CHARACTERIZATION OF METHICILLIN-RESISTANT STAPHYLOCOCCUS AUREUS ISOLATES FROM APPARENTLY HEALTHY INDIVIDUALS IN MALETE, KWARA STATE, NIGERIA


1Department of Biosciences and Biotechnology, College of Pure and Applied Sciences, Kwara State of University, Malete, Kwara State, Nigeria; 2Department of Biological Sciences, Faculty of Science, Kings University, Odeomu, Osun State, Nigeria

*Correspondence: Dr. Shola Kola Babatunde, Department of Biological Sciences, Faculty of Science, Kings University, Odeomu, Osun State, Nigeria. E-mail: sk.babatunde@kingsuniversity.edu.ng or sholakemibab@yahoo.com Phone: 0803503550: 08074390025

ABSTRACT

Background: Methicillin-resistant Staphylococcus aureus (MRSA) is a common and continuously growing cause of nosocomial and community-acquired staphylococcal infections around the world. Screening for colonization with MRSA is a major aspect of control and limiting the spread of infections caused by this organism. We investigated the carriage of MRSA among apparently healthy individuals in four rural villages: Eleburu, Tapa, Atere and Apo all around semi-urban town-Malete, in Moro Local Government of Kwara State, Nigeria.

Methods: Nasal swabs were collected from volunteered individuals and were cultured on mannitol salt agar and blood agar for isolation and identification of Staph aureus using standard microbiological techniques. Susceptibility to cefoxitin disc (30 µg) was used to determine MRSA status of the isolates. Molecular method was used to detect the gene responsible for resistance among MRSA isolates. Antimicrobial susceptibility test to commonly prescribed antibiotics was carried out using discs diffusion method.

Results: Total number of individuals carrying Staph aureus in their nostrils was 42 (37.2 %). Antibiotics susceptibility profile of Staph aureus isolates showed 100 % resistance to cefuroxime, cefotaxime, cloxacillin and augmentin, and were 87 %, 81 %, 69 % and 23.8 % and 19 % resistant to tetracycline, ceftriaxone, erythromycin, ofloxacin and gentamicin respectively. A total of 6 (100%) Community-acquired MRSA (CA-MRSA) isolates were recovered from individuals living in these villages. Molecular method detected nuc and mecA genes in all the 6 (100%) CA-MRSA isolates and lukS-lukF was detected in 3 (50%) of the isolates.

Conclusion: Detection of CA-MRSA strains among these rural dwellers indicates that they are harbouiring enhance virulence organism that may manifest a more severe disease condition. The danger associated with high prevalence of multidrug resistant Staph aureus and CA-MRSA and its consequential effects of poor drug administration in Nigeria was discussed. There is need to establish a more strict legislation and enforcement on drug control; and a body that would monitor production and appropriate use of antibiotics in the Nigeria.

KEYWORDS: CA-MRSA, Staph aureus, Antibiotics, Rural Villages and Molecular Characterization

CARACTÉRISATION DES ISOLATS DE STAPHYLOCOCCUS AUREUS RÉSISTANTS À LA MÉTHICILLINE PROVENANT DE PERSONNES APPAREMMENT SAINS À MALETE, ÉTAT DE KWARA (NIGÉRIA)


1 Département des biosciences et de la biotechnologie, Collège des sciences pures et appliquées, Université de l’État de Kwara, Malete, État de Kwara, Nigéria; 2Département des sciences biologiques, faculté des sciences, université Kings, Odeomu, État d’Osun, Nigéria

* Correspondance: Dr. Shola Kola Babatunde, Département des sciences biologiques, Faculté des sciences, Université Kings, Odeomu, État d’Osun, Nigéria. Adresse électronique: sk.babatunde@kingsuniversity.edu.ng ou sholakemibab@yahoo.com Téléphone: 0803503550: 08074390025

ABSTRACT

Contexte: Le Staphylococcus aureus résistant à la méthicillin (SARM) est une cause commune et en augmentation constante des infections staphylocoocciques nosocomiales et d’origine communautaire dans le monde. Le dépistage de la colonisation par le SARM est un aspect majeur du contrôle et de la limitation de la propagation des infections causées par cet organisme. Nous avons enquêté sur le transport de MRSA chez des individus apparemment en bonne santé dans quatre villages ruraux: Eleburu, Tapa, Atere et Apo tout autour d’une ville semi-urbaine-Malete, dans le gouvernement local de Moro dans l’État de Kwara, au Nigéria.

