

<http://dx.doi.org/10.4314/ajtcam.v10i6.19>

ANTISPASMODIC EFFECTS OF YARROW (*ACHILLEA MILLEFOLIUM* L.) EXTRACT IN THE ISOLATED ILEUM OF RAT

Mohammad-Taghi Moradi¹, Mahmoud Rafeieian-Koupaei², Reza Imani-Rastabi³, Jafar Nasiri⁴, Mehrdad Shahrani^{*5}, Zahra Rabiei¹, Zahra Alibabaei¹

¹Medical Plants Research Center, Shahrekord University of Medical Science, Shahrekord, Iran

²Professor, Medical Plants Research Center, Shahrekord University of Medical Science, Shahrekord, Iran

³Assistant Professor, Infectious Diseases Dept., Shahrekord University of Medical Science, Shahrekord, Iran, ⁴Assistant Professor, Internal Medicine Dept., Shahrekord University of Medical Science, Shahrekord, Iran, ⁵Assistant Professor, Cellular and Molecular Research Center, Shahrekord University of Medical Science, Shahrekord, Iran & Physiology Research Center, Ahvaz Jundishapur University of Medical Science, Ahvaz, Iran.

*E-mail: mehrdadeshahrani2000@gmail.com

Abstract

Achillea millefolium L. is cultivated in Iran and widely used in traditional medicine for gastrointestinal disorders. The aim of this study was to determine the effect of hydroalcoholic extract of *A. millefolium* on the contraction and relaxation of isolated ileum in rat. In this experimental study, aerial parts of *A. millefolium* were extracted by maceration in ethanol 70% for 72h. Terminal portion of ileum in 100 male Wistar rats was dissected and its contractions were recorded isotonicly in an organ bath containing Tyrode solution (37 °C, pH 7.4) under one gram tension. Acetylcholine (1mM) and KCl (60mM) were used to create isotonic contractions. Propranolol and N ω -Nitro-L-arginine methylester hydrochloride (L-NAME) were used to investigate the mechanisms of action prior to giving the extract to the relevant groups. Data were compared by ANOVA and Turkey's post hoc test. The results showed that the ileum contraction was induced by KCl and acetylcholine induced contraction was significantly reduced by *A. millefolium* extract. The cumulative concentrations of *A. millefolium* relaxed the KCl and acetylcholine induced contractions (n=14, p<0.001). The inhibitory effect of extract on contraction induced by KCl and acetylcholine was not significantly affected neither by propranolol (1 μ M) nor by L-NAME (100 μ M). There was no significant difference in the rate of relaxation by propranolol and L-NAME between the two groups. In conclusion, *A. millefolium* can inhibit contraction of smooth muscle of ileum in rat, and it can be used for eliminating intestinal spasms. These results suggest that the relaxatory effect of *A. millefolium* on ileum contractions can be due to the blockade of voltage dependent calcium channels. In addition, the β -adrenoceptors, cholinergic receptors and nitric oxide production are not powerful actors in inhibitory effect of *A. millefolium*. So, the nitric oxide and adrenergic systems may also be involved in the antispasmodic effect of *A. millefolium*.

Keywords: *Achillea millefolium*, ileum, rats, contraction and relaxation, L-NAME, propranolol, smooth muscle

Introduction

A common functional disorder of the intestine that affects many people is irritable bowel syndrome (IBS). The factors inducing IBS are not known. Patients suffer from IBS episodes with alternating constipation and diarrhea. Abdominal pain and cramps are among the most common symptoms of IBS. Nowadays, many people have resorted to the use of natural medicines for the treatment of intestinal disorders.

Medicinal plants have been used for more than 2000 years. Increasing attention has been paid to herbal products because of their capability and cheaper cost in recent years (World et al., 2002, Uprety et al., 2012). One of the common natural medicines used for the treatment of IBS is peppermint oil (Hills, and Aaronson, 1991) which has a spasmolytic effect. There are various herbal medicines traditionally used for intestinal disorders and diarrhea (Sadraei et al., 2003). A famous characteristic of these medicinal plants is their spasmolytic activity. Therefore, the selection of the most effective herbs in this field is of particular interest.

