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CHEMICAL COMPOSITION, CYTOTOXIC ACTIVITY AND ANTIMICROBIAL ACTIVITY OF ESSENTIAL OILS OF LEAVES AND BERRIES OF *JUNIPERUS PHOENICEA* L. GROWN IN EGYPT.

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Abstract

Hydrodistillation of berries and leaves of *Juniperus phoenicea* grown in Sinai yielded volatile oils in the yield of 0.36 and 1.96%, respectively. Using gas chromatography/mass spectrometry technique, fifty eight compounds were identified in berry oil representing 99.2% of the oil composition. α -Pinene was the major compound in berry oil (39.30%) followed by sabinene (24.29%). Berry oil composed mainly of monoterpenoids which amounted to 90.53%, of which 72.85% was monoterpene hydrocarbons. The sesquiterpenoids accounted for about 8% of the total oil composition. Leaf oil was composed of about 66 compounds representing 99.16% of the total composition of the oil. α -Pinene was the major constituent of leaf oil at concentration of 38.22%, followed by α -cedrol (31.23%). The monoterpene hydrocarbon was the predominant chemical group (41.29%) followed by the oxygenated sesquiterpenes (32.21%). Both oils showed very high cytotoxic activities against all cell line tested. They showed equal activities against brain (0.6 μ g/ml) and cervix (5.0 μ g/ml) human cell lines, while berry oil was slightly more active than leaf oil against lung (0.6 and 0.7 μ /ml, respectively), liver (0.7 and 0.9 μ g/ml, respectively) and breast human cell lines (0.8 and 1. μ g/ml, respectively). The antimicrobial activity and minimum inhibitory concentration (MIC) of leaf and berry oils were also determined. The oils showed high activity against most of the tested strains.

Key words: *Juniperus phoenicea*, Cupressaceae, essential oils, berries, leaves, antimicrobial, cytotoxic, GC-MS analysis.

Introduction

The most famous of the oils of junipers is *Juniperus communis* L. They are used in aromatherapy, through inhalation, massage, bathing, or ingestion to create good health and beauty. Oil of juniper, distilled from the wood and leaves of several species, is used in perfumes. It is noted by German authorities that the tea is diuretic and urinary antiseptic (Hagar, 1979). The diuretic activity is thought to be largely due to the tea's content of terpinen-4-ol, a non irritating terpene. Juniper berry tea is listed in the German Pharmacopoeia as a digestive aid, both stimulating appetite as well as relieving flatulence. The oils also help increase the flow of digestive fluids, improve digestion and eliminate gas and stomach cramping (Uphof, 1968). Juniper has been used for centuries as a steam inhalant for bronchitis, and to control arthritis. The oil is also irritating to microbes, so much so that it kills many of them (Watt and Breyer-Brandwijk, 1962; Stassi *et al.*, 1996). Traditional formulas are in combination with ginger and dong quai or with goldenseal or with *Uva ursi*. Juniper berry is probably best known as the unique flavouring agent of gin, an important component of the dry martini, a popular intoxicant and a putative calmative revered by western culture for over 300 years (Duke, 1980).

J. phoenicea L. (Fam. Cupressaceae) is the species found in Egypt. It is grown in Sinai (Yelleg, Halal and Maghara mountains) in rocky ridges. It extends to Central Arabia (Boulos, 1999). The leaf essential oil of *J. phoenicea* has been reported in varying details from Saudi Arabia (Dawidar *et al.*, 1991), France (Tabacik and LaPorte, 1971; Tabacik and Poisson, 1971) and from Greece and Spain (Adams *et al.*, 1996). Also, there are some reports on the analysis of fruit essential oils (Ramic and Murko, 1983; De Pascual *et al.*, 1981; Delitala, 1980). Only one report dealt with composition of leaf and berry essential oils of *J. phoenicea* grown in Egypt (Afifi *et al.*, 1992) in which 18 compounds in both oils were identified. In this paper, we report on the chemical

composition of the essential oils of leaves and berries of *J. phoenicea* L grown in Egypt and their cytotoxic and antimicrobial activities.

Experimental

Plant materials

J. phoeniceae leaves and berries were collected from Sinai, Egypt from fully grown plants when berries were at the ripening stage. They were identified in the Herbarium Department, Faculty of Sciences, Cairo University. A voucher specimen (voucher number J-3) was deposited in Pharmacognosy department, National Research Centre, Cairo.

