.
- Álvarez P, Santamaría R, Blanco C, Granda M (2005). Thermal degradation of lignocellulosic materials treated with several acids. J. Analyt. Appl. Pyrolysis 74(1-2): 337-343. doi:10.1016/j.jaap.2004.11.030.
- Ammary BY (2004). Nutrients requirements in biological industrial wastewater treatment. Afr. J. Biotechnol.3 (4): 236-238.
- Anozie AN, Layokun SK, Okeke CU (2005). An evaluation of a batch pilot-scale digester for gas production from agricultural wastes. Energy Sources, Part A: Recovery, Util. Environ. Effects 27(14):1301-1311. doi: 10.1080/009083190519023.
- Anupama A, Ravindra P (2000). Value-added food: Single cell protein. Biotechnol. Adv. 18(6): 459-479. doi: 10.1016/S0734-9750(00)00045-8.
- Aranda E, Sampedro I, Ocampo JA, García-Romera I (2004). Contribution of hydrolytic enzymes produced by saprophytic fungi to the decrease in plant toxicity caused by water-soluble substances in olive mill dry residue. J. Appl. Microbiol. Biotechnol. 64(1): 132-135. doi 10.1007/s00253-003-1368-6.
- Arora A, Nain L, Gupta JK (2005). Solid-state fermentation of wood residues by *Streptomyces griseus* B1, a soil isolate, and solubilization of lignins. World J. Microbiol. Biotechnol. 21(3): 303-308. doi 10.1007/s11274-004-3827-3.
- Arvanitoyannis IS, Kassaveti A (2007a). Current and potential uses of composted olive oil waste. Int. J. Food Sci. Technol. 42(3): 281 – 295. doi: 10.1111/j.1365-2621.2006.01211.x.
- Arvanitoyannis IS, Kassaveti A, Stefanatos S (2007b). Current and potential uses of thermally treated olive oil waste. Int. J. Food Sci. Technol. 42(7): 852 867. doi: 10.1111/j.1365-2621.2006.01296.x.
- Asada C, Nakamura Y, Kobayashi F (2005). Waste reduction system for production of useful materials from un-utilized bamboo using steam explosion followed by various conversion methods. Biochem. Eng. J. 23(2): 131-137 doi:10.1016/j.bej.2004.11.004.
- Asgher M, Asad MJ, Legge RL (2006). Enhanced lignin peroxidase synthesis by Phanerochaete chrysosporium in solid state bioprocessing of a lignocellulosic substrate. World J. Microbiol. Biotechnol. 22(5): 449-45. doi 10.1007/s11274-005-9055-7.
- Ashori A (2008). Wood–plastic composites as promising greencomposites for automotive industries. Bioresour. Technol. 99(11): 4661-4667. doi:10.1016/j.biortech.2007.09.043.
- Baig MMV, Baig MLB, Baig MIA, Yasmeen M (2004). Saccharification of banana agro-waste by cellulolytic enzymes. Afric. J. Biotechnol. 3(9): 447-450.
- Banik S (2004). Jute caddis: A new substrate for biogas production. J. Sci. Ind. Res. 63(9): 747-751.
- Baroulaki I, Karakasi O, Pappa G, Tarantili PA, Economides D, Magoulas K (2006). Preparation and study of plastic compounds containing polyolefins and post used newspaper fibers. Composites Part A: Appl. Sci. Manuf. 37(10): 1613-1625. doi:10.1016/j.compositesa.2005.10.012.
- Basso MC, Cerrella EG, Cukierman AL (2004). Cadmium uptake by lignocellulosic materials: Effect of lignin content. Sep. Sci. Technol. 39(5): 1163-1175.
- Basta A, Abd El-Sayed ES, Fadl NA (2002). Lignocellulosic materials in

building elements. Part III. Recycled newsprint waste paper in manufacturing light-weight agro-gypsum panels. Pigment & Resin Technol. 31(3): 160-170. doi: 10.1108/03699420210428523.

- Batzias FA, Sidiras DH (2007). Dye adsorption by prehydrolysed beech sawdust in batch and fixed-bed systems. Bioresour. Technol. 98(6): 1208-1217. doi:10.1016/j.biortech.2006.05.020.
- Bederina M, Marmoret L, Mezreb K, Khenfer MM, Bali A, Quéneudec M (2007). Effect of the addition of wood shavings on thermal conductivity of sand concretes: Experimental study and modeling. Construction and Building Materials 21(3): 662-668. doi:10.1016/j.conbuildmat.2005.12.008.
- Belewu MA (2006). Conversion of masonia tree sawdust and cotton plant by product into feed by white rot fungus (*Pleurotus sajor caju*). Afric. J. of Biotechnol. 5(6): 503-504.
- Benitez E, Sainzh H, Nogales R (2005). Hydrolytic enzyme activities of extracted humic substances during the vermicomposting of a lignocellulosic olive waste. Bioresour. Technol. 96(7): 785-790 doi:10.1016/j.biortech.2004.08.010.
- Bhattacharyya D, Jayaraman K (2003). Manufacturing and evaluation of woodfibre-waste plastic composite sheets. Polymers & polymer composites. 11(6): 433-440
- Bhavsar K, Shah P, Soni SK, Khire JM (2008). Influence of pretreatment of agriculture residues on phytase production by *Aspergillus niger* NCIM 563 under submerged fermentation conditions. Afr. J. Biotechnol. 7(8): 1101-1106.
- Bohlmann GM (2006). Process economic considerations for production of ethanol from biomass feedstocks. Ind. Biotechnol. 2(1): 14-20. doi:10.1089/ind.2006.2.14.
- Bonelli PR (2003). Slow pyrolysis of nutshells: characterization of derived chars and of process kinetics . Energy Sources, 25(8): 767-778.
- Botella C, de Ory I, Webb C, Cantero D, Blandino A (2005). Hydrolytic enzyme production by *Aspergillus awamori* on grape pomace. Biochem. Eng. J. 26(2-3): 100-106 doi:10.1016/j.bej.2005.04.020.
- Botella C, Diaz A, Ignacio de Ory I, Webb C, Blandino A (2007). Xylanase and pectinase production by *Aspergillus awamori* on grape pomace in solid state fermentation. Process Biochem. 42(1): 98-101. doi:10.1016/j.procbio.2006.06.025.
- Bourne PJ, Bajwa SG, Bajwa DS (2007). Evaluation of cotton gin waste as a lignocellulosic substitute in woodfiber plastic composites. For. Products J. 57:127-131
- Brienzo M, Silva EM, Milagres AMF (2007). Degradation of eucalypt waste components by *Lentinula edodes* strains detected by chemical and near-infrared spectroscopy methods. J. Appl. Biochem. Biotechnol. 141(1): doi 10.1007/s12010-007-9209.
- Ça lar A, Demirba A (2002). Hydrogen rich gas mixture from olive husk via pyrolysis. Energy Conversion Manag. 43(1): 109-117 doi:10.1016/S0196-8904(01)00012-7.
- Cara C, Ruiza E, Ballesteros M, Manzanares P, Negro MJ, Castro E (2008). Production of fuel ethanol from steam-explosion pretreated olive tree pruning. Fuel 87(6): 692-700. doi:10.1016/j.fuel.2007.05.008.
- Cardona CA, Śáncheza ÓJ (2007). Fuel ethanol production: Process design trends and integration opportunities. Bioresour. Technol. 98(12): 2415-2457. doi:10.1016/j.biortech.2007.01.002.
- Carlo N Hamelinck CN, Hooijdonk G, Faaij APC (2005). Ethanol from lignocellulosic biomass: Techno-economic performance in short-, middle- and long-term. Biomass Bioenergy 28(4): 384-410. doi:10.1016/j.biombioe.2004.09.002.
- Carrillo F, Lis MJ, Colom X, López-Mesas M, Valldeperas J (2005). Carrillo F, Lisa MJ, Colom X, López-Mesas M, Valldeperas J (2005). Effect of alkali pretreatment on cellulase hydrolysis of wheat straw: Kinetic study. Process Biochem. 40(10): 3360-3364. doi:10.1016/j.procbio.2005.03.003.
- Cayuela ML, Sánchez-Monedero MA, Roig A (2006). Evaluation of two different aeration systems for composting two-phase olive mill wastes. Process Biochem. 41(3): 616-623. doi:10.1016/j.procbio.2005.08.007.
- Champagne P (2007). Feasibility of producing bio-ethanol from waste residues. Resourc. Conserv. Recycling 50(3) 211-230. doi:10.1016/j.resconrec.2006.09.003.
- Chatterjee P, Metiya G, Saha N, Haldar M, Mukherjee D (2005).

Methods and substrate oriented composting of lignocellulosic materials for the production of humus rich composts. Environ. Ecol. 23(1): 55-60.