Achillea millefolium L. (Asteraceae) grows wildly in Asia, Europe, North Africa, and North America. Its different types have been used by local people as folk or traditional medicinal plants. Bumadaran is a known name for several species of *Achillea* in Iran. It is cultivated in Iran and widely used in traditional medicine for gastrointestinal disorders (Nemeth and Bernath, 2008).

Several effects such as antibacterial (Candan et al., 2003, Stojanovic et al., 2005), anti-inflammatory (Benedek et al., 2007), antihypertensive and antihyperlipidemia (Asgary et al., 2000), and antitumor (Tozyo et al., 1994, Csupor-Loffler et al., 2009) have been reported for *Achillea*. Also, effects on the gastrointestinal tract such as antiulcer (Cavalcanti et al., 2006), antispasmodic (Lemmens-Gruber et al., 2006, Yaesh et al., 2006, Karamenderes and Apaydin, 2003), antibacterial (*Helicobacter pylori*) (Mahady et al., 2005), choleric (Benedek et al., 2006), and hepatoprotective (Lemmens-Gruber et al., 2006) have been reported. Flavonoids, alkaloids (achilleine), cineol, borneol, α and β pinen, camphor, caryophyllene, thujene, rutin, monoterpenoids and sesquiterpenoids (Dokhani et al., 2005, Afsharypour et al., 1996, Gherase et al., 2003, Javidian K et al., 2004) are among its chemical compounds.

Propranolol is a non-selective beta-adrenergic receptor blocking agent (Dobarro et al., 2013). It has no other autonomic nervous system activity. Propranolol is a competitive antagonist which specifically competes with beta-adrenergic receptor stimulating agents for available beta-receptor sites (Pereira-Leite et al., 2013).

NG-nitro-L-arginine methyl ester (L-NAME) is a non-selective inhibitor of nitric oxide synthesis (NOS) (Zhao et al., 2013). In this study, we investigated the effect of hydroalcoholic extract of floriated shoot of *A. millefolium* on the smooth muscle relaxation of isolated ileum in rats to find out their antispasmodic activity possibility.

<http://dx.doi.org/10.4314/ajtcam.v10i6.19>

Materials and Methods

Plant material

Aerial parts of *A. millefolium* were collected from plants in Tangesayad, Caharmahal va Bakhtiari province (Iran). The plant was identified by the staff of Medical Plants Research Center of Shahrekord University of Medical Sciences, (Iran). A voucher specimen was deposited at herbarium of the Medical Plants Research Center (voucher No. 304).

Extraction method

Hydroalcoholic extract was obtained by the Maceration method. The crude *A. millefolium* was reduced to pieces of suitable size, mixed thoroughly with the specified extracting solvent (ethanol 70%), and allowed to stand at room temperature in a closed container for an appropriate time (72h) with frequent agitation until soluble matter was dissolved. The mixture was filtered, the insoluble material was washed with the same solvent used for maceration, and the filtrates were combined and concentrated, usually under pressure reduced by the vacuum pump, to isolate the solvent from the extraction by lower temperature (40°C) to the desired consistency (Rashidi et al., 2005, Moreno-Loaiza and Paz-Aliaga, 2010).

Experimental procedure

This experimental study was conducted using 84 male Wistar rats (weighing 185–250 g). The animals were kept in a 20-24°C temperature with a 12h light/dark cycle and fed with standard diet, and drinking water was made easily accessible for them. The rats were deprived of food 24h before the experiment but they had free access to water. Animals were randomly assigned to six experimental groups: 1) KCl+saline; 2) KCl + *A. millefolium*; 3) acetylcholine (Ach) + saline; 4) Ach + *A. millefolium*; 5) L-NAME + KCl + *A. millefolium*; and 6) propranolol + KCl + *A. millefolium*. All procedures were in accordance with the guide for the care and use of laboratory animals adopted by Iran's National Institute of Health and federation of Iranian societies for experimental biology. This study was approved by the Ethics Committee of Shahrekord University of Medical Sciences, Sahrekord, Iran (approval number 89-1-3) and the study was conducted in accordance with the Ethical and Practical Principles of Laboratory Animal guidelines.

