

Full Length Research Paper

Effects of Chloroquine and Coartem on Haematological Parameters in Rats

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ABSTRACT

Coartem has recently replaced chloroquine as the first-line treatment for uncomplicated *Plasmodium falciparum* malaria due to resistance in many regions worldwide. This study was aimed to compare the effect of chloroquine and coartem on haematological parameters in rats. Thirty (30) albino Wistar rats were randomly assigned into 2 batches of 15 rats. Each batch was further divided into 3 groups of 5 rats each. Group 1 was control, groups 2 and 3 received coartem (1.6mg/100g body weight) and chloroquine (0.875mg/100g body weight) orally and once daily. The feeding regimen lasted for 3 and 7 days for batches 1 and 2 respectively. Complete blood count was done using automatic counter. Results revealed that administration of chloroquine and coartem for 3 days did not significantly altered the levels of RBC, Hb, PCV, total & differential WBC, platelet count and platelet indices, but led to significant reductions in MCV and MCH in chloroquine recipient (60.80 ±0.98fL and 19.14 ±0.31pg) compared with control (68.64 ±2.12fL and 20.80 ±0.60pg; p<0.05) and coartem (68.16 ±1.73fL and 20.36 ±0.14pg; p<0.01) groups respectively. However, administration of these drugs for 7 days caused significant reduction in RBC, Hb and PCV in coartem recipients compared with control (p<0.05) and chloroquine (p<0.01) groups. RDW was also significantly reduced in chloroquine recipients. In conclusion, administration of coartem and chloroquine at their recommended doses and durations would not pose any deleterious effect on haematological parameters in rats.

Keywords: Blood, antimalaria, chloroquine, coartem, blood, rat.

INTRODUCTION

Malaria is a mosquito borne disease, which has posed a serious health challenge to human. It is an infectious disease caused by a mosquito parasite called *Plasmodium*. The vast majority of deaths by mosquitoes are caused by *P*. falciparum, the other species of malaria parasite are *P*. vivax, *P*. malariae, *P*. ovale, and *P*.

knowlesi and are less effective in causing threaten to life (WHO, 2006; Cox-Singh, 2008; Fairhust and Wellems, 2010).

The *plasmodium* affects blood cells, malaria symptoms are accompanied by fever, shaking chills, sweating, pains etc which is widely affecting people in the tropic region, tormenting about 400-600 million of people yearly, with over 3-5 million death (Joy *et al.*,

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Bioline International, African Journals online (AJOL), Index Copernicus, African Index Medicus (WHO), Excerpta medica (EMBASE), CAB Abstracts, SCOPUS, Global Health Abstracts, Asian Science Index, Index Veterinarius, , African Journals online 2003, Snow *et al.*, 2005; Mulenga *et al*, 2006; Hay *et al.*, 2010; World Malaria Report, 2010).

To prevent the menace of malaria among the populace, anti-malarial drugs have been produced to either prevent or cure the occurrence of malaria. The earliest drugs which have long been used in the treatment and prevention of malaria were the aminoquinolines of which chloroquine was the mainstay from 1934 (Greenwood, 1995; WHO, 2008; Kraft *et al.*, 2012; White, 2004), but the emergence and spread of resistance in a large population of the African continent had led to the introduction of artemisinin based combination therapy (ACTs) in the 21st century. Zambia was the first African country to adopt ACT treatments in its policy, (Spilanyambe *et al.*, 2008). Among the ACT drugs of choice is coartem, (Nosten and White, 2007; Katzung, 2007).

Coartem (Arthermether + Lumefantrine) is currently the only fixed dose artemisinin based combination therapy prequalified by the WHO for procurement by United Nations agencies for the treatment of acute uncomplicated plasmodium Falciparum, (Falade et al., 2005; WHO, 2010). Artemether is one of the semi synthetic derivatives of artemisinin, Artemisinin is a natural anti-malarial derived from the Chinese medicinal plant Artemisia annua. The artemisinin derivatives are the most effective anti-malarial drugs available today and they have been used with success in areas with multidrug resistant Plasmodium falciparum malaria (Adjuik , 2004; Sinclair et al., 2009; Kokwaro, 2007; Koram et al., 2012). Lumefantrine (also known as benflumetol and CGP 56695 during development) is purely synthetic, (Makenga et al., 2002; Novartis, 2005).

