

Full Length Research Paper

# Exposure to red light, temperature and exogenous gibberellins influenced germination of some winter weeds

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Red light, temperature and gibberellins are well known for their capacity to induce higher germination in dormant seeds of several plant species. In the current study, we investigated the effect of various temperature (10, 13, 16, 19, 22 and 25 °C) and gibberellic acid (GA<sub>3</sub>) and GA<sub>4+7</sub> concentrations (0.1, 1, 10, and 100 μM) on germination of *Poa annua*, *Alopecurus aequalis* and *Stellaria aquatica* seeds incubated under continuous dark or treated with red light pulse of 80 μmol m<sup>-2</sup> s<sup>-1</sup> red light (660 nm) for 10 min after 12 h of dark period. We found that incubation temperature and gibberellins (GAs) significantly enhanced seed germination in red light treated seeds, as maximum germination was observed for *A. aequalis* (67.3% at 13 °C), *P. annua* (61.3% at 13 °C), and *S. aquatica* (42% at 19 °C) in such treatments. Exogenous GA<sub>4+7</sub> significantly promoted germination of *P. annua* and *A. aequalis* seeds, while GA<sub>3</sub> induced higher germination in *S. aquatica* seeds. It was observed that GAs were more effective in the presence of red light as least germination stimulation was recorded for GAs under dark condition and no germination was found in weed seeds at control and 0.1 mM GA<sub>3</sub> and GA<sub>4+7</sub> treatments. It can be concluded that lower temperature (13 °C) and GA<sub>4+7</sub> in conjunctions with red light pulse, induced maximum germination in dormant weed seeds.

**Key words:** Germination, temperature, nitrogen-containing compounds, red light, gibberellins.

## INTRODUCTION

Seed dormancy could be considered simply as a block to the completion of germination of an intact viable seed under favorable conditions, although to some researchers, it is one of the least understood phenomena in the field of seed biology (Hilhorst, 1995; Bewley, 1997a). In

the last years, significant progress has been made in understanding seed dormancy; however, there have also been some potential sources of confusion that have been reported in the literature (Finch-Savage and Leubner-Metzger, 2006). In ecological studies, there has been confusion reported between seed dormancy and persistence in soil (Thompson et al., 2003; Fenner and Thompson, 2005; Walck et al., 2005). This has resulted in part from different views on dormancy, such as whether light terminates dormancy or induces germination.

Environmental factors are believed to regulate seed dormancy and germination. Temperature substantially affects the dormancy level of weed seeds (Forcella,

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**Abbreviation:** GA<sub>3</sub>, Gibberellic acid.

1998), as diurnal and seasonal temperature fluctuation and mean temperature affect the breaking of dormancy and thus the germination rate. Some weeds require periods in which the temperature either increases or decreases slowly, whereas other species require daily temperature fluctuations or chilling to break dormancy (Baskin and Baskin, 2004). Likewise temperature, light is another environmental factor regulating seed dormancy and germination. Such regulation is mediated by photoreceptors called phytochromes, which initiate or stop physiological processes in the seed depending on the characteristics of the light to which it is exposed. The phytochromes change their form depending on the wavelength of the light to which they are exposed. The phytochrome Pr absorbs red light (R) and is biologically inactive, while Pfr absorbs far-red light (FR) and is considered the active form that promotes germination (Benech-Arnold et al., 2000). The wavelength, flux density, and exposure duration of light determine the extent of germination induction and breaking dormancy (Ballare and Casal, 2000).

Plant-hormones are naturally occurring organic substances, which influence physiological processes at very low concentrations either in distant tissues to which they are transported or in the tissue where synthesis occurred (Davies, 1995). Due to their structural simplicity, plant hormones are not specific enough to match the variety of controlled reactions (Canny, 1985). Contrary to this, it has been suggested that hormones only provide "turn on" or "turn off" signals and that the actual information's are provided by the cell. Gibberellins are considered important in the regulation of seed dormancy. Gibberellins are tetracyclic diterpenes, which are reported as necessary for seed germination in many plant species (Sawada et al., 2008; Calvo et al., 2004). Gibberellic acid ( $GA_3$ ) is also reported to reprogram plants to higher growth and significantly ameliorate the adverse effects of salt stress and rescue the productivity and quality of soybean (Hamayun et al., 2010).

