



Research Paper

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ACUTE AND SUBACUTE TOXICITY OF *ASPILIA AFRICANA* LEAVES

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Abstract

This study was designed to evaluate the toxicity of the aqueous extract of *Aspilia africana* leaves. Oral doses of 500 mg/kg and 1000 mg/kg were administered for 28 days to rats after every 2 days for sub-acute toxicity. For acute toxicity, 5 doses of 2, 4, 8, 12 and 16g/Kg body weight were investigated in mice. The control groups consisted of mice or rats administered with distilled water. The signs of toxicity fluctuated lightly from one mammal to another throughout the experiment. The liver, kidneys and heart weight of rats revealed no significant differences between the test groups and the control. The results indicated that the medium lethal dose (LD₅₀) was found to be greater in females than males with an average of 6.6g/Kg body weight for both sexes. Regardless of the significant differences observed at certain points in some biochemical parameters (ALT, AST, ALP, Creatinine and Glutathione); none showed any linear dose responsiveness. On the other hand, most of the parameters investigated were found to be gender dependent. These results suggested that *A Africana* can be classified among substances with low toxicity.

Key words: *Aspilia africana*, Asteraceae, toxicities, dose responsiveness.

Introduction

Natural products have been, and have remained, the cornerstone of health care. Present estimates show that, 80% of the world's population still rely on traditional medicine for their health care needs (Farnsworth et al., 1985). Unfortunately, most of those who use these plants in our society have not undergone adequate training. Therefore, in order to have standard natural plant products, preliminary studies have to be done in order to evaluate possible risks such as, undesirable effects, overdose or poisoning. *Aspilia africana* (Pers.) C.D. Adams is not an exception. It is a perennial herb which belongs to the Asteraceae family.

Aspilia africana is widely used in African folk medicine to stop bleeding, remove corneal opacities, induce delivery and in the treatment of anaemia and various stomachs complains (Iwu, 1993; Adjanohoun et al., 1996). Phytochemical studies revealed the presence of saponines and tannins as the most abundant compounds in the plant while flavonoids were the least (Obadoni and Ochuko, 1998). Other studies showed that essential oils from the leaves of *Aspilia africana* were rich in sesquiterpenes and monoterpenes. Also, the presence of precocene1 was found (Kuate et al., 1999). The medicinal plant contained ascorbic acid riboflavin thiamine, and niacinm. These herbs are good sources of minerals such as Ca, P, K,Mg, Na, Fe and Zn (Okwu and Josiah, 2006). Despite the popular use of this plant, no toxicological study has been reported.

The present study was undertaken to determine the toxicity of the aqueous extract of *Aspilia africana* leaves.

Materials and Methods

Plant material

Fresh leaves of *Aspilia africana* were collected from Simbock, a village in neighbourhood of the capital city of Cameroon. The sample was identified at the National Herbarium where some specimen was already available with the number 6555/SRF/Cam.

Preparation of the extract

The leaves collected were dried in an air-circulating oven at $38^{\circ}\text{C} \pm 2^{\circ}\text{C}$. The dry material (300 g) was macerated in 6 liters of distilled water for 48 h at 4°C in a refrigerator. The extract was sieved and the juice was filtered using whatman N°1 filter paper. The filtrate was put in a stainless-steel tray, and concentrated in an air-circulating oven at 42°C until total dryness. The gummy extract was put into small glass dishes and stored at 28°C in an incubator for further studies.

Bioassay

Young male and female *Wistar* albino rats (90-110 g) and *Swiss* albino mice (12-25 g) were bred at the Department of Biochemistry, in the breeding house of the Microbiology and Pharmacology Laboratory of the Faculty of Science. They were all clinically healthy and were kept under standard environmental conditions of temperature ($27^{\circ}\text{C} \pm 2^{\circ}\text{C}$). The animals had free access to water and standard diet. The principles of laboratory animal care were approved by the Department's ethical committee.

Acute toxicity

The bioassay was conducted according to the World Health Organisations guideline for the evaluation of the safety and efficiency of herbal medicines (O.M.S., 2000). For the study, *Swiss* albino mice were divided into six groups of 10 animals each (5 males and 5 females). Animals were deprived of food but not water (16-18 h) prior to administration of the extract. Five groups were given single oral doses of 2, 4, 8, 12 and 16 g/kg of the aqueous extract. The last group used as control received distilled water per os in the experiment.