Distillation of the volatile oils

The volatile oils of the dry leaves and berries were prepared by hydrodistillation for three hours according to Guenther (1961). The prepared volatile oils were dehydrated over anhydrous sodium sulphate and stored in refrigerator until analyzed.

Gas chromatography-Mass spectrometry

A Trace GC 2000 (produced by Thermo) was used. The Mass Spectrometer model SSQ 7000 produced by Finnigan equipped with library software Wiley 138 and NBS 75 was used. Capillary DB5 (Methylpolysiloxane containing 5% phenyl groups) column 25m X 0.25 mm i.d., Temp. program was; 2 min. at 60° C, 60-100° C (2° C/min) and 100-250° C (5° C /min). Carrier gas was helium at flow rate of 1.0 ml/min. Injection voltage 70 eV.; scan mass range 40-450.

Identification of compounds

Compounds were identified by matching their mass spectra with those recorded in the MS library and further confirmed by injecting the authentic samples of different available compounds with the volatile oil and by comparing the mass spectra with those of reference compounds or with the published data (Adams, 1995).

Measurement of potential cytotoxicity by SRB assay

Potential cytotoxicity of leaf and berry essential oils was tested using the method of Skehan and Storeng (1990). Cells were plated in 96-multiwell plate (10^4 cell/well) for 24 hours before treatment with the oils to allow attachment of cell to the wall of the plate. Different concentrations of the oils (0, 1, 2.5, 5 and 10 µg/ml) were added to the cell monolayer. Triplicate wells were prepared for each individual dose. Monolayer cells were incubated with the oils for 48 hours at 37° C and in atmosphere of 5% CO₂. After 48 h, cells were fixed, washed and stained with sulforhodamine B stain. Excess stain was washed with acetic acid and attached stain was recovered with Tris EDTA buffer. Color intensity was measured in an ELISA reader. The relation between surviving fraction and drug concentration was plotted to get the survival curve of the tumor cell line after the specified compound.

Antimicrobial activities:

Microbial strains

The antimicrobial activity of the essential oils was tested against four bacterial strains, six fungal strains, and six yeast strains. Test organisms used were obtained from the Faculty of Agriculture, Cairo University.

Agar diffusion method

This method was carried out according to Collins and Lyne (1985). Nutrient agar (NA) was used for cultivation of bacteria and yeast, and Czapek-Dox's for cultivation of fungi. In this method, presterilized paper discs (0.5 mm in diameter) were impregnated with 100 µl of each oil and applied on the surface of agar plates freshly seeded with standard inoculums of young cultures, 24 h old bacteria and yeast, and 7 days old fungi. The plates of test organisms were then incubated at 27° C for 24 h for bacteria and yeast and 48 h for fungi. At the end of the incubation period, the inhibition zones were measured (the experiment were designed, the results are averages of triplicates).

Determination of minimum inhibitory concentration (MIC)

MIC values were determined by testing representative organisms in liquid NA (for bacteria and yeast) and Czapek-Dox's solutions, using the method reported by Gardener and Provine (1984). Stock solutions of known concentrations of the tested extracts were prepared and serial dilutions (ranging from 333.33 to 0.016 mg/ml) were made. The assay tubes were inoculated with standard inocula prepared from young cultures. The minimum dilution in which no visible growth was observed is the lowest inhibitory concentration of the extract. MIC were exhibited in terms of $\mu\text{g/ml}$ media after 24 hrs incubation at $27\pm 1^\circ\text{C}$ for bacteria and yeast and 48-72 h for fungi.

Results and discussion

Chemical composition

Oils from leaves and berries of *J. phoenicea* were obtained in comparatively high yield (0.36 and 1.96% v/w, respectively). Berry oil was characterized by spicy aroma, while that of the leaves was characterized by pleasant odor. Results of GC/MS analysis of essential oils of leaves and berries are listed in Table 1. In contrast to Afifi *et al.* (1992) who stated that the major constituent of both leaf and berry oils of the Egyptian *Juniperus* was sabinene, it was found that α -pinene was the major constituent of both oils recording 39.3% for berry oil and 38.22% for leaf oil, while sabinene was the second major constituent in berry oil recording 24.29%. β -pinene was also found in considerable amounts in both oils (2.45% and 0.48% in berry and leaf oils, respectively). These results are in agreement with those reported on the analysis of other juniper oils (Adams *et al.*, 1996) in which pinane derivatives are more common. On the other hand, this report agreed with Afifi *et al.* (1992) in that α -cedrol was the second major component in the leaf oil (31.23%) and its amount was far less in berry oil (0.47%).

