Egg temperature and embryonic metabolism of A- and B-eggs of macaroni and rockhopper penguins

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Macaroni and rockhopper penguins lay two eggs but rear only one chick to independence. The eggs are markedly dimorphic in size and, although the smaller A-egg is laid several days before the B-egg, in nests where both eggs are incubated, the B-egg always hatches first. Incubation temperatures and embryonic oxygen consumption were measured to determine whether the observed hatching sequence could be accounted for by differences in egg temperatures or rate of embryonic development. Lowest egg temperatures were recorded from A-eggs incubated in the less favourable anterior position in the brood patch and highest temperatures from A-eggs incubated singly and B-eggs. Differences, however, were not significant. Levels of embryonic oxygen consumption of A-eggs of the same age showed a similar pattern to egg temperatures, but differences were slight. A-eggs incubated singly, and those incubated for several days after laying in a hot-room, still had incubation periods longer than B-eggs, suggesting that egg temperature alone does not account for differences in the hatching sequence. Consequently, it appears that there are inherent differences in embryonic metabolism of A- and B-eggs that result in the B-egg, which represents the greater parental investment, hatching first.

Macaroni- en geelkuifpikkewyne lê twee eiers op 'n keer maar net een kuiken word grootgemaak. Die eiers is opvallend dimorf in grootte en al word die kleiner A-eier verskeie dae voor die B-eier gelê, in neste waar altwee eiers bebroei word, broei die B-eier altyd eerste uit. Broeitemperature en embrioniese suurstofverbruik is gemeet om te bepaal of die waargenome uitbroeivolgorde toegeskryf kan word aan verskillende broeitemperature of embrioniese ontwikkelingstempo's. Die laagste eiertemperature is gemeet by A-eiers wat in die minder gunstige voorste posisie in die broeikol bebroei is en die hoogste temperature is by A-eiers wat alleen bebroei is en by B-eiers gemeet, maar die verskille was nie betekenisvol nie. Embrioniese suurstofverbruik het 'n soortgelyke patroon as eiertemperature getoon, en die verskille was eweneens baie klein. A-eiers wat alleen bebroei is en A-eiers wat vir verskeie dae nadat hulle gelê is in 'n broeikamer bebroei is, het nog broeiperiodes gehad wat langer was as dié van B-eiers. Dit dui aan dat die temperatuur van die eiers alleen nie vir die verskil in die uitbroeivolgorde verantwoordelik is nie. Dit blyk dus asof daar inherente verskille in die embrioniese metabolisme van A- en A-eiers is, wat verseker dat die B-eier, wat die grootste ouerlike bydrae verteenwoordig, altyd eerste uitbroei.

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Crested penguins of the genus Eudyptes lay two eggs which are markedly dimorphic in size (Gwynn 1953). This dimorphism is most pronounced in macaroni penguins E. chrysolophus and rockhopper penguins E. chrysocome in which the first laid A-eggs average 59% and 44% lighter, respectively, than the second laid B-eggs (Williams 1980a). Although both eggs are viable, only one chick is reared. In macaroni penguins brood reduction usually occurs through loss or ejection of the A-egg from the nest before the B-egg is laid (Warham 1963, 1971; Williams 1980b). However, in rockhopper penguins, both A- and B-eggs are incubated in about 30% of nests (Williams 1980b). The laying interval between A- and B-eggs of both species averages 4-5 days, but the B-egg hatches before or on the same day as the A-egg (Williams 1981a). Williams (1980a) suggested two possible reasons for this. Firstly, in nests where both eggs are incubated, incubation of the Aegg does not begin until the B-egg has been laid. Also, because eggs are positioned one behind the other in the elongated brood patch and the birds incubate in a semiupright position, the anterior egg is more exposed. A-eggs are more frequently placed in the anterior position. Consequently, A-eggs are incubated for the same period as B-eggs but generally experience lower and less steady temperatures during incubation (cf. Burger & Williams 1979) which might retard embryonic growth. Secondly,

the time required for embryonic development is different for A-and B-eggs.

