

Afr. J. Biomed. Res. Vol. 25 (January, 2022); 83 - 87

Research Article

# Effects of Exogenously Administered Cortisol on Lipolysis in the Common African Toad, *Bufo regularis*

Isehunwa G.O\*, Agboola, M.O, Shittu, S. T, Alada, A.R.A

Department of Physiology, College of Medicine, University of Ibadan. Ibadan, Nigeria

#### ABSTRACT

The role of cortisol on lipolysis in amphibians is not known. This study was designed to investigate the effects of cortisol on lipolysis in the common African toad, *Bufo regularis*. Adult toads were collected and used for the study. The animals were fasted 24h and anaesthetized by sodium thiopentone 50mg/kg intraperitoneally. Blood was collected from truncus arteriosus for estimation of blood glucose and blood free fatty acids levels. Cortisol caused significant increase in blood free fatty acids and glucose levels in the common African toad. Pretreatment with prazosin 0.2mg/kg produced significant reduction in blood free fatty acids levels. Propranolol 0.5mg/kg pretreatment caused significant increase in blood free fatty acids and significant reduction in glucose levels. The combination of both blockers abolished the cortisol-induced hyperglycemia and caused significant reduction in blood free fatty acids. The results of this study confirmed that cortisol caused lipolysis from toad adipose tissue. Thus, cortisol administration caused increase in blood free fatty levels and induced hyperglycemia. The alpha-adrenergic receptors are involved in the release of free fatty acids in the common African toad *Bufo regularis*.

Keywords: Cortisol, Free fatty acids, hyperglycemia, prazosin, propranolol, common African toad

\*Author for correspondence: Email: funmisehunwa@yahoo.com; Tel: +234 8064322920

Received: February 2021; Corrected version accepted: September 2021

# INTRODUCTION

Free fatty acids (FFA) are the dominant oxidative fuel for all major tissues during fasting except the brain which depends largely on glucose derived mainly from gluconeogenesis (Chen et al., 1999). Lipoprotein lipase (LPL) hydrolyses triglycerides of chylomicrons and very low-density lipoproteins (VLDL). The LPL activity determines triglyceride concentration and causes release of free fatty acid into tissues that utilize them (Eckel, 1989; Fielding and Frayn, 1998). The role of FFA in gluconeogenesis is species specific. For instance, free fatty acids stimulate gluconeogenesis in rats but inhibits in dogs, cats, and guinea pigs (Corredor et al., 1969; Arinze and Hanson, 1973; Jomain-Baun and Hanson, 1975). There is discrepancy in results in humans. For instance, the studies of (Clore et al., 1991; Puhakainen et al., 1993) reported that FFA stimulated gluconeogenesis while Frye et al (1996) reported FFA inhibited gluconeogenesis.

Cortisol increases fuel substrates through mobilization of glucose (Rizza *et al.*, 1982; Dinneen *et al.*, 1993), free fatty acids (Djurhuus *et al.*, 2002) and amino acids (Horber and Haymond, 1990; Berneis *et al.*, 1997) from endogenous stores. Chronic exposure to cortisol is associated with impaired metabolism and insulin action which results in hyperglycemia and dyslipidemia. Cortisol induces insulin resistance (Rizza *et al.*, 1982; Dinneen *et al.*, 1993). It has