The first objective of the current study was to investigate the effect of red light and temperature on seed germination rate and dormancy release of *Poa annua*, *Alopecurus aequalis* and *Stellaria aquatica*. These are common weeds of agriculture fields during winter season. The second objective was to observe the stimulating effect of bioactive GAs on seed germination, and compare the effectiveness of  $GA_3$  and  $GA_{4+7}$  in the regulation of seed germination of these three weeds.

## MATERIALS AND METHODS

### General procedure

Seeds of *P. annua*, *A. aequalis* variety *amurensis*, and *S. aquatica* were collected from agriculture fields in Suwen, South Korea. Seeds were then properly dried for two weeks at green house and kept in air-tight polyethylene containers at cold room at 4°C. The seeds were surface sterilized by immersion in 0.5% (w/v) NaClO for 5 min and washed with sterile distilled water. Fifty seeds of each

weed species were placed in 9 cm Petri plates with two layers of Whatman No. 1 filter paper and moistened with 4 ml of distilled water. The Petri plates were wrapped in aluminum foil, covered with a dark cloth and transferred to incubator set at 25°C. Seeds germination was observed after one week of red light treatment and emergence of radical (2 mm) was the criterion for germination (Andersson et al., 2002). Each treatment comprised 3 replicates and was repeated twice.

### Application of red light, temperature and gibberellins

For red light treatment, seeds were irradiated with red light for 10 min after 12 h of imbibition. Light emitting diode (LED) lamp (Mil-R18: Sanyo Biomedical) with irradiation of 660 nm, with irradiation of 660 nm ( $80 \mu \text{mol.m}^{-2} \text{s}^{-1}$ ) was used as light source for seed germination throughout the experiment (Tang et al., 2008). Treated seeds were transferred back to incubator. The temperature treatment comprised temperatures ranging from 10 to 30°C at 3°C intervals. For investigating GAs effect, seeds were incubated in the dark at 25°C and treated with 0.1, 1, 10 and 100  $\mu\text{M}$  of  $GA_3$  and  $GA_{4+7}$ .  $GA_3$  and  $GA_{4+7}$  are commercially available bioactive gibberellins. GAs ( $GA_3$  and  $GA_{4+7}$ ) were dissolved in 1 N NaOH and then solutions were made with distilled  $H_2O$ .

### Statistical analysis

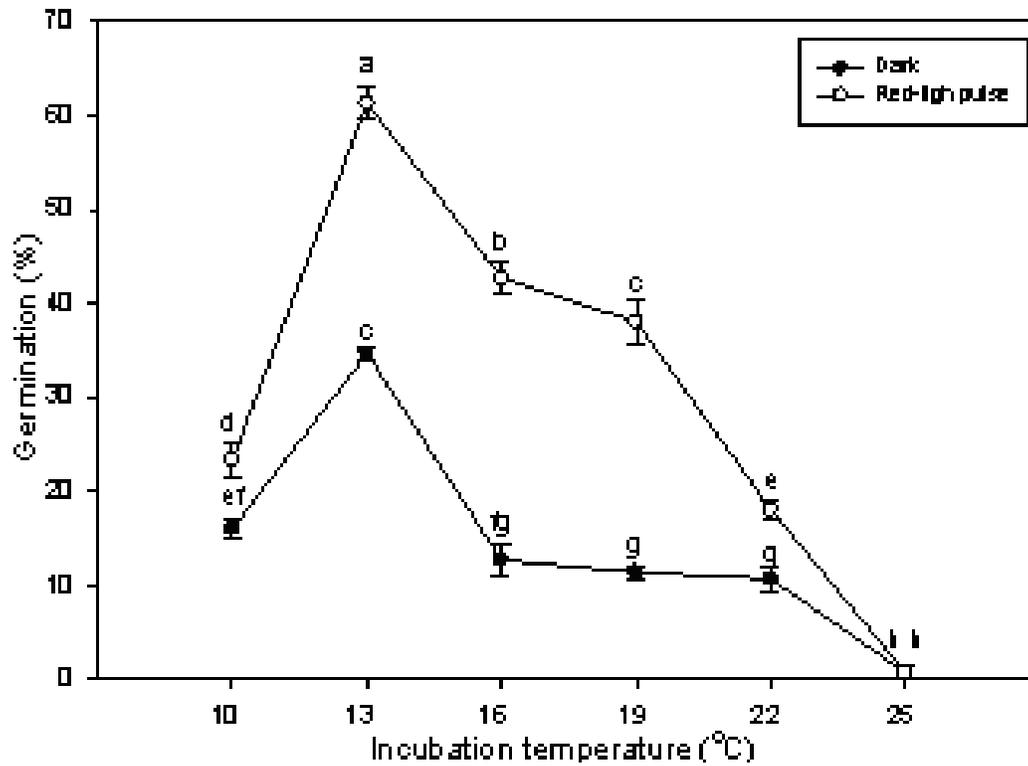
The means and standard errors for all treatments were compared using analysis of variance (ANOVA SAS release 9.1; SAS, Cary, NC, USA) in order to define whether the differences were significant. Mean percentage germination was graphically compared through Sigma Plot software (Sigmaplot 9.0, Systat Software Inc., 2004). Values included in the same group indicate that the differences among the values are not significant. But, values divided into the different groups means that the differences are significant otherwise.