In this study I investigated egg temperatures and embryonic oxygen consumption of A- and B-eggs of macaroni and rockhopper penguins. In order to test the suggestions of Williams (1980a), egg temperatures and embryonic oxygen consumption were measured on (i) Aeggs incubated normally with B-eggs, (ii) A-eggs incubated alone and (iii) A-eggs which were incubated at temperatures above 30°C from day of laying.

Methods

The study was carried out at sub-Antarctic Marion Island (46°52'S / 37°51'E), between October and December 1985. Selected nests of both macaroni and rockhopper penguins were numbered prior to egg laying and nests were checked daily for eggs, whose laying dates were recorded. Because the A-egg of macaroni penguins is usually lost before the B-egg is laid, A-eggs were substituted for B-eggs in a number of nests at the beginning of incubation. In order to obtain rockhopper penguin nests in which A-eggs were incubated alone, B-eggs were removed from several nests on the day they were laid. Because A-eggs of rockhopper penguins are not normally incubated until the B-egg is laid, A-eggs were

removed from four nests and 'pre-incubated' at 33-35°C in a hot-room. These eggs were then replaced in their nests once the B-eggs were laid.

Measurement of egg temperatures

Egg temperatures of B-eggs of macaroni penguins and Aand B-eggs of rockhopper penguins were measured using previously collected and blown eggs of the respective species filled with water and containing previously calibrated Model T or Model L minimitters (Minimitter Co., Indianapolis). Minimitters were held in place in the centre of the eggs by cut off, perforated, plastic syringe barrels inserted into the eggs through a hole in the blunt end and glued into place in the long axis of the eggs. Water and the minimitter were introduced into each egg through the hole in the blunt end which was then sealed using the rubber plunger from the syringe.

Eggs containing transmitters were substituted for fertile eggs in the field when the latter were removed for measurements of oxygen consumption and were readily accepted and incubated by the birds. Because the macaroni penguin colony was about 2,5 km from the research station, temperatures of the dummy eggs were measured prior to replacement of the fertile eggs, between 3-4 h and 24 h after their initial removal. Temperatures were then recorded at 5-min intervals over a period of 15-30 min. Rockhopper penguins, however, nest under some of the buildings of the research station and on the adjacent point. Consequently, a series of egg temperature measurements was usually made, when convenient, over periods of 12-24 h. The mean egg temperature over each period was subsequently used.

Embryonic oxygen consumption

Measurements of embryonic oxygen consumption were made at approximately 5-day intervals from 6–10 days into incubation. Dummy eggs containing transmitters were substituted for fertile eggs in the nests and fertile eggs taken to the laboratory, a trip of less than 5 min for eggs of rockhopper penguins, but 25 min for those of macaroni penguins, which were wrapped in cotton wool for the trip. In the laboratory, eggs were weighed to the nearest 0,01 g on a Mettler analytical balance and placed in a hot-room at 33–35°C, within the range of egg temperatures previously measured for penguins (Burger & Williams 1979 and references therein).

Embryonic oxygen consumption was measured in closed respiratory systems consisting of plexiglass syringes of 2 800 ml (macaroni penguin eggs) or 1 400 ml (rockhopper eggs) maximum volume. Chamber volume depended on how far back the plunger was drawn and, because this was varied depending on the size of the egg and the age of the embryo, chamber volume was calculated for each measurement. The net volume of air in the chamber was calculated by subtracting the volume of each egg (measured by water displacement) from the calculated chamber volume. Eggs were left for 1–3 h to come into thermal equilibrium with the hot-room air before being placed in a chamber. Chambers were then flushed with air and the open ends sealed off with three-

way stopcocks. Eggs were left in the chambers for periods estimated to reduce the oxygen concentration in the chamber by 0,5–2,0%. This required as long as 24 h for young embryos (7–10 days) to 5–10 min for pipped eggs and hatchlings. Final oxygen concentration in the chambers was measured by injecting the chamber air through a tube of soda asbestos and silica gel into a Taylor Servomex OA 570 paramagnetic oxygen analyser. Initial oxygen concentration in the hot-room was measured by pumping hot-room air through the analyser with a handheld aspirator. Embryonic oxygen consumption (VO_2) was calculated from Equation 1 of Vleck, Hoyt & Vleck (1979) and corrected to STPD. Eggs were returned to their nests after measurement.

