

Short communication

# SCREENING OF SOME NIGERIAN MEDICINAL PLANTS FOR ANTIOXIDANT ACTIVITY USING 2, 2, DIPHENYL-PICRYL-HYDRAZYL RADICAL

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As part of a screening program for biologically active compounds in plants, twenty two medicinal plants were extracted and screened for anti-oxidant activity using the 2, 2,diphenyl-picryl-hydrazyl radical.

Key words:- 2,2- diphenyl-picyl-hydrazyl, flavonoids, antioxidants

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#### INTRODUCTION

Nature is and will still serve as the man's primary source for the cure of his ailments. However, the potential of higher plants as sources for new drugs is still largely unexplored. Consequently, in consultation with some prominent Nigerian native medicine practitioners, twenty two (22) Nigerian medicinal plant materials (Table 1) were selected for this study. These medicine practitioners have claimed that these plants materials are effective in the cure of central nervous system (C.N.S.) diseases e.g. psychiatric disorders, inflammation Recent interest in the study of and pains. antioxidants may not be unconnected with the efficacy of these compounds to cure most diseases of man particularly the C.N.S. ailments.

Antioxidants are radical scavengers which protect the human body against free radicals that may cause pathological conditions such as asthma. ischemia, anaemia. arthritis. neuro-degenertion, Parkinson's inflammation. diseases, mongolism, ageing process and perhaps dementias (Polterat 1997) Flavonoids and flavones are widely distributed secondary metabolites with antioxidant and antiradical (Nakayoma and Yamada 1995). properties.

This paper reports the phyto-chemical screening and isolation of the antioxidant components of these selected Nigerian medicinal plant materials.

## MATERIALS AND METHODS Plant Materials

All the plant materials (shown on Table 1) are collected around the Southern Western part of Nigeria and then their voucher specimen deposited at Forestry Research Institute of Nigeria (FRIN) Ibadan, Nigeria.

#### **Extraction Procedure**

50gm of the powdered specimen of each of the plant materials shown in Table 1 was extracted

with 500mls of methanol for 3 days respectively. The coloured solution obtained from each of the plant material was concentrated by vacuum evaporation. The concentrate (extract) obtained was preserved for further use.

**Table1**Plants tested for antioxidant activity

Name	lame Family	
		No.
Alstonia bonei, cortex	Apocynaceae	105370
Alstonia bonei, folium	Apocynaceae	105356
Alstonia bonei, radix	Apocynaceae	105350
Byrsocarpus coccineus, folium	Connraceae	105362
Byrsocarpus coccineus, raidx	Connraceae	105361
Cnestis ferruginea, folium	Connraceae	105373
Cnestis ferruginea, lignum	Connraceae	105357
Cnmestis ferruginea, radix	Connraceae	105375
Crinum purpurascens, bulbus	Liliaceae	105340
Crinum purpurascens, folium	Liliaceae	105341
Funtumia elastical, folium	Apocynaceae	105369
Hedranthera batteri folium	Apocynaceae	10563
Icacina trichantha, folium	Icacinaceae	105359
Icacina trichantha, lignum	Icacinaceae	105357
Icacina trichantha, radix	Icacubaceae	105360
Landolfia owariensis, folium	Apocynaceae	105368
Leea guinensis, lignum	Leeceae	105346
Leea guinensis, radix	Leeceae	105358
Sphenocentrum jollyanum,	Menispermacea	105372
folium	е	
Sphenocentrum jollyanum,	Menispermacea	105074
radix	е	
Voacanga africana folium	Liliaceae	105341
Canarium(spp), cortex	Burseraceae	105387

#### **Procedure**

1 mg of each extract was weighed into a small test tube and 10mls of methanol added. The mixture was shaken together and by use of capillary was spotted carefully on the aluminum-coated plate.