Méthodes: Des écouvillons nasaux ont été prélevés sur des individus volontaires et ont été cultivés sur gélose au sel de mannitol et sur gélose au sang en vue de l’isolement et de l’identification de Staph aureus à l’aide de techniques microbiologiques standard. La sensibilité à la céfoxitine (30 µg) a été utilisée pour déterminer l’état de SARM des isolats. La méthode moléculaire a été utilisée pour détecter le gène responsable de la résistance parmi les isolats de SARM. Un test de sensibilité aux antimicrobiens aux antibiotiques couramment prescrits a été réalisé à l’aide de la méthode de diffusion sur disques. Résultats: Le nombre total d’individus porteurs de Staph aureus dans leurs narines était de 42 (37,2%). Le profil de sensibilité aux antibiotiques couramment prescrits a été réalisé à l’aide de la méthode de diffusion sur disques.

Copyright ©2017 AJCEM. This work is licensed under the Creative Commons Attribution 4.0 International License CC-BY
INTRODUCTION

Methicillin-resistance *Staph aureus* (MRSA) refers to isolates of *Staph aureus* that are resistant to antibiotics with β-lactam ring such as penicillins, cephalosporins and their derivatives from healthy individuals or patients. *Staph aureus* is an opportunistic pathogen that asymptptomatically colonizing the nostrils, skin, glands and mucous membranes of both human and animal. It possesses a wide range of virulent factors and are responsible for a wide range of infections such as rashes, endovascular infections, inflammations of bones and the meninges, pneumonia, skin soft tissue infection (SSTIs), septic arthritis, endocarditis, sepsis, bloodstream infection and osteomyelitis [1], [2].

The first recorded emergence of methicillin resistance was in England in 1961 two years after introduction of methicillin [3]. MRSA organisms are also found to be resistant to other methicillin related antibiotics such as oxacillin, amoxicillin and penicillin. These organisms also acquire resistance to other groups of antibiotics that include clindamycin, cotrimoxazole, erythromycin and gentamicin [4]. The resistance is as a result of acquisition of resistant genes known as mec A gene which is a membrane-bound enzymes that catalyzes the transeptidation reaction and encodes the production of PBP-2a a penicillin-binding protein-2a which is essential for cross-linkage of peptidoglycan layer [5]. Lately, a new methicillin resistance gene known as mecC gene has been reported and MRSA carrying this gene has been isolated from both human and animal [6], [7], [8].

Hospital acquired MRSA (HA-MRSA) has always been associated with prior exposure to hospitals and healthcare centres, however, recently MRSA infection in healthy individuals in community is on increase and it has been confirmed to be responsible for morbidity and mortality in various parts of the world [9], [10], and is referred to as community acquired methicillin-resistant *Staph aureus* (CA-MRSA) [11], [12].

HA-MRSA strains are genetically distinct from CA-MRSA, CA-MRSA strains are always resistant to the B-lactam and sometimes to erythromycin but largely remain susceptible to several other antibiotics. Many outbreaks of CA-MRSA have been reported in healthy populations and institutions such as Daycare centers, Military quarters and prisons [13].

Malete, a semi-urban town is surrounded by four rural villages; all are in Moro Local Government and Kwara North Senatorial District of Kwara State of Nigeria. These villages: Apo, Eleburu, Tapa and Atere have population of less than 500 each [14]. The residents are mainly farmers and herders. There are no schools or healthcare facilities in the villages. The pupils attend primary and secondary schools at Malete and the sick ones are cared for at Cottage Hospital, Malete.

In Nigeria, there is paucity of information on MRSA, particularly CA-MRSA in rural and semi-urban populations with its antecedent danger of ignorance. In other to meet with this need, we investigated the carriage of CA-MRSA stains in the nostril of some apparently healthy individuals in rural communities of Malete, Kwara State. We also screened to detect genes responsible for MRSA in individuals that were positive to MRSA.

MATERIALS AND METHODS

Sample Collection

Visits were made to each village heads (Baale) and his elders in council and obtained consent to collect nasal swab from only volunteered members of the villages. Further approval to carry out this research was obtained from Kwara State University research and ethical committee under the Community Development Centre (CDC). All subjects that have been admitted into the Hospital or Healthcare centre and those that were on any antimicrobial treatment in the past three months prior to sample collection were excluded from study. Samples were collected with sterile cotton swab pre-wetted with sterile saline; the swab was inserted into the two anterior nares and gently rotated. Samples were collected randomly from 113 apparently healthy individuals in the four rural villages of Malete that included Apo, Eleburu, Atere and Tapa between first and third weeks of August, September and October, 2017.