Observations were made and recorded systematically as 1, 2, 4 and 24 h after substance administration. The visual observations included motility, respirations, sensitivity to sound and pinch, smelling of food and faeces consistence. The numbers of survivors were recorded after 24 h and the animals were observed daily for the next 7 days. The LD_{50} was determined based on Behrens and Kaber (1983), and Schorderet (1992) methods.

Sub-acute toxicity

Three groups of 10 rats each (5 males and 5 females) were given intra-gastric intubation of 500 mg/kg and 1000 mg/kg of the aqueous extract or distilled water (for control) every 48 h for 26 days. Food and water intake as well as body weight were monitored during the period of administration. After 26 days, all surviving animals were allowed to fast overnight and sacrificed by decapitation after anaesthetising with petroleum ether. Blood samples were collected from these mammals into heparinized tubes for haematological analyses and non heparinized centrifuge tubes. The liver, kidneys and heart were collected and weighed. Part of the liver or kidney tissues was also washed and kept in a freezer (-20°C) for further analysis of biochemical parameters. Another part was preserved in 10% formalin for histopathological studies.

Biochemical analysis

The blood collected into non heparinized tubes was centrifuged at 3000 rpm for 10 min. The serum was separated and liver and kidney homogenates (20%) were analysed for enzymes. Alanine aminotransferase (ALT) and Aspartate aminotransferase (AST) were assayed using the Reitman and Frankel (1957) method; Alkaline phosphatase (ALP) was analysed by the method of Bessey *et al.*, 1946; Protein by the method of Gornall *et al.*, 1949; Creatinine and Glutathione by Barterls *et al.*, (1972) and Ellman (1959) methods respectively.

Haematological studies

The blood samples that were collected into heparinized tubes were used for the estimation of white blood cells (WBC), red blood cells (RBC) and platelets by visual methods (Dacie, 1991).

Histopathological studies

Histopathological analyses of the liver and kidneys were done according to the conventional haematoxylin-eosin technique.

Statistical Analysis

Statistical analysis was done using ANOVA. The Duncan's test was used to locate significant differences between means'. Significant differences treatments were accepted at $P < 0.05$. Data were expressed as means \pm standard deviation.

Results and Discussions

In acute toxicity, a slight but non significant increase in body weight was registered for mice treated with the aqueous extract of *Aspilia africana* at the dose 2 g/kg (Figure 1). This increase in body weight, which is higher than that of the control group, may be due to stimulation of appetite by the extract, leading to increased food consumption and the body weight observed. Behavioural changes in the mice were linear dose responsiveness. These changes included increased aggressiveness and motility. The changes greatly reduced within 48 h without completely turning to normal and this continued throughout the experimental period with all the animals that survived. This enabled us to suggest that there was an irritating compound in the aqueous extract.