The use of these drugs to treat malaria may be associated with possible side effects on the blood cells, it was therefore the aim of this study to undertake a comparative effect of chloroquine and coartem on haematological parameters (RBC count, PCV, Hb, MCV, MCH, MCHC, RDW, WBC count, Platelets count, platelet indices-MPV, P-LCR, PDW and differential WBC count).

MATERIALS AND METHODS

Animals

Thirty (30) albino Wistar rats (initially weighing between 120-160g) were obtained from animal house of the Department of Physiology, University of Calabar and housed in cages in the laboratory of the Department of Physiology, College of Medical Sciences, University of Calabar.

The animals were allowed to acclimatize for one week in a well aerated room at room temperature under natural lighting condition of 12 hour light and 12 hour dark cycle. All the animals were handled under standard guidelines for care and use of laboratory animals as promulgated by Canadian Council of Animal Care (2009).

Drugs

Coartem: Coartem manufactured by Beijing Norvatis Pharma LTD, Beijing, China for Norvatis Pharmabarle, Switzerland under license from the PRC was obtained from the Bez Pharmacy, Calabar, Nigeria. One tablet (140mg) of coartem was crushed using a glass mortar and it was dissolved in a total of 10ml of distilled water to give a concentration of 14mg/ml stock. The drug was administered to the animals at a dose of 1.6mg/100g body weight [equivalent to human (70kg) daily dose], i.e. 0.11mL of stock /100g body weight After every administration the remaining drug was poured away and a new one prepared following the next administration.

Chloroquine: Chloroquine was obtained from Turtle Bay Pharmacy, Calabar-Nigeria. One tablet (150mg) was ground to powder and dissolved in a total of 20ml of distilled water to give a stock concentration of 7.5mg/ml. The drug was first administered at a dose of 0.875mg/100g body weight (i.e. 0.12mL of stock/100g body weight) for the first 2 days. On the 3rd day, 4.38mg/kg body weight (i.e. 0.06mL of stock/100g body weight) was administered.

After every administration, the remaining drug was poured out and a new one was prepared following the next administration

Experimental Design

Thirty (30) albino Wistar rats were assigned into 2 batches of 15 rats each. Each batch was divided into 3 groups of 5 rats each and fed thus:

- Group 1 (control) received normal rat chow + water.
- Group 2 (coartem treated): in addition to control diet received coartem treatment orally and once daily,
- Group 3 (chloroquine treated): in addition to the control diet received chloroquine treatment orally and once daily.

Treatment lasted for 3 days and 7 days for batches 1 and 2 respectively.

Collection of Blood Samples

The animals were made unconscious inhaling chloroform anesthesia (3.5% soaked in cotton wool) and blood collected via cardiac puncture (blood was drawn from the heart) a modification of the method by Ohwada

(1986). The samples were collected by the help of 5mls syringe attached to needle (21 SWG) into plain capped bottles containing ethylene diamine tetraacetate (EDTA). The samples were immediately used for the estimation of the different variables.

Measurement of blood parameters

Blood samples were analyzed using automated cell counter (Coulter Electronics, Luton, Bedfordshire, UK) calibration according with standard to the manufacturer's instruction (Coulter Electronic, 1979) using normal human blood and with complete profile for red blood cell (RBC) count, total white blood cell (WBC) count, differential WBC count, haemoglobin (Hb), packed cell volume (PCV), mean corpuscular volume (MCV), mean corpuscular haemoglobin (MCH), mean corpuscular haemoglobin concentration (MCHC), red blood cell distribution width (RDW), mean platelet volume (MPV), platelet distribution width (PDW) and platelet large cell ratio (P-LCR)

Statistical Analysis

Data were presented as mean \pm SD. Data were analysed using a one way analysis of variance (ANOVA) then followed with post hoc test (Least Square Deviation). P value of less than 0.05 was declared as significant statistically.