## RESULTS AND DISCUSSION

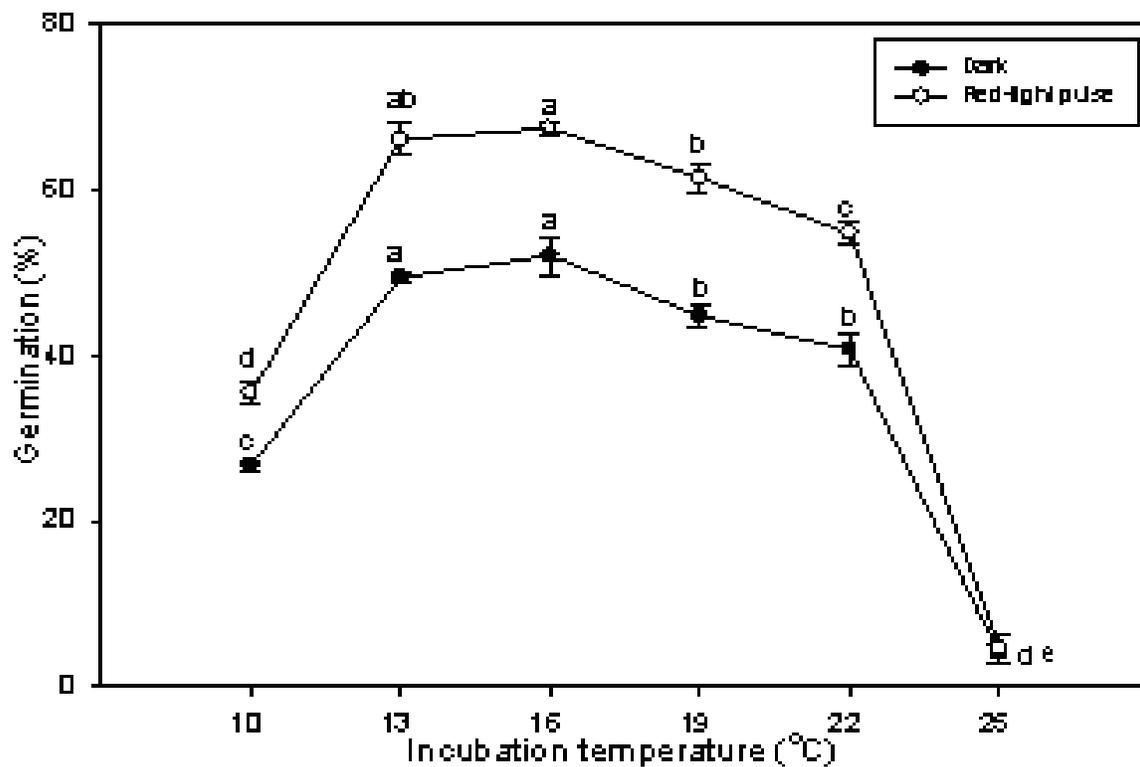
### Influence of varying temperatures on germination of winter weeds

Viable seeds are dormant when their germination is prevented by a block within the seed, so that they cannot germinate in an otherwise favorable environment (Bewley and Black, 1982). Temperature has a double effect on seed germination. Both the final germination percentage and the rate at which this percentage is reached are affected by temperature. Generally, the maximum final germination percentage is reached in a broad temperature range, whereas the germination rate has a sharp optimum temperature (Garcia-Huidobro et al., 1982).