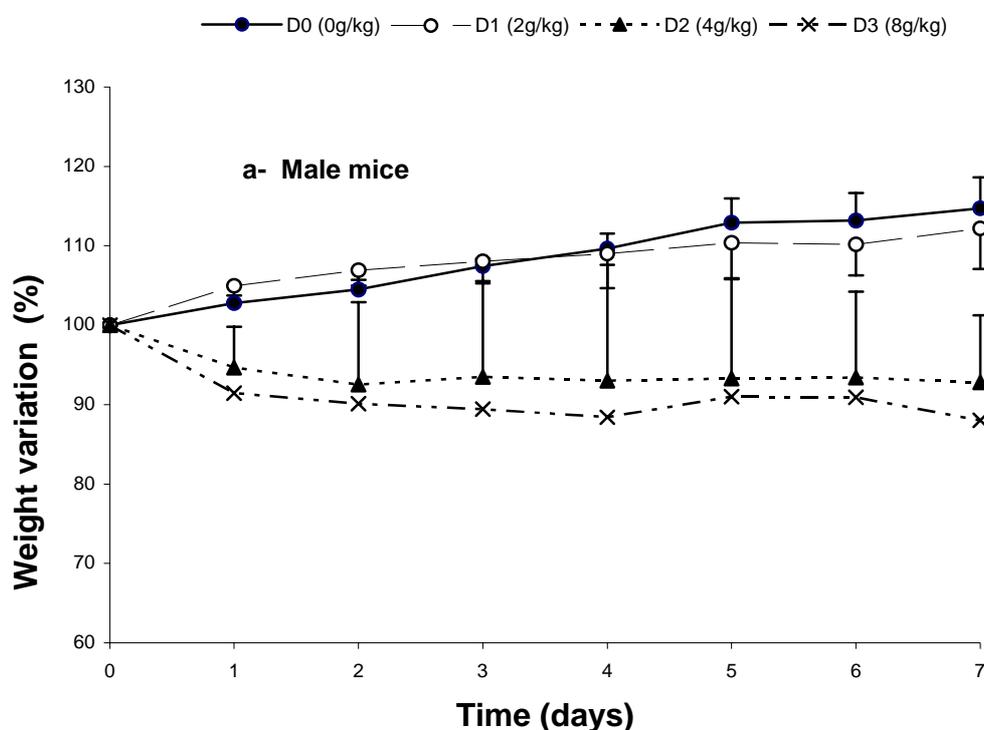


Figure 1a: Body weights of male mice in acute toxicity of aqueous extract of *Aspilia africana* leaves.

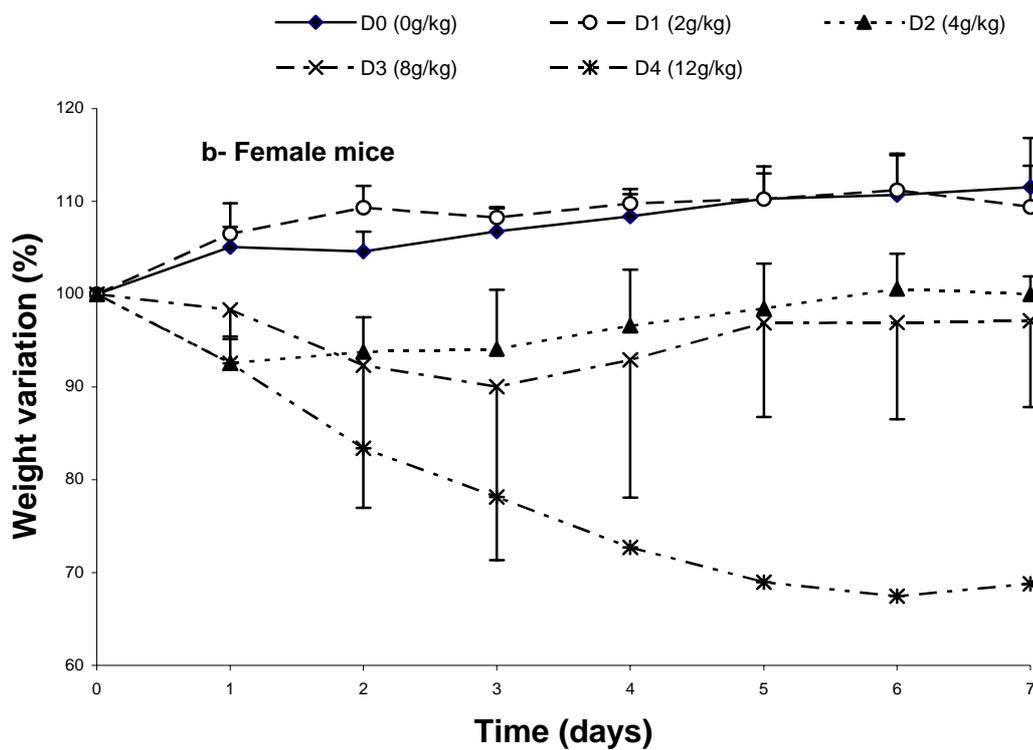


Figure 1b: Body weights of female mice in acute toxicity of aqueous extract of *Aspidia africana* leaves