been reported that cortisol contributes to the metabolic syndrome and cardiovascular disease (Andrews and Walker, 1999; Darmon et al., 2006). There are many but conflicting reports on the effects of cortisol on lipolysis. For instance, in vitro studies in adipocytes showed that cortisol increased (Djurhuus et al., 2002; Xu et al., 2009), unchanged (Lee et al., 2012), and decreased (Ottosson et al., 2000) adipose tissue lipolysis. The effects of cortisol on lipolysis varies with species, the specific adipose tissue depot (Lee et al., 2008), concentration and duration of exposure and effects of other hormones in the medium (Geer et al., 2014; Stimson et al., 2017) thus accounting for the conflicting reports. In vivo studies in humans have shown that cortisol acutely increased whole-body lipolysis (Divertie et al., 1991; Djurhuus et al., 2002, 2004). However, recent study shows that acute supraphysiological concentration of cortisol and in the presence of insulin and adrenaline is required to cause lipolysis in human subcutaneous adipose tissue and not visceral adipose tissue (Stimson et al., 2017). Adrenocotropic hormone (ACTH) and catecholamines have been reported to cause release of free fatty acids from rat adipose tissue while addition of phentolamine an alpha-adrenergic blocking agent inhibited the release of free fatty acid (Schote and Page, 1960). Cortisol and other glucocorticoids have been documented to be secreted by amphibians in different stress conditions (Narayan et al., 2013; Jones et al., 2016; Fonner et al., 2017;

Gabor et al., 2018) and several non-invasive methods of cortisol sampling in amphibians have been developed (Santymire et al., 2018; Forsburg et al., 2019). There is paucity of information on the effect of cortisol on lipolysis in amphibians. Therefore, it is important to know the metabolic response of amphibian species to cortisol in view of the earlier reported differences in responses of mammals to cortisol. This study was designed to investigate the effects of cortisol on blood free fatty acids and glucose levels in the Common African toad *Bufo regularis*. Furthermore, the role of adrenergic receptors on the effects of cortisol on lipolysis was investigated.

#### MATERIALS AND METHODS

**Animals:** One hundred adult toads (70-100g) were used for the study. The toads were collected randomly from around ponds and banks of slow-moving streams around University of Ibadan, Ibadan, Oyo state, South-western Nigeria.

Experimental procedure: The toads were fasted 24h and anaesthetized with sodium thiopentone 50mg/kg intraperitoneally. The truncus arterious was dissected free from surrounding tissue for blood sample collection while the anterior abdominal vein was cannulated for drug injection. Fluidity of blood was maintained by heparin injection. Each toad was heparinised (170 units/0.1ml) and allowed 30mins to stabilize. After stabilization, toads in group 1 (control) received 0.7% amphibian saline intravenously (i.v) through anterior abdominal vein cannula while toads in group 11 (untreated) received cortisol (50µg/kg i.v). Toads in groups 111, 1V, and V were pretreated with propranolol 0.5mg/kg i.v, prazosin 0.2mg/kg i.v, combined propranolol 0.5mg/kg i.v and prazosin 0.2mg/kg i.v respectively 30mins before given cortisol 50µg/kg i.v. Each drug injection was in a volume between 0.1ml and 0.12ml given intravenously.

**Biochemical analysis:** Blood samples were collected at 0mim, 30min, 60min, and 90min post-injection period for estimation of blood glucose and blood free fatty acid levels. Blood glucose was estimated by modified glucose oxidase method (Trinder, 1969). Blood free fatty acids was measured using modified colorimetric determination of free fatty acids in biological fluids (Itaya and Ui, 1965). Owing to the small size of the toad, animals were sampled only once in each experiment and then sacrificed.

**Statistical analysis:** All values given are mean  $\pm$  S.E.M of the variables measured. Differences between two groups were compared using student t-test while one-way analysis of variance (ANOVA) was used to compare mean values in multiple groups. P< 0.05 was taken as statistically significant

#### RESULTS

The results of the experiments are shown in Figure 1, and Tables 1 and 2. Injection of 0.7% amphibian saline had no effect on blood glucose and blood free fatty acids levels. However, injection of cortisol 50µg/kg caused significant increase in blood glucose and free fatty acids levels (Figure 1). Pretreatment of toads with propranolol blocked the increase in glucose levels caused by cortisol injection and caused rise in blood free fatty acids compared with untreated toads (Tables 1 and 2). In toads pretreated with prazosin, cortisol injection did not produce any significant effect on blood glucose levels but caused significant reduction in blood free fatty acids (Tables 1 and 2) compared with untreated animals. Combined pretreatments with propranolol and prazosin abolished the increase in blood glucose levels caused by cortisol injection and caused reduction in blood free fatty acids (Tables 1 and 2).