- Chaudhary N, Sharma, CB (2005). Production of citric acid and single cell protein from agrowaste. Nat. Acad. Sci. Lett. 28(5/6): 189-193.
- Chen G, Leung DYC (2003). Experimental investigation of biomass waste, (rice straw, cotton stalk, and pine sawdust), pyrolysis characteristics. Energy Sourc. Part A: Recovery, Util. Environ. Effects 25(4): 331- 337. doi: 10.1080/00908310390142361.
- Chen F, Dixon RA (2007). Lignin modification improves fermentable sugar yields for biofuel production. Nat. Biotechnol. 25: 759-761. doi:10.1038/nbt1316.
- Chu BCH, Lee H (2007). Genetic improvement of *Saccharomyces cerevisiae* for xylose fermentation. Biotechnol. Adv. 25(5): 425-441. doi:10.1016/j.biotechadv.2007.04.001.
- Chundawat SP, Venkatesh B, Dale BE (.2007). Effect of particle size based separation of milled corn stover on AFEX pretreatment and enzymatic digestibility. Biotechnol. Bioeng. 96(2): 219-31.
- Coulibaly L, Gourene G, Agathos NS (2003). Utilization of fungi for biotreatment of raw wastewaters. Afric. J. Biotechnol.2 (12): 620-630.
- Couto SR, Domínguez A, Sanromán A (2001). Utilisation of lignocellulosic wastes for lignin peroxidase production by semi-solidstate cultures of *Phanerochaete chrysosporium*. Biodegradation 12(5): 283-289. doi: 10.1023/A:1014392810649.
- Couto SR, Rodríguez R, Gallego PP, Sanromán A (2003). Biodegradation of grape cluster stems and ligninolytic enzyme production by *Phanerochaete chrysosporium* during semi-solid-state cultivation. Acta Biotechnological 23(1): 65-74. doi 10.1002/abio.200390010.
- Couto SR, Sanromán MA (2005). Application of solid-state fermentation to ligninolytic enzyme production. Biochem. Eng. J. 22(3): 211-219 doi:10.1016/j.bej.2004.09.013.
- Couto SR, Sanromán MÁ (2006). Effect of two wastes from groundnut processing on laccase production and dye decolourisation ability. J. of Food Eng. 73(4): 388-393 doi:10.1016/j.jfoodeng.2005.02.018.
- Couto SR, López E, Sanromán MA (2006). Útilisation of grape seeds for laccase production in solid-state fermentors. J. Food Eng. 74(2): 263-267 doi:10.1016/j.jfoodeng.2005.03.004.
- Damisa D, Ameh J, Umoh, VJ (2008). Effect of chemical pretreatment of some lignocellulosic wastes on the recovery of cellulase from Aspergillus niger AH3 mutant. Afr. J. Biotechnol. 7 (14): 2444-2450.
- Daroit DJ, Silveir ST, Hertz PF, Brandelli A (2007). Production of extracellular β-glucosidase by *Monascus purpureus* on different growth substrates. Process Biochem. 42(5): 904-908. doi:10.1016/j.procbio.2007.01.012.
- Das M, Hossain, SKM (2000). Studies on lignin biodegration of nonconventional lignocellulosic agro-waste material by using *P. chrysosporium* - A pollution control biopulping process. J. Ind. Pollut. Control 16(2): 195-200.
- Das H, Singh S (2004). Useful by-products from cellulosic wastes of agriculture and food industry A critical appraisal. Crit. Rev. Food Sci. Nutr. 44(2): 77-89 DOI: 10.1080/10408690490424630.
- Das N, Mukherjee M (2007). Cultivation of *Pleurotus ostreatus* on weed plants. Bioresour. Technolo. 98(14): 2723-2726. doi:10.1016/j.biortech.2006.09.061.
- Digabel FL, Boquillon N, Dole P, Monties B, Averous (2004). Properties of thermoplastic composites based on wheat-straw lignocellulosic fillers. J. Appl. Poly. Sci. 93(1): 428 436. doi: 10.1002/app.20426.
- Draude KM, Kurniawan CB, Duff JB (2001). Effect of oxygen delignification on the rate and extent of enzymatic hydrolysis of lignocellulosic material. Bioresourc. Technol. 79(2): 113-120 doi:10.1016/S0960-8524(01)00055-4.
- De Sousa MV, Monteiro SN, d'Almeida JRM (2004). Evaluation of pretreatment, size and molding pressure on flexural mechanical behavior of chopped bagasse–polyester composites. Poly. Testing 23 (3): 253-258. doi:10.1016/j.polymertesting.2003.09.002.
- Del Campo I, Alegría I, Zazpe M, Echeverría M, Echeverría I (2006). Diluted acid hydrolysis pretreatment of agri-food wastes for bioethanol production. Industrial Crops Products 24(3): 214-221. doi:10.1016/j.indcrop.2006.06.014.
- Demirbas A (2002). Utilization of urban and pulping wastes to produce synthetic fuel via pyrolysis. Energy Sources Part A: Recovery, Util.

Environ. Effects, 24(3): 205-213. doi:10.1080/009083102317243593.

- Demirbas A, Ozturk T (2005). Anaerobic digestion of agricultural solid residues. Int. J. Green Energy 1(4): 483-494 doi: 10.1081/GE-200038719.
- Demirbas A (2008). Products from lignocellulosic materials via degradation processes. Energy Sources Part A: Recovery, Utilization, and Environmental Effects 30(1): 27 37. doi: 10.1080/00908310600626705.
- Dobrev GT, Pishtiyski IG, Stanchev VS, Mircheva R (2007). Optimization of nutrient medium containing agricultural wastes for xylanase production by *Aspergillus niger* B03 using optimal composite experimental design. Bioresour. Technol. 98(14): 2671-2678. doi:10.1016/j.biortech.2006.09.022.
- Dupont L, Bouanda J, Dumonceau J, Aplincourt M (2005). Biosorption of Cu(II) and Zn(II) onto a lignocellulosic substrate extracted from wheat bran. J. Environ. Chem. Lett. 2(4): 165-168. doi 10.1007/s10311-004-0095-2.
- El-hawary FI, Mostafa YS (2001). Factors affecting cellulase production by *Trichoderma koningii* J. Acta Alimentaria, 30(1): 3-13. doi: 10.1556/AAlim.30.2001.1.2.
- Elisashvili V, Penninckx, M Kachlishvili E, Asatiani M, Kvesitadze G (2006). Use of *Pleurotus dryinus* for lignocellulolytic enzymes production in submerged fermentation of mandarin peels and tree leaves. Enz. Microbial. Technol. 38(7): 998-1004.doi:10.1016/j.enzmictec.2005.08.033.
- Elisashvili V, Kachlishvili E, Penninckx MJ (2008). Lignocellulolytic enzymes profile during growth and fruiting of *Pleurotus ostreatus* on wheat straw and tree leaves. J. Acta Microbiologica et Immunologica Hungarica 55(2): 157-168. doi: 10.1556/ AMicr.55.2008.2.7.
- Emtiazi G, Nahvi I (2000). Multi-enzyme production by *Cellulomonas* sp. grown on wheat straw. Biomass Bioenergy, 19 (1): 31-37. doi:10.1016/S0961-9534(00)00015-5.
- Emtiazi G, Naghavi N, Bordbar A (2001). Biodegradation of lignocellulosic waste by *Aspergillus terreus*. Biodegradation 12(4): 257-261. doi: 10.1023/A:1013155621336.
- Eyheraguibel B, Silvestre J Morard P (2008). Effects of humic substances derived from organic waste enhancement on the growth and mineral nutrition of maize. Bioresour. Technol. 99(10: 4206-4212. doi:10.1016/j.biortech.2007.08.082.
- Fan M (2003). Evaluation of producing fuel and chemicals from corn stover pre-treated with flue gas. Int. J. Environ. Technol. Manag. 3 (3-4): 290-294.
- Fan YT, Zhang YH, Zhang SF, Hou HW, Ren BZ, (2006). Efficient conversion of wheat straw wastes into biohydrogen gas by cow dung compost. Bioresour. Technol. 97(3): 500-505. doi: 10.1016/j.biortech.2005.02.049.
- Fox M, Noike T (2004). Wet oxidation pretreatment for the increase in anaerobic biodegradability of newspaper waste. Bioresour. Technol. 91(3): 273-281 doi:10.1016/j.biortech.2003.06.001.
- Foyle T, Jennings L Mulcahy P (2007). Compositional analysis of lignocellulosic materials: Evaluation of methods used for sugar analysis of waste paper and straw. Bioresour. Technol. 98(16): 3026-3036. doi:10.1016/j.biortech.2006.10.013.
- Gaitán-Hernández R, Salmones D (2008). Obtaining and characterizing *Pleurotus ostreatus* strains for commercial cultivation under warm environmental conditions. Scientia Horticult. 118(2): 106-110. doi: 10.1016/j.scienta.2008.05.029.
- Galbe M, Zacchi G (2007). Pretreatment of lignocellulosic materials for efficient bioethanol production. Adv. Biochem. Eng. Biotechnol. doi: 10.1007/978-3-540-73651-6 108: 41-65.
- Gan Q, Allen SJ, Matthews R (2004). Activation of waste MDF sawdust charcoal and its reactive dye adsorption characteristics. Waste Management. doi: 10.1016/j. wasman. 2004.02.010. 24(8): 841-848.
- Gao J, Weng H, Zhu D, Yuan M, Guan F, Xi Y (2008). Production and characterization of cellulolytic enzymes from the thermoacidophilic fungal *Aspergillus terreus* M11 under solid-state cultivation of corn stover. Bioresour. Technol. doi: 10.1016/j.biortech.2008.02.005. 99(16): 7623-7629.
- García-Gómez A, Bernal MP, Roig A (2005). Organic matter fractions involved in degradation and humification processes during composting. Compost Sci. Util. 13(2) 127-135.
- Georgieva TI, Hou X, Hilstrøm1 T, Ahring BK (2008). Enzymatic hydro-

lysis and ethanol fermentation of high dry matter wet-exploded wheat straw at low enzyme loading. J. Appl. Biochem. Biotechnol. doi: 10.1007/s12010-007-8085-z. 148(1-3): 35-44.