Oxygen consumption was measured throughout incubation or until the embryo died. Measurements from embryos which died were only used from that portion of incubation in which oxygen consumption was similar to that of eggs which hatched. All means are given ± 1 standard deviation.

Results

A-eggs of rockhopper penguins incubated alone had slightly shorter incubation periods (mean = 38.8 ± 1.7 days; range 36-41; n = 6) than did those incubated in twoegg clutches (mean = $39,9 \pm 1,3$ days; range 38-42; n = 7), although the difference was not significant (t = 1,383; p > 0,02). Two A-eggs that were incubated above 30°C from laying hatched in 39 days; the remaining two pre-incubated eggs were found next to their nests once the B-eggs had hatched and were presumably ejected from their nests (Williams 1980b). Overall incubation periods and fresh egg masses of A- and B-eggs. of macaroni and rockhopper penguins, recorded during the present study (Table 1), were similar to those previously measured for the species at Marion Island (Williams 1980, 1981a) and at other localities (Warham 1963, 1971; Strange 1982).

Egg temperatures

Egg temperatures of rockhopper penguins measured over 12–24 h fluctuated by as little as 1°C to as much as 7°C,

| Table 1 | incub | ation peric | ods and | fresh egg | mass of | A - |
|-----------|---------------|-------------|---------|-------------------|----------|------------|
| and B-eg | igs of | macaroni | and ro | ckhop pe r | penguins | at |
| Marion is | land | | | | | |

| | | Macaron | i penguins | Rockhopper penguins' | | | |
|---------------|--------------|------------|-------------|----------------------|------------|--|--|
| | - | A-eggs | B-eggs | A-eggs | B-eggs | | |
| Incubation | Mean | 38,8 | 34,6 | 39,1 | 33,6 | | |
| period (days) | S.D | 1,3 | 1,8 | 1,3 | 1,1 | | |
| | Range | 37-41 | 32-39 | 38-42 | 31–35 | | |
| | N | 4 | 12 | 14 | 20 | | |
| Egg mass (g) | Mean | 99,2 | 163,3 | 77,5 | 111,9 | | |
| | <i>S.D</i> . | 7,7 | 16,0 | 8,5 | 12,9 | | |
| | Range | 87,7-109,3 | 138,7–179,0 | 63,4-91,4 | 98,1–131,8 | | |
| | N | 8 | 9 | 8 | 7 | | |

| - | | Egg temperature (°C) | | | | | | | | | | |
|---------------|------|----------------------|-------------|------|---------------------|-------|-------------|--------|------|-------|-------------|------|
| Day of | | | | | Rockhopper penguins | | | | | | | |
| incubation | | Macaroni penguins | | | A-eggs | | | B-eggs | | | | |
| 1- 7 | 27,9 | | | [1] | | | _ | | | | _ | |
| 8-14 | 33,6 | ± 0,8 | (33,0–34,9) | [5] | 31,0 | ± 0,6 | (30,4–31,6) | [3] | 31,2 | • | | [1] |
| 1521 | 34,0 | ± 1,8 | (31,1–35,9) | [6] | 33,9 | | (33,7–34,0) | [2] | 33,8 | ± 0,7 | (33,0-34,4) | [4] |
| 22-28 | 35,4 | ± 1,1 | (33,2-36,1) | [6] | 32,8 | | (31,9-33,6) | [2] | 33,5 | ± 0,7 | (32,7–34,0) | [3] |
| 29– 35 | 34,1 | | | [1] | | | _ | | 33,7 | | (32,8–34,5) | [2] |
| Overall | 34,0 | ± 2,1 | (27,9–36,1) | [19] | 32,3 | ± 1,4 | (30,4–34,0) | [7] | 33,5 | ± 1,2 | (31,2–34,5) | [10] |

Table 2 Egg temperatures of macaroni and rockhopper penguins during natural incubation at MarionIsland. Figures are means \pm standard deviation, range and sample size

presumably as a result of egg turning, changes in adult attentiveness, nest ventilation and the position of the egg in the nest.