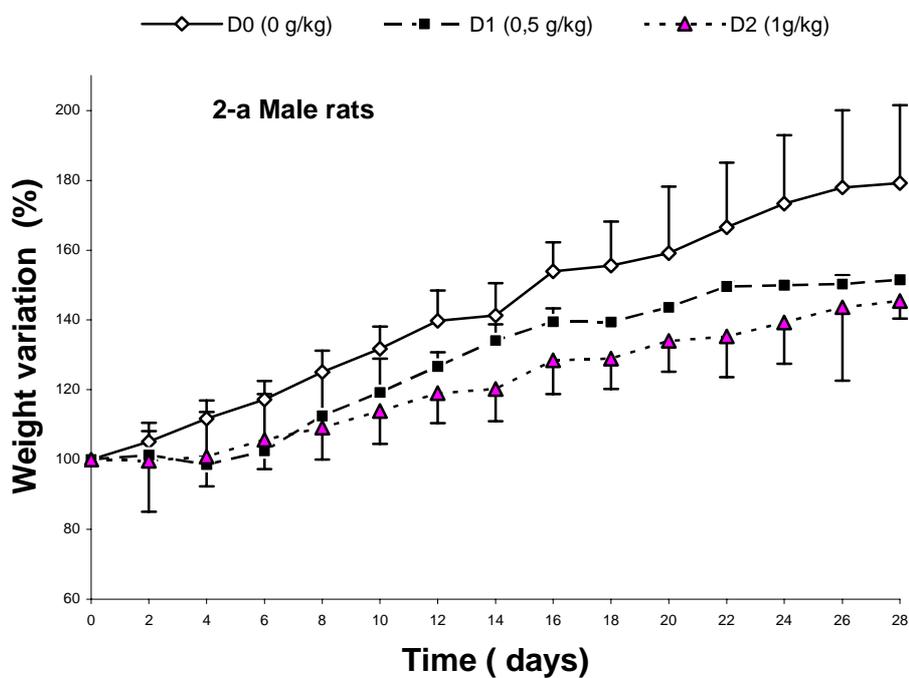


Figure 2a: Body weights of male rats in subacute toxicity of aqueous extract of *Aspidia africana* leaves.

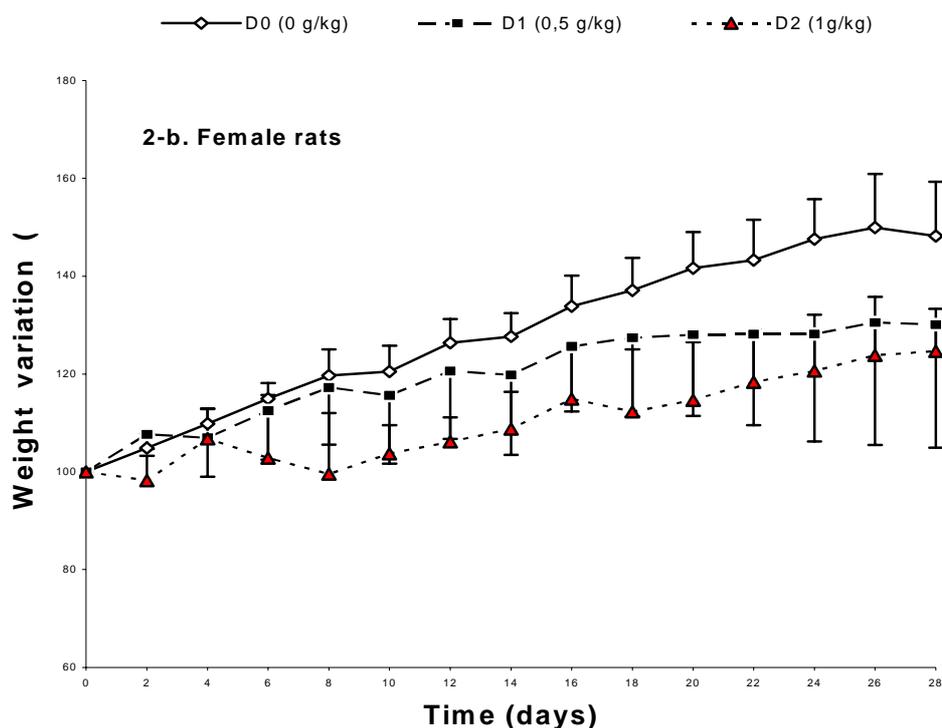


Figure 2b: Body weights of female rats in subacute toxicity of aqueous extract of *Aspilia africana* leaves.

The medium lethal dose value (LD_{50}) was 6.1 g/kg and 7.5 g/kg body weight for males and females respectively with an average of 6.6 g/kg body weight. According to Schorderet (1992), substances with LD_{50} values greater than 5000 mg/kg body weight are classified as substances with low toxicity. Thus, the aqueous extract of *Aspilia africana* can be considered as a substance with low toxicity. Furthermore, all the male mice died when treated with 12 g/kg body weight dose while the female mice treated with 16 g/kg body weight produced the same result. This indicates that the female mice are less sensitive to toxicity than the males. This may be due to the hormonal status which is different in males and females. Similar results were obtained by Solomon *et al.*, (1993) when studying the ethanolic extract of the roots of *Plumbago rosea* (Plumbaginaceae).