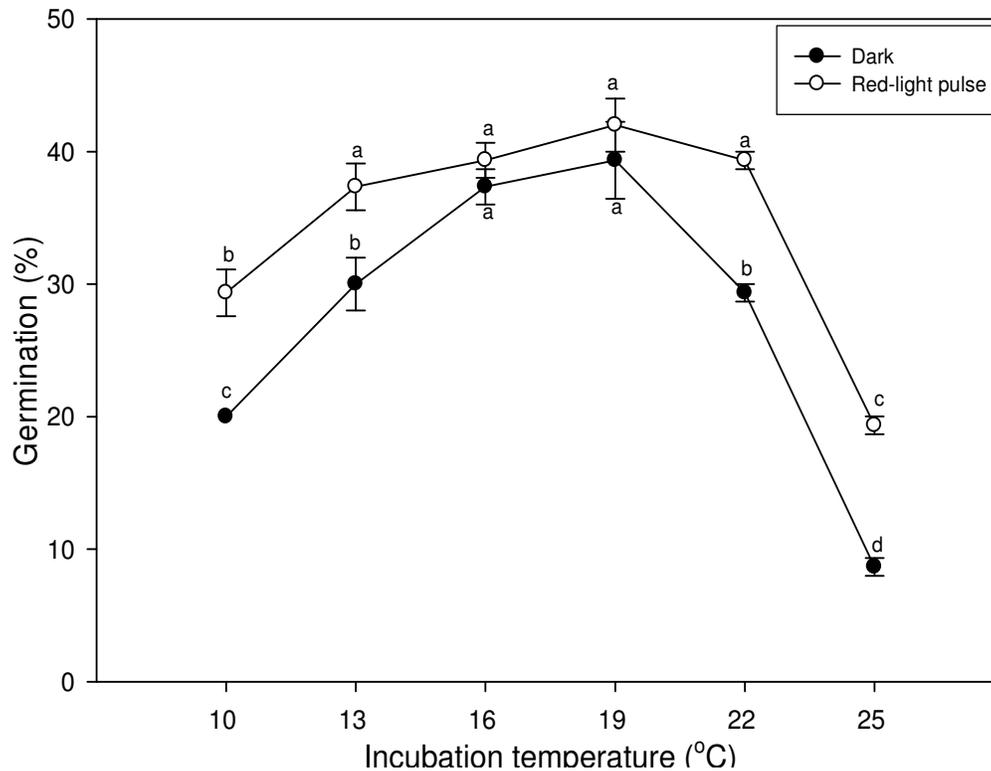
We observed that the seeds of *P. annua* and *A. aequalis* variety *amurensis* germinated at 10°C and the germination rate increased with temperature and maximum seed were germinated at 13°C in both red light and control treatments. However, with a further increase in temperature, the germination gradually declined and least germination was observed in seeds kept at 25°C. Current results showed that *P. annua* seeds failed to germinate below 10°C and above 25°C (Figures 1 and 2). Addition



**Figure 1.** Germination of *P. annua* seeds at different temperatures. Seeds were treated with red light for 10 min after 12 h of dark, while the control seeds were kept under dark throughout.



**Figure 2.** Germination of *A. aequalis* var. *amurensis* seeds at different temperatures. Seeds were treated with red light for 10 min after 12 h of dark, while the control seeds were kept under dark throughout.



**Figure 3.** Germination of *S. aquatica* seeds at different incubation temperatures. Seeds were treated with red light for 10 min after 12 h of dark, while the control seeds were kept under dark throughout.