There were slight differences in temperatures of dummy eggs placed in different positions within rockhopper penguin nests, with single eggs $(33,4 \pm 1,0^{\circ}\text{C}; n = 9)$ being incubated at a higher temperature than eggs placed in the posterior position in a two-egg clutch $(32,7 \pm 1,1^{\circ}\text{C}; n = 4)$ and lowest temperatures being recorded from eggs placed anteriorly in a two-egg clutch $(31,9 \pm 1,9^{\circ}\text{C}; n = 3)$. Differences, however, were not significant (p > 0,05), probably because of the small sample sizes involved, and results were pooled. A single macaroni penguin B-egg recorded prior to 7 days after



Figure 1 Embryonic oxygen consumption of A- and B-eggs of macaroni penguins in relation to age. Solid circles = unpipped eggs, stars = fractured eggs, open circles = pipped eggs (holed) and diamond = hatchlings ± 1 S.D. Data for A-eggs based on 76 measurements on seven eggs and those of B-eggs on 98 measurements on nine eggs. Equations relate to unpipped eggs only.



Figure 2 Embryonic oxygen consumption of A- and B-eggs of rockhopper penguins in relation to age. Symbols as for Figure 1. Data for A-eggs based on 149 measurements on 17 eggs and those of B-eggs based on 68 measurements on nine eggs. Equations relate to unpipped eggs only.

laying had a temperature of 27,9°C. However, from the second week of incubation until hatching there was no consistent trend in dummy egg temperatures of either macaroni penguins or A- and B-eggs of rockhopper penguins (Table 2). Overall egg temperature of macaroni penguins averaged $34,0 \pm 2,1^{\circ}$ C and those of rockhopper penguins A-eggs ($32,2 \pm 1,4^{\circ}$ C) averaged slightly, but not significantly, lower than those of B-eggs ($33,5 \pm 1,2^{\circ}$ C; t = 1,90; p > 0,20) (Table 2).

Embryonic oxygen consumption

Oxygen consumption (VO_z) increased throughout incubation in A- and B-eggs of macaroni and rockhopper penguins (Figures 1 & 2). Mean VO_2 prior to fracture of the egg shell averaged 821 and 1 337 ml day⁻¹ for A- and B-eggs of macaroni penguins, respectively (Table 3). In the first 10–25 days of incubation, A-eggs of rockhopper penguins which were pre-incubated for the first 3–4 days in the hot-room had rates of VO_2 similar to those

Table 3Mean pre-pipping oxygen consumption (V_{O_2}) ofmacaroni and rockhopper penguin embryos at MarionIsland

| | Pre-pipping Vo ₂ ml O ₂ /day | Day of incubation | No. of eggs |
|---------------------|---|-------------------|----------------|
| Macaroni penguins | | | |
| A-eggs | 820,6 ± 90,3 | 38 | 7 |
| B-eggs | 1337,0 ± 295,0 | 33 | 4 |
| Rockhopper penguins | | | |
| A-eggs | 592,6 ± 55,0 | 35 | 12 |
| B-eggs | 732,3 ± 78,7 | 30 | 7 |

incubated alone on the nest, both of which were slightly higher than A-eggs incubated in two-egg clutches. However, differences in mean VO_2 on comparable days of incubation were not significant, nor were differences in the slopes of the semilog transformed data relating VO_2 to incubation time (p > 0.05). Consequently, results for all A-eggs were pooled, overall pre-pipping VO_2 averaging 593 ml day⁻¹ (Table 3). Pre-pipping VO_2 of rockhopper penguin B-eggs averaged 732 ml day⁻¹ (Table 3) but was, however, measured about three days prior to initial fracture of the shell and is thus probably a slight underestimate of actual pre-pipping VO_2 .

Mean embryonic VO_2 of both macaroni and rockhopper penguin B-eggs was always significantly greater than that of A-eggs measured on the same day of incubation (macaroni penguins p < 0,02; rockhopper penguins p < 0,001), although there was no difference in the slopes of the semilog transformed data relating embryonic VO_2 to incubation time (macaroni penguins t = 1,527; p > 0,10; rockhopper penguins t = 1,183; p > 0,20).