Animals were anesthetized by ether and 1.5-2cm of ileum terminal portion was isolated in post laparotomy and placed in oxygenated Tyrode's solution (Pickering et al., 2009, Sadraei et al., 2011, Ince et al., 2011, Brankovic et al., 2011) of the following composition (mM): NaCl 136.9, KCl 2.68, CaCl₂ 1.8, MgCl₂ 1.05, NaHCO₃ 11.9, NaH₂PO₄ 0.42, and glucose 5.55. The ileum was mounted in a 10 ml organ bath containing Tyrode solution, which was maintained at 37°C and constantly aerated with oxygen (pH=7.4). The tissues were subjected to a resting tension of one gram. The period of adaptation was 60min and bath solutions were changed every 15 minutes. Isotonic contractions of tissue were recorded using isotonic transducer (Harvard Isotonic Transducer, UK) and displayed on a Harvard Universal Oscillograph pen recorder device. At the end of adaptation period, smooth muscle of ileum with KCl (60 mM) (Camilleri, 2001) or acetylcholine (1µM) (Hasler and Owyang, 1999) was contracted. After reaching the plateau of contraction, cumulative concentrations of extract (1.5 mg/mL) was added to the organ bath. The percentage change in contractile force was compared with the plateau of contraction. The effects of concentrations of *A. millefolium* extracts on contraction induced by KCl and acetylcholine were measured. After 30min incubation of the tissue with 1µM propranolol at concentration (Vanner et al., 1999) and 20min with 100 µM L-NAME at concentration (Hamm et al., 1999) the previous steps were repeated. To investigate the role of extracellular calcium in the extracts, the tissues in calcium-free Tyrode's solution containing KCL concentration (120mM) were placed and the cumulative CaCl₂ (0.225-3.6 mM) was added to the bathroom. After 5min presence of extracts in different tissues with 2-4 mg/ml concentrations, the previous steps were repeated.

Statistical analysis

Mean and standard error of mean (SEM) values were calculated for each group of results and the significant difference between the means was calculated by one way analysis of variance (ANOVA) and Turkey's post hoc test. Differences were considered statistically significant when P<0.05.

Results

In all experiments, the addition of KCl solution (60mM) and Acetylcholine (1mM) increased the initial sudden contraction in the ileum rapidly. The contractions returned to the plateau state immediately. At this point, the contractions of the ileum were considered 100%. Adding the extracts to bath tissue decreased KCl- and acetylcholine-induced contractions.

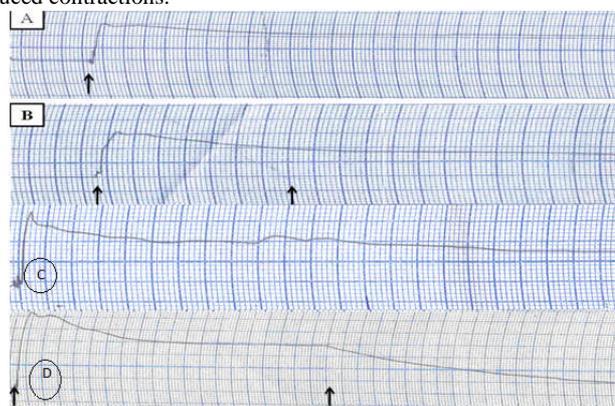


Figure 1: The recorded samples taken from the ileum of rats Response to (A) added acetylcholine, (B) KCl (left arrow) or saline (right arrow), (C) KCl + extracts and (D) acetylcholine (left arrow) + extract (right arrow)

<http://dx.doi.org/10.4314/ajtcam.v10i6.19>

According to Figure 2, significant differences between groups Ach + saline with KCl + saline were seen ($P < 0.01$). Acetylcholine caused more contraction of ileum compared to KCl. Significant difference was observed between groups Ach + saline, KCl + *A. millefolium*, L-NAME + KCl + *A. millefolium*, and propranolol + KCl + *A. millefolium* ($P < 0.05$). In the acetylcholine group, contraction rate was higher compared to those of others.

According to Figure 3 antispasmodic effect of *A. millefolium* extract was greater than those of the other groups ($P < 0.001$).

