



## EXPERIMENTAL ASSESSMENT ON PREPARATION OF BIODEGRADABLE POLYETHYLENE/POLYLACTIC ACID-CHITOSAN COMPOSITE FROM RENEWABLE RESOURCES

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### ABSTRACT

*Due to poor degradability and contamination risk of synthetic plastics, utilization of renewable resources is encouraged. Biobased thermoplastic polymers from renewable resource that is inexpensive, biodegradable, compostable and renewably non-toxic, is focused. In this paper mixtures of synthetic and natural polymers were used as a potential option to reduce pollution by plastic waste. The study is aimed at assessing utilization of sweet potato waste as a source of bioplastic for package application, the polymer was modified with a biopolymer chitosan to obtain polylactic acid-chitosan plastic. The developed polymer matrix was blended with polyethylene to obtain biodegradable packaging material. The bioplastic was characterized using Fourier Transformed Infra-Red Spectroscopy (FTIR) and scanning electron microscope (SEM). Physical and mechanical properties of the composites were evaluated by measuring enzymatic degradation, tensile strength, and elongation at break. The results have shown that the film obtained have homogeneous surface by (SEM). Mechanical properties of the bio plastics revealed that tensile strength increases with increases in the concentration of chitosan and hence, the elongation at break decreases with increase in chitosan content. While the fastest enzymatic degradation was observed to have high microbial growth on the bio plastics with high content of Chitosan-Polylactic acid.*

### KEY WORDS

*Polyethylene, sweet potato waste, chitosan, poly lactic acid, composite.*

### INTRODUCTION

Petroleum based plastics such as polyethylene (PE) polyvinyl chloride (PVC), polypropylene (PP), polyethyleneterephthalate (PET) poly styrene (P.S) and polyamide (P.A) have been widely used as packaging materials due to their availability, flexibility and affordability (Akbar et al.,2019). However, large production and utilization of those synthetic polymers led to the accumulation of plastics wastes that create a serious environmental threat due to their poor biodegradability. Hence the need for plastics from renewable sources which are biodegradable and non-toxic (Formin et al.,2017).

Biodegradable plastics are produced from biopolymers obtained from biomass, such as starch, cellulose, or proteins. Among those biopolymers, starch would be suitable for the manufacture of bioplastics, because it is renewable, biodegradable, inexpensive and compostable (Akbar et al., 2019)

However, starch-based plastics have shown some drawn back, such as brittleness, high

sensitivity to moisture and poor mechanical strength compared to polyethylene (Akbar et al., 2019).

Blends of synthetic and natural polymers can yield a new class of materials with improved biodegradability and mechanical strength that can alternatively contribute to the reduction of environmental damages caused by plastic wastes (Formin et al., 2017)

Sweet potato is one of the sources for starch, because it contains (50-80%) starch on a dry basis and the starch comprises of (70 – 80%) of highly branched amylopectin and (20 – 30%) of linear and slightly branched amylose (Akbar et al.,2019)

Chitosan is amino polysaccharide cationic polymer that is obtained from chitin by alkaline deacetylation. Chitosan is one of the polysaccharide commonly found in nature and it's film have great potential to be used as packaging materials due to it's antimicrobial activity, non-toxicity and biodegradability (Ke-ke et al.,2007).

Poly(lactic acid) is a highly versatile and biodegradable aliphatic polyester derived from 100% renewable resources such as corn, potato, sugar cane etc (Layla *et al.*, 2018). Poly(lactic acid) is a thermoplastic that offers great promise in a wide range of commodity applications (Mohd *et al.*, 2015). Chitosan is a rigid material that can be used to modify the thermal, plastic and antimicrobial properties of poly(lactic acid) without affecting its biodegradability.

Thus, poly(lactic acid) can be modified with chitosan by amidation reaction between the amino group of the main chain of chitosan and the terminal carboxyl group of the poly(lactic acid) Nurshahida and Zainoha (2015)

However, the co-polymer blends of chitosan-poly(lactic acid) shows some deficiencies in used as a substitute for packaging application, mechanical properties, such as tensile strength, elongation to break and thermal stability. Hence, to improve these properties polyethylene (low density polyethylene) can be used as a composite with the co-polymer blends as reported by Vincento (2013)

The aim of this study, is to prepare film based on blends of synthetic polymers with natural polymers, so as to have biodegradable, biocompatible, and ecofriendly packaging materials, using simple hand lay-up technique.

Hand lay-up method is the oldest and most widely used method with acceptable mechanical properties and environmental friendly Tarek and Bader (2016).

## **MATERIAL AND METHODS**

**Materials:** Sweet potato waste used for the synthesis of poly(lactic acid) was obtained from Yankaba Market Nassarawa Local Government Area of Kano state; *Aspergillus niger* and potato dextrose agar (loxoid UK), were obtained from Department of Microbiology, Bayero University Kano;

Short horned grasshopper used for the preparation of chitosan was obtained from Maigatari international market, Jigawa State, Nigeria.

All other chemicals, like, Hydrochloric acid, (Aldrich), Acetic acid, glycerol (Aldrich), Sodium hydroxide were analytical reagents grades

obtained from analytical laboratory, Bayero University, Kano.