After initial pipping, VO_2 increased markedly in all eggs measured and reached a peak in hatchlings at levels 2–3 times pre-pipping levels (Figures 1 & 2). Although hatchlings from B-eggs of both species had higher metabolic rates than hatchlings from A-eggs, differences were not significant (macaroni penguins t = 2,04; p > 0,05; rockhopper penguins t = 1,67; p > 0,01). Total oxygen consumption during embryonic development, estimated from graphical integration of the areas under the curves in Figures 1 & 2, was 12,6 and 14,5 ℓ O₂ for A- and B-eggs of macaroni penguins,

Table 4Total embryonic oxygen consumption ofmacaroni and rockhopper penguins

| | Total | VO ₂ | Pre-ninning VO- | | Pip-to-hatch VO. | | |
|---------------------|--------|-----------------|-----------------|---------|------------------|---------|--|
| | ml | ml/g egg | ml | % total | ml | % total | |
| Macaroni penguins | | | | | | | |
| A-eggs | 12 558 | 126,6 | 9 314 | 74,2 | 3 244 | 25,8 | |
| B-eggs | 14 540 | 89,0 | 10 700 | 73,6 | 3 840 | 26,4 | |
| Rockhopper penguins | 6 | | | | | | |
| A-eggs | 6 985 | 90,1 | 4 955 | 70,9 | 2 030 | 29,1 | |
| B-eggs | 10 552 | 94,2 | 7 788 | 73,8 | 2 764 | 26,2 | |

respectively, and 7,0 and 10,6 ℓ O₂ for those of rockhopper penguins (Table 4), of which 70–74% was consumed prior to pipping.

Discussion

Egg temperature

Incubation temperatures measured for both A- and Beggs of macaroni and rockhopper penguins in the present study were within the range measured for several other species of penguins (Table 5). In particular, those of rockhopper penguins were similar to those measured for the same species by Burger & Williams (1979) during the second half of incubation. Although Burger & Williams (1979) reported low egg temperatures in both macaroni and rockhopper penguins during the first half of incubation, egg temperatures of both species in the present study were maintained above 30°C from eight and 10 days after laying, respectively (see Table 2).

Consistent with Burger & Williams (1979), B-eggs of rockhopper penguins were maintained at a higher temperature than were A-eggs, although in the present

Table 5 Egg temperatures of penguins during incubation, measured with transmitters (M) and thermistors or thermocouples (T) on dummy (D), infertile (I) or in the centre (L) or air space (A) of live eggs

| | Egg tempe | rature (°C) | | |
|-----------------------------|----------------|-------------|-------------|-----------|
| Species | Mean | Range | - Method | Reference |
| Emperor penguin | 32,6 ± 0,7 | | MD | 1 |
| | $32,7 \pm 0,4$ | | | |
| Adélie penguin | 33,7 | (29,2-36,8) | MD | 2 |
| | 35,2 | (30,0-38,0) | TL | 3 |
| | 35,9 ± 1,1 | (34,7–37,2) | TL | 4 |
| Chinstrap penguin | $37,4 \pm 0,5$ | (34,8-38,0) | ΤΙ | 5 |
| | 34,5 ± 2,9 | (29,0-38,0) | | |
| Gentoo penguin | 32,9 ± 4,0 | (16,8–37,9) | ТА | 6 |
| Yellow-eyed | | | | |
| penguin | 35,2° | | - | 7 |
| Macaroni A-egg ^b | 11,7 | (1,3–33,0) | ТА | 6 |
| penguin B-egg | 23,4 | (17,2–32,5) | TA | |
| | 37,3 ± 0,6 | (34,0–37,8) | ТІ | 5 |
| | 34,0 ± 2,1 | (27,9-36,1) | MD | 8 |
| Rockhopper A-egg | 32,9 | (22,8-37,9) | ТА | 6 |
| penguin | 32,3 ± 1,4 | (30,4–34,0) | MD | 8 |
| B-egg | 25,9 ± 10,1 | (8,4–37,9) | TA | 6 |
| | 33,5 ± 1,2 | (31,2–34,5) | MD | 8 |
| Jackass penguin | 34,9° | (14,0-36,0) | - | 9 |
| | 34,5 ± 1,5 | (31,9-36,0) | TA | 6 |

^aEstimated as the midpoint between the egg-brood patch interface and the egg-nest substrate interface.