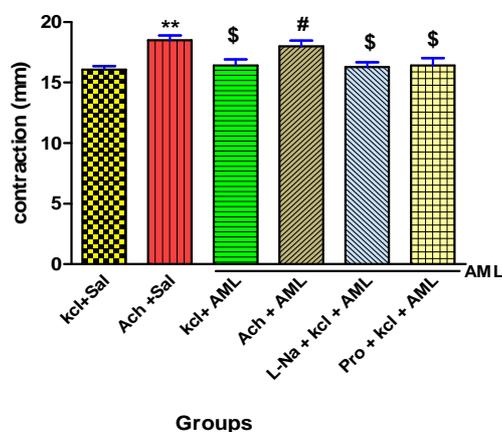


Figure 2: Effect of *A. millefolium* on ileum contractions of rats in different groups

Values are expressed as mean \pm S.E.M. ($n = 14$). ** $p < 0.01$ as compared to kcl+Sal group; # $p < 0.05$ as compared to kcl+Sal group; \$ $p < 0.05$ as compared to Ach +Sal group; (One-way ANOVA followed by Tukey's test) Sal= saline, AML= *A. millefolium* L., Ach= acetylcholine, L-Na= L-NAME, Pro= propranolol

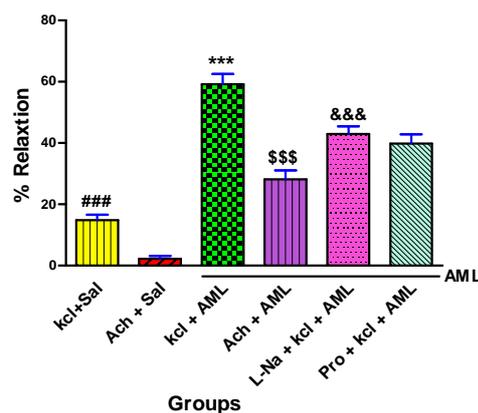


Figure 3: Effect of *A. millefolium* on ileum relaxation of rats in different groups

Values are expressed as mean \pm S.E.M. ($n = 14$). *** $p < 0.001$ as compared to all other groups; ### $p < 0.001$ as compared to Ach +Sal group; \$\$\$ $p < 0.001$ as compared to Ach +Sal and kcl + Sal groups; &&& $p < 0.001$ as compared to Ach +Sal, kcl + Sal and Ach+ *A. millefolium* groups (one-way ANOVA followed by Tukey's test) Sal= saline, AML= *A. millefolium* L., Ach= acetylcholine, L-Na= L-NAME, Pro= propranolol.

Discussion

The purpose of this study was to investigate the pharmacological effects of *A. millefolium* on rat ileum contractions. As obtained in the results, the alcoholic extract of *A. millefolium* reduced KCl and Acetylcholine-induced contraction of ileum. Smooth muscle membrane contains more voltage-dependent calcium channels compared to skeletal muscle, but much less voltage-dependent sodium channels. Therefore, calcium ions flow into fiber through slow calcium-sodium channels thanks to KCl effect is responsible for the contraction (Ratz et al., 2005). Substances that inhibit KCl-induced contraction of the smooth muscle are referred to as voltage-dependent calcium channel blockers (Berridge, 2008). Acetylcholine causes the contraction through the effect on the receptors of muscarinic M4, decrease in cAMP (Cyclic adenosine monophosphate), increase in potassium, the activation of muscarinic M3 receptors and calcium channels, increase in intracellular calcium, or by affecting nicotinic receptor and opening the ion channels and depolarization (Nausch et al., 2010, Wright and Huddart, 2002). The inhibitory effects of *A. millefolium* extract stimulated by KCl and acetylcholine are not fully reversible. These effects only decline and will not be completely eliminated through washing or replacing the tissue. The relative decline of the effects of extract after washing the tissue is probably attributable to the removal of extract from surface receptors that were reversibly bound to extract. Also, the results of long term ileum contraction by KCl and acetylcholine without using extracts revealed that ileum can stay contracted in long term with no decline in contractile force. These results indicated that the observed decrease in contractile force was due to the performance of extract rather than muscle fatigue. Since the nitric oxide synthase inhibitor drug that is normally responsible for the production of nitric oxide in the body could not block the

<http://dx.doi.org/10.4314/ajtcam.v10i6.19>

antispasmodic effects of the extract, the extract could not act by affecting the body's levels of nitric oxide. Also, contractile effect is not completely inhibited with the presence of propranolol that is an inhibitor of adrenergic system. So, the extract can affect through adrenergic as well as nitric oxide receptors. Increasing the intracellular calcium and potassium is the controlling factor of tension of gastrointestinal smooth muscle (Wells et al., 2008). So, the existence of extract may have been inhibiting the normal function of calcium and potassium, through which the present extract could have inhibited the contraction.