In sub-acute toxicity, the behavioural changes (aggressiveness) observed in acute toxicity were noted with rats during the experiment. As shown in Table 1, there were no significant variations in the weight of the organs of treated animals compared to the control. In the animals tested with the extract there was a slight and time dependent decrease in body weight compared to the control group. This decrease in body weight could be due to reduced appetite or impairment of some nutrients as a result of the extract.

Table 1: weights of organ in subacute toxicity of *Wistar* albino rats given the aqueous extract of *Aspilia africana* leaves (g/100g of body weight)

	D0		D1		D2	
	M	F	M	F	M	F
Liver (g)	4.452 ^a (0.080)	3.816 ^a (0.034)	3.989 ^a (0.079)	4.141 ^a (0.218)	4.377 ^a (0.173)	3.760 ^a (0.046)
Heart (g)	0.395 ^a (0.046)	0.408 ^a (0.118)	0.436 ^a (0.048)	0.445 ^a (0.095)	0.384 ^a (0.076)	0.422 ^a (0.104)
Kidney (g)	0.612 ^a (0.099)	0.613 ^a (0.171)	0.727 ^a (0.148)	0.730 ^a (0.279)	0.740 ^a (0.361)	0.656 ^a (0.040)

Means with the same letter superscript within the same row are not significant ($p < 0.05$) different. Values in bracket represent the Standard Deviation ($n = 5$). M (males), F (females).

Table 2: Haematological values in subacute toxicity of *Wistar* albino rats given the aqueous extract from *Aspilia africana* leaves (quantities / ml)

Test	D0		D1		D2	
	M	F	M	F	M	F
RBC (10 ⁶)	12.20 ^a (0.746)	12.36 ^a (0.684)	13.80 ^a (0.740)	12.75 ^a (0.608)	11.25 ^a (0.552)	16.12 ^b (1.278)
WBC (10 ⁶)	14.72 ^a (0.656)	16.76 ^{ab} (0.110)	16.20 ^a (0.440)	17.30 ^b (0.828)	13.40 ^a (1.344)	14.12 ^a (0.904)
PL (10 ³)	0.96 ^a (0.176)	1.16 ^a (0.182)	1.90 ^b (0.174)	2.00 ^b (0.204)	0.80 ^a (0.166)	1.52 ^{ab} (0.192)

Means with the same letter superscript within the same row are not significantly ($p < 0.05$) different. Values in bracket represent the Standard Deviation (n=25)

RBC: red blood cells, WBC: white blood cells, PL: platelets, M: males, F: females,

D0 : control (H₂O) D1 : 500 mg/kg, D2 : 1000 mg/kg

Table 3: Blood chemistry values of *Wistar* albino rats given the aqueous extract of *Aspilia africana* leaves in subacute toxicity.

	Parameters		Control (H ₂ O)	500 mg	1000 mg
Females	AST (U/L)	L	380.14 ^a (0.92)	381.44 ^a (1.70)	380.05 ^a (0.22)
		S	383.52 ^a (0.93)	385.01 ^a (1.21)	385.35 ^a (1.78)
	ALT (U/L)	L	390.07 ^a (0.14)	389.10 ^a (0.39)	389.27 ^a (1.01)
		S	388.01 ^a (8.78)	389.40 ^{ab} (1.86)	387.82 ^b (0.15)
	ALP (U/L)	L	106.34 ^a (9.50)	112.70 ^a (23.68)	123.96 ^a (20.52)
		S	298.30 ^a (34.11)	294.04 ^a (39.27)	315.65 ^a (19.17)
	Glutathione (mmol/mg of liver)	L	4.16 ^a (1.29)	6.61 ^{ab} (1.53)	8.48 ^b (2.36)
Creatinine (mg/dl)	K	0.66 ^a (0.08)	0.75 ^{ab} (0.017)	0.83 ^b (0.013)	
	S	0.80 ^a (0.08)	0.96 ^a (0.73)	1.05 ^a (0.070)	
Proteins (mg/ml)	L	3.72 ^a (0.87)	3.85 ^a (0.45)	3.76 ^a (0.47)	
	S	10.22 ^a (1.54)	10.23 ^a (1.33)	10.44 ^a (1.58)	
Males	AST (U/L)	L	377.38 ^a (0.98)	379.70 ^{ab} (2.26)	381.82 ^b (2.45)
		S	387.56 ^a (3.48)	389.07 ^a (2.64)	385.77 ^a (5.45)
	ALT (U/L)	L	389.72 ^a (0.39)	389.09 ^{ab} (0.71)	388.70 ^b (0.65)
		S	388.22 ^a (0.27)	388.56 ^a (0.82)	390.09 ^b (1.16)
	ALP (U/L)	L	134.98 ^a (29.47)	127.46 ^a (6.37)	145.84 ^a (20.10)
		S	337.18 ^a (6.83)	338.66 ^a (1.22)	336.96 ^a (2.19)
	Glutathione (mmol/mg of liver)	L	3.99 ^a (1.00)	7.19 ^b (1.23)	8.23 ^b (1.17)
Creatinine (mg/dl)	K	0.55 ^a (0.026)	0.59 ^a (0.025)	0.94 ^a (0.043)	
	S	0.76 ^a (0.023)	0.73 ^a (0.028)	1.32 ^a (0.072)	
Proteins (mg/ml)	L	3.31 ^a (0.62)	3.30 ^a (0.23)	3.49 ^a (0.41)	
	S	15.83 ^b (2.67)	14.34 ^{ab} (4.59)	9.48 ^a (0.93)	