- Georgopoulos ST, Tarantili PA, Avgerinos E, Andreopoulos AG, Koukios EG (2005).Thermoplastic polymers reinforced with fibrous agricultural residues. Poly. Degrad. Stability doi:10.1016/j.polymdegradstab.2005.02.020. 90(2): 303-312.
- Gray KA, Zhao L, Emptage M (2006). Bioethanol. Curr. Opin. Chem. Biol. doi:10.1016/j.cbpa.2006.02.035. 10(2): 141-146.
- Guerra A, Filpponen I, Lucia LA, Saquing C, Baumberger S, Argyropoulos DS (2006). Toward a better understanding of the lignin isolation process from wood. J. Agric. Food Chem., doi: 10.1021/jf060722vS0021-8561(06)00722-9. 54(16): 5939-5947.
- Gürses A, Doğar Ç, Karaca S, Açikyildiz, M, Bayrak, R. (2006). Production of granular activated carbon from waste *Rosa canina* sp. seeds and its adsorption characteristics for dye. J. Hazard. Mater. doi:10.1016/j.jhazmat.2005.09.014. 131(1-3): 254-259.
- Haan RD, Rose SH, Lynd LR, van Zyl WH (2007). Hydrolysis and fermentation of amorphous cellulose by recombinant *Saccharomyces cerevisiae*. Metabolic Eng. doi:10.1016/j.ymben.2006.08.005. 9(1): 87-94.
- Habibi Y, El-Zawawy WK, Ibrahim MM, Dufresne A (2008). Processing and characterization of reinforced polyethylene composites made with lignocellulosic fibers from Egyptian agro-industrial residues. Composites Sci. Technol. 68(7-8): 1877-1885. doi: 10.1016/j.compscitech.2008.01.008.
- Hahn-Hägerdal B, Galbe M, Gorwa-Grauslunda MF, Lidén G, Zacchi G (2006). Bio-ethanol- the fuel of tomorrow from the residues of today. Trends Biotechnol. doi: 10.1016/j.tibtech.2006.10.004. 24(1): 549-556.
- Han SK, Shin HS (2002). Enhanced acidogenic fermentation of food waste in a continuous-flow reactor. Waste Manag. Res. 20(2): 110-118.
- Hanchar RJ, Farzaneh Teymouri F, Nielson CD, McCalla D, Stowers MD (2007). Separation of glucose and pentose sugars by selective enzyme hydrolysis of AFEX-treated corn fiber. J. Appl. Biochem. Biotechnol. doi: 10.1007/s12010-007-9061-3. 137-140(1-12): 313-325.
- Hao JJ, Tian XJ, Song FQ, He XB, Zhang ZJ Zhang P (2006). Involvement of lignocellulolytic enzymes in the decomposition of leaf litter in a subtropical forest. J. Eukaryotic Microbiol. 53(3): 193-198.
- Haq IU, Javed M, Khan TS (2006). An innovative approach for hyperproduction of cellulolytic and hemicellulolytic enzymes by consortium of *Aspergillus niger* MSK-7 and *Trichoderma viride* MSK-10. Afr. J. Biotechnol. 5 (8): 609-614.
- Harman G, Patrick R, Spittler T (2007). Removal of heavy metals from polluted waters using lignocellulosic agricultural waste products. Ind. Biotechnol. doi:10.1089/ind.2007.3.366. 3(4): 366-374.
- Hart TD, Lynch JM,De Leij FAAM (2003). Production of *Trichurus spiralis* to enhance the composting of cellulose-containing waste. Enz. Microbial. Technol. doi: 10.1016/S0141-0229(03)00049-8. 32(6): 745-750.
- Hassan EM, Shukry N (2008): Polyhydric alcohol liquefaction of some lignocellulosic agricultural residues. Ind. Crops Products. doi: 10.1016/j.indcrop.2007.07.004. 27(1): 33-38.
- Hendriks ATWM, Zeeman G (2009). Pretreatments to enhance the digestibility of lignocelluloses biomass. Bioresour. Technol. doi:10.1016/j.biortech.2008.05.027. 100(1)10-18.
- Henrique M, Baudel HM, Zaror C, de Abreu CAM (2005). Improving the value of sugarcane bagasse wastes via integrated chemical production systems: An environmentally friendly approach. Ind. Crops Products, doi: 10.1016/j.indcrop.2004.04.013. 21 (3): 309-315.
- Herrera A, Téllez-Luis SJ, González-Cabriales JJ, Ramírez JA, Vázquez M (2004): Effect of the hydrochloric acid concentration on the hydrolysis of sorghum straw at atmospheric pressure. J. Food Eng. doi: 10.1016/S0260-8774(03)00288-7. 6(1): 103-109.
- Howard RL, Abotsi E, Jansen van Rensburg EL, Howard S (2003). Lignocellulose biotechnology: Issues of bioconversion and enzyme production. Afr. J. Biotechnol. 2(12): 602-619.
- Hu ZH, Yu HQ (2005). Application of rumen microorganisms for enhanced anaerobic fermentation of corn stover. Process Biochem. doi: 10.1016/j.procbio.2004.09.021. 40(7): 2371-2377

- Hu ZH, Yu HQ (2006). Anaerobic digestion of cattail by rumen cultures. Waste Manag. doi: 10.1016/j.wasman.2005.08.003. 26(1): 1222-1228.
- Hu ZH, Liu SY, Yue ZB, Yan LF, Yang MT, Yu HQ (2008). Microscale analysis of *in vitro* anaerobic degradation of lignocellulosic wastes by rumen microorganisms. Environ. Sci. Technol. doi: 10.1021/es071915h. 42(1): 276–281.
- Huang DL, Zeng GM, Feng CL, Hu S, Jiang XY, Tang L, Su FF, Zhang Y, Zeng W, Liu HL (2008). Degradation of lead-contaminated lignocellulosic waste by *Phanerochaete chrysosporium* and the reduction of lead toxicity. Environ. Sci. Technol. 42(13): 4946–4951. doi: 10.1021/es800072c.
- Ibrahim MNM, Nadiah MYN, Norliyana MS, Sipaut CS, Shuib S (2008). Separation of vanillin from oil palm empty fruit bunch lignin. Acta hydrochimica et hydrobiologica 36(3): 287-291.
- Iñiguez-Covarrubias G, Lange SE, Rowell RM (2001). Utilization of byproducts from the tequila industry: part 1: Agave bagasse as a raw material for animal feeding and fiberboard production. Bioresour. Technol. 77(1): 25-32. doi:10.1016/S0960-8524(00)00137-1.
- Inoue H, Yano S, Endo T, Sakaki T, Sawayama S (2008). Combining hot-compressed water and ball milling pretreatments to improve the efficiency of the enzymatic hydrolysis of eucalyptus. Biotechnol Biofuels. 2008; 1(2): 1-9. doi: 10.1186/1754-6834-1-2.
- Iranmahboo J, Nadim F, Monemi S (2002). Optimizing acid-hydrolysis: a critical step for production of ethanol from mixed wood chips. Biomass Bioenergy 22(5): 401-404. doi: 10.1016/S0961-9534(02)00016-8.
- Isil S, Nilurér A (2005). Investigation of factors affecting xylanase activity from *Trichoderma harzianum* 1073 D3. Braz. Arch. Biol. Technol. 48(2): 1516-8913. doi: 10.1590/S1516-89132005000200004.
- Israilides C, Philippoussis A (2003). Bio-technologies of recycling agroindustrial wastes for the production of commercially important polysaccharides and mushrooms. Biotechnol. Gen. Eng. Reviews 20(2003), pp. 247–259.
- Ito T, Nakashimada Y, Senba K, Matsui T, Nishio N (2005). Hydrogen and ethanol production from glycerol-containing wastes discharged after biodiesel manufacturing process. J. Biosci. Bioeng. 100(3): 260-265. doi: 10.1263/jbb.100.260.
- Itoh H, Wada M, Honda Y, Kuwahara M, Watanabe T (2003). Bioorganosolve pretreatments for simultaneous saccharification and fermentation of beech wood by ethanolysis and white rot fungi. J. Biotechnol. 103(3): 273-280. doi: 10.1016/S0168-1656(03)00123-8.
- Jacob M, Thomas S, Varughese KT (2006). Novel woven sisal fabric reinforced natural rubber composites tensile and swelling characteristics. J. Comp. Mater. 40(16): 1471-1485. doi: 10.1177/002199830605973.
- Jarboe LR, Grabar TB, Yomano LP, Shanmugan KT, Ingram LO (2007). Development of ethanologenic bacteria. Adv Biochem Eng Biotechnol. 108: 237-261. doi: 10.1007/10_2007_068.
- Jecu L (2000). Solid state fermentation of agricultural wastes for endoglucanase production. Ind. Crops and Products. 11(1): 1-5. doi: 10.1016/S0926-6690(99)00022-9.
- Jeoh T, Agblevor FA (2001). Characterization and fermentation of steam exploded cotton gin waste. Biomass and Bioenergy 21(2): 109-120. doi:10.1016/S0961-9534(01)00028-9.
- Jin F, Zheng J, Enomoto H, Moriya T, Sato N, Higashijima H (2002. Hydrothermal process for increasing acetic acid yield from lignocellulosic wastes. Chemistry Lett. 31(5): p. 504. doi:10.1246/cl.2002.504.
- Jin F, Cao J, Zhou Z, MoriyaT, Enomoto H (2004). Effect of lignin on acetic acid production in wet oxidation of lignocellulosic. wastes. Chem. Lett. 33: p. 910 doi:10.1246/cl.2004.910.
- Jin F, Zhou Z, Kishita A, Enomoto H (2006). Hydrothermal conversion of biomass into acetic acid. J. Mater. Sci. 41(5): 1495-1500. doi: 10.1007/s10853-006-7493-8.
- John MJ, Francis B, Varughese KT, Thomas S (2008). Effect of chemical modification on properties of hybrid fiber biocomposites. Composites Part A: Appl. Sci. Manuf. 39(2): 352-363. doi:10.1016/j.compositesa.2007.10.002.
- Kádár Z, Réczey K (2004). Simultaneous saccharification and fermentation (SSF) of industrial wastes for the production of ethanol. Ind. Crops Prod. 20(1): 103-110. doi:10.1016/j.indcrop.2003.12.015.