^bMeasured on day of laying.

^cCalculated indirectly from water-vapour pressure difference between the egg and the nest microclimate.

1 Bucher et al. (1986); 2 Eklund & Charlton (1959); 3 Derksen (1977); 4 Rahn & Hammel (1982); 5 Haftorn (1986); 6 Burger & Williams (1979); 7 Farner (1958); 8 This study; 9 Yom-Tov et al. (1986). study the differences were not very marked. Burger & Williams (1979) attributed differences in temperature to the position of the eggs in the nest, B-eggs being found more frequently in the posterior position where they are more covered by the brood patch and less exposed to cool ambient temperatures than are A-eggs. Differences in temperatures of eggs placed singly or in different positions in a two-egg clutch in the present study, although slight, support this, temperatures of A-eggs incubated alone and those placed posteriorly in two-egg clutches being close to those of B-eggs.

Egg temperatures of macaroni penguins measured by Haftorn (1986) were made with thermistors placed near the egg surface and are probably more representative of brood patch temperatures than of central egg temperatures measured in the present study (see Farner 1958; Yom-Tov, Wilson & Ar 1986).

Embryonic metabolism

Low egg temperatures in penguins are known to retard embryonic development (Weinrich & Baker 1978) and the slightly higher rates of embryonic oxygen consumption of pre-incubated A-eggs of rockhopper penguins, and those of A-eggs incubated singly, are consistent with the slightly higher incubation temperatures and slightly shorter incubation periods observed in these eggs compared to those incubated in two-egg clutches. However, even Aeggs which were incubated singly or at temperatures similar to those of B-eggs from day of laying had incubation periods longer than B-eggs. Furthermore, the incubation period of macaroni penguin A-eggs was 2-4 days longer than those of B-eggs, even though the A-eggs were substituted for B-eggs on the day of laying and consequently occupied the posterior position in the brood patch throughout incubation (Williams 1981a; this study). Similarly, a comparison of macaroni penguin A-eggs and rockhopper penguin B-eggs, both of which are similar in size (see Table 1), are incubated posteriorly in the brood patch and produce similarly sized hatchlings (Williams 1980a, Table 6), shows that A-eggs of macaroni penguins have rates of oxygen consumption significantly lower than those of rockhopper penguin B-eggs at equivalent stages of incubation between 10 and 30 days (p < 0.005). From the above observations, it is evident that the different

temperatures experienced by A- and B-eggs of, in particular rockhopper penguins, do not, by themselves, account for the differences in incubation periods which result in B-eggs hatching earlier than A-eggs.

Embryonic metabolism of A-eggs could be retarded if oxygen conductance across the eggshell was limiting. Because conductances of oxygen and water across eggshells are proportional to their respective diffusion coefficients, oxygen conductances of eggshells at 38°C (GO_2^{38}) can be calculated from their water vapour conductances, measured at 25°C (GH2O25), using the relationship $GO_2^{38} = 1,08$ (GH₂O²⁵) (Hoyt, Board, Rahn & Paganelli 1979). Water vapour conductances of A- and B-eggs of both macaroni and rockhopper penguins have been measured (C.R.Brown unpubl. data) and oxygen conductances calculated from these have values of 19,5 and 27,0 ml d⁻¹ torr⁻¹ for A- and B-eggs of macaroni penguins, respectively, and 15,9 and 19,2 ml d⁻¹ torr⁻¹ for of rockhopper penguins. Higher those oxygen conductances of B-eggs are consistent with their larger size and larger functional pore area, but oxygen conductances of rockhopper penguin B-eggs, which had oxygen uptakes significantly greater than macaroni penguin A-eggs of similar size, were the same. Consequently, it must be concluded that oxygen conductance of the shell does not limit embryonic metabolism in A-eggs. This suggests that Williams' second suggestion, that the time required for embryonic development of A- and B-eggs is different, is more likely. Presumably, embryos have to reach a particular level of development and metabolism before hatching can occur. The generally higher levels of metabolism in B-eggs result in this level being attained earlier than in A-eggs. A-eggs have proportionately less albumen than do B-eggs (Williams, Siegfried & Cooper 1982). Since the proportion of albumen has a controlling effect on posthatching development (cf. Nisbet 1978), Williams (1980a) suggested that it might also affect embryonic development.