Culturing method

Samples collected were transported in ice pack box within one hour of collection to the Microbiology laboratory, Department of Biosciences and Biotechnology, Kwara State University, Malete. The samples were cultured on mannitol salt agar and blood agar plates. They were incubated aerobically at 35-37 °C for 18-24 hours. On blood agar, colonies that were light yellow to whitish cream and were 1-2 mm in diameter with some producing β-haemolysis or colonies on mannitol salt agar that were pinkish-yellow and were 0.5-1 mm in diameter were picked for further
tests. Further tests carried out to confirm that the isolates were *Staph aureus* are Gram staining, catalase, coagulase and DNase tests as previously described [15], [16].

**Antimicrobial Susceptibility Test**

Antimicrobial susceptibility profile of each isolated *Staph aureus* was determined using standard disc diffusion method on Mueller-Hinton agar (Mast Diagnostics, Mast Group Ltd, Merseyside, UK) incubated aerobically at 37 °C for 18-24 hours using the following antibiotics discs; Cefuroxime (CXM₃₀), Cefotaxime (CF₃₀), Cloxacillin (CX₉), Augmentin (AU₃₀), Ceftriaxone (CRO₃₀), Tetracycline (TE₉), Erythromycin (E₁₅), Gentamicin (GEN₁₀) and Ofloxacin (OF₅). Zones of inhibition were measured in millimeters and interpreted following criteria of the Clinical and Laboratory Standard Institute [17].

**Detection of Methicillin Resistance**

Resistance to methicillin was determined using disc diffusion susceptibility method as described by guideline of the [18]. This was performed with Cefoxitin₃₀µg antibiotics disc (Oxoid, UK) on Mueller-Hinton agar plate (Mast Diagnostics, Mast Group Ltd, Merseyside, UK); where there are no zone of inhibition or zone radius measures 2 mm or less, isolates were regarded as resistant [18].

**Molecular Screening and Characterization**

The genomes of the isolates were extracted using QIAamp DNA Mini Kit according to manufacturer’s instructions. Methicillin resistant *Staph aureus* isolated that were phenotypically identified were subjected to molecular screening for the presence of muc gene [19] mecA gene, Panton valentine leucocidin (PVL) gene and lukS-lukF genes [19] using the appropriate primers as listed in Table 1. The PCR condition were initiated for denaturation at 95 °C for 2 minutes followed by 30 cycles of amplification with 94 °C for 30 minutes, annealing at 50 °C for 30 seconds, extension at 72 °C for 30 seconds and final extension at 72 °C for 4 minutes. The PCR products (10 µg each for muc, mec and PVL genes) were analyzed in 1.5% agarose gel. Electrophoresis was performed in TBE buffer at 180 volts for 1 hour, and gel was subsequently stained with ethidium bromide (10 mg/ml) (Sigma, UK). DNA bands were visualized using UV-illuminator and photographed.

**RESULTS**

**Distribution of Staph aureus among villages**

*Staph aureus* was recovered from the volunteered individuals from four rural villages studied. The total numbers of apparently healthy individuals carrying *Staph aureus* in their nostrils were 42 (37.2%) n=113; however the numbers recovered varied from one village (Table 2) to the other.

<table>
<thead>
<tr>
<th>Villages</th>
<th>Total Sample Collected</th>
<th>Staph aureus isolated</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Number</td>
<td>Percentage (%)</td>
</tr>
<tr>
<td>Apo</td>
<td>38</td>
<td>34.0</td>
</tr>
<tr>
<td>Eleburu</td>
<td>27</td>
<td>24.0</td>
</tr>
<tr>
<td>Tapa</td>
<td>23</td>
<td>20.0</td>
</tr>
<tr>
<td>Atere</td>
<td>25</td>
<td>22.0</td>
</tr>
<tr>
<td>Total</td>
<td>113</td>
<td>100</td>
</tr>
</tbody>
</table>

**TABLE 1: SEQUENCE OF PCR PRIMER SET FOR IDENTIFICATION OF STAPHYLOCOCCUS AUREUS, METHICILLIN RESISTANT GENES AND PVL GENE**