5ml of each of the mixture was spotted on the coated Aluminum plate about 10mm away from the bottom of the plate. The point of the spot was clearly labeled and the plate allowed to dry in air and developed in a tank containing the mobile phase: (Ethyl acetate: fomic Acid: water 85: 15: 10:)

The above was allowed to dry and viewed in the UV light at 365 and 254nm. The efflorescent points were marked at the wavelength after which the slide was sprayed with Diphenyl-Picrylhydraxyl (DPPH) reagent in methanol (10mg in 10mls). After this, the plate was left to dry and the colouration produced on the plate was noted.

The DPPH reagent in this case was used to detect the presence of antioxidants. This reagent form complexes with the free hydroxyl group present in the crude extract. Examples of such compounds are the flavonoids, flavones etc. Thus DPPH on forming these complexes show the observed coloration (yellow coloration) on the TLC plate.

#### **RESULTS**

Table 2 shows the result of the anti-oxidant assay. All the medicinal plant materials used contain varying degrees of anti-oxidants. The colours of the efflorescence observed by the spots under the UV are a suggestion of the type of polyphenolic compounds present in the sample (Wagner 1996). Although phytochemical screening showed that some of the plant materials contain other metabolites e.g. alkaloids, the major components contained in the plant material under investigation are poly phenolic compounds as confirmed by the D.P.P.H. spray reagent.

#### **DISCUSSION**

Flavonoids are groups of naturally occurring compounds widely distributed, as secondary metabolites in the plant kingdom. These Flavonoids have also been reported to possess anti oxidant and anti radical properties (Nakayoma and Yamada 1995).

The DPPH test (Wagner, 1996) provided information on the reactivity of test compounds with a stable free radical. Because of its odd electron, 2, 2- diphenyl-picryl-hydrazyl radical (DPPH) gives a strong absorption band at 517nm in visible spectroscopy (deep violet colour). As the electron becomes paired off in the presence of a free radical scavenger, the absorption varnishes, thus the resulting decolorization is stoictiometric

with respect to the number of electrons taken up.

The scavenging properties of anti- oxidants are often associated with their ability to form stable radicals. Also, it is well known that aromatic compounds containing hydroxyl groups, especially those having ortho-di-or trihydroxyl functions can give rise to radical stable enough to be detected by ESR spectroscopy. The above is the general principle of the phytochemical screening for anti-oxidants.

TABLE 2:
Result of anti-oxidant assay

Result of anti-oxidant as		
Name of Medical plant	Result of antioxidant assay	Colour of efflorescent spot of the plant material
Alstonia bonei, cortex	++	
Alstonia bonei, folium	++	Bluish, yellow
Alstonia bonei, radix	++	
Byrsocarpus coccineus, folium	++	
Byrsocarpus coccineus, radix	++	
Canarium(spp), cortex	++++	Deep purple
Cnestis ferruginea, folium	++++	Yellow
Cnestis ferruginea, lignum	++++	Blue, yellow
Cnmestis ferruginea, radix	++++	Bluish yellow
Crinum purpurascens, bulbus	++	
Crinum purpurascens, folium	++	
Funtumia elastical, folium	+++	Purple
Hedranthera batteri folium	++	
Icacina trichantha, folium	++	
Icacina trichantha, lignum	++	
Icacina trichantha, radix	++	
Landolfia owariensis, folium	+++	Purple- bluish-
IOIIUIII		purple
Leea guinensis, lignum	++	
Leea guinensis, radix	++	
Sphenocentrum jollyanum, folium	+++	Orange, yellow
Sphenocentrum jollyanum, radix	++++	Orange, yellow
Voacanga africana folium	++++	Purple

+ + - antioxidant in low quantity; + ++- antioxidant in moderate quantity; ++++- antioxidant in large quantity.

These spots are isolated by the preparative low pressure chromatographic method can be used for future structural elucidation of these compounds by the help of spectrophotoscopic methods (Merbry et al, 1970). Diphenyl boryloxyl ethyl—amine is another regent similarly used in the laboratory to detect the presence of flavonoids in natural products. This method of screening also allows for a bio-assay guided study of natural products.

This study reveals the presence of antioxidant activity in varying degrees in all the plant materials under study. Bio-assay guided studies on all these medicinal plants and their structural elucidation of the active isolates are progressing in our laboratories.

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