Williams (1980a) hypothesized that ancestral *Eudyptes* penguins were inshore foragers which laid two eggs of similar size and were capable of rearing two chicks, as do present inshore-foraging species. He speculated that the ability to raise only a single chick resulted from a move to

Hatchling Total embryonic Fresh egg Pre-pipping V_{0_2} Hatchling VO₂ mass Vo, mass Species (ml day⁻¹) (ml) $(ml g^{-1} egg)$ (ml day⁻¹) $(ml.g day^{-1})$ (g) (g) Emperor penguin^a 43 997 465,5 1 621 94,5 Adélie penguin^a 114,2 830 11 865 99,7 81,0 1 640 20,2 Macaroni A-eggs^b 99.2 821 12 558 126,6 84.2 2 393 28.4 penguin B-eggs 163,3 1 337 14 540 89.0 106,7 2 946 27,6 Rockhopper A-eggs^b 77,5 593 6 985 90,1 30,8 53,1 1 634 732 10 552 penguin 111,9 94.2 81.0 1 9 3 4 23,9 **B-eggs**

 Table 6
 Metabolic rates of penguin embryos during development

^a Bucher et al. (1986). ^b This study.

offshore-foraging, characteristic of present Eudyptes penguins. Under these circumstances, where feeding frequency is reduced, early brood reduction would allow all energy delivered by the adults to be channelled into the single chicks which would survive, rather than wasting it on a second chick which could not be raised. Chicks from large eggs grow larger, survive better and, in penguins, are fed preferentially (Williams 1981b). Consequently, an inherently slower rate of embryonic metabolism, and presumably slower embryonic development, brought about through differences in egg composition, ensures that hatching of eggs in Eudyptes penguins is such that the larger B-egg, which represents the greater parental investment, always hatches first. Clearly, however, the relationship between egg composition and embryonic development in these and other species of Eudyptes penguins needs to be investigated further.

Embryonic metabolism has previously been measured only for emperor penguins *Aptenodytes forsteri* and Adélie penguins (Bucher, Bartholomew, Trivelpiece & Volkman 1986). Relevant results are compared with those from macaroni and rockhopper penguins in Table 6. Adélie penguin eggs and hatchlings have similar masses to those of B-eggs of rockhopper penguins and pre-pipping VO_2 , total embryonic VO_2 and hatchling VO_2 for the two species are also similar (Table 6). Overall, despite the large range of egg sizes (78 g for rockhopper penguin Aeggs to 466 g for emperor penguin eggs), total embryonic VO_2 per gram fresh egg mass was, with the single exception of macaroni penguin A-eggs, very similar for all species.

On the basis of egg composition and the ability of their chicks to leave the nest and fend for themselves (Nice 1962; Williams et al. 1982), penguins have generally been classified as semi-altricial species. However, Bucher et al. (1986) compared measured embryonic and hatchling metabolic rates with those predicted for altricial and precocial species and concluded that embryos and hatchlings of emperor and Adélie penguins had metabolic rates more characteristic of semi-precocial and precocial species than with those of semi-altricial species. Although embryonic oxygen consumption of macaroni and rockhopper penguins shows no indication of a plateau in the days immediately preceding pipping, a feature characteristic of precocial species (Vleck et al. 1979), prepipping, total, and hatchling VO_2 of both species were, as with emperor and Adélie penguins, more closely predicted by equations for precocial species than by those for altricial species. However, it is notable that post-natal metabolism of macaroni and rockhopper penguins is consistent with that of semi-altricial chicks (Brown 1987).