RESULTS

Effect of chloroquine and coartem on RBC, PCV and Hb on days 3 and 7 of administration in rats

On day 3 of the experiment, no significant differences were observed in the RBC count, Hb and PCV of the different groups, Figs. 1 to 3. The RBC count $(x10^6)$ cells/µL), Hb (g/dL) and PCV (%) of control group on day 3 were 6.22±0.51, 12.82±0.77 and 42.32±2.51 respectively. But on day 7, the red blood cell count of coartem recipients (6.05±0.31) was significantly lower when compared with the control $(7.20\pm0.21, p<0.05)$ and chloroquine (7.40±0.26, p<0.01) groups. Hb concentrations (g/dL) on day 7 was also significantly lower in the coartem (11.70 ± 0.33) recipients when compared with the control $(13.12\pm0.32, p<0.05)$ and chloroquine (13.90±0.42, p<0.01). Same results were obtained for the PCV (%) on day 7, it was significantly lower (p<0.05) in coartem (38.24 ± 1.64) recipients compared with control (44.46±1.21) and chloroquine (49.68±3.21) groups.

Effect of chloroquine and coartem on platelet count and total WBC count on days 3 and 7 of administration in rats

As shown in Fig. 5, the mean platelet counts $(x10^{3}cells/\mu L)$ in the control, coartem and chloroquine groups at day 3 were 558.20±110.94, 817.20±53.01 and 703.40±84.10 respectively, while their respective values at day 7 were 847.00±68.24, 674.40±53.41 and 851.00±56.43 respectively. The mean platelet counts were not statistically significant among the different groups both at day 3 and day 7 of administration.



Fig. 1

Erythrocyte counts in control rats and those treated with Choloroquine and Coartem 3 and 5 days after treatment. Values are Mean SEM of five animals *p<0.05 (vs control); $^{b}P<0.05$ (vs Coartem)



Fig. 2 Comparison of Hb concentration in the differen experimental groups after 3 and 7 days of treatment. Values are mean <u>+</u> SEM, n = 5. *p<0.05 vs control; b = p<0.01 vs Coartem</p>

Fig. 2

Hemoglobin concentration in control rats and those treated with Choloroquine and Coartem 3 and 5 days after treatment. Values are Mean SEM of five animals *p<0.05 (vs control); $^{a}P<0.05$ (vs Coartem)



Fig. 3

Packed Cell Volume in control rats and those treated with Choloroquine and Coartem 3 and 5 days after treatment. Values are Mean SEM of five animals *p<0.05 (vs control); $^{a}P<0.05$ (vs Coartem)



Fig. 4

Leucocyte counts in control rats and those treated with Choloroquine and Coartem 3 and 5 days after treatment. Values are Mean SEM of five animals *p<0.05 (vs control); $^{a}P<0.05$ (vs Coartem)

Effect of Chloroquine and Coartem on RBC indices on days 3 and 7 of administration in rats

Results on the effects of chloroquine and coartem on RBC indices (MCV, MCH, MCHC, RDW-SD and RDW-CV) are summarized in table 1.

On day 3, the MCV (fL) for control was 68.64 ± 2.12 , values for coartem, and chloroquine groups were 68.18 ± 1.73 , and 60.80 ± 0.98 respectively, showing significant decrease in chloroquine compared with control (p<0.05) and coartem (p<0.01) groups. Also, the MCH was significantly lower in chloroquine compared with control (p<0.05) and coartem (p<0.01) groups, while MCHC for control, coartem, and chloroquine were not significantly different.

On day 7, MCV (fL), MCH (pg) and MCHC (g/dL) for the control were 61.56 ± 0.70 , 18.26 ± 0.29 and 29.72 ± 0.59 respectively. No significant differences were observed in MCV, MCH and MCHC of the different groups on day 7.