Fifty eight compounds were identified in berry oil representing 99.20% of the composition of the oil while sixty five compounds were identified in leaf oil representing 99.12 % of the oil. Berry oil composed mainly of monoterpenoid which amounted to 90.53% of which 72.85% were monoterpene hydrocarbons. β -phellandrene, β -pinene and δ -3-carene were the most abundant monoterpene hydrocarbons in the berry oil after α -pinene and sabinene, while trans-pinocarveol, α -terpinyl acetate, p-mentha-1, 5-diene-8-ol, camphor, α -campholene aldehyde, cis-pipretol and verbenone accounted for most of the oxygenated monoterpenes. Although the highest group in the leaf oil was monoterpene hydrocarbons (41.29%) predominated by α -pinene, δ -3-carene, limonene, β -pinene and myrcene, yet the sesquiterpenes predominated in leaf oil (50.54%) of which 32.21% was oxygenated sesquiterpenes (mostly α -cedrol) and 18.33% was sesquiterpene hydrocarbons, the most abundant were cedr-8-ene, δ -cadenene, β -cadenene, γ -muurolene and α -humulene. Very low amounts of diterpenes were found in both berry and leaf oils (0.08% and 0.35%, respectively).

Cytotoxic activities

Both oils possess very high activities against all tumor cell lines tested. Activities of leaf and berry oils are shown in Table 2 and Figure 1. Berry oil possess the highest activities against brain tumor and lung carcinoma cell line (IC_{50} = 0.6 $\mu\text{g/ml}$), followed by liver carcinoma cell line and breast carcinoma cell line (IC_{50} = 0.7 and 0.8 $\mu\text{g/ml}$, respectively), while the least activity was recorded against cervix carcinoma cell line (5.0 $\mu\text{g/ml}$). The activities of leaf oil resemble that of berry oil against both brain tumor and cervix carcinoma cell lines, while the activities of the leaf oil showed an ascending order from lung carcinoma, liver tumor and breast carcinoma cell lines (IC_{50} = 0.7, 0.9 and 1.0 $\mu\text{g/ml}$, respectively). Figure 1 showed that the activities of the berry oil against lung carcinoma, liver tumor and breast carcinoma cell lines were higher than those of leaf oil, while both oils had the same activities against brain tumor and cervix carcinoma cell lines.

The high activities of these oils may be accounted for by the presence of high ratio of monoterpenes in their composition ((Buhagiar *et al.*, 1999). Setzer *et al.* (1999) referred the activity of the essential oil of *Myrcianthes sp.* against liver tumor cell line to the presence of α -pinene, β -pinene and limonene. This may explain the higher activities of berries oils than leaf oil against lung carcinoma, liver tumor and breast carcinoma cell lines since the former contained higher percentages of total monoterpenes, α -pinene and β -pinene. Sylvestre *et al.* (2006) attributed the activity of *Croton flavens* leaf oil against lung carcinoma cell line to its contents of elemene and α -humulene which are constituents of both tested oils. Elemene was found to inhibit proliferation, stimulate apoptosis and induce cell cycle arrest in malignant cell. Limonene and perilla alcohol have been shown to inhibit protein prenylation in cultured cells (Crowell *et al.*, 1992). A minor metabolite of both limonene and perillyl alcohol, perillic acid methyl ester, is a potent inhibitor of both protein farnesyl transferase (PFT) and protein geranylgeranyl transferase (PGGT) *in vivo* (Gelb *et al.*, 1995). Limonene and perilla alcohol inhibited

chemically induced carcinogenesis from chemical carcinogens (Elegbede et al.,1984; Elegbede et al. ,1986; Elson et al. ,1988; Maltzman et al.,1989).

Antimicrobial activity

Results of antimicrobial activity of *Juniperus* leaf and berry oils are shown in Tables 3, 4. By comparing the activities of oils, leaf oil was found to have higher activity than berry oil. Berry oil was inactive against two bacterial strains; *S. aureus* and *B. cereus*, while it showed highest activity against *B. subtilis* and *E. coli*. In case of fungi, the highest activity of berry oil was against *T. veridius*, while it was inactive against *F. oxysporium* and *A. niger*. Leaf oil showed high activity against *Candida spp.*, the least activity against *R. minuta* and no activity against *C. albicans* and *S. chevallii*. Similarly, leaf oil showed highest activity against *B. subtilis* and inactive against *S. aureus*, but in contrast to berry oil, it was active against *B. cereus*. The activity of leaf oil against fungi was higher than that of berry oil and the leaf oil was also active against *A. niger*. In general, the highest activities of leaf oil were against yeasts (lowest MIC value) except for *C. carlspergensis* and *S. chevallii*, which were insensitive to leaf oil. In contrast to Stassi et al. (1996), who stated that the antimicrobial activity of *J. oxycedrus* berry oil was due mainly to its content of terpineol, *J. phoenicea* berry oil did not contain terpineol, yet it had a high antimicrobial activity.