- Kalm E, Sargın S (2004). Cultivation of two *Pleurotus* species on wheat straw substrates containing olive mill waste water. Int. Biodeter. Biodegrad. 53(1): 43-47. doi:10.1016/j.ibiod.2003.08.002.
- Karimi K, Kheradmandinia S, Taherzadeh MJ (2006). Conversion of rice straw to sugars by dilute-acid hydrolysis. Biomass Bioenergy 30(3): 247-253. doi:10.1016/j.biombioe.2005.11.015.
- Keller FA, Hamilton JE, Nguyen QA (2003). Microbial pretreatment of biomass: Potential for reducing severity of thermochemical biomass pretreatment. J. Appl. Biochem. Biotechnol. 105(1-3): 27-41. doi: 10.1385/ABAB:105:1-3:27.
- Kemppainen AJ, Shonnard DR (2005). Comparative life-cycle assessments for biomass-to-ethanol production from different regional feedstocks. Biotechnol. Prog. 21 (4): 1075-1084. doi: 10.1021/bp049548q S8756-7938(04)09548-7.
- Kim J, Park C, Kim TH, Lee M, Kim S, Kim SW, Lee J (2003). Effects of various pretreatments for enhanced anaerobic digestion with waste activated sludge. J. Biosci. Bioeng. 95(3): 271-275. doi: 10.1016/S1389-1723(03)80028-2.
- Kim Y, Hendrickson R, Mosier NS, Ladisch MR, Bals B, Balan V, Dale BE (2008). Enzyme hydrolysis and ethanol fermentation of liquid hot water and AFEX pretreated distillers' grains at high-solids loadings. Bioresour. Technol. 99(12): 5206-5215. doi: 10.1016/j.biortech.2007.09.031.
- Kishore KK, Parimala V, Gupta BP, Azad IS, Meng X, Abraham M (2006). Bagasse-assisted bioremediation of ammonia from shrimp farm wastewater. Water Environ. Res. 78(9): 938-950(13).
- Kobayashi M, Asano T, Kajiyama M, Tomita B (2004). Analysis on residue formation during wood liquefaction with polyhydric alcohol. J. Wood Sci. 50(5): 407-414. doi: 10.1007/s10086-003-0596-9.
- Krishnani KK, Ayyappan S (2006). Heavy metals remediation of water using plants and lignocellulosic agrowastes. Rev. Environ. Contam. Toxicol. 188: 59-84.
- Kumar R, Singh S, Singh OV (2008). Bioconversion of lignocellulosic biomass: Biochemical and molecular perspectives. J. Ind. Microbiol. Biotechnol. 35(5): 377-391. Doi: 10.1007/s10295-008-0327-8.
- Lapuerta M, Hernández JJ, Rodríguez J (2004). Kinetics of devolatilisation of forestry wastes from thermogravimetric analysis. Biomass Bioenergy. 27(4): 385-391 doi: 10.1016/j.biombioe.2003.11.010.
- Lertsutthiwong P, Khunthon S, Siralertmukul K, Noomun K, Chandrkrachang S (2008). New insulating particleboards prepared from mixture of solid wastes from tissue paper manufacturing and corn peel. Bioresour. Technol. 99(11): 4841-4845. doi:10.1016/j.biortech.2007.09.051.
- Li A, Antizar-Ladislao B, Khraisheh M (2007). Bioconversion of municipal solid waste to glucose for bio-ethanol production. Bioprocess Biosystems Eng. 30(3): 189-196. doi: 10.1007/s00449-007-0114-3.
- Li W, Xu J, Wang J, Yan YJ, Zhu XF, Chen MQ, Tan ZC (2008). Studies of monosaccharide production through lignocellulosic waste hydrolysis using double acids. Energy Fuels, 22(3): 2015–2021. doi: 10.1021/ef700762h.
- Lim SJ, Kim BJ, Jeong CM, Choi J, Ahn YH, Chang HN (2008). Anaerobic organic acid production of food waste in once-a-day feeding and drawing-off bioreactor. Bioresour. Technol. 99(16): 7866-7874. doi: 10.1016/j.biortech.2007.06.028.
- Lin Y, Tanaka S (2006). Ethanol fermentation from biomass resources: Current state and prospects. J. Appl. Microbiol. Biotechnol. 69 (6): 627-642.
- Lissens G, Klinke H, Verstraete W, Ahring B, Thomsen AB (2004). Wet oxidation pre-treatment of woody yard waste: Parameter optimization and enzymatic digestibility for ethanol production. J. Chem. Technol. Biotechnol. 79(8): 889-895.
- Liu HW, Walter HK, Vogt GM, Vogt HS, Holbein BE (2002). Steam pressure disruption of municipal solid waste enhances anaerobic digestion kinetics and biogas yield. Biotechnol. Bioeng. 77(2): 121-130. DOI: 10.1002/bit.10130.
- Liu B, Zhao ZK (2007). Biodiesel production by direct methanolysis of oleaginous microbial biomass. J. Chem. Technol. Biotechnol. 82(8): 775-780. doi: 10.1002/jctb.1744.
- Locci E, Laconi S, Pompei R, Scano P, Lai A, Marincola FC (2008). Wheat bran biodegradation by *Pleurotus ostreatus*: A solid-state