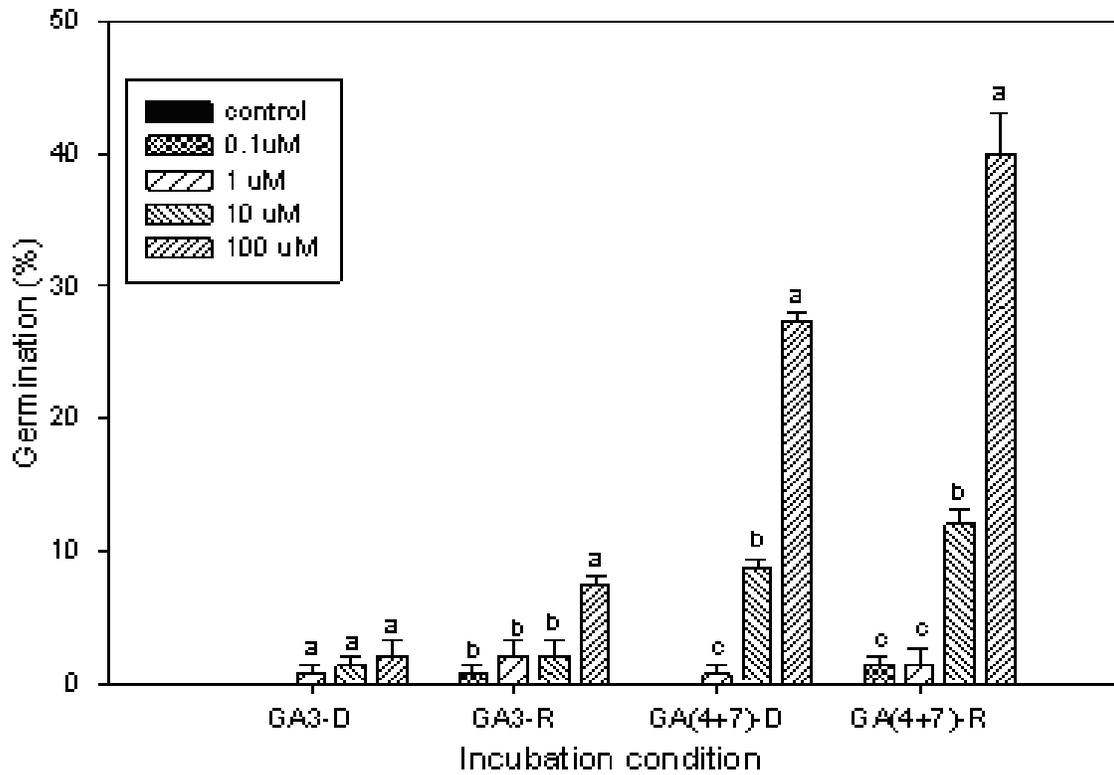
of red light significantly stimulated seed germination at temperatures ranging from 10 to 22°C as compared to dark treatments, but it failed to induce germination below 10°C and above 25°C. The overall germination capacity of *A. aequalis* variety *amurensis* was found higher than *P. annua* under similar environmental condition. It was previously reported that at temperatures below the optimum, germination rate increases linearly with temperature (Hegarty, 1973; Garcia-Huidobro et al., 1982); however, the current study showed that in weeds, at temperatures above the optimum, germination rate decreases linearly an increase in temperature.

Seeds of *S. aquatica* germinated at low temperature (10°C), and further increase in temperature regime stimulated germination till maximum germination was attained at 16°C. However, a gradual further increase in temperature caused marked reduction in germination of *S. aquatica* seeds (Figure 3). Similar to the other two weed studied, red light provided an additive stimulation for seed germination, although at extreme lower and higher temperatures, the red light failed to induce any germination. Current results proved that treatment of weed seeds with red light for a brief period significantly enhance germination percentages of treated seeds. It is because the red light promotes phytochrome biosynthesis which induces seed germination. Our current findings confirm previous report on favorable role of red light in