Apigenin flavonoid has been reportedly present in the extract (Yaesh et al., 2006) and can take part in an antispasmodic activity (Takzare et al., 2011). As a result, antispasmodic effects observed in the present work could be attributed to the apigenin flavonoid of *A. millefolium*. However, more research is needed in this regard. By methods such as immunohistochemistry and binding, the precise mechanisms of *A. millefolium* antispasmodic effects could be more clarified. Decrease in small intestinal contractions could suggest that *A. millefolium* may be helpful in the treatment of diarrhea predominant irritable bowel syndrome (IBS).

Conclusion

A. millefolium can inhibit contraction of smooth muscle of ileum in rat, and it can be used for eliminating intestinal spasms. These results suggest that the relaxatory effect of *A. millefolium* on ileum contractions can be due to the blockade of voltage dependent calcium channels. In addition, the β -adrenoceptors, cholinergic receptors and nitric oxide production are not powerful actors in inhibitory effect of *A. millefolium*. So, the nitric oxide and adrenergic systems may also be involved in the antispasmodic effect of *A. millefolium*.

Acknowledgements

This study was conducted with the assistance of Medical Plants Research Center of Shahrekord University of Medical Science and was approved by the Ethics Committee of Shahrekord University of Medical Sciences. Authors also would like to thank Research and Technology Deputy of Shahrekord University of Medical Sciences for their financial support.

