Antimicrobial activities of medicinal plants used in folklore remedies in south-western

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Accepted 18 May, 2006

In south-western part of Nigeria Psidium guajava and Mangifera indica are commonly used for herbal preparations in the treatment of toothache, gastrointestinal disorders, dysentery, diarrhoea, sore gums and sore throats. This has, therefore, led to the investigation of the antimicrobial activities of methanolic extracts of P. guajava and M. indica. Fifteen different bacterial isolates comprising of both Gram negative and Gram positive organisms were used. The results show that P. guajava and M. indica extracts exhibited antimicrobial activities at a concentration of 20 mg/ml. The zones of inhibition exhibited by P. guajava extract ranged between 12 mm and 30 mm while that of M. indica varied between 11 and 28 mm. The minimum inhibitory concentration (MIC) exhibited by P. guajava extract against the tested organisms ranged between 0.313 and 0.625 mg/ml. On the other hand MIC exhibited by M. indica extract varied between 1.25 and 10.0 mg/ml. Overall, P. guajava extract show more antimicrobial activity than M. indica extract against tested organisms.

Key words: Psidium guajava, Mangifera indica, antimicrobial activity.

INTRODUCTION

Many works have been done which aim at knowing the different antimicrobial and phytochemical constituents of medicinal plants and using them for the treatment of microbial infections (both topical and systemic applications) as possible alternatives to chemically synthetic drugs to which many infectious microorganisms have become resistant. During the last ten years the pace of development of new antimicrobial drugs has slowed down while the prevalence of resistance (especially multiple) has increased astronomically (Hugo and Russell, 1984). The increase in number of antibiotic resistant bacteria is no longer matched by expansion in the arsenal of agents available to treat infections. Literature reports and ethnobotanical records suggest that plants are the sleeping giants of pharmaceutical industry (Hostettmann and Hamburger, 1991). They may provide natural source of antimicrobial drugs that will/or provide novel or lead compounds that may be employed in controlling some infections globally.

Psidium guajava is a shrub known as “guofa” in Yoruba and “goba” in Hausa tribes in Nigeria. P. guajava typically has very thin skins; the leaves are evergreen, opposite, short petioled, oval or oblong. The plant is used in folk medicine to treat fevers, diarrhoea, and as tonic in psychiatry (Oliver-Bever, 1986; Iwu, 1993). The hydroalcoholic extract was shown to decrease motor activity in mice (Iwu, 1993). Leaf extract from P. guajava was also shown to possess anti-inflammatory effect in rats (Olajide et al, 1999) as well as exhibiting antidiabetic effects in mice (Oh et al., 2005). The methanolic extract of P. guajava was also shown to possess antibacterial effect on Bacillus subtilis, Staphylococcus aureus, Escherichia coli and Pseudomonas aeruginosa (Abdelrahim et al., 2002). Clinical studies on phytodrugs from leaves of P. guajava on some volunteers with gastrointestinal ailments were found to be effective (Lazoya et al., 2002). Bark extract of P. guajava is also used for diarrhoea, stomach ache and diabetes (Tanaka et al., 1992).
Phytochemical constituents isolated from the leaves of *P. guajava* includes triterpenoids, approximately 10% tannin as well as quercetin (Begun et al., 1974; Osman et al., 1974). Other phytochemical compounds obtained from *P. guajava* are guavins A, C and D (Okuda et al., 1987). Ester of hexahydroxydiphenic acid with L-arabinose, leucocyanidin and oxalates were obtained from unripe fruits of *P. guajava* while ripe fruits contained only ellagic acid (Misra and Seshadri, 1968). A bio-antimutagenic compound (+)-gallocatechin was identified in *M. indica* leaves (Matsu et al., 1999). In Guatemala, compound (+)-gallocatechin was identified in *P. guajava* acid (Misra and Seshadri, 1968). A bio-antimutagenic properties (Garcia et al., 2003). It was also observed that *M. indica* aqueous leaf extract produce a reduction of blood glucose level in glucose-induced diabetic mice (Aderibigbe et al., 2001). Among the compounds isolated from *M. indica* extract are two new terpenoidal saponins (Khan et al., 1993). Other phytochemical compounds obtained from *M. indica* are polygalacturonase I, II, III from mango pulp (Prasanna et al., 2005) and a new triterpenoid, 29-hyroxymangiferonic acid (Anjaneyulu et al., 1994). Tetracyclic triterpenoid (Anjayulu et al., 1993) and pentacyclic triterpenoid (Anjaneyulu et al., 1989) were isolated from stem bark of *M. indica*. Another phytochemical compound isolated from unripe mango fruit was fructose-1,6-diphosphatase (Rao and Modi, 1976).