# Figure 1

Effect of cortisol (50  $\mu$ g/kg) on blood free fatty acids and blood glucose levels in the common African toad *bufo regularis* 

Afr. J. Biomed. Res. Vol. 25, No.1 (January) 2022

Table 1:

Comparison of Blood Glucose of Control, Cortisol, and Pretreated groups (Prazosin and Propranolol groups)

Treatment	Blood glucose (mg/dl)				
Time	0min	30min	60min	90min	
Amphibian saline 0.7% (control)	$65.2 \pm 1.4$	53 ±5.4	64±5.5	55.2±3.2	
Cortisol 50µg/kg	$54.4 \pm 1.3$	$*102 \pm 4.6$	$*109 \pm 4.6$	$*103.4 \pm 2.9$	
Propranolol 0.5mg/kg + cortisol 50µg/kg	$64.2\pm6.9$	$76.8\pm3.4$	$88 \pm 2.7$	$47 \pm 9.3$	
Prazosin 0.2mg/kg + cortisol 50µg/kg	$53.6\pm7.7$	$94.2\pm3.9$	$100.6\pm3.4$	$83.6\pm5.1$	
Prazosin 0.2mg/kg + propranolol 0.5mg/kg + cortisol 50µg/kg	$56.8\pm6.1$	$**69.8\pm6.2$	$**59.4\pm3.4$	$**58.8\pm8.1$	

\*P< 0.01 significant increase compared with basal 0 min and control

\*\*P< 0.01 significant decrease compared with cortisol group

#### Table 2:

Comparison of Blood Free Fatty Acids of Control, Cortisol, and Pretreated groups (Prazosin and Propranolol groups)

	Blood free fatty acids (mg/dl)				
Time	0min	30mins	60mins	90mins	
Treatment					
Amphibian saline 0.7% control	$70.0\pm2.0$	$66.2\pm$ 6.5	$58.6\pm5.6$	67.7±9.8	
Cortisol 50µg/kg	$54.8\pm3.4$	$86.6\pm8.4*$	$*81.2 \pm 3.4$	$*110.5 \pm 4.6$	
Propranolol 0.5mg/kg + cortisol 50µg/kg	$54.5\pm3.3$	$157.9 \pm 10.9 **$	$**137.3 \pm 10.1$	$**121.2 \pm 4.0$	
Prazosin 0.2mg/kg + cortisol 50µg/kg	$52.5\pm3.4$	$31.9 \pm 11.1^{***}$	$71.2 \pm 6.7$	$78.5\pm6.6$	
Prazosin (0.2mg/kg) + propranolol (0.5mg/kg) +	$56.2 \pm 2.1$	$51.8\pm1.0$	$92.3\pm6.6$	$80.5\pm6.2$	
cortisol $(50\mu g/kg) (0.5mg/kg) + \text{cortisol} (50\mu g/kg)$					

\*P < 0.05 significant increase compared with basal 0min; \*\*P < 0.01 significant increase compared with cortisol group \*\*\*P < 0.01 significant decrease compared with cortisol group

Cortisol 50µg/kg caused significant increase in blood glucose levels 30mins, 60mins, and 90mins post-injection period. Pretreatment with propranolol 0.5mg/kg blocked the increase in blood glucose caused by 50µg/kg cortisol injection. In prazosin pretreated toads, cortisol injection (50µg/kg) produced no significant difference in blood glucose levels compared with cortisol group

Combined pretreatment with propranolol 0.5 mg/kg and prazosin 0.2 mg/kg abolished the increase in blood glucose level caused by cortisol injection ( $50\mu$ g/kg).

Cortisol injection  $50\mu$ g/kg caused significant increase in blood free fatty acids compared with basal (0min). Pretreatment with propranolol 0.5mg/kg caused significant increases in blood free fatty acids compared with cortisol group. Prazosin 0.2mg/kg pretreatment caused reduction in blood free fatty acids compared with cortisol group. Combined pretreatment with propranolol 0.5mg/kg and prazosin 0.2mg/kg caused reduction in blood free fatty acids compared with cortisol group.