References

1. Afsharypour, S., Asgary, S. and Lockwood, G. B. (1996). Constituents of the essential oil of *Achillea wilhelmsii* from Iran. *Planta medica*, 62, 77-8.
2. Asgary, S., Naderi, G. H., Sarrafzadegan, N., Mohammadifard, N., Mostafavi, S. and Vakili, R. (2000). Antihypertensive and antihyperlipidemic effects of *Achillea wilhelmsii*. *Drugs under experimental and clinical research*, 26, 89-93.
3. Benedek, B., Geisz, N., Jager, W., Thalhammer, T. and Kopp, B. (2006). Choleric effects of yarrow (*Achillea millefolium* s.l.) in the isolated perfused rat liver. *Phytomedicine*, 13, 702-6.
4. Enekek, B., Kopp, B. and Melzig, M. F. (2007). *Achillea millefolium* L. s.l. -- is the anti-inflammatory activity mediated by protease inhibition? *Journal of Ethnopharmacology*, 113, 312-7.
5. Berridge, M. J. (2008) Smooth muscle cell calcium activation mechanisms. *The Journal of physiology*, 586, 5047-61.
6. Brankovic, S., Kitic, D., Radenkovic, M., Veljkovic, S., Jankovic, T., Savikin, K. and Zdunic, G. (2011) Spasmolytic activity of the ethanol extract of *Sideritis raeseri* spp. *Raeseri* Boiss. & Heldr. on the isolated rat ileum contractions. *Journal of medicinal food*, 14, 495-8.
7. Camilleri, M. (2001) Review article: tegaserod. *Alimentary pharmacology and therapeutics*, 15, 277-89.
8. Candan, F., Unlu, M., Tepe, B., Daferera, D., Polissiou, M., Sokmen, A. and Akpulat, H. A. (2003) Antioxidant and antimicrobial activity of the essential oil and methanol extracts of *Achillea millefolium* subsp. *millefolium* Afan. (Asteraceae). *J Ethnopharmacol*, 87, 215-20.
9. Cavalcanti, A. M., Baggio, C. H., Freitas, C. S., Rieck, L., De sousa, R. S., Da silva-santos, J. E., Mesia-vela, S. and Marques, M. C. (2006) Safety and antiulcer efficacy studies of *Achillea millefolium* L. after chronic treatment in Wistar rats. *J Ethnopharmacol*, 107, 277-84.
10. Csupor-loffler, B., Hajdu, Z., Zupko, I., Rethy, B., Falkay, G., Forgo, P. and Hohmann, J. (2009) Antiproliferative effect of flavonoids and sesquiterpenoids from *Achillea millefolium* s.l. on cultured human tumour cell lines. *Phytother Res*, 23, 672-6.
11. Dobarro, M., Orejana, L., Aguirre, N. and Ramirez, M. J. (2013) Propranolol restores cognitive deficits and improves amyloid and Tau pathologies in a senescence-accelerated mouse model. *Neuropharmacology*, 64, 137- 44.
12. Dokhani, S., Cottrell, T., Khajeddin, J. and Mazza, G. (2005) Analysis of aroma and phenolic components of selected *Achillea* species. *Plant foods for human nutrition*, 60, 55-62.
13. Gherase, F., Spac, A., Dorneanu, V., Stanescu, U. and Grigorescu, E. (2003) [Pharmacognostic research of some species of *Achillea*. Note 1. Volatile oils analysis]. *Revista medico-chirurgicala a Societatii de Medici si Naturalisti din Iasi*, 107, 188-91.
14. Hamm, L. R., Sorrells, S. C., Harding, J. P., Northcutt, A. R., Heath, A. T., Kapke, G. F., Hunt, C. M. and Mangel, A. W. (1999) Additional investigations fail to alter the diagnosis of irritable bowel syndrome in subjects fulfilling the Rome criteria. *The American journal of gastroenterology*, 94, 1279-82.
15. Hasler, W. L. and Owyang, C. (1999) Irritable bowel syndrome. In: Yamada, T., Alpers, D. H., Laine, L., Owyang, C., Powell, D. W., editors. *Text book of gastroenterology*. 3rd ed. Philadelphia: Lippincott Williams and Wilkins.
16. Hills, J. M. and Aaronson, P.I. (1991) The mechanism of action of peppermint oil on gastrointestinal smooth muscle. *Gastroenterology*, 101,55-65.
17. Ince, S., Turkmen, R. and Yavuz, H. (2011) The effect of boric acid on acetylcholine, bethanechol and potassium-evoked responses on ileum of rat. *Autonomic and autacoid pharmacology*, 31, 50-6.
18. Javidian, K., Miri, R., and H. S. (2004) Composition of the volatile oil of *Achillea wilhelmsii* C. Koch from Iran. *Daru*, 12, 63-66.
19. Karamenderes, C. and Apaydin, S. (2003) Antispasmodic effect of *Achillea nobilis* L. subsp. *sipylea* (O. Schwarz) Bassler on the rat isolated duodenum. *Journal of Ethnopharmacology*, 84, 175-9.