<table>
<thead>
<tr>
<th>Target genes</th>
<th>primer set</th>
<th>Primer sequence (5'→3')</th>
<th>Amplicon Size (bp)</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Staphylococcus aureus</em></td>
<td>Nuc</td>
<td>F-TCAGCAAATGCACTACAAACAG</td>
<td>200bp</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R-CGTAATGCACTTGCCTCAAG</td>
<td></td>
</tr>
<tr>
<td>Methicillin resistance</td>
<td>Mec A</td>
<td>F-AAAAATCGATGTAAAGTTGTCGCC</td>
<td>500</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R-AGTTCCTGACATGGAGATTGCC</td>
<td></td>
</tr>
<tr>
<td>Panton valentine</td>
<td>lukS-lukF</td>
<td>F-ATCATTAGTTAAAATGTCGACATGATCCA</td>
<td>433</td>
</tr>
<tr>
<td></td>
<td></td>
<td>R-GCATCAATGTTATGGATAGCAAAGC</td>
<td></td>
</tr>
</tbody>
</table>
Resistance Profile of CA-MRSA isolates to antibiotics

The percentage distribution of CA-MRSA showed that 42 (100 %) were resistance to cefuroxime, cefotaxime, cloxacillin and augmentin, while 34 (80.9 %) were resistant to ceftriaxone, 35 (87.5 %) are resistant to tetracycline, 29 (69.1 %) were resistant to erythromycin, 8 (19.1 %) were resistant to gentamicin and 10 (23.8 %) were resistant to ofloxacin.

Molecular analysis of the isolated MRSA

Molecular analysis on the 6 isolates that were methicillin resistant Staph aureus showed that muc and mec A genes were detected in all the 6 isolates (Plates 1; 2). While only 3 of the six Methicillin resistant isolates had lukS-lukF (Plate 3) and only 3 of the six methicillin resistant Staph aureus isolates had PVL gene detected (Plates 3).

---

PLATE 1: PCR PRODUCTS AFTER GEL ELECTROPHORESIS SHOWING AMPLICONS DETECTION OF THE MUC GENES OF S. AUREUS ISOLATES.

PLATE 2: PCR PRODUCTS AFTER GEL ELECTROPHORESIS SHOWING AMPLICONS FOR MEC A (535BP). DNA MOLECULAR WEIGHT MARKER (M) (535BP)
DISCUSSION

Community acquired methicillin-resistant *Staphylococcus aureus* (CA-MRSA) has emerged as an important human pathogen in various parts of the world. This pathogen has varied distribution in both urban and rural community settings. In this study, we reported a prevalence of 37.2 % of *Staph aureus* in nostrils of apparently healthy individuals in rural communities of Malete. This prevalence is lower than a prevalence of 56.3 % reported by Chibuike and colleagues [20] in rural communities of Ururu among healthy individuals. Also a lower prevalence of 36.5 and 13.9% were reported by Egwuata et al. [21] in health workers in Lagos and Okwu et al. [22] among healthy individuals in Okada, Edo State Nigeria respectively. Various authors have recommended eradication of nasal carriage of *Staph aureus* to reduce transmission to others with the use of systemic antimicrobials, normal bacterial flora augmentation, antiseptic washes and topical antimicrobials [23], [24].

Antimicrobial resistance is one of the major threat posed by microorganisms to infection management in this twenty first century and *Staph aureus* has always been one of the major pathogen that possess ability to develop resistance to newly developed antimicrobial agents [25]. In this study, *Staph aureus* isolates were resistant to commonly available and recommended antibiotics such as cefuroxime, cefotaxime, cloxacillin and augmentin, but susceptible to ceftriaxone, tetracycline, gentamicin and ofloxacin in varying degrees. Susceptibility of erythromycin ranges from 5 to 64% in different geographic areas [26], while the susceptibility in this study is 30.9 %. There are many factors that may be adduced to the high resistance of this organism to antibiotics in these rural communities. Such factors included self-medication, availability and use of antibiotics without Doctor’s prescription, irrational consumption rate of antibiotics, noncompliance to prescription and sales of fake or substandard drugs and transmission of resistant strains between individuals within the community.

Methicillin resistance is mediated among *Staph aureus* by the penicillin-binding protein (PBP)- 2a encoded by meca gene. It was reported that all β-lactam antibiotics have poor affinity when PBP is altered, and such microbe would not be killed when exposed to therapeutic concentration. The meca gene is found to be located on a mobile genetic element of staphylococcal chromosome cassette (SCCmec) [27]. In this study, the prevalence of methicillin resistance *Staph aureus* was 14.3 %. This is prevalence is lower when compared with what other researchers reported among apparently healthy individuals. Prevalence of 22.0, 47.6 and 60.6 % were reported by Akerele et al. [28] in Ekosundun, Benin City; Adelowo et al. [29] in Maiduguri and Chibuike et al. [21] in Uturu rural community respectively.