- Carbon-13 NMR study. Bioresour. Technol. 99(10): 4279-4284. doi:10.1016/j.biortech.2007.08.048.
- Lopez MJ, Carmen M, Vargas-García C, Suárez-Estrella F, Moreno J (2006). Biodelignification and humification of horticultural plant residues by fungi. Int. Biodeterioration Biodegradation. 57(1): 24-30. doi:10.1016/j.ibiod.2005.10.005.
- López DGJ, Balart R, Ruseckaite RA, Stefani PM (2007). Composites based on sintering rice husk–waste tire rubber mixtures. Materials Design 28(7): 2234-2238. doi:10.1016/j.matdes.2006.06.001.
- Lu Y, Mosier NS (2007). Biomimetic catalysis for hemicellulose hydrolysis in corn stover. Biotechnol. Prog., 23(1): 116-123. doi: 10.1021/bp060223e S8756-7938(06)00223-2.
- Mabrouk MEM, El-Ahwany AMD (2008). Production of β-mannanase by *Bacillus amylolequifaciens* cultured on potato peels. Afr. J. Biotechnol. 7(8): 1123-1128.
- Madani M, Altaf HB, Abdo AE, Houssni E (2004). Utilization of waste paper in the manufacture of natural rubber composite for radiation shielding. Progress in rubber, plastics Recycling Technol. 20(4): 287-310.
- Mahro B, Timm M (2007). Potential of Biowaste from the Food Industry as a Biomass Resource. Eng. Life Sci. 7: 457-468. doi:10.1002/elsc.200620206.
- Mani S, Tabil LG, Sokhansanj S (2004). Grinding performance and physical properties of wheat and barley straws, corn stover and switchgrass. Biomass Bioenergy, 27(4): 339-352. doi:10.1016/j.biombioe.2004.03.00.
- Marques S, Alves L, Roseiro JC, Gírio FM (2008). Conversion of recycled paper sludge to ethanol by SHF and SSF using *Pichia stipitis*. Biomass Bioenergy, 32(5): 400-406. doi:10.1016/j.biombioe.2007.10.011.
- Martín C, Marce M, Thomsen AB (2008). Comparison of wet oxidation and steam explosion as pretreatment methods for bioethanol production from sugarcane bagasse. BioResour. 3(3): 670-683.
- Melamane X, Tandlich R, Burgess J (2007). Anaerobic digestion of fungally pre-treated wine distillery wastewater. Afr. J. Biotechnol. 6(17): 1990-1993.
- Merino ST, Cherry J (2007). Progress and challenges in enzyme development for biomass utilization. Adv. Biochem. Eng. Biotechnol. 108: 95-120. doi: 10.1007/10_2007_066.
- Miller S, Hester R, Mississippi H (2007). Concentrated acid conversion of pine sawdust to sugars. Part II: High-temperature batch reactor kinetics of pretreated pine sawdust Chem. Eng. Communications, 194(1):103-116. doi: 10.1080/00986440600715854.
- Minussi RC, Miranda MA, Silva JA, Ferreira CV, Aoyama H, Marangoni S, Rotilio D, Pastore GM, Durán N (2007). Purification, characterization and application of laccase from *Trametes versicolor* for colour and phenolic removal of olive mill wastewater in the presence of 1-hydroxybenzotriazole. Afr. J. Biotechnol. 6(10): 1248-1254.
- Mishra S, Mohanty AK, Drzal LT, Misra M, Hinrichsen G (2004). A review on pineapple leaf fibers, sisal fibers and their biocomposites. Macromolecular Mater. Eng. 289(11): 955-974. doi: 10.1002/mame.200400132.
- Mishra A, Kumar S (2007). Cyanobacterial biomass as N-supplement to agro-waste for hyper-production of laccase from *Pleurotus ostreatus* in solid state fermentation. Process Biochem. 42(4): 681-685. doi:10.1016/j.procbio.2006.09.022.
- Mohana S, Shah A, Divecha J, Madamwar D (2008). Xylanase production by *Burkholderia* sp. DMAX strain under solid state fermentation using distillery spent wash. Bioreasour. Technol. 99(16): 7553-7564. doi:10.1016/j.biortech.2008.02.009.
- Moldes D, Gallego PP, Rodríguez S, Couto SR, Sanromán A (2003). Grape seeds: the best lignocellulosic waste to produce laccase by solid state cultures of *Trametes hirsute*. J. Biotechnol. Lett. 25(6): 491-495. DOI: 10.1023/A:1022660230653
- Moldes D, Lorenzo M, Sanromán MA (2004). Different proportions of laccase isoenzymes produced by submerged cultures of *Trametes versicolor* grown on lignocellulosic wastes. J. Biotechnol. Lett. 26(4): 327-330 doi: 10.1023/B:BILE.0000015452.40213.bf.
- Morais MH, Ramos AC, Matos N (2000). Production of shiitake mushroom (*Lentinus edodes*) on lignocellulosic residues. Food Sci. Technol. Int. 6(2): 123-128. doi: 10.1177/108201320000600206.
- Mosier N, Wyman C, Dale B, Elander R, Lee YY, Holtzapple M, Ladisch

M (2005). Features of promising technologies for pretreatment of lignocellulosic biomass. Bioresour. Technol. 96(6): 673-686. doi:10.1016/j.biortech.2004.06.025.

- Mshandete A, Björnsson L, Kivaisi AK, Rubindamayugi ST, Mattiasson B (2005). Enhancement of anaerobic batch digestion of sisal pulp waste by mesophilic aerobic pre-treatment. Water Res. 39(8): 1569-1575. doi:10.1016/j.watres.2004.11.037.
- Mshandete AM, Björnsson L, Kivaisi AK, MST Mattiasson B (2008). Effect of aerobic pre-treatment on production of hydrolases and volatile fatty acids during anaerobic digestion of solid sisal leaf decortications residues. Afric. J. Biochem. Res. 2(5):111-119.
- Mtui G, Nakamura Y (2005): Bioconversion of lignocellulosic waste from selected dumping sites in Dar es Salaam, Tanzania Biodegradation, 16(6): 493-499 doi: 10.1007/s10532-004-5826-3.
- Mtui G (2007). Trends in industrial and environmental biotechnology research in Tanzania. Afr. J. Biotechnol. 6(25): 2860-2567.
- Mtui G, Nakamura Y (2007). Characterization of lignocellulosic enzymes from white-rot fungus *Phlebia crysocreas* isolated from a marine habitat. J. Eng. Appl. Sci. 2: 1501-1508.
- Mtui G, Nakamura Y (2008). Lignocellulosic enzymes from *Flavodon flavus*, a fungus isolated from Western Indian Ocean off the Coast of Dar es Salaam, Tanzania. Afr. J. Biotechnol. 7(17): 3066-3072.
- Mtui G, Masalu R (2008). Extracellular enzymes from Brown-rot fungus *Laetioporus sulphureus* isolated from mangrove forests of Coastal Tanzania. Sci. Res. Essay 3: 154-161.
- Mukherjee R, Nandi B (2004). Improvement of in vitro digestibility through biological treatment of water hyacinth biomass by two *Pleurotus* species. Int. Biodeterioration Biodegrad. 53(1): 7-12 doi: 10.1016/S0964-8305(03)00112-4.
- Murphy JD, McCarthy K (2005). Ethanol production from energy crops and wastes for use as a transport fuel in Ireland. Appl. Energy 82(2): 148-166 doi:10.1016/j.apenergy.2004.10.004.
- Muruke MHS, Hosea KM, Palangyo A, Heijthuijsen JHFG (2006). Production of lactic acid from waste sisal stems using a *Lactobacillus* isolate. Discovery Innov., 18(1): 1-5.
- Muthuvelayudham R, Viruthagiri T (2006). Fermentative production and kinetics of cellulase protein on *Trichoderma reesei* using sugarcane bagasse and rice straw. Afr. J. Biotechnol. 5 (20): 1873-1881.
- Najafpour G, Ismail KSK, Younesi H, Mohamed AR, Kamaruddin AH (2004). Hydrogen as clean fuel via continuous fermentation by anaerobic photosynthetic bacteria, *Rhodospirillum rubrum*. Afr. J. Biotechnol. (10): 503-507.
- Namasivayam C, Sureshkumar MV (2006). Anionic dye adsorption characteristics of surfactant-modified coir pith, a waste lignocellulosic polymer. J. Appl. Poly. Sci. 100(2): 1538-1546.
- Nazareth SW, Sampy JD (2003). Production and characterisation of lignocellulases of *Panus tigrinus* and their application. Int. Biodeterioration Biodegradation, 52(4): 207-214. doi: 10.1016/S0964-8305(03)00051-9.
- Ndazi B, Tesha JV, Bisanda TN (2006). Some opportunities and challenges of producing bio-composites from non-wood residues. J. Mater. Sci. 41(21): 6984-6990. doi 10.1007/s10853-006-0216-3.
- Neves L, Oliveira R, Alves MM (2006). Anaerobic co-digestion of coffee waste and sewage sludge. Waste Manag. 26(2): 176-181. doi: 10.1016/j.wasman.2004.12.022.
- Ngah WSW, Hanafiah MAKM (2008). Removal of heavy metal ions from wastewater by chemically modified plant wastes as adsorbents: A review. Bioresour. Technol. 99(10): 3935-3948. doi: 10.1016/j.biortech.2007.06.011.
- Nikolov T, Bakalova N, Petrova S, Benadova R, Spasov S, Kolev D (2000). An effective method for bioconversion of delignified wastecellulose fibers from the paper industry with a cellulase complex. Bioresour. Technol. 71(1): 1-4 doi:10.1016/S0960-8524(99)00059-0.
- Ögel ZB, Yarangümeli K, Dündar H, Ifrij I (2001). Submerged cultivation of *Scytalidium thermophilum* on complex lignocellulosic biomass for endoglucanase production. Enzyme Microbial. Technol. 28(7-8): 689-695 doi: 10.1016/S0141-0229(01)00315-5.
- Öhgren K, Vehmaanperä J, Siika-Aho M, Galbe M, Viikari L, Zacchi G (2007). High temperature enzymatic prehydrolysis prior to simultaneous saccharification and fermentation of steam pretreated corn stover for ethanol production. Enzyme Microbial. Technol. 40(4): 607-613. doi: 10.1016/j.enzmictec.2006.05.014.