Table 1: Composition of the volatile oils of berries and leaves of *Juniperus Phoenicea* L grown in Egypt.

Compound	KI	berries	leaves
		Area%	Area%
Terpenoids			
(a) Monoterpene hydrocarbons			
α -Pinene	939	39.30	38.22
Camphene	953	-	0.02
Sabinene	976	24.29	-
2,4 (10)Thujadiene	957	0.14	-
β -Pinene	980	2.45	0.48
Myrcene	991	-	0.45
α -Phellandrene	1005	1.01	-
δ -3-Carene	1011	1.25	0.99
α -Terpinene	1018	0.05	0.03
Limonene	1031	-	0.74
β -Phellandrene	1031	4.13	-
Cis-Ocimene	1040	-	0.01
γ -Terpinene	1062	0.07	0.14
Terpinolene	1088	0.16	0.21
(b) Oxygenated monoterpenes			
Camphenone	1093	0.50	-
α -Pinene oxide	1095	-	0.14
Linalool	1098	-	0.28
Camphenol	1109	0.06	-
β -Campholene aldehyde	1125	1.34	0.24
Trans-pinocarveol	1139	4.22	-
Camphor	1143	1.93	0.54
Trans-verbenol	1144	0.04	-
p-Mentha-1,5-diene-8-ol	1166	2.88	-
Isopinocampone	1173	-	0.34
Terpineol	1177	-	0.53
Myrtenal	1193	0.30	-
Cis-pipretol	1193	1.10	0.55
Myrtenol	1194	0.22	-
Verbenone	1204	0.86	0.17

Myrtenyl acetate	1235	-	0.07
Carvacrol methyl ether	1244	0.27	-
Carvotanacetone	1246	0.07	-
Piperitone	1252	0.28	-
Linalyl acetate	1257	-	0.51
Methyl citronellate	1261	-	0.06
Isopulegol acetate	1273	-	1.46
Bornyl acetate	1285	0.05	0.59
Perilla alcohol	1295	0.03	0.01
Trans pinocarvyl acetate	1297	0.12	-
Dehydro-elsholtziane	1298	-	0.01
Isothujyl acetate	1301	-	0.03
Trans-carvyl acetate	1337	0.15	0.26
α -Terpinyl acetate	1350	3.36	1.01

(c) Sesquiterpene hydrocarbons

δ -Elemene	1339	0.15	-
α -Copaene	1376	0.09	0.24
β -Elemene	1391	0.58	-
Italicene	1401	-	0.65
Junipene	1402	0.02	0.24
Cedr-8-ene	1409	-	5.11
β -Caryophyllene	1418	0.50	-
Widderene	1429	-	0.61
α -Himachalene	1447	0.30	-
α -Humulene	1454	0.30	1.11
Cis muurolo-4 (14),5-diene	1460	-	0.95
Germacrene D isomer	1462	-	0.63
β -Cadinene	1473	0.80	2.59
β -Chamigrene	1475	-	0.01
γ -Muurolole	1477	-	1.24
α -Selinene	1494	0.28	-
α -Muurolole	1499	0.50	-
δ -Cadinene	1524	-	3.70
Cadina-1,4-diene	1532	0.20	0.41
α -Calacorene	1542	0.11	0.14
Selina-3,7(11)1diene	1542	0.18	0.06
Germacrene B	1556	-	0.14

(d) Oxygenated sesquiterpenes

α -Agarofuran	1545	0.30	-
Occidentalol	1549	0.11	-
Cis-isoeugenol acetate	1563	-	0.04
β -Copen-4- α -ol	1584	0.35	-
Guaiol	1595	-	0.43
α -Cedrol	1596	0.47	31.23
1, 10-Di-epi-cubenol	1614	0.30	0.11
Trans-isolongifolanone	1618	0.41	-
γ -Eudesmol	1630	-	0.31
Trans dihydro-occidentalol	1645	-	0.01
β -Eudesmol	1649	-	0.17
α -Eudesmol	1652	0.32	-
Khusinol	1674	0.06	0.21
Methyl epi-jasmonate	1676	-	0.01
Acorenone	1685	-	0.01
α -Epi-bis alolol	1686	-	0.01
Cedr-8-en-4-ol	1688	0.07	0.01