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References

- BROWN, C.R. 1987. Energetic requirements for growth and maintenance in macaroni and rockhopper penguins. *Polar Biol.* 8: 95–102.
- BUCHER, THERESA L., BARTHOLOMEW, G.A., TRIVELPIECE, W.Z. & VOLKMAN, N.J. 1986.
 Metabolism, growth and activity in Adélie and emperor penguin embryos. Auk 103: 485–493.
- BURGER, A.E. & WILLIAMS, A.J. 1979. Egg temperatures of the rockhopper penguin and some other penguins. *Auk* 96: 100-105.
- DERKSEN, D.V. 1977. A quantitative analysis of the incubation behavior of the Adélie penguin. *Auk* 94: 552–566.
- EKLUND, C.R. & CHARLTON, F.E. 1959. Measuring the temperature of incubating penguin eggs. Amer. Sci. 47: 80-86.
- FARNER, D.S. 1958. Incubation and body temperatures in the yellow-eyed penguin. Auk 75: 249–262.
- GWYNN, A.M. 1953. Egg laying and incubation periods of rockhopper, macaroni and gentoo penguins. *Austrl. Nat. Antarct. Res. Expedn. Rep.*, Series B: 1–29.
- HAFTORN, S. 1986. A quantitative analysis of the behaviour of the chinstrap penguin *Pygoscelis antarctica* and macaroni penguin *Eudyptes chrysolophus* on Bouvetoya during the late incubation and early nestling periods. *Pol. Res.* 4: 33-45.
- HOYT, D.F., BOARD, R.G., RAHN, H. & PAGANELLI, C.V. 1979. The eggs of the Anatidae: conductance, pore structure and metabolism. *Physiol. Zool.* 52: 438–450.
- NICE, M.M. 1962. Development of behavior in precocial birds. *Trans. Linn. Soc. New York* 8: 1–211.
- NISBET, I.C.T. 1978. Dependence of flegding success on egg size, parental performance and egg composition among common terns and roseate terns, *Sterna hirundo* and *S. dougalli. Ibis* 120: 207–215.
- RAHN, H. & HAMMEL, H.T. 1982. Incubation water loss, shell conductance, and pore dimensions in Adélie penguin eggs. *Polar Biol.* 1: 91–97.
- STRANGE, I.J. 1982. Breeding ecology of the rockhopper penguins (*Eudyptes crestatus*) in the Falkland Islands. *Gerfaut* 72: 137–188.
- VLECK, CAROL M., HOYT, D.F. & VLECK, D. 1979. Metabolism of avian embryos: patterns in altricial and precocial birds. *Physiol. Zool.* 52: 363–377.
- WARHAM, J. 1963. The rockhopper penguin *Eudyptes* chrysocome at Macquarie Island. Auk 80: 229-256.
- WARHAM, J. 1971. Aspects of breeding behaviour in the royal penguin *Eudyptes chrysolophus schlegeli*. Notornis 18: 91–115.
- WEINRICH, J.A. & BAKER, J.R. 1978. Adélie penguin (*Pygoscelis adeliae*) embryonic development at different temperatures. Auk 95: 569–576.
- WILLIAMS, A.J. 1980a. The breeding biology of *Eudyptes* penguins with special reference to egg-size dimorphism. Ph.D. thesis, University of Cape Town.
- WILLIAMS, A.J. 1980b. Offspring reduction in macaroni and rockhopper penguins. Auk 97: 754–759.
- WILLIAMS, A.J. 1981a. The laying interval and incubation

1

period of rockhopper and macaroni penguins. Ostrich 52: 226-229.

WILLIAMS, A.J. 1981b. Growth and survival of artificially twinned rockhopper penguin chicks. *Cormorant* 9: 8-12.
WILLIAMS, A.J., SIEGFRIED, W.R. & COOPER, J. 1982. Egg composition and hatchling precocity in seabirds. *Ibis* 124: 456–470.

YOM-TOV, Y., WILSON, R.P. & AR, A. 1986. Water loss from jackass penguin *Spheniscus demersus* eggs during natural incubation. *Ibis* 128: 1–8.