# DISCUSSION

Cortisol injection caused increase in blood free fatty acids levels in the toads. This is consistent with studies in humans (Samra *et al.*, 1998; Djurhuus *et al.*, 2002, 2004., Campbell *et al.*, 2011; Stimson *et al.*, 2017) and rats (Schote and Page, 1960). Cortisol promotes lipolysis by mobilizing free fatty acids from adipose tissue and enhanced hepatic gluconeogenesis (Grav Holt *et al.*, 2002; Geer *et al.*, 2014). Studies in humans (Ottosson *et al.*, 1994; Stimson *et al.*, 2017) and rats (Xu *et al.*, 2009; Campell *et al.*, 2011) showed that cortisol regulates lipolytic pathway through increase in transcription of key lipases enzymes, adipose triglyceride lipase and hormone sensitive lipase. The observation of the present study in which cortisol injection caused increase in blood free fatty acids confirmed its lipolytic activity in the toad *bufo regularis* and may be due to increase in transcription of lipases enzymes (Stimson *et al.*, 2017). The lipoprotein lipase activity hydrolyses triglycerides and releases free fatty acids to tissues that utilize them. (Zechner, 1997; Fielding an Frayn, 1998, Kovar *et al.*, 2004). It has been reported that adrenocorticotropic hormone (ACTH) injection caused mobilization of free fatty acids from rat adipocytes (Schote and Page, 1960).

The increase in glucose level following cortisol injection confirms its hyperglycemic effect (Broughton and Deroos, 1984; Leach and Taylor, 1982; Pretty et al., 2009; Isehunwa et al., 2013). Cortisol causes muscle protein breakdown, lipolysis of adipose tissue, hepatic gluconeogenesis, impairs glucose uptake in muscle thereby increase circulating glucose levels in mammals (Geer et al., 2014). The rise in glucose level and concomitant increase in free fatty acids level observed in the present study suggests that cortisol injection most probably suppressed secretion and action of insulin. As a result of the insulin release suppression, there was decrease in peripheral glucose uptake and increase in circulating glucose levels (Dinneen et al., 1993; Macfarlane et al., 2008). It has also been shown that an increase in plasma free fatty acids inhibits insulin-stimulated glucose uptake in man (Ferrannini et al., 1983). Thus, the findings of the present study revealed that under stress cortisol administration caused increase in blood free fatty acids and induced hyperglycemia. The decrease in glucose levels observed in toads pretreated with propranolol seems to confirm that the beta adrenergic

receptors are involved in cortisol-induced hyperglycemia in the toad bufo regularis (Isehunwa et al., 2013) while the significant rise in blood FFA in propranolol pretreated toads is most probably an indication that the beta adrenergic receptors may not be involved in the release of FFA from toad adipose tissue. During beta-blockade, there was unopposed alpha stimulation which could result in activated lipoprotein lipase enzyme causing increase in the level of blood FFA. Therefore, pretreatment of toads with propranolol did not block the release of free fatty acids hence the significant rise in blood FFA compared with untreated toads. This observation contrasts the findings in humans by Imura et al (1971) which reported that beta adrenergic blockage inhibited fat mobilization whereas alpha blockage stimulated fat mobilization from adipose tissue.

In toads pretreated with prazosin, an alpha blocking agent, there was significant reduction in blood FFA suggesting that the alpha-adrenergic receptors are probably involved in the release of blood FFA from the adipose tissue. As a result, pretreatment with prazosin blocked release of FFA from adipose tissue. This is consistent with the findings in rats (Schote and Page, 1960) which reported that prazosin blocked the increase in blood free fatty acids caused by ACTH. The reduction in blood FFA in toads pretreated with prazosin in the present study could be that prazosin inhibited lipoprotein lipase enzyme activity with subsequent reduction in blood FFA. Lipoprotein lipase activity hydrolyses triglycerides and releases free fatty acids to tissues that utilize them. (Zechner, 1997; Fielding an Frayn, 1998; Kovar et al., 2004). The findings of the present study contrast the observations in humans (Day et al., 1982; Shaw et al., 1978) that prazosin caused reduction in triglycerides while propranolol increased triglycerides levels. Combined pretreatment with both blockers abolished the increase in glucose levels caused by cortisol and resulted in reduction of free fatty acid levels compared with untreated animals. This observation seems to suggest that the beta and alpha- adrenergic receptors are involved cortisol-induced hyperglycemia and increased release of free fatty acids in the toads. The findings of this study revealed that cortisol caused lipolysis and induced hyperglycemia in the toads.