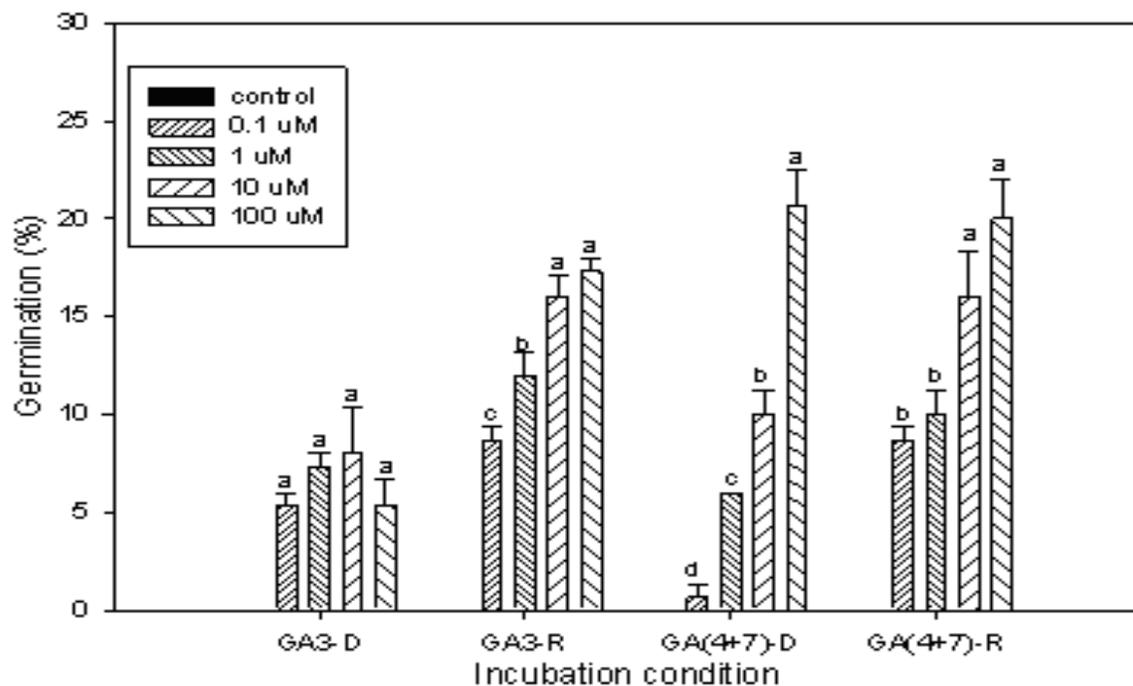
seed germination (Tang et al., 2008).