The RDW-SD (fL) in the control, coartem and chloroquine at day 3 were 44.64 ± 4.46 , 42.08 ± 2.28 and 34.32 ± 1.66 respectively, showing significant decrease (p<0.05) in chloroquine recipients compare to coartem recipients. But at day 7, no significant differences were observed among the different groups.

The RDW-CV values in the control, coartem and chloroquine groups at day 3 were 15.44 ± 1.50 , 16.24 ± 0.7 and 14.54 ± 0.81 (fL) respectively, showing no significant differences. At day 7 their respective values were 17.98 ± 1.38 , 15.62 ± 1.05 and 14.68 ± 0.25 (fL), showing significant (p<0.05) decrease in chloroquine recipients compared with control..

Table 1

Effect of Chloroquine and Coartem on RBC and platelet indices on day 3 and 7 of administration in rats

^		Day 3			Day 7		
	Control	Coartem	CQ	Control	Coartem	CQ	
MCV (fL)	68.64±2.12	68.18±1.73	60.80±0.98*, ^b	61.56±0.70	63.34±0.90	59.50±2.52	
MCH (pg)	20.80±0.60	20.36±0.14	19.14±0.31*, b	18.26±0.29	19.46±0.60	18.82±0.23	
MCHC (g/dL)	30.32±0.53	29.96±0.71	31.52±0.38	29.72±0.59	30.68±0.57	30.46±0.39	
RDW-SD (fL)	44.64±4.46	42.08±2.28	34.32±1.66ª	39.42±2.34	37.36±2.00	34.84±0.88	
RDW-CV (%)	15.44 ± 1.51	16.24±0.71	14.54 ± 0.81	17.98 ± 1.38	15.62 ± 1.05	14.68±0.25*	
PDW (fL)	9.94±1.11	8.82±0.18	8.92±0.43	8.46±0.43	8.56±0.34	8.62±0.32	
MPV (fL)	7.52±0.40	7.28±0.06	7.30±0.24	6.90±0.24	7.06±0.14	7.04±0.15	
P-LCR (%)	11.44±2.51	9.10±0.39	9.32±1.44	6.90±0.99	7.84±0.67	7.54 ± 1.08	

Values are mean \pm SEM, n = 5. *p<0.05 vs control; a = p<0.05, b = p<0.01 vs coartem

Effect of Childfordune and Coartem on anterential whice count on day 5 and 7 of administration in fats.									
	Day 3			Day 7					
	Control	Cortem	CQ	Control	Cortem	CQ			
Lymphocytes (%)	83.40±5.85	84.86±4.61	81.60±4.24	76.00±8.06	77.80±4.82	74.20±3.60			
Eosinophils (%)	1.00 ± 0.45	2.20 ± 0.97	2.00 ± 0.45	2.00±0.55	1.80 ± 0.58	1.60 ± 0.51			
Neutrophils (%)	15.40±5.41	11.00±3.52	16.20±3.98	21.20±7.27	19.60±4.58	23.60±3.04			
Monocytes (%)	0.20±0.20	0.20±0.20	0.20±0.20	0.80±0.37	0.80±0.37	0.60±0.40			

 Table 2:

 Effect of Chloroquine and Coartem on differential WBC count on day 3 and 7 of administration in rats

Values are mean \pm *SEM, n* = 5.

No significant differences were observed between days 3 and 7



Fig. 4

Platelet counts in control rats and those treated with Choloroquine and Coartem 3 and 5 days after treatment. Values are Mean SEM of five animals p<0.05 (vs control); aP<0.05 (vs Coartem)

Effect of chloroquine and coartem on platelet indices on days 3 and 7 of administration in rats

The effects of chloroquine and coartem platelet indices (MPV, P-LCR and PDW) are also summarized in table 1. The PDW in the control, coartem and chloroquine at day 3 were 9.94 ± 1.11 , 8.82 ± 0.18 and 8.92 ± 0.43 (fL) respectively, while respective values at day 7 were 8.46 ± 0.43 , 8.56 ± 0.34 and 8.62 ± 0.32 respectively. No significant differences were also observed among the different experimental groups at both days 3 and 7 of administration. No significant differences were also observed in MPV and P-LCR following administration of chloroquine and coartem

Effect of chloroquine and coartem on differential WBC counts on days 3 and 7 of administration in rats

Results on differential WBC count are shown in table 2. No significant differences were observed in the lymphocytes, neutrophils, eosinophils and monocytes counts following administration of chloroquine and coartem on days 3 and 7.