- Ohkouchi Y, Inoue Y (2007). Impact of chemical components of organic wastes on I(+)-lactic acid production. Bioresour. Technol. 98(3): 546-553. doi:10.1016/j.biortech.2006.02.005.
- Ojumu TV, Solomon BO, Betiku E, Layokun SK, Amigun B (2003). Cellulase Production by *Aspergillus flavus* Linn Isolate NSPR 101 fermented in sawdust, bagasse and corncob. Afr. J. Biotechnol. 2(6): 150-152.
- Okafoagu CU, Nzelibe, HC (2006). Effect of acid hydrolysis of *Garcinia kola* (bitter kola) pulp waste on the production of CM-cellulase and βglucosidase using *Aspergillus Niger*. Afr. J. Biotechnol. 5 (10): 819-822.
- Oku T (2004). Sawdust-based cultivation of some mushrooms using unutilized wood resources Bulletin Utsunomiya Univ. Forests 40: 1-67.
- Okuda N, Ninomiya K, Takao M, Katakura Y, Shioya S (2007). Microaeration enhances productivity of bioethanol from hydrolysate of waste house wood using ethanologenic *Escherichia coli* KO11. J. Biosci. Bioeng. 103(4): 350-357. doi: 10.1263/jbb.103.350.
- Oliveira LA, Porto ALF, Tambourgi EB (2006). Production of xylanase and protease by *Penicillium janthinellum* CRC 87M-115 from different agricultural wastes. Bioreasour. Technol. 97(6): 862-867 doi: 10.1016/j.biortech.2005.04.017.
- Olofsson K, Bertilsson M, Lidén G (2008). A short review on SSF, an interesting process option for ethanol production from lignocellulosic feedstocks. Biotechnol Biofuels 1(7): 1-14. doi: 10.1186/1754-6834-1-7. PMCID: PMC2397418.
- Olson ES (2001). Conversion of lignocellulosic material to chemicals and fuels. National Energy Technology Lab., Pittsburgh, PA (US); National Energy Technology Lab., Morgantown, WV(US). Report No. FC26-98FT40320-19. doi: 10.2172/786842.
- Orlando US, Baes AU, Nishijima W, Okada M (2002). A new procedure to produce lignocellulosic anion exchangers from agricultural waste materials. Bioreasour. Technol. 83(3): 195-198. doi: 10.1016/S0960-8524(01)00220-6.
- Otero JM, Panagiotou G, Olsson L (2007). Fueling industrial biotechnology growth with bioethanol. Adv. Biochem. Eng. Biotechnol. 108:1-40. doi. 10.1007/978-3-540-73651-6
- Özçelika E, Pekşen A (2007). Hazelnut husk as a substrate for the cultivation of shiitake mushroom (*Lentinula edodes*). Bioresour. Technol. 98(14): 2652-2658. doi: 10.1016/j.biortech.2006.09.020.
- Pandey A, Soccol CR, Poonam Nigam P, Soccol VT (2000). Biotechnological potential of agro-industrial residues. I: sugarcane bagasse. Bioresour. Technol. 7(1): 69-80. doi:10.1016/S0960-8524(99)00142-X.
- Pandey P, Pandey AK (2002). Production of cellulase-free thermostable xylanases by an isolated strain of *Aspergillus Niger* PPI, utilizing various lignocellulosic wastes World J. Microbiol. Biotechnol. 18(3): 281-283. doi: 10.1023/A:1014999728406.
- Pasha C, Nagavalli M, Rao LV (2007). *Lantana camara* for fuel ethanol production using thermotolerant yeast. Lett. Appl. Microbiol. 44 (6): 666-672. doi:10.1111/j.1472-765X.2007.02116.x.
- Patel SJ, Onkarappa R, Shobha KS (2007). Study of ethanol production from fungal pretreated wheat and rice straw. The Internet. J. Microbiol. 4(1): 1-4.
- Peker H, Baysal E, Yigitbasi ON, Simsek H, Colak M, Toker H (2007). Cultivation of *Agaricus bisporus* on wheat straw and waste tea leaves based compost formulas using wheat chaff as activator material. Afr. J. Biotechnol. 6(4): 400-409.
- Pérez J, Muñoz-Dorado J, de la Rubia T, Martínez J (2002). Biodegradation and biological treatments of cellulose, hemicellulose and lignin: an overview. J. Int. Microbiol. 5(2): 53-63. doi: 10.1007/s10123-002-0062-3.
- Peterson JD, Ingram LO (2008). Anaerobic respiration in engineered *Escherichia coli* with an internal electron acceptor to produce fuel ethanol. Ann. NY Acad. Sci. 1125: 363-372.
- Phan AN, Ryu C, Sharifi VN, Swithenbank J (2008). Characterisation of slow pyrolysis products from segregated wastes for energy production. J. Analyt. Appl. Pyrolysis, 81(1). 65-71. doi: 10.1016/j.jaap.2007.09.001.
- Philippoussis A, Zervakis G, Diamantopoulou P (2001). Bioconversion of agricultural lignocellulosic wastes through the cultivation of the edible mushrooms Agrocybe aegerita, Volvariella volvacea and Pleu-

rotus spp. World J. Microbiol. Biotechnol. 17(2): 191-200 DOI 10.1023/A:1016685530312.

- Philippoussis A, Diamantopoulou P, Israilides C (2007). Productivity of agricultural residues used for the cultivation of the medicinal fungus *Lentinula edodes*. Int. Biodeterioration Biodegradation, 59(3): 216-219. doi:10.1016/j.ibiod.2006.10.007.
- Pothan LA, George CN, Jacob M, Thomas S (2007). Effect of chemical modification on the mechanical and electrical properties of banana fiber polyester composites. J. Comp. Materials, 41(19): 2371-2386. doi: 10.1177/0021998307075456.
- Pothiraj C, Balaji P, Eyini M (2006). Enhanced production of cellulases by various fungal cultures in solid state fermentation of cassava waste. Afr. J. Biotechnol. 5(20): 1882-1885.
- Prasad S, Singh A, Joshi H (2007). Ethanol as an alternative fuel from agricultural, industrial and urban residues. Reasourc. Conserv. Recycling, 50(1): 1-39 doi: 10.1016/j.resconrec.2006.05.007.
- Puértolas R, Gea G, Murillo MB, Arauzo J (2001). Pyrolysis of black liquors from alkaline pulping of straw. Influence of a preoxidation stage on the char characteristics. J. Analyt. Appl. Pyrolysis, pp. 58-59 (1): 955-966. doi:10.1016/S0165-2370(00)00175-3.
- Qi BC, Aldrich C, Lorenzen L, Wolfaardt GW (2005). Acidogenic fermentation of lignocellulosic substrate with activated sludge. Chem. Eng. Communications, 192(9): 1221-1242. doi: 10.1080/009864490515676.
- Rahman SHA, Choudhury JP, Ahmad AL, Kamaruddin AH (2007): Optimization studies on acid hydrolysis of oil palm empty fruit bunch fiber for production of xylose. Bioreasour. Technol. 98(3): 554-559. doi:10.1016/j.biortech.2006.02.016.
- Ragunathan R, Swaminathan K (2004). Bioconversion of lignocellulosic agro-wastes by fungus, *Pleurotus spp.* Biological Memoirs, 30(1): 1-6.
- Raj K, Singh R (2001). Semi-solid-state fermentation of *Eicchornia crassipes* biomass as lignocellulosic biopolymer for cellulase and β-glucosidase production by co-cultivation of Aspergillus niger RK3 and *Trichoderma reesei* MTCC164. Appl. Biochem. Biotechnol. 96(1-3): 71-82.
- Rani P, Kalyani N, Prathiba K (2008). Evaluation of Lignocellulosic Wastes for Production of Edible Mushrooms. J. Appl. Biochem. Biotechnol (in Press) (2008). doi: 10.1007/s12010-008-8162-y.
- Reis JML (2006). Fracture and flexural characterization of natural fiberreinforced polymer concrete. Con. Build. Mat. 20(9): 673-678. doi:10.1016/j.conbuildmat.2005.02.008.
- Rezende MI, Barbosa AM, Vasconcelos AFD, Endo AS (2002). Xylanase production by *Trichoderma harzianum* Rifai by solid state fermentation on sugarcane bagasse. Braz. J. Microbiol. 33(1): 67-72. doi: 10.1590/S1517-83822002000100014.
- Robinson T, Nigam P (2003). Bioreactor design for protein enrichment of agricultural residues by solid state fermentation. Biochem. Eng. J. 13(2-3): 197-203. doi: 10.1016/S1369-703X(02)00132-8.
- Rodríguez-Chonga A, Ramírez JA, Garrote G, Vázquez M (2004). Hydrolysis of sugar cane bagasse using nitric acid: a kinetic assessment. J. Food Eng. 61(2): 143-152. doi: 10.1016/S0260-8774(03)00080-3.
- Rodríguez G, Lama A, Rodríguez R, Jiménez A, Guilléna R, Fernández-Bolaños J (2008). Olive stone an attractive source of bioactive and valuable compounds. Bioreasour. Technol. 99(13): 5261-5269. doi: 10.1016/j.biortech.2007.11.027.
- Roig A, Cayuela ML, Sánchez-Monedero MA (2006). An overview on olive mill wastes and their valorisation methods. Waste Manag. 26(9): 960-969. doi:10.1016/j.wasman.2005.07.024.
- Roman HJ, Burgess JE, Pletschke BI (2006). Enzyme treatment to decrease solids and improve digestion of primary sewage sludge. Afr. J. Biotechnol. 5(10): 963-967.
- Rosales E, Couto SR, Sanromán MA (2007). Increased laccase production by *Trametes hirsuta* grown on ground orange peelings. Enz. Microbial. Technol. 40(5): 1286-1290. doi:10.1016/j.enzmictec.2006.09.015.
- Rubin EM (2008). Genomics of cellulosic biofuels. Nat. 454(14): 841-845. doi: 10.1038/nature07190.
- Saha BC (2003). Hemicellulose bioconversion. J. Ind. Microbiol. Biotechnol. 30: 279-29. doi: 10.1007/s10295-003-0049-x.
- Salètes S, Siregar FA, Caliman JP, Liwang T (2004). Lignocellulose

composting: Case study on monitoring oil palm residuals. Compost. Sci. Util. 12(4): 372-382.