### Influence of GA<sub>3</sub> and GA<sub>4+7</sub> on germination of winter weeds

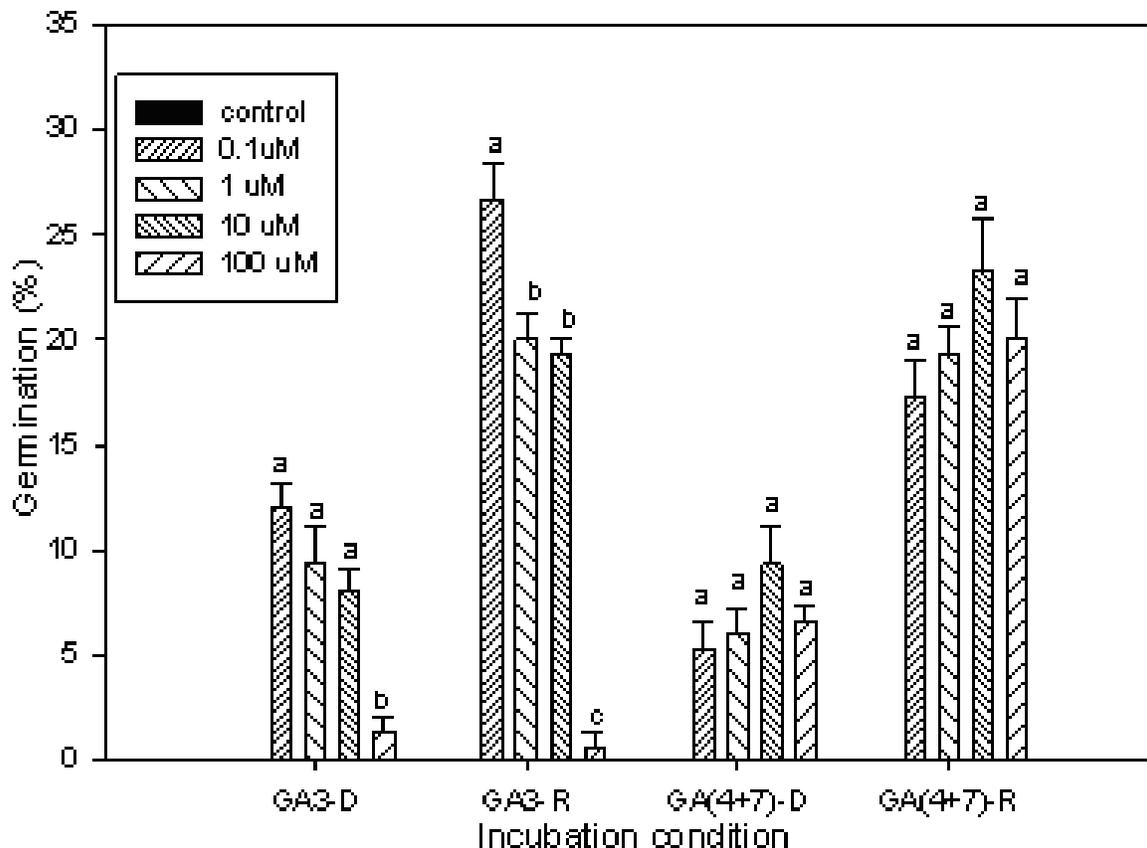
Physiologically active GAs is known for inducing seed germination in a wide range of plant species, as they induce hydrolytic enzymes. The hydrolytic enzymes weaken the barrier tissues such as the endosperm or seed coat, inducing mobilization of seed storage reserves, and stimulating expansion of embryo (Bewley and Black, 1994). In the current study, exogenous GA<sub>3</sub> and GA<sub>4+7</sub> applications induced higher germination in *P. annua*, *A. aequalis* var. *amurensis*, and *S. aquatica*, while higher temperature favored seed dormancy (Figure 4). Moreover, red light exposure showed an additive effect to seed germination of weed species in-conjunction with these GAs. However, the effect of GA<sub>3</sub> and GA<sub>4+7</sub> varied for different weed species as GA<sub>3</sub> seems to show lesser effect on *P. annua* seeds as compared to GA<sub>4+7</sub> treatments (Figure 4). The stimulatory effect of GAs was enhanced when there was an increase in GAs concentration from 0.1 to 100 µM (Figures 4, 5 and 6). In case of *A. aequalis* var. *amurensis*, either of GA<sub>3</sub> and GA<sub>4+7</sub> significantly prompted seed germination (Figure 5), which was similar to that of *P. annua*. On the contrary, lower



**Figure 4.** Germination (%) of *P. annua* seeds treated with elevated GA<sub>3</sub> and GA<sub>4+7</sub>. Seeds were treated with red light for 10 min after 12 h of dark, while the control seeds were kept under dark throughout. D stands for dark, while R stands for red light.



**Figure 5.** Germination (%) of *A. aequalis* var. *amurensis* treated with elevated GA<sub>3</sub> and GA<sub>4+7</sub>. Seeds were treated with red light for 10 min after 12 h of dark, while the control seeds were kept under dark throughout. D stands for dark, while R stands for red light.



**Figure 6.** Germination (%) of *S. aquatica* treated with elevated GA<sub>3</sub> and GA<sub>4+7</sub>. Seeds were treated with red light for 10 min after 12 h of dark, while the control seeds were kept under dark throughout. D stands for dark, while R stands for red light.

concentrations of GA<sub>3</sub> and GA<sub>4+7</sub> induced seed germination of *S. aquatica*, as maximum germination was achieved at 0.1 μM GA<sub>3</sub> and 10 μM GA<sub>4+7</sub> (Figure 6).

## Conclusion

Weed eradication is a difficult task as weed seeds possess differential dormancy levels and can survive under the soil for many years. An efficient eradication of these undesired plants can be achieved by adopting a two pronged strategy, that is, either to induce permanent dormancy in these seeds or to break the existing varying degrees of seed dormancy, so that all seeds germinate at the same season and can be effectively eradicated by applying chemicals or through traditional weeding practices. Our study is linked to the latter part and narrates about the favorable role of red light and N-compounds in germination induction or breaking of seed dormancy. The current study explains how interactions between environmental factors have a great impact on the germination of weeds. The variable responses of weeds to environmental factors and their interaction could account for differences in seed germination and seedling emergence

timing, which represent differences in adaptation and survival strategies. This study would help in understanding the favorable role of red light, temperature and exogenous GA<sub>3</sub> and GA<sub>4+7</sub> in germination induction of weed seeds, as weed eradication from crop fields is highly desirable, which will increase crop production and reduce the use of herbicides. However, further research is needed on the topic in order to understand and elaborate the germination induction mechanism of weed seeds.