<http://dx.doi.org/10.4314/ajtcam.v10i6.19>

20. Lemmens-gruber, R., Marchart, E., Rawnduzi, P., Engel, N., Benedek, B. and Kopp, B. (2006) Investigation of the spasmolytic activity of the flavonoid fraction of *Achillea millefolium* s.l. on isolated guinea-pig ilea. *Arzneimittelforschung*, 56, 582-8.
21. Mahady, G. B., Pendland, S. L., Stoia, A., Hamill, F. A., Fabricant, D., Dietz, B. M. and Chadwick, L. R. (2005) *In vitro* susceptibility of *Helicobacter pylori* to botanical extracts used traditionally for the treatment of gastrointestinal disorders. *Phytotherapy Research: PTR*, 19, 988-91.
22. Moreno-loaiza, O. and Paz-aliaga, A. (2010) [Vasodilator effect mediated by nitric oxide of the *Zea mays* L. (andean purple corn) hydroalcoholic extract in aortic rings of rat]. *Revista peruana de medicina experimental y salud publica*, 27, 527-31.
23. Nausch, B., Heppner, T. J. and Nelson, M. T. (2010) Nerve-released acetylcholine contracts urinary bladder smooth muscle by inducing action potentials independently of IP3-mediated calcium release. *American journal of physiology. Regulatory, integrative and comparative physiology*, 299, R878-88.
24. Nemeth, E. and Bernath, J. (2008) Biological activities of yarrow species (*Achillea* spp.). *Current pharmaceutical design*, 14, 3151-67.
25. Pereira-leite, C., Carneiro, C., Soares, J. X., Afonso, C., Nunes, C., Lucio, M. and Reis, S. (2013) Biophysical characterization of the drug-membrane interaction: the case of propranolol and acebutolol. *European journal of pharmaceuticals and biopharmaceutics : official Journal of Arbeitsgemeinschaft fur Pharmazeutische Verfahrenstechnik e.V.*
26. Pickering, J. D., White, E., Duke, A. M. and Steele, D. S. (2009) DHPR activation underlies SR Ca²⁺ release induced by osmotic stress in isolated rat skeletal muscle fibers. *The Journal of general physiology*, 133, 511-24.
27. Rashidi I, T.M.M. and Ar, M. (2005) Study of anti-inflammatory and healing effects of *Achillea Millefolium* in the treatment of Indomethacin-induced gastric ulcer in rat. *Journal of Ghazvin University of Medical Sciences and Health Services*, 33, 9-13.
28. Ratz, P. H., Berg, K. M., Urban, N. H. and Miner, A. S. (2005) Regulation of smooth muscle calcium sensitivity: KCl as a calcium-sensitizing stimulus. *American journal of physiology. Cell physiology*, 288, C769-83.
29. Sadraei, H., Asghari, G. and Poorkhosravi, R. (2011) Spasmolytic effect of root and aerial parts extract of *Pycnocycla spinosa* on neural stimulation of rat ileum. *Research in pharmaceutical sciences*, 6, 43-50.
30. Sadraei, H., Ghannadi, A. and Malekshahi, K. (2003) Relaxant effect of essential oil of *Melissa officinalis* and citral on rat ileum contractions. *Fitoterapia*, 74, 445-52.
31. Stojanovic, G., Radulovic, N., Hashimoto, T. and Palic, R. (2005) *In vitro* antimicrobial activity of extracts of four *Achillea* species: the composition of *Achillea clavennae* L. (Asteraceae) extract. *Journal of Ethnopharmacology*, 101, 185-90.
32. Takzare, N., Hosseini, M. J., Hamideh M. S., Safaie, S. and Moradi, R. (2011) The effect of *Achillea millefolium* extract on spermatogenesis of male Wistar rats. *Human and Experimental Toxicology*, 30, 328-34.
33. Tozyo, T., Yoshimura, Y., Sakurai, K., Uchida, N., Takeda, Y., Nakai, H. and Ishii, H. (1994) Novel antitumor sesquiterpenoids in *Achillea millefolium*. *Chem Pharm Bull (Tokyo)*, 42, 1096-100.
34. Uprety, Y., Asselin, H., Dhakai, A. and Julien, N. (2012) Traditional use of medicinal plants in the boreal forest of Canada: review and perspectives. *Journal of Ethnobiology and Ethnomedicine*, 8, 7.
35. Vanner, S. J., Depew, W. T., Paterson, W. G., Dacosta, L. R., Groll, A. G., Simon, J. B. and Djurfeldt, M. (1999) Predictive value of the Rome criteria for diagnosing the irritable bowel syndrome. *The American Journal of Gastroenterology*, 94, 2912-7.
36. Wells, R. W., Lourenssen, S. and Blennerhassett, M. G. (2008) Impaired acetylcholine-induced smooth muscle contraction in colitis involves altered calcium mobilization and AKT phosphorylation. *Pflugers Archiv : European journal of physiology*, 456, 507-17.
37. World, Health Organization (2002) Traditional medicine growing needs and potential. Geneva World Health Organization, 2.
39. Wright, T. J. and Huddart, H. (2002) The nature of the acetylcholine and 5-hydroxytryptamine receptors in buccal smooth muscle of the pest slug *Deroceras reticulatum*. *Journal of comparative physiology. B, Biochemical, systemic, and environmental physiology*, 172, 237-49.
40. Yaesh, S., Jamal, Q., Khan, A. U. and Gilani, A. H. (2006) Studies on hepatoprotective, antispasmodic and calcium antagonist activities of the aqueous-methanol extract of *Achillea millefolium*. *Phytother Research*, 20, 546-51.
41. Zhao, Y., Vanhoutte, P. M. and Leung, S. W. (2013) Endothelial nitric oxide synthase-independent release of nitric oxide in the aorta of the spontaneously hypertensive rat. *The Journal of Pharmacology and Experimental Therapeutics*, 344, 15-22.