In conclusion, this study showed that administration of cortisol caused lipolysis and induced hyperglycemia in the common African toad *bufo regularis*. The results also suggest that the alpha-adrenergic receptors are most probably involved in cortisol-induced increase in blood free fatty acid levels whereas the beta-adrenergic receptors are involved in cortisol hyperglycemia in the common African toad.

# REFERENCES

Andrews R.C., and Walker B.R. (1999). Glucocorticoids and insulin resistance: old hormones, new targets. Clin. Sci. (Lond) 96:513 – 523

Arinze I., and Hanson R. (1973). Mitochondrial redox state and the regulation of gluconeogenesis in the isolated, perfused cat liver. FEBS Lett. 31: 280-282

Berneis K., Ninnis R., Girard J., Frey B. M., Keller U. (1997). Effects of insulin-like growth factor I combined with growth hormone on glucocorticoid-induced whole-body

protein catabolism in man. J. Clin. Endocrinol. Metab. 82: 2528 – 2534.

**Broughton R. E., and Deroos R. C. (1984).** Temporal effects of infused corticosterone and aldosterone on plasma glucose levels in the American Bullfrog (Rara Castebiana). Gen. Comp. Endocrinol. 35:205 - 215

Campbell J.E., Peckett A. J., D 'Souza A. M, Hawke T. J., and Riddell M.C. (2011). Adipogenic and lipolytic effects of chronic glucocorticoid exposure, Am. J. Physiol. Cell. Physiol. 2011; 300(1):C198-209. [Pubmed: 20943959]

**Chen X., Igbal N., and Boden G. (1999).** The effects of free fatty acids on gluconeogenesis and glycogenolysis in normal subjects. J. Clin. Invest. 103: 365-372

Clore J. N., Glickman P. S., Helm S.T., and Nestler J. E., and Blackard W. G. (1991). Evidence for dual control mechanism regulating hepatic glucose output in nondiabetic men. Diabetes 40: 1033-1040.

**Corredor C., Brandel K., and Bressler R. (1969).** Effects of 4-pentenoic acid on carbohydrate metabolism in pigeon liver homogenate. J. Biol. Chem. 244: 1212-1219

**Darmon P., Dadoun. F., Boullu-Ciocca S., Grino M., Alessi M. C., and Dutour A. (2006).** Insulin resistance induced by hydrocortisone is increased in patients with abdominal obesity. Am J. Physiol. Endocrinol. Metab. 291: E995 – E1002.

Day J. L., Metcalfe J., Simpson C. N. (1982). Adrenergic mechanisms in control of plasma lipid concentrations Br. Med. J. 284: 1145-118

**Dinneen S., Alzaid A., Miles J., and Rizza R. (1993).** Metabolic effects of the nocturnal rise in cortisol on carbohydrate metabolism in normal humans. J. Clin. Invest. 92: 2283-2290.