DISCUSSION

Chloroquine is one of the oldest known anti-malaria drugs but due to resistance of malaria parasites, especially *P. falciparum* to chloroquine, new therapeutic regimens have been compounded. Recently, WHO recommends the use of artemisinin-based combination (ACTs) as first-line treatment of therapies uncomplicated Plasmodium falciparum malaria, (WHO, 2010). ACTs therapy is currently regarded more effective relative to non-artemisinin regimens like chloroquine, and also yielding rapid symptomatic improvement and parasite clearance and a reduction in gametocyte carriage, which could help to reduce malaria transmission (Targett et al., 2001; Kokwaro et al., 2007; Premji, 2009).

These drugs when administered are carried in the blood stream where their actions are executed. Obviously, these drugs are without side effects, hence this study aimed to elucidate comparatively the effect of chloroquine and coartem on haematological parameters after 3 and 7 days of administration in rats.

Results obtained from this study reveal that these drugs do not have tremendous or serious adverse effects on blood parameter after 3 days of administration, the red blood cell count, Hb and PCV were not significantly altered following administration of the drugs after 3 days, but after 7 days of administration, the RBC count, Hb and PCV were significantly reduced in coartem recipients compared with control. Coartem appeared to have no deleterious effect on red blood cell, Hb and PCV when administered at the recommended dose for 3 days. But there is an indication that prolonged administration of coartem would possibly lead to anaemia. This is in consonance with reports from preclinical data suggesting that repeated exposure to coartem may affect blood cell counts and predispose to anaemia (Obianine et al, 2011).

However, chloroquine did not have any significant influence on the RBC, Hb and PCV both after

3 and 7 days of administration but it rather lead to a reduction in MCV and MCH, a possible indication that administration of chloroquine for 3 days could lead to microcytic anaemia.

The RDW-SD was also significantly reduced in chloroquine recipients relative to coartem administered group. RDW-SD is a numerical measure of the variability in size (anisocytosis) of circulating erythrocytes (Perkins, 2003). This parameter is routinely reported as part of the complete blood count but its use is generally restricted to narrowing the differential diagnosis of anaemia (McKenzie, 2003). There is also a strong correlation between RDW and risk of adverse outcome of heart failure (Felker et al., 2007). It is also elevated in thrombotic thrombocytopenic purpura, a disease of unknown origin, characterised by abnormally low levels of platelets in the blood, formation of blood clot in the arterioles and capillaries of many organ and neurological damage.

Results obtained from this study on platelet counts and platelet indices indicate that neither chloroquine nor coartem has any tremendous effect on platelets, no significant alterations in platelet count and platelet indices were observed following administration of these drugs for 3 and 7 days. This observation is contrary to earlier report by Ashley (2008), that coartem drug may cause low platelet count which may lead to bleeding tendencies in patients, the drugs may also cause symptoms that are related to low platelet count. Nevertheless, Low mean platelet volume measurements are relatively rare and may be associated with serious illnesses such as leukemia.

Platelets play an important role in the integrity of normal homeostasis and mean platelet volume (MPV) is the indicator for its function (Jakubowski et al., 1983), including aggregation, release of thromboxane A₂, platelet factor 4, beta-thromboglbulin (Martin and Bath, 1991; Sharp et al., 1995) and expression of glycogen 1b and glycogen IIb/IIIa receptors 'Tschoepe et al., 1990; Giles et al., 1994).