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## REFERENCES

- Andersson L, Miberg P, Schutz W, Steinmetz O (2002). Germination characteristics and emergence time of annual *Bromus* species of differing weediness in Sweden. *Weed Res.* 42: 135-147.
- Ballaré CL, Casal JJ (2000). Light signals perceived by crop and weed plants. *Field Crops Res.* 67: 149-160.

- Baskin JM, Baskin CC (2004). A classification system for seed dormancy. *Seed Sci. Res.* 14: 1-16.
- Benech-Arnold RL, Sanchez RA, Forcella F, Kruk BC, Ghera CM (2000). Environmental control of dormancy in weed seed banks in soil. *Field Crops Res.* 67: 105-122.
- Bewley JD (1997). Seed germination and dormancy. *Plant Cell.* 9: 1055-1066.
- Bewley JD, Black M (1994). *Seed: physiology of development and germination.* New York, Plenum press.
- Bewley JD, Black M (1982). *Physiology and biochemistry of seeds: 2. Viability, dormancy, and environmental control.* Springer-Verlag, Berlin, Germany.
- Calvo AP, Nicolas C, Lorenzo O, Niclas G, Rodriguez D (2004). Evidence for positive regulation by gibberellins and ethylene of ACC oxidase expression and activity during transition from dormancy to germination in *Fagus sylvatica* L. seeds. *J. Plant Growth Reg.* 23: 44-53.
- Canny MJ (1985). Ashby's law and the pursuit of plant hormones: A critique of accepted dogmas, using the concept of variety. *Aust. J. Plant Physiol.* 12: 1-7.
- Davies PJ (1995). The plant hormones: their nature, occurrence, and functions. In: Davies, P.T. (ed.): *Plant Hormones.* Kluwer Academic Publishers, Netherlands. pp. 1-12.
- Fenner M, Thompson K (2005). *The Ecology of Seeds.* Cambridge University Press. UK.
- Finch-Savage WE, Leubner-Metzger G (2006). Seed dormancy and control of germination. *New Phytologist.* 171: 501-523.
- Forcella F (1998). Real-time assessment of seed dormancy and seedling growth for weed management. *Seed Sci. Res.* 8: 201-210.
- Garcia-Huidobro J, Monteith JL, Squire GR (1982). Time, temperature and germination of pearl millet (*Pennisetum typhoides* S and H). I. Constant temperature. *J. Exp. Bot.* 33: 288-296.
- Hamayun M, Khan SA, Khan AL, Shin JH, Ahmad B, Shin DH, Lee IJ (2010). Exogenous gibberellic acid reprograms soybean to higher growth and salt stress tolerance. *J. Agric. Food Chem.* 58(12): 7226-32.
- Hegarty TW (1973). Temperature sensitivity of germination in carrots: its frequency of occurrence and response to seed advancement. *J. Hort. Sci.* 48: 43-50.
- Hilhorst HWM (1995). A critical update on seed dormancy. I. Primary dormancy. *Seed Sci. Res.* 5: 61-73.
- Sawada Y, Aoki M, Nakaminami K, Mitsuhashi W, Tatematsu K, Kushiro T, Koshiro T, Kamiya Y, Inoue Y, Nambara E, Toyomasu T (2008). Phytochrome and gibberellin mediated regulation of abscisic acid metabolism during germination of photoblastic lettuce seeds. *Plant Physiol.* 146: 1386-1396.
- Tang DS, Hamayun M, Ko YM, Zhang YP, Kang SM, Lee IJ (2008). Role of red light, temperature, stratification and nitrogen in breaking seed dormancy of *Chenopodium album* L. *J. Crop Sci. Biotechnol.* 11: 199-204.
- Thompson K, Ceriani RM, Bakker JP, Bekker RM (2003). Are seed dormancy and persistence in soil related? *Seed Sci. Res.* 13: 97-100.
- Walck JL, Baskin JM, Baskin CC, Hidayati SN (2005). Defining transient and persistent seed banks in species with pronounced seasonal dormancy and germination patterns. *Seed Sci. Res.* 15: 189-196.