Juniper camphor	1691	-	0.01
Cedryl acetate	1762	-	0.01
Cedr-8(15)-ene- α -ol acetate	1739	0.01	-
14, α -Hydroxy muurolene	1775	0.01	-
14, Hydroxy δ -cadinene	1799	0.01	-
Oblopanonyl acetate	1881	0.01	-
(e) Diterpene hydrocarbons			
Abietatriene	2054	-	0.03
Abeitadiene	2080	-	0.03
(f) Oxygenated diterpenes			
Manool	1961	-	0.01
Manoyl oxide	1989	-	0.28
Sclareol	2220	0.08	-
Miscellaneous compounds			
a)Hydrocarbons			
Tricycline	926	-	0.01
a)Oxygenated			
Hexanal	800	-	0.01
Diacetone alcohol	840	0.17	0.06
Oroselone	2146	0.33	-
Total			
No of compounds		58	65
Monoterpene hydrocarbons		72.85	41.29
Oxygenated monoterpenes		17.78	6.80
Sesquiterpene hydrocarbons		5.56	18.33
Oxygenated sesquiterpenes		2.43	32.21
Diterpene hydrocarbons		-	0.06
Oxygenated diterpenes		0.08	0.29
Miscellaneous compounds			
a) Hydrocarbon		-	0.07
b) Oxygenated		0.50	0.06

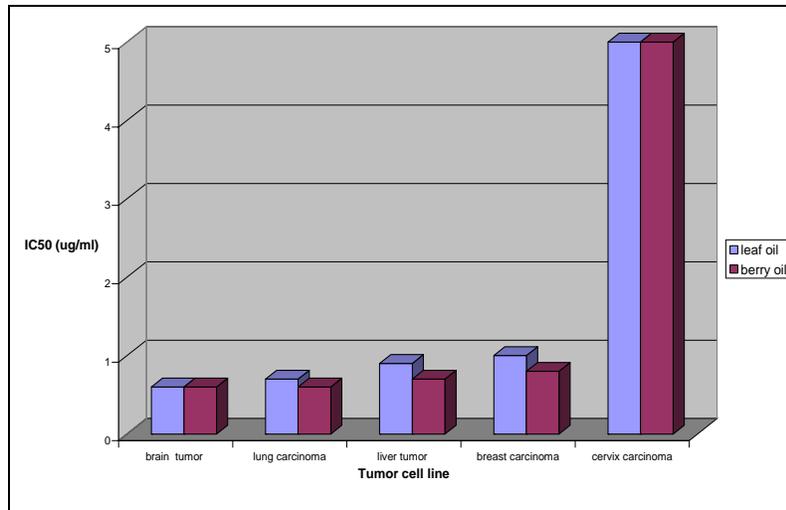
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Table 2: Percent cell survival of different human tumor cell lines against different concentrations of essential oils of berries and leaves of *J. phoenicea* L grown in Egypt.

Tumor cell line	Oil conc. μ g/ml	% cell survival \pm SEM		
		Leaf oil (100 μ l/disc)	Berry oil (100 μ l/disc)	Cisplatin
^b U251	0	100 \pm 0.06	100 \pm 0.10	100 \pm 0.07
	1	13.5 \pm 0.03	17.2 \pm 0.07	60.9 \pm 0.07
	2.5	14.0 \pm 0.03	7.7 \pm 0.06	56.4 \pm 0.05
	5	3.3 \pm 0.01	5.2 \pm 0.07	55.5 \pm 0.07
	10	7.5 \pm 0.04	7.0 \pm 0.04	49.4 \pm 0.05
^c H460	0	100 \pm 0.08	100 \pm 0.08	100 \pm 0.07
	1	34.0 \pm 0.05	23.5 \pm 0.04	79.6 \pm 0.02
	2.5	21.6 \pm 0.06	27.1 \pm 0.03	66.5 \pm 0.05
	5	16.7 \pm 0.02	19.9 \pm 0.06	48.8 \pm 0.03
	10	9.8 \pm 0.03	18.0 \pm 0.02	40.1 \pm 0.0
^d Hepg2	0	100 \pm 0.09	100 \pm 0.07	100 \pm 0.10