MPV was not significantly altered in this study following administration of the two anti-malaria drugs. MPV a determinant of platelet function; is a newly emerging risk factor for atherothrombosis. Increase in MPV has been documented in patients with metabolic syndrome, stroke and diabetes mellitus (DM) (O'Malley and Langhorne, 1995; Tavil et al., 2007). Many studies have shown that increased MPV is one of the risk factors for myocardial infarction, cerebral ischaemia and transient ischaemic attacks (Khandekar et al., 2006; Kiliçli-Camur et al., 2005; Nadar et al., 2004; McCabe et al., 2004) and chronic vascular disease (Endler et al., 2002).

The total white blood cell (leucocyte) and differential counts were not significantly altered following administration of the chloroquine and coartem throughout the duration of the experiment. Our finding is at variance with earlier report by Adeleye *et al.*, (2012) that coartem increases total WBC and lymphocyte count but decreases neutrophils count, they attributed these changes to immunological response induced by the drug (Guyton and Hall, 2006).

In conclusion, administration of chloroquine and coartem at their recommended doses for 3 days does not adversely alter the levels of RBC, total & differential WBC, Hb, PCV, platelet count and platelet function indices, except for MCV and MCH reductions in chloroquine recipients. However, after 7 days of administration, coartem causes reduction in RBC count, Hb and PCV. Hence, this drugs should be used as prescribed by a certified Physician.

REFERENCES

Adeleye, G.S., Nneli R., Nwozor, C.M and Emesiana, M.C (2012). Effects of Coartem and Artesunate on some haematological and biochemical parameters in albino rats. Afr. J. Biomed. Res. 15: 55 - 58

Adjuik M, Babiker A, Garner P, Olliaro P, Taylor W, White N. (2004). Artesunate combinations for treatment of malaria: meta-analysis. Lancet.;363: 9-17. International Artemisinin Study Group

Ashley, E. A., Gready, M. C., Tarso, R. (2008). A randomized controlled trails of artemether-lumefantrine versus arteunate for uncomplicated *plasmodium falciparum* treatment. Parasitol Rev. 5-25.

Canadian Council of Animal Care (2009). The CCAC guidelines on: the care and use of farm animals in research, teaching and testing. Coulter Electronics. Instruction manual for the Coulter Model S-plus, 2nd edition. Bedfordshire, UK, 1979.

Cox-Singh J, Davis TM, Lee KS, Shamsul SS, Matusop A, Ratman S, et al. (2008). Plasmodium knowlesi malaria in humans is widely distributed and potentially life threatening. Clin Infect Dis.46:165-71.

Endler, G., Klimesch, A., Sunder-Plassmann, H. (2002). Mean platelet volume is an independent risk factor for myocardial infarction but not for coronary artery disease. British Journal of Haematology. 117, 399–404.

Fairhurst, R. M, Wellems, T. E (2010). Plasmodium species (malaria). In GL Mandell et al., eds., Mandell, Douglas, and Bennett's Principles and Practice of Infectious Diseases, 7th ed., vol. 2, pp. 3437-3462. Philadelphia: Churchill Livingstone Elsevier.

Falade C, Makanga M, Premji Z, Ortmann CE, Stockmeyer M, de Palacios PI (2005). Efficacy and safety of artemether-lumefantrine (Coartem) tablets (six-

regimen) in African infants and children with acute, dose

uncomplicated falciparum malaria. *Trans R Soc Trop Med Hyg*, 99:459-467.

Felker, G. M., Allen, L. A., Pocock, S., Shaw, L. K., McMurray, J. J. V., Pfeffer, M. A., Swedberg, K., Wang, D., Yusuf, Y., Michelson, E. L., Granger, C. (2007): CHARM Investigators. Red cell distribution width as a novel prognostic marker in heart failure: data from the CHARM Program and the Duke Databank for Cardiovascular Diseases. *Journal of American College of Cardiology*. 50, 40-47.