**MATERIALS AND METHODS**

**Plant materials**

Fresh bark of *P. guajava* and *M. indica* were collected from Abeokuta, Nigeria in the month of July, 2004 and were identified by the Botany Department, Obafemi Awolowo University, Ile-Ife, Nigeria. Voucher samples were prepared and deposited in the Herbarium of the Botany Department, Obafemi Awolowo University, Ile-Ife, for reference. The bark were later air-dried, powdered and stored in an air-tight container for further use.

**Preparation of extracts**

Exactly 150 g each of the powdered bark of the two plants were separately extracted in cold using 60% methanol for 4 days. The mixture was then filtered and the filtrate was dried in vacuo using a rotatory evaporator. The yield collected from *P. guajava* was 17% (w/w) while that of *M. indica* was 13.93% (w/w).

**Preparation of microorganisms for experiment**

The following microorganisms were used *Bacillus polymyxa* (LIO, locally isolated organism; isolated from pus and wound infection), *Bacillus anthracis* (LIO), *Bacillus cereus* (NCIB 6349), *Bacillus stearothermophilus* (NCIB 8222) *Bacillus subtilis* (NCIB 3610), *Clostridium sporogenes* (NCIB 532), *Corynecbacterium pyogenes* (LIO), *Escherichia coli* (NCIB 86), *Klebsiella pneumoniae* (NCIB 418), *Pseudomonas aeruginosa* (NCIB 950), *Pseudomonas fluorescens* (NCIB 3756), *Serratia marcescens* (NCIB 1377), *Shigella dysenteriae* (LIO), *Staphylococcus aureus* (NCIB 8588) and *Streptococcus faecalis* (LIO). For use in experiments, the organisms were sub-cultured in nutrient broth and nutrient agar (Oxoid Ltd.) while diagnostic sensitivity test agar (DST) (Oxoid Ltd.) was used in antibiotic sensitivity testing.

**Phytochemical analysis of the extract**

A small portion of the dry extract was used for phytochemical screening test (Treuse et al., 1983; Harbourne, 1973). Dragendorffs reagents were used to test for alkaloids, ferric chloride for tannins, while Benedict’s solution was used to test for saponins.

**Sensitivity testing**

The sensitivity testing of the extracts were determined using agar-well diffusion method (Russell and Furr, 1977; Irobi et al., 1996). The MIC of the extracts was also determined using a two-fold dilutions method (Russell and Furr, 1977). The bacterial isolates were first grown in nutrient broth for 18 h before use. The inoculum suspensions were standardized and then tested against the effect of the two plant extracts at a concentration of 20 mg/ml each in DST medium. The plates were later incubated at 37°C ± 0.5°C for 24 h after which they were observed for zones of inhibition (Table 1). The effects were compared with that of the standard antibiotic streptomycin at a concentration of 1 mg/ml (Khan and Omotoso, 2003).

**RESULTS AND DISCUSSION**

In this study the results of the investigations show that the two extracts from the bark of *P. guajava* and *M. indica* possess antimicrobial activities against some of the tested organisms at a concentration of 20 mg/ml (Table 1). The two extracts compared favourably with the standard antibiotic streptomycin. *P. guajava* extract showed more activity than *M. indica* extract. Both plant extracts exhibited a broad spectrum of activity.

The MIC of *P. guajava* extract against the tested organisms varied between 0.313 mg/ml and 0.625 mg/ml while that of *M. indica* ranged between 1.25 mg/ml and 10.0 mg/ml. The standard streptomycin had MIC values varying between 0.0313 mg/ml and 0.500 mg/ml. The results indicated that standard antibiotic streptomycin has stronger activity than the two plant extracts as shown in Table 2.

The phytochemical analysis of the *P. guajava* extract revealed the presence of tannins while that of *M. indica* showed the presence of alkaloids, saponins and tannins. These compounds are known to be biologically active. Tannins have been found to form irreversible complexes with proline-rich proteins (Hagerman and Butler, 1981) resulting in the inhibition of the cell protein synthesis. This activity was exhibited against test organisms with the two plant extracts.
Table 1. Antimicrobial activities of *P. guajava* and *M. indica* bark extracts.