**Divertie G. D., Jensen M. D., and Miles J. M. (1991).** Stimulation of lipolysis in humans by Physiological hypercortisolemia. Diabetes 40: 1228-1232

**DJurhurus C. B., Gravholt C. H., Nielsen S., Mengel A., Christiansen J. S., Schmitz O. E., and Moller N. (2002).** Effects of cortisol on lipolysis and regional interstitial glycerol levels in humans. AM. J. Physiol. Endocrinol. MetaE172-: 177

Djurhuus C. B., Gravhol C. H., Nielsen S., Pedersen S. B., Moller N., and Schmitz O. (2004) Additive effects of cortisol and growth hormone on regional and systemic lipolysis in humans. Am. J. Physiol. Endocrinol. Metab. 286:E488-494. [Pubmed: 14600073]

**Eckel R. H.** (1989). Lipoprotein lipase. A multifunctional enzyme relevant to common metabolic diseases. N. Engl. J. Med. 320 : 1060-1068

Ferrannini E., Barrett E. J., Bevilacoua S., and Defronzo R. A. (1983). Effect of fatty acids on glucose production and utilization in man. J. Clin. Invest. 72: 1737-1747

**Fery F., Plat L., Melot C., and Balasse E. O. (1996).** Role of fat derived substrates in the regulation of gluconeogenesis during fasting. Am. J. Physiol, 270: E822-E830

Fielding B. A., and Frayn K. N. (1998). Lipoprotein lipase and the disposition of dietary fatty acids. Br. J. Nutr. 80:495-502.

Fonner C.W., Patel S.A., Boord S.M., Venesky M.D. and Woodley S.K. (2017). Effects of corticosterone on infection

and disease in salamanders exposed to the amphibian fungal pathogen *Batrachochytrium dendrobatidis*. Dis Aquat Org 123:159-171. https://doi.org/10.3354/dao03089

**Forsburg Z.R., Goff C.B., Perkins H.R., Robicheaux J.A., Almond G.F. and Gabor C.R. (2019).** Validation of waterborne cortisol and corticosterone in tadpoles: Recovery rate from an acute stressor, repeatability, and evaluating rearing methods. General and Comparative Endocrinology, 281: 145-152

Gabor C.R., Knutie S.A., Roznik E.A. and Rohr J.R.

(2018). Are the adverse effects of stressors on amphibians mediated by their effects on stress hormones? Oncologia 186: 393-404

**Geer E. B., Islam J., and Buettner C. (2014).** Mechanism of Glucocorticoid-induced insulin resistance. Endocrinol. Metab. Clin. North. AM. 43 (1): 75 – 102

Gravoholt C. H., Dall R., Christiansen J. S., Moller N., Schmitz O. (2002). Preferential stimulation of abdominal subcutaneous lipolysis after prednisone exposure in humans. Obes. Res. 10:774-781

Horber F. F., Haymond M. W. (1990). Human growth hormone prevents the protein catabolic side effects of prednisone in humans. J. Clin. Invest. 86:265 – 272

**Imura H., Kato Y., Ikeda M., Morimoto M., and Yawata M. (1971).** Effect of adrenergic blocking or stimulating agents on plasma growth hormone, immune reactive insulin, and blood free fatty acids levels in man. J. Clin. Invest. 50:1069-1079.

**Isehunwa G. O., Olaniyan O. T., and Alada A.R.A. (2013).** The role of alpha- and beta-adrenergic receptors in cortisolinduced hyperglycemia in the common African toad (*Bufo regularis*). Afri. J. Biotech. Vol. 12 (36), pp.5554-5558

Itaya K., and Ui M. (1965). Colorimetric determination of free fatty acids in biological fluids. J. Lipid Res, 6:16 – 20.

Jomain-Baum M., and Hanson R. (1975). Regulation of hepatic gluconeogenesis in the gunea pig by fatty acids and ammonia. J. Biol. Chem. 250: 8978-8985

Jones B.C., Smith A.D., Bebus S.E. and Schoech S.J. (2016). Two seconds is all it takes: European starlings (*Sturnus vulgaris*) increase levels of circulating glucocorticoids after witnessing a brief raptor attack. Hormones and Behavior 78: 72-78

Kovar J., Fejfarova V., Pelikonova T., Poledne R. (2004). Hyperglycemia down regulates total lipoprotein lipase activity in humans Physiol. Res. 53:61-68

**Leach G. J., and Taylor M. H. (1982).** The effects of cortisol treatment on carbohydrate and protein metabolism in Fundulus heteroclitus. Gen. Comp. Endocrinol. 48: 76 - 83