Giles, H., Smith, R. E. A., Martin, J. F. (1994). Platelet glycoprotein IIb/IIIa and size are increased in acute myocardial infarction. *European Journal of Clinical Investigation*. 24:69-72.

Greenwood D. (1995) Conflicts of interest: the genesis of synthetic antimalarial agents in peace and war. *J Antimicrob Chemother* 36: 857–872.

Jakubowski, J. A., Thompson, C. B., Vaillancourt, R., Valeri, C. R., Deykin, D. (1983). Arachidonic acid metabolism by platelets of differing size. *British Journal of Haematology*. 983(53): 503-511.

Joy, D. A., Feng X, Mu J, Furuya T, Hotivanich K, Kretti A. U. Krettli. Ho M, Wang A, White N. J, Suh E, Beerli P, Su X Z. (2003). Early origin and recent expansion of plasmodium falciparum. Science, 300 (5617): 318-321

Katzung, G. (2010). *Basic and clinical pharmacology* 11th ed. Lange Medical.

Khandekar, M. M., Khurana, A. S., Deshmukh, S. D. (2006). Platelet volume indices in patients with coronary artery disease and acute myocardial infarction: an Indian scenario. *Journal of Clinical Pathology*. 59: 146-149.

Kiliçli-Camur, N., Demirtunç, R., Konuralp, C., Eskiser, A., Başaran, Y. (2005). Could mean platelet volume be a predictive marker for acute myocardial infarction? *Medical Science Monitor*. 11: CR387-392.

Kokwaro G, Mwai L, Nzila A. (2007). Artemether/lumefantrine in the treatment of uncomplicated falciparum malaria. *Expert Opin Pharmacother*. 8:75–94.

Koram KA, Abuaku B, Duah N, Quashie N. (2005). Comparative efficacy of antimalarial drugs including ACTs in the treatment of uncomplicated malaria among children under 5 years in Ghana. Acta Trop. 95:194–203.

Krafts, K., Hempelmann, E., Skorskastania, H. A (2012). Methylene blue to chloroquine: A brief review of the development of an antimalaria therapy. *Parasitol Rev.* 11 (1):1-6.

Makanga M, Premji Z, Falade C, Karbwang J, Mueller EA, Andriano K, Hunt P, De Palacios PI. (2006). Efficacy and safety of the six-dose regimen of arthemeter-lumefrantrine in pediatrics with uncomplicated *Plasmodium falciparum* malaria: a pooled analysis of individual patient data. *Am J Trop Med Hyg.* 74:991–998.

Martin, J. F., Bath, P. M. W. (1991). Platelets and megakaryocytes in vascular disease. In: Herman AG, ed. Antithrombotics: pathophysiological rationale for pharmacological inventions. Dordrecht, Boston. Kluwer Academic Publishers. 1:49–62.

McCabe, D. J., Harrison, P., Sidhu, P. S., Brown, M. M., Machin, S. J. (2004). Circulating reticulated platelets in the early and late phases after ischaemic stroke and transient ischaemic attack. *British Journal of Haematology*. 126: 861-869

McKenzie, S. D. (2003). Introduction to anemia. In: McKenzie SD, ed. *Clinical Laboratory Hematology*.

Saddle River, NJ: Pearson Prentice-Hall. Pp. 161–188.

Mueller *et al*, (2006). Efficacy and safety of the six-dose regimen of artemether–lumefantrine for treatment of uncomplicated Plasmodium falciparum malaria in adolescents and adults: A pooled analysis of individual patient data from randomized clinical trials; *Acta Tropica* 100:41–53

Mulenga, M., Vangheertruyden, J. P., Mwanayanda, L,
Chalwe, V., Moerman, F.Chilengi,
Chilengi,
R.,
Vandremai, C., Durjardin, J. C., Dalessando, U., (2006).Safety and
(coartem) for the treatment oflumefantrine-artemether
uncomplicated

plasmodium falciparum malaria in Zambian adult. Malaria Journal, 5:73

Nadar, S. K., Lip, G. Y., Blann, A. D. (2004). Platelet morphology, soluble P selectin and platelet P-selectin in acute ischaemic stroke. The West Birmingham Stroke Project. Thrombosis and Haemostasis. 92:1342-1348.