<table>
<thead>
<tr>
<th>Microorganism</th>
<th><em>P. guajava</em> (20 mg/ml)</th>
<th><em>M. indica</em> (20 mg/ml)</th>
<th>Streptomycin (1 mg/ml)</th>
<th>Ampicillin (10 μg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Bacillus anthracis</em></td>
<td>20</td>
<td>0</td>
<td>18</td>
<td>16</td>
</tr>
<tr>
<td><em>Bacillus cereus</em></td>
<td>15</td>
<td>14</td>
<td>28</td>
<td>25</td>
</tr>
<tr>
<td><em>Bacillus polymyxa</em></td>
<td>0</td>
<td>0</td>
<td>15</td>
<td>12</td>
</tr>
<tr>
<td><em>Bacillus subtilis</em></td>
<td>18</td>
<td>17</td>
<td>20</td>
<td>0</td>
</tr>
<tr>
<td><em>Clostridum sporogenes</em></td>
<td>18</td>
<td>0</td>
<td>25</td>
<td>20</td>
</tr>
<tr>
<td><em>Corynbacterium pyogenes</em></td>
<td>15</td>
<td>12</td>
<td>20</td>
<td>36</td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>16</td>
<td>12</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td><em>Klebsiella pneumoniae</em></td>
<td>12</td>
<td>14</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em></td>
<td>13</td>
<td>12</td>
<td>21</td>
<td>0</td>
</tr>
<tr>
<td><em>Pseudomonas fluorescens</em></td>
<td>15</td>
<td>12</td>
<td>30</td>
<td>22</td>
</tr>
<tr>
<td><em>Serratia marcescens</em></td>
<td>0</td>
<td>0</td>
<td>19</td>
<td>16</td>
</tr>
<tr>
<td><em>Shigella dysenteriae</em></td>
<td>23</td>
<td>28</td>
<td>22</td>
<td>23</td>
</tr>
<tr>
<td><em>Staphylococcus aureus</em></td>
<td>25</td>
<td>12</td>
<td>21</td>
<td>0</td>
</tr>
<tr>
<td><em>Streptococcus faecalis</em></td>
<td>30</td>
<td>11</td>
<td>23</td>
<td>16</td>
</tr>
</tbody>
</table>

mm* = Mean of three replicates
LIO = Locally isolated organism
NCIB = National Collection of Industrial Bacteria
O = Resistant.

Table 2. The MIC of the *P. guajava* and *M. indica* bark extracts and streptomycin against the bacterial isolates.

<table>
<thead>
<tr>
<th>Microorganism</th>
<th><em>P. guajava</em> (mg/ml)</th>
<th><em>M. indica</em> (mg/ml)</th>
<th>Streptomycin (mg/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Bacillus cereus</em></td>
<td>0.313</td>
<td>1.25</td>
<td>0.0313</td>
</tr>
<tr>
<td><em>Bacillus anthracis</em></td>
<td>1.25</td>
<td>-</td>
<td>0.0313</td>
</tr>
<tr>
<td><em>Bacillus subtilis</em></td>
<td>1.25</td>
<td>5.00</td>
<td>0.0625</td>
</tr>
<tr>
<td><em>Bacillus stearothermophilus</em></td>
<td>1.25</td>
<td>1.25</td>
<td>0.0625</td>
</tr>
<tr>
<td><em>Corynbacterium pyogenes</em></td>
<td>0.625</td>
<td>2.50</td>
<td>0.0313</td>
</tr>
<tr>
<td><em>Escherichia coli</em></td>
<td>-</td>
<td>10.00</td>
<td>-</td>
</tr>
<tr>
<td><em>Klebsiella pneumoniae</em></td>
<td>0.313</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td><em>Pseudomonas aeruginosa</em></td>
<td>0.313</td>
<td>5.00</td>
<td>0.25</td>
</tr>
<tr>
<td><em>Shigella dysenteriae</em></td>
<td>0.625</td>
<td>5.00</td>
<td>0.25</td>
</tr>
<tr>
<td><em>Staphylococcus aureus</em></td>
<td>0.625</td>
<td>2.50</td>
<td>0.50</td>
</tr>
<tr>
<td><em>Streptococcus faecalis</em></td>
<td>2.50</td>
<td>2.50</td>
<td>0.0625</td>
</tr>
</tbody>
</table>

Apart from antimicrobial activity exhibited by tannins, they also react with proteins to provide the typical tanning effect. Medicinally, this is important for the treatment of inflamed or ulcerated tissues (Mota et al., 1985). Tannins have important roles such as stable and potent antioxidants (Trease et al., 1983). Herbs that have tannins as their main component are astringent in nature and are used for treating intestinal disorders such as diarrhoea and dysentery (Dharmananda, 2003), thus exhibiting antimicrobial activity. One of the largest groups of chemical produced by plants are the alkaloids and their amazing effect on humans has led to the development of powerful pain killer medications (Raffauf, 1996).

*P. guajava* and *M. indica* are used among the Yorubas of the southwestern part of Nigeria in preparing decoction, hence different formulations could be prepared for clinical trials. It is hoped that this study would lead to the establishment of some compounds that could be used to formulate new and more potent antimicrobial drugs of natural origin. Studies are in progress to further evaluate the mechanisms of action of *P. guajava* and *M. indica* extracts on some organisms associated with human diseases.

ACKNOWLEDGEMENT

We wish to acknowledge Dr. H.C. Illoh of Botany Department, Obafemi Awolowo University, Ile-Ife, Nigeria for identifying the plants used in this study.
REFERENCES