^e MCF7	1	44.2 ± 0.06	24.7 ± 0.0	60.1 ± 0.04
	2.5	32.8 ± 0.05	29.9 ± 0.03	59.3 ± 0.06
	5	22.0 ± 1.0	27.0 ± 0.05	52.4 ± 0.04
	10	29.6 ± 0.0	37.9 ± 0.09	51.8 ± 0.05
	0	100 ± 0.10	100 ± 0.10	
	1	52.1 ± 0.04	34.6 ± 0.04	
	2.5	42.5 ± 0.07	33.8 ± 0.03	NT
	5	37.1 ± 0.06	33.5 ± 0.04	
	10	30.7 ± 0.01	20.6 ± 0.01	
	^f Hela	0	100 ± 0.09	100 ± 0.08
1		73.4 ± 0.08	83.5 ± 0.07	90.3 ± 0.06
2.5		70.9 ± 0.04	70.6 ± 0.08	42.4 ± 0.02
5		51.5 ± 0.02	48.7 ± 0.06	17.0 ± 0.04
10		11.0 ± 0.0	38.2 ± 0.02	5.9 ± 0.02

^aOil concentration (µg/ml) required to reduce cell survival by 50%, ^bbrain tumor cell line, ^clung carcinoma cell line, ^dliver carcinoma cell line, ^ebreast carcinoma cell line, ^fcervix carcinoma cell line. NT: not testes. Cisplatin: standard antitumor drug.



IC₅₀= extract concentration (µg/ml) required to reduce cell survival by 50%.

Figure 1: Comparison between the cytotoxic activities of both leaf and berry essential oils of *Juniperus Phoenicea* L on human tumor cell lines.

Table 3. Antimicrobial activities of volatile oils of leaves and berries of *Juniperus Phoenicea* L grown in Egypt.

Test organism	Berry v. oil	leaf v. oil	Control
	(100µl/disc)	(100µl/disc)	(100µg/disc)
Inhibition zone in mm ± SE			
Bacteria			
(Gram negative)			
<i>Escherichia coli</i>	20± 0.3	15± 0.3	16± 0.6
(Gram positive)			
<i>Staphylococcus aureus</i>	R	R	22± 0.8
<i>Bacillus subtilis</i>	25± 1.1	28± 0.3	24± 0.5
<i>Bacillus cereus</i>	0	20± 0.9	10± 0.0

Fungi

<i>Fusarium oxysporium</i>	R	R	18± 0.5
<i>Aspergillus niger</i>	R	20± 0.8	9± 0.3
<i>Aspergillus flavus</i>	25± 1.2	18± 0.7	20± 0.5
<i>Macrophomina phaseoli</i>	10± 0.5	20± 0.3	21± 0.7
<i>Botrytis allii</i>	18± 0.3	20± 0.0	NT
<i>Trichoderma veridus</i>	30± 1.3	30± 0.9	NT

Yeast

<i>Candida pseudotropicalis</i>	20± 1.0	40± 1.3	NT
<i>Candida albicans</i>	R	28± 1.1	NT
<i>Candida carlspergensis</i>	30± 0.8	R	NT
<i>Saccharomyces cerevisiae</i>	20± 0.8	25± 0.9	11± 0.2
<i>Saccharomyces chevallii</i>	R	R	12± 0.1
<i>Rhodotorula minuta</i>	11± 0.5	22± 1.2	NT -

R: resistant. NT: not tested. Control: amoxicillin for bacteria and canestatin for fungi and yeast.

Table 4. Minimal inhibitory concentration (MIC) of volatile oils of leaves and berries of *Juniperus Phoenicea* L grown in Egypt.

Test organism	Berry v. oil	leaf v. oil
	mg/ml	
Bacteria		
<i>Escherichia coli</i>	12.34	37.03
<i>Staphylococcus aureus</i>	-	-
<i>Bacillus subtilis</i>	12.34	0.46
<i>Bacillus cereus</i>	-	12.34
Fungi		
<i>Fusarium oxysporium</i>	-	-
<i>Aspergillus niger</i>	-	4.11
<i>Aspergillus flavus</i>	37.03	4.11
<i>Macrophomina phaseoli</i>	12.34	4.11
<i>Botrytis allii</i>	12.34	4.11
<i>Trichoderma veridus</i>	12.34	12.34

Yeast

<i>Candida pseudotropicalis</i>	37.03	0.15
<i>Candida albicans</i>	-	0.46
<i>Candida carlspergensis</i>	37.03	-
<i>Saccharomyces cerevisiae</i>	4.11	4.11
<i>Saccharomyces chevallii</i>	-	-
<i>Rhodotorula minuta</i>	111.11	0.46

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