- Sánchez OJ, Cardona CA (2008). Trends in biotechnological production of fuel ethanol from different feedstocks. Bioresour. Technol. 99(13): 5270-5295 doi:10.1016/j.biortech.2007.11.013.
- Sassner P, Galbe M, Zacchi G (2008). Techno-economic evaluation of bioethanol production from three different lignocellulosic materials. Biomass Bioenergy 32(5): 422-430. doi:10.1016/j.biombioe.2007.10.014.
- Schenk PM, Thomas-Hall SR, Stephens E, Marx UC, Mussgnug Posten JHC, Kruse O, Ben Hankamer B (2008). Second Generation Biofuels: High-Efficiency Microalgae for Biodiesel Production. J. BioEnergy Res. 1(1): 20-43. doi: 10.1007/s12155-008-9008-8.
- Schilling CH, Tomasik P, Karpovich DS, Hart B, Garcha J, Boettcher PT (2004). Preliminary studies on converting agricultural waste into biodegradable plastics Part III: Sawdust. J. Poly. Environ. 13: 177-183. doi: 10.1007/s10924-005-2948-6.
- Schneider UA, McCar BA (2003). Economic potential of biomass based fuels for greenhouse gas emission mitigation. J. Environ. Resour. Economics, 24(4): 291-312. doi: 10.1023/A:1023632309097.
- Šćiban M, Klašnja M, Škrbić B (2008). Adsorption of copper ions from water by modified agricultural by-products. Desalination 229(1-3): 170-180. doi: 10.1016/j.desal.2007.08.017.
- Scott M, Roger H (2007). Concentrated acid conversion of pine sawdust to sugars. Part II: High-temperature batch reactor kinetics of pretreated pine sawdust. Chem. Eng. Communications 194(1): 103-116. doi: 10.1080/00986440600715854.
- Sedlak M, Ho NWY (2004). Production of ethanol from cellulosic biomass hydrolysates using genetically engineered *Saccharomyces* yeast capable of cofermenting glucose and xylose. J. Appl. Biochem. Biotechnol. 114(1-3): 403-416. doi: 10.1385/ABAB:114:1-3:403.
- Sepúlveda-Huerta E, Tellez-Luis SJ, Bocanegra-García V, RamírezJA, Vázquez M (2006). Production of detoxified sorghum straw hydrolysates for fermentative purposes. J. Sci. Food Agr. 86(15): 2579-2586. doi:10.1002/jsfa.2651.
- Shaji J, Kuruvilla J, Sabu T (2006). Green composites from natural rubber and oil palm fiber: physical and mechanical properties. Int. J. Poly. Mater. 55(11): 925-945.
- Shi J, Chinn MS, Sharma-Shivappa RR (2008). Microbial pretreatment of cotton stalks by solid state cultivation of *Phanerochaete chrysosporium*. Bioresour. Technol. 99(14). 6556-6564. doi:10.1016/j.biortech.2007.11.069.
- Shide EG, Wuyep PA, Nok A (2004). Studies on the degradation of wood sawdust by *Lentinus squarrosulus* (Mont.) Singer. Afr. J. Biotechnol. 3(8): 395-398.
- Silva D, Tokuioshi K, Martins ED, Da Silva R, Gomes E (2005). Production of pectinase by solid-state fermentation with *Penicillium viridicatum.* RFC3 Process Biochem. 40(8): 2885-2889. doi:10.1016/j.procbio.2005.01.008.
- Silva EM, Machuca A, Milagres AMF (2005). Effect of cereal brans on *Lentinula edodes* growth and enzyme activities during cultivation on forestry waste. Letters in Appl. Microbiol. 40(4): 283-288.
- Sinegani AAS, Emtiazi G, Hajrasuliha S, Shariatmadari H (2005). Biodegradation of some agricultural residues by fungi in agitated submerged cultures. Afr. J. Biotechnol. (10): 1058-1061.
- Singh SP, Asthana RK, Singh (2007). Prospects of sugarcane milling waste utilization for hydrogen production in India. Energy Policy 35(8): 4164-4168. doi:10.1016/j.enpol.2007.02.017.
- Singh P, Suman A, Tiwari P, Arya N, Gaur A, Shrivastava AK (2008). Biological pretreatment of sugarcane trash for its conversion to fermentable sugars. World J. Microbiol. Biotechnol. 24(5): 667-673. doi: 10.1007/s11274-007-9522-4.
- Sjöde A, Alriksson B, Jönsson LJ, Nilvebrant NO (2007). The potential in bioethanol production from waste fiber sludges in pulp mill-based biorefineries. J. Appl. Biochem. Biotechnol. pp.137-140(1-12): 327-337. doi: 10.1007/s12010-007-9062-2.
- Songulashvili G, Elisashvili V, Penninckx M, Metreveli E, Hadar Y, Aladashvili N, Asatiani M (2005). Bioconversion of plant raw materials in value-added products by *Lentinus edodes* and *pleurotus* spp. Int. J. of Med. Mushrooms 7(3): 467-468.
- Songulashvili G, Elisashvili V, Wasser SP, Nevo E, Hadar Y (2007). Basidiomycetes laccase and manganese peroxidase activity in sub-

merged fermentation of food industry wastes. Enz. Microbial. Technol. 41(1-2): 57-61 doi:10.1016/j.enzmictec.2006.11.024.

- Sørensen A, Teller PJ, Hilstrøm T, Åhring BK (2008). Hydrolysis of *Miscanthus* for bioethanol production using dilute acid presoaking combined with wet explosion pre-treatment and enzymatic treatment. Bioresour. Technol. 99(14): 6602-6607. doi: 10.1016/j.biortech.2007.09.091.
- Sun Y, Cheng J (2002). Hydrolysis of lignocellulosic materials for ethanol production: A review. Bioresour. Technol. 83(1): 1-11. doi: 10.1016/S0960-8524(01)00212-7.
- Suthar S (2007). Production of vermifertilizer from guar gum industrial wastes by using composting earthworm *Perionyx sansibaricus* (Perrier). J. Environmentalist 27(3): 329-335. doi: 10.1007/s10669-007-9032.
- Tabka MG, Herpoël-Gimbert I, Monod F, Asther M, Sigoillot JC (2006). Enzymatic saccharification of wheat straw for bioethanol production by a combined cellulase xylanase and feruloyl esterase treatment. Enz. Microbial. Technol. 39(4,2): 897-902. doi:10.1016/j.enzmictec.2006.01.021.
- Tahezaden MJ, Karimi K (2007). Enzyme-based ethanol: A review: Bioresour. 2(4): 707-738.
- Taiwo LB, Oso BA (2004). Influence of composting techniques on microbial succession, temperature and pH in a composting municipal solid waste. Afr. J. Biotechnol. 3(4): 239-243.
- Taşeli, BK. (2008). Fungal treatment of hemp-based pulp and paper mill wastes. Afric. J. Biotechnol. 7 (3): 286–289.
- Tengerdy RP, Szakacs G (2003). Bioconversion of lignocellulose in solid substrate fermentation. Biochem. Eng. 13(2-3): 169-179. doi: 10.1016/S1369-703X(02)00129-8.
- Thomsen MC, Belinda Á (2007). Wet oxidation pretreatment of lignocellulosic residues of sugarcane, rice, cassava and peanuts for ethanol production. J. Chem. Technol. Biotechnol. 82(2): 174-181. doi: 10.1002/jctb.1648.
- Torre P, Aliakbarian B, Rivas B, Domínguez JM Converti A (2008). Release of ferulic acid from corn cobs by alkaline hydrolysis. Biochem. Enging J. 40(3): 500-506. doi: 10.1016/j.bej.2008.02.005.
- Torres ML, Lloréns MCE (2008). Effect of alkaline pretreatment on anaerobic digestion of solid wastes. Waste Manag. 28(11): 2229-2234. doi:10.1016/j.wasman.2007.10.006.
- Tsai WT, Chang CY, Wang SY, Chang CF, Chien SF, Sun HF (2001). Utilization of agricultural waste corn cob for the preparation of carbon adsorbent J. Environ. Sci. Health Part B 36(5): 677-686. doi: 10.1081/PFC-100106194.
- Tserki V, Matzinos P, Zafeiropoulos NE, Panayiotou C (2006). Development of biodegradable composites with treated and compatibilized lignocellulosic fibers. J. Appl. Poly. Sci. 100(6): 4703-4710. doi: 10.1002/app.23240.
- Ubalua, AU. (2007). Cassava wastes: Treatment options and value addition alternatives. Afric. J. Biotechnol. 6 (18): 2065-2073.
- van Maris AJA, Abbott DA, Bellissimi E, van den Brink J, Kuyper M, Luttik MAH, Wisselink HW, Scheffers WA, van Dijken JP, Pronk JT (2006). Alcoholic fermentation of carbon sources in biomass hydrolysates by *Saccharomyces cerevisiae*: Current status. Antonie van Leeuwenhoek, 90(4): 391-418. doi: 10.1007/s10482-006-9085-7.
- van Wyk JPH (2001). Biotechnology and the utilization of biowaste as a resource for bioproduct development. Trends Biotechnol. 19(50): 172-177. doi: 10.1016/S0167-7799(01)01601-8.
- Vargas-Garcı MC, Suárez-Estrella F, López MJ, Moreno J (2007). Effect of inoculation in composting processes: Modifications in lignocellulosic fraction. Waste Manag. 27(9): 1099-1107. doi: 10.1016/j.wasman.2006.06.013.
- Velázquez-Cedeño MA, Farnet AM, Ferré E, Savoie JM (2004). Variations of lignocellulosic activities in dual cultures of *Pleurotus* ostreatus and *Trichoderma longibrachiatum* on unsterilized wheat straw. Mycologia, 96(4): 712-719.
- Vertès AA, Inui M, Yukawa H (2008). Technological options for biological fuel ethanol. J. Mol. Microbiol. Biotechnol. 15(1): 16-30. doi: 10.1159/000111989.
- von Blottnitz H, Curran MA (2007). A review of assessments conducted on bio-ethanol as a transportation fuel from a net energy, greenhouse gas, and environmental life cycle perspective. J. Cleaner Prod. 15(7): 607-619 doi: 10.1016/j.jclepro.2006.03.002.