The major objective of screening for meca gene is to compare results of antibiotic susceptibility by disc diffusion method with gene analysis results in *Staph aureus* isolates. The phenotypic expression of antibiotic resistant genes has been reported to be affected by different conditions such as the incubation temperature and time, medium inoculated, inoculum size, test agents, pH, and ionic strength of NaCl [30]. The meca gene may be heterogeneously expressed and, therefore, all MRSA strains may not be detectable with phenotypical methods and the test require at least 24 h for evaluation of the results. However, the detection of meca gene by PCR techniques is considered the gold standard method. All the six isolates that were cefoxitin resistant were also positive for this gene detection method;
confirming that they are all methicillin-resistant *Staph\ aureus*.

The present study showed strong correlation between genotypic and phenotypic analysis, the results were consistent with previous studies which showed a perfect and strong correlation between the results obtained by the phenotypic antibiotic resistance determination and PCR-based assays [31], [32]. The critical parameters for success of a PCR-based assay for the detection of multidrug resistant bacteria like MRSA are the reliability, accuracy, fast and sensitivity and results were obtained within hours [33]. However the cefoxitin disc has been shown to have specificity 97-100 % to detected mec A gene [34].

Previous reports on MRSA in North-central, Nigeria were based phenotypic method, to our best knowledge this is the first report based on the detection of mec A gene in CA-MRSA among healthy individuals in rural communities of North-central, Nigeria. However, mec A gene has been discovered from other different part of Nigeria such reports included, a study in Benin city, Nigeria, 4 isolates representing 11 % were confirmed to carry mec A gene using molecular technique [35], Esan et al. [36] confirmed only one MRSA isolate from health care institutions from Ekiti and Ondo states and Shittu et al. [37], detected two MRSA isolates with mecA gene were detected in Ile-Ife, one from Lagos and two from Ibadan (all in South-western Nigeria)

In the North-eastern Nigeria, five MRSA isolates with mecA gene were detected in Maiduguri by Okon et al. [38] reported the detection of 12.5 % MRSA from clinical specimens from six tertiary hospitals.

PCR result for the detection PVL (lukS-lukF) gene in this research showed that out of 6 MRSA isolates screened only 3 of them were PVL positive, representing a percentage of 50 % (n=6). This finding was higher compared to the report of Sani et al. [39] who detected PVL gene prevalence of 3.25 and 8.13 % amongst MRSA from hospitalized and non-hospitalized patients in Benin. However, this difference may be due to the size of our samples. Another report by Breurec et al. [40] detected a 57 % lukS-lukF genes encoding Panton-Valentine Leukocidin (PVL) prevalent in MRSA isolates from Cameroon, Niger, and Senegal (West and Central Africa).

It appears that misuse and overuse of these antibiotics could have contributed to increase in antibiotics resistance in these rural communities as it has been observed in other part of the world. Therefore, to prevent resistance and treatment failures, we advocate proper monitoring of antibiotics usage and collection of data on antibiotic susceptibility testing pattern, public enlightenment on the effectiveness of various antibiotics and the enactment of drug policies in Nigeria; are urgently needed to move in line with the global trends.

Multidrug resistant organisms are defined as organisms that have acquired non-susceptibility to at least one agent in three or more classes of antimicrobial [41], [42], [43], [44]. By this definition, all the six CA-MRSA isolated in this community are multidrug resistant *Staph\ aureus*.

### Conclusion:

The detection of methicillin-resistant *Staph\ aureus* carriage among individuals in these remote rural communities of Malete indicated the extensive distribution of this organism and also indicated that the abuse of drug is on high side in Nigeria. Unlicensed patent medicine dealer and drug hawkers may be the main source of drugs distribution to these villages.

### Acknowledgment:

We acknowledged the cooperation of members of Apo, Eleburu, Atare and Tapa communities, Director of Community Development Centre (CDC), Kwara State University, Malete and technical staff of Microbiology laboratory for their support.

### REFERENCES

1. Aklilu E, Zunita Z, Hassan L, Chen HC. Phenotypic and genotypic characterization of methicillin-resistant *Staphylococcus aureus* (MRSA) isolated from dogs and cats at University Veterinary Hospital, Universiti Putra Malaysia. Tropical Biomedical 2010; 27(3): 483-492.


29. Adelowo KA, Okon K O, Denue BA, Ladan J, Tahir F, Uba A. Methicillin-Resistant Staphylococcus Aureus (MRSA) Colonisation level among Patients seen at a


43. CDC-Centers for Disease Control and Prevention (2013). 1600 Clifton Rd. Atlanta, GA 30333, USA 800-4-CDC-INFO (800-232-4636) TTY 2013; (888) 232-6348.