Lee M. J., and Fried S. K. (2012). Glucocorticoids antagonize tumor necrosis factor alpha- stimulated lipolysis and resistance to the antilipolytic effect of insulin in human adipocytes. Am. J. Physiol. Endocrinol. Metab.303: E1126-E1133

Lee M. J., Fried S. K., Mundt S. S., et al., (2008). Depotspecific regulation of the conversion of cortisone to cortisol in human adipose tissue. Obesity (Silver Spring) 16(6) 1126 – 33. **Macfarlane D. P., Forbes S., and Walker B. R. (2008).** Glucocorticoids and fatty acid metabolism in humans: fueling fat redistribution in the metabolic syndrome. J. Endocrinol. 197: 189 – 204

Narayan E.J., Cockrem J.F. and Hero J-M (2013). Sight of a Predator Induces a Corticosterone Stress Response and Generates Fear in an Amphibian. PLoS ONE 8(8): e73564. https://doi.org/10.1371/journal.pone.0073564

Ottosson M., Lonnroth P., Bjorntorp P., and Eden S. (2000). Effects of cortisol and growth hormone on lipolysis in human adipose tissue. J. Clin. Endocrinol. Meta. 85:799-803. Ottosson M., Vikman-Adolfsson K., Enerback S., Olivecrona G., Bjorntorp P. (1994). The effects of cortisol on the regulation of lipoprotein Lipase activity in human adipose tissue. J. Clin. Endocrinol. Metab. 79(3): 820 – 5

**Pretty C., Chase J. G., Lim J., Shaw G., Compte A. L., Razak N., and Parente J. (2009).** Corticosteroids and insulin resistance in the ICU proceedings of the 7<sup>th</sup> IFAC symposium on modelling and control in biomedical systems, Aalborg, Denmark

**Puhakainen I., and Yki-Jarvinen H. (1993).** Inhibition of lipolysis decreasing lipid oxidation and gluconeogenesis from lactate but not fasting hyperglycemia or total hepatic glucose production in NIDDM. Diabetes. 42: 1694-1699

**Rizza R. A., Mandarino L. J., Gerich J. E. (1982).** Cortisolinduced insulin resistance in man: impaired suppression of glucose production and stimulation of glucose utilization due to a postreceptor detect of insulin aaction. J. Clin. Endocrinol. Metab. 54: 131 -138

Samra J. S., Clark L., Humphreys S. M., MacDonald I. A., Banister P. A., and Frayn N K.N. (1998) Effects of Physiological hypercortisolemia on the regulation of lipolysis in subcutaneous adipose tissue. J. Clin. Endocrinol. Metab. 83:626-631. [Pubmed: 9467584]

Santymire, R. M., Manjerovic, M. B. and Sacerdote-Velat, A. (2018). A novel method for the measurement of glucocorticoids in dermal secretions of amphibians. *Conservation physiology*, 6(1), coy008. https://doi.org/10.1093/conphys/coy008

Schote M. C and Page I. H. (1960). Effect of adrenergic blocking agents on the release of free fatty acids from rat adipose tissue. J. Lip. Res. Vo11(5) 466-468

Shaw J., England J. D. F., and Hua A. S. P. (1978). Betablockers and plasma tryglycerides. Br. Med. J. 486

Stimson R. H, Macfarlane D. P., Andrew R., Walker B. R., Anderson A. J. et al. (2017). Acute physiological effects of glucocorticoids on fuel metabolism in human are permissive but not direct. Diabetes Ches metab. 19:883-891.

**Trinder E. (1969).** Determination of blood glucose using 4 amino phenazone as oxygen acceptor. J. Chem. Pathol. 22: 246-248

Xu C., He J., Jiang H. et al. (2009). Direct effect of glucocorticoids on lipolysis in adipocytes. Mol. Endocrinol. 23: 1161-1170

**Zechner R.** (1997). The tissue-specific expression of lipoprotein lipase: implications for energy and lipoprotein metabolism. Curr. Opin. Lipidol 8: 77 – 88.