- Wang H, Chen HZ (2007). A novel method of utilizing the biomass resource: Rapid liquefaction of wheat straw and preparation of biodegradable polyurethane foam (PUF). J. Chinese Inst. Chem. Eng. 38(2): 95-102. doi:10.1016/j.jcice.2006.10.004.
- Wen Z, Liao W, Chen S (2005). Production of cellulase by *Trichoderma* reesei from dairy manure. Bioresour. Technol. 96(4): 491-499 doi:10.1016/j.biortech.2004.05.021.
- Wen-Jing L, Hong-Tao W, Yong-Feng N, Zhi-Chao W, De-Yang H, Xiang-Yang Q, Jin-Chun C (2004). Effect of inoculating flower stalks and vegetable waste with ligno-cellulolytic microorganisms on the composting process. J. Environ. Sci. Health Part B: Pesticides, Food Contaminants, Agric. Wastes 39: 871-887.
- Wisselink HW, Toirkens MJ, Berriel MDRF, Winkler AA, van Dijken JP, Pronk JT, van Maris AJA (2007). Engineering of *Saccharomyces cerevisiae* for efficient anaerobic alcoholic fermentation of Larabinose. Appl. Environ. Microbiol. (15): 4881-4891. doi: 10.1128/AEM.00177-07.
- Wuyep PA, Khan AU, Nok AJ (2003). Production and regulation of lignin degrading enzymes from *Lentinus squarrosulus* (mont.). Afr. J. Biotechnol. 2(11): 444-447.
- Wyman CE (2003). Potential synergies and challenges in refining cellulosic biomass to fuels, chemicals and power. Biotechnol. Progress 19(2): 254-262. DOI: 10.1021/bp025654I S8756-7938(02)05654-0.
- Xiao C, Bolton R, Pan WL (2007). Lignin from rice straw kraft pulping: Effects on soil aggregation and chemical properties. Bioresour. Technol. 98(7): 1482-1488. doi:10.1016/j.biortech.2005.11.014.
- Yan L, Shuya T (2006). Ethanol fermentation from biomass resources: Current state and prospects. Appl. Microbiol. Biotechnol. 69(6): 627-642. doi: 10.1007/s00253-005-0229-x.
- Yáñez R, Alonso JL, Parajó, JC (2004). Production of hemicellulosic sugars and glucose from residual corrugated cardboard. Proc. Biochem. 39(11): 1543-1551. doi: 10.1016/S0032-9592(03)00283-8.
- Yang HS, Kim DJ, Lee YK, Kim HJ, Jeon JY, Kang WC (2004a). Possibility of using waste tire composites reinforced with rice straw as construction materials. Bioresour. Technol. 95(1): 61-65. doi: 10.1016/j.biortech.2004.02.002.
- Yang HS, Kim HJ, Son J, Park HJ, Lee BJ, Hwang TS (2004b). Ricehusk flour filled polypropylene composites; mechanical and morphological study. Composite Structures 63(3-4): 305-312. doi:10.1016/S0263.
- Yang HS, Kim HJ, Park HJ, Lee BJ, Hwang TS (2007). Effect of compatibilizing agents on rice-husk flour reinforced polypropylene composites. Composite Structures, 77(1): 45-55. doi:10.1016/j.compstruct.2005.06.005.
- Yang C, Shen Z, Yu G, Wang J (2008). Effect and after effect of γ radiation pretreatment on enzymatic hydrolysis of wheat straw. Bioresour. Technol. 99(14): 6240-6245. doi: 10.1016/j.biortech.2007.12.008.
- Yuan X, Jayaraman K, Bhattacharyya D (2004). Effects of plasma treatment in enhancing the performance of woodfibre-polypropylene composites. Composites Part A: Appl. Sci. Manuf. 35(12): 1363-1374. doi: 10.1016/j.compositesa.2004.06.023.
- Yue ZB, Yu HQ, Hu ZH, Harada H, Li YY (2008). Surfactant-enhanced anaerobic acidogenesis of *Canna indica* L. by rumen cultures. Bioresour. Technol. 99(9): 3418-3423. doi: 10.1016/j.biortech.2007.08.010.
- Yildiz S, Yildiz UC, Gezer ED, Temiz A (2002). Some lignocellulosic wastes used as raw material in cultivation of the *Pleurotus ostreatus* culture mushroom. Proc. Biochem. 38(3): 301-306(6). DOI: 10.1016/S0032-9592 (02)00040-7
- Zabaniotou AA, Kantarelis EK, Theodoropoulos DC (2008). Sunflower shells utilization for energetic purposes in an integrated approach of energy crops: Laboratory study pyrolysis and kinetics. Bioresour. Technol. 99(8): 3174-3181. doi: 10.1016/j.biortech.2007.05.060.
- Zhang YHP, Evans BR, Mielenz JR, Hopkins, RC, Adams MWW (2007). High yield hydrogen production from starch and water by a syntheric enzymatic pathway. Plos One 5: e456. <u>www.plosone.org</u>.
- Zhang M, Cui SW, Cheung, PCK, Wang Q (2007). Antitumor polysaccharides from mushrooms: A review on their isolation process, structural characteristics and antitumor activity. Trends Food Sci. Technol. 18(1): 4-19. doi: 10.1016/j.tifs.2006.07.013.

- Zhao Y, Wang Y, Zhu JY, Ragauskas A, Deng Y (2007). Enhanced enzymatic hydrolysis of spruce by alkaline pretreatment at low temperature. Biotechnol. Bioeng. 99(6). 1320-1328.
- Zhu S, Yu Z, Wu Y, Zhang X, Li H, Gao M (2005a). Enhancing enzymatic hydrolysis of rice straw by microwave pretreatment. Chem. Eng. Communications, 192(12): 1559-1566(8). doi: 10.1080/009864491007750.
- Zhu S, Wu Y, Ziniu Z, Liao J, Zhang Y (2005b). Pretreatment by microwave/alkali of rice straw and its enzymic hydrolysis. Process Biochem. 40(9): 3082-3086 doi:10.1016/j.procbio.2005.03.016.
- Zhu S, Wu Y, Yu Z, Zhang X, Li H, Gao M (2006). The effect of microwave irradiation on enzymatic hydrolysis of rice straw. Bioresour. Technol. 97(15): 1964-1968. doi: 10.1016/j.biortech.2005.08.008.
- Zubair A, Bhatti HN, Hanif MA, Shafqat F (2008). Kinetic and equilibrium modeling for Cr (iii) and Cr(vi) removal from aqueous solutions by *Citrus reticulata* waste Biomass. J. Water Air Soil Poll. 191(1-4): 305-318 doi: 10.1007/s11270-008-9626-y.