Nosten, F. and White, N. J. (2007). Artemisinin-based combination treatment of falciparum malaria. *Am Trop. Med. Hyg.* 77 (6) 181-192.

Novartis, (2005). *Coartem product monograph*. Novartis AG 5th ed.

Obianime, A.W. and J.S. Arioku (2011). Mechanism of action of Artemisinin on biochemical, hematological and reproductive parameters. Int J. pharmacology 7: 84-95.

Ohwada, K. (1986). Improvement of cardiac puncture in mice. *Jikken Dodutsu*, 35(3): 353-355.

O'Malley, T., Langhorne, P., Elton, R. (1995). Platelet size in stroke patients. *Stroke*. 26: 995-999.

Perkins, S. L. (2003). Examination of blood and bone marrow. In: Greer, J. P., Foerster, J., Lukens, J. N., Rodgers, G. M., Paraksevas, F., Glader, B. E, eds. *Wintrobe's Clinical Hematology*. 11th ed. Salt Lake City, Utah: Lippincott Wilkins & Williams. pp 5–25.

Premji ZG. (2009). Coartem: the journey to the clinic. *Malar J.* 8 (Suppl 1):S3.

Sharp, D. B., Bath, P. M. W., Martin, J. F. (1995). Cigarette smoking sensitizes and desensitizes impedance-measured ADP-induced platelet aggregation in whole blood. *Thrombosis and Haemostasis*. 74: 730–735.

Sheikh, M. A., Ahmed, S., Diju, I. U., Dur, E., Yakta, J. A. (2011). Med. Coll. Abbottabad. *Rev.* 23 (1):143-145.

Sipilanyambe, N., J. L. Simon, P. Chanda, P. Olumese, R. W. Snow, and D. H. Hamer. (2008). From chloroquine to artemether-lumefantrine: the process of drug policy change in Zambia. *Malar. J.* 7:25.: 25.

Sinclair D, Zani B, Donegan S, Olliaro P, Garner P. (2009). Artemisinin-based combination therapy for treating uncomplicated malaria. Cochrane Database Syst Rev. 8:CD007483. **Snow RW, Guerra CA, Noor AM, Myint HY, Hay SI** (2005) The global distribution of clinical episodes of Plasmodium falciparum malaria. *Nature* 434: 214–217.

Targett G, Drakeley C, Jawara M, von Seidlein L, Coleman R, Deen J, Pinder M, Doherty T, Sutherland C, Walraven G, Milligan P. (2001). Artesunate reduces but does not prevent posttreatment transmission of *Plasmodium falciparum* to *Anopheles gambiae*. J Infect Dis. 183:1254– 1259.

Tavil, Y., Sen, N., Yazici, H. U. (2007). Mean platelet volume in patients with metabolic syndrome and its relationship with coronary artery disease. *Thrombosis Research.* 120: 245-250.

Tschoepe, D., Rosen, P., Kaufmann, L. (1990). Evidence for abnormal platelet glycoprotein expression in

diabetes mellitus. European Journal of Clinical Investigation. 20:166-170.

White, N. J. (2004). Antimalarial drug resistance *J. Clin. Invest Rev.* 113 (8) 1084-1092.

WHO (2008): *World Malaria Report.* World Health Organization, Geneva; 2008.

WHO/Regional Office for Europe (2006) Regional strategy: from malaria control to elimination in the WHO European Region 2006–2015. Copenhagen: World Health Organization Regional Office for Europe. 41 p. WHO-EUR/06/5061322.

WHO (2010): World Health Organization Guidelines for the Treatment of Malaria. Second Edition. World Health Organization; 2010. <u>http://www.who.int/malaria/publications</u>/<u>atoz/9789241547925/en/index.html</u>. Accessed January 26, 2013