Means with the same letter superscript within the same row are not significantly ($P < 0.05$) different. Values in brackets show the standard Deviation (n=5). S: serum, K: kidney, L: liver

The haematological status of the rats, after 28 days of oral administration of the aqueous extract of *Aspilia africana* is shown in Table 2. Compared to the control, an increase in values of all the parameters studied was registered with the dose of 500 mg/kg body weight. This increase was statistically significant ($P < 0.05$) for males as far as the platelets are concerned. An increase in platelet numbers led to: chronic myeloproliferative diseases, carcinoma, chronic inflammatory diseases, haemorrhage, sickle cell diseases associated with a non-functioning spleen or after splenectomy, iron deficiency anaemia associated with active bleeding (Cheesbrough, 2001). In females, a significant ($P < 0.05$) increase was noted with WBC and platelets at the dose of 500 mg/kg body weight. The main causes of increased WBC count are: metabolic disorders, poisoning, acute haemorrhage, leukaemia and myeloproliferative disorders, stress, menstruation and strenuous exercises (Cheesbrough, 2001). At the dose of 1g/kg, a decrease in number of WBC, RBC and platelets was registered for males compared to the control at the dose of 500 mg/kg. The decrease was not significantly different compared to the control. There was a significant difference ($P < 0.05$) compared to that of the 500 mg dose for platelets only. Causes of raised level of erythropoietin can occur in renal disease, cyanotic heart disease, shock and acute alcohol poisoning (Cheesbrough, 2001). In females, a significant ($P < 0.05$) increase was noted for platelets and WBC, compared to the control, at 500 mg/kg body weight.

For biochemical parameters (Table 3), no significant variations in ALP activity were noted. A significant increase in AST activity was noted in the liver at a dose of 1 g/kg in males only. For serum ALT, a significant ($P < 0.05$) increase was registered in males. Although AST and ALT are common liver enzymes because of their high concentrations in hepatocytes, only ALT is remarkably specific for the liver function since AST is mostly present in the myocardium, skeletal muscle, brain and kidneys (Sacher and McPherson, 1991, Witthawasku *et al.*, 2003). Despite the increased level of creatinine observed in the kidneys and serum in both sexes of all the animals with respect to the control groups, a significant increase ($P < 0.05$) was registered only in females. According to Harper (1971), an increase in creatinine level can be observed in some kidney illnesses, due to loss of its normal excretive function of creatinine, when there is muscular cells damage or following an incompatible medication interfering with normal functioning of the kidneys. The level of glutathione significantly ($P < 0.05$) increased at the dose of 500 mg/kg in males only, and at 1000mg/kg body weight the dose in males and females. The high concentration of glutathione observed may be due to the effectors effects of some components present in the extract.

A histopathological examination of the liver and kidneys showed the presence of necrosis, oedema and inflammatory infiltrations.

Conclusion

From the results obtained, we can say that, although aqueous extract of *Aspilia africana* leaves can be classified among substances with low toxicity, dosages of 500mg/kg body-weight or more, can be toxic for a long term treatment when taken orally.

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