## **METHODS:**

### ➤ **Starch extraction and preparation of poly(lactic acid)**

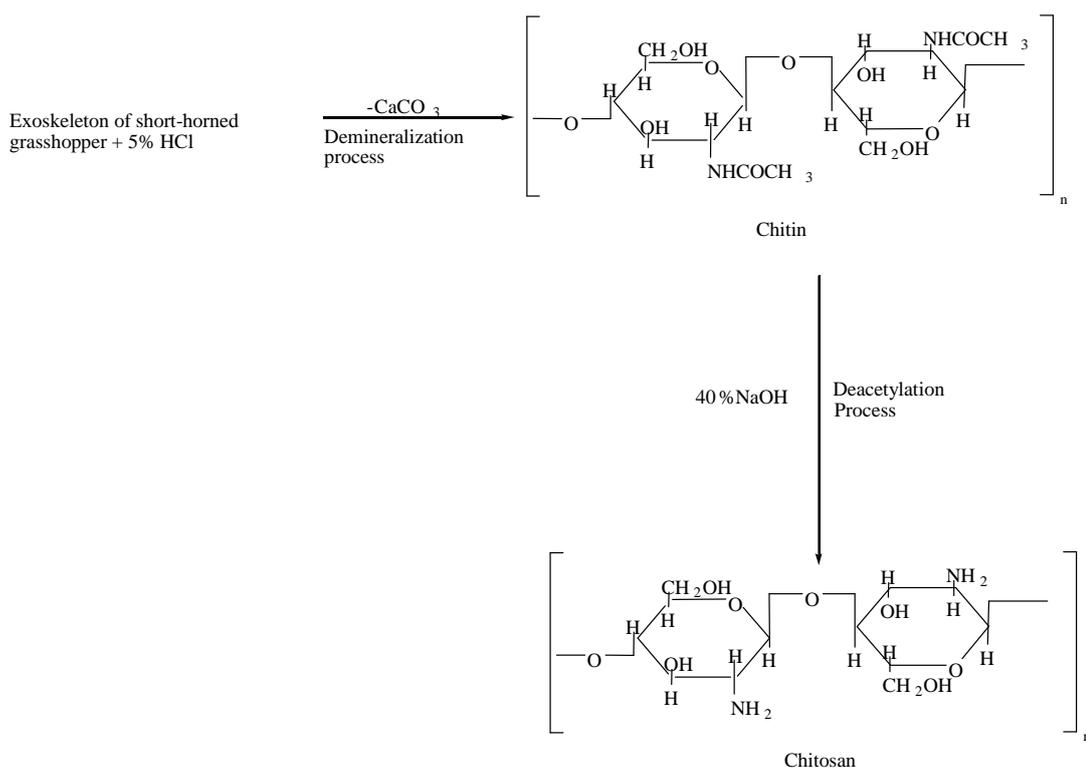
The potato peel (waste) was ground using mortar and pestle to obtain fine powder. The potato waste powder was dissolved in distilled water with solid to liquor ratio of (1:2, w/v). The mixture was filtered with Muslin cloth. The starch was separated from the liquid by decantation. The starch was collected and oven dried at 50°C for 24 hours. The dried powder was kept in an airtight container (Suryani *et al.*, 2017)

### ➤ **Extraction of chitosan from the exoskeleton of grasshopper**

Chitosan was extracted from the exoskeleton of short horned grasshopper by chemical treatments.

150g of the exoskeleton of grasshopper was treated with 5% HCl to remove the mineral content (demineralization process) at 30°C for 1h with solid to liquid ratio, (1:15 w/v). The mixture was washed in a running tap water and then rinsed 3 times with distilled water to neutrality. The paste was then treated with 3% NaOH at 30°C for 1 hour with material to liquid ratio (1:50) to remove the protein content (deproteinization). The resultant chitin was then treated with 50% NaOH solution at 120°C for 2hrs with solid to liquor ratio of (1:50 w/v). The mixture was stirred periodically for homogeneity. The resultant chitosan was washed to neutrality and oven dried at 50°C for 24hrs (Yaret *et al.*, 2018).

The chitosan was then purified by dissolving in 2% acetic acid for 30min. at 60°C under constant stirring and then re-precipitated using 20% NaOH with solid to liquid ratio (1:20 w/v). The pure chitosan was then washed to neutrality and oven dried at 50°C for 24hrs. The pure chitosan was then milled into fine powder with an electrical blender and kept in an airtight container.

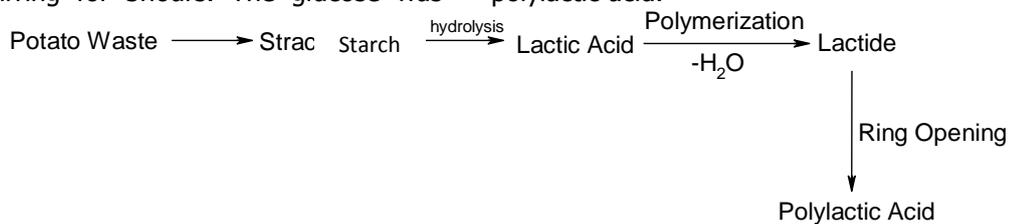


Scheme 1: Schematic representation of chitosan synthesis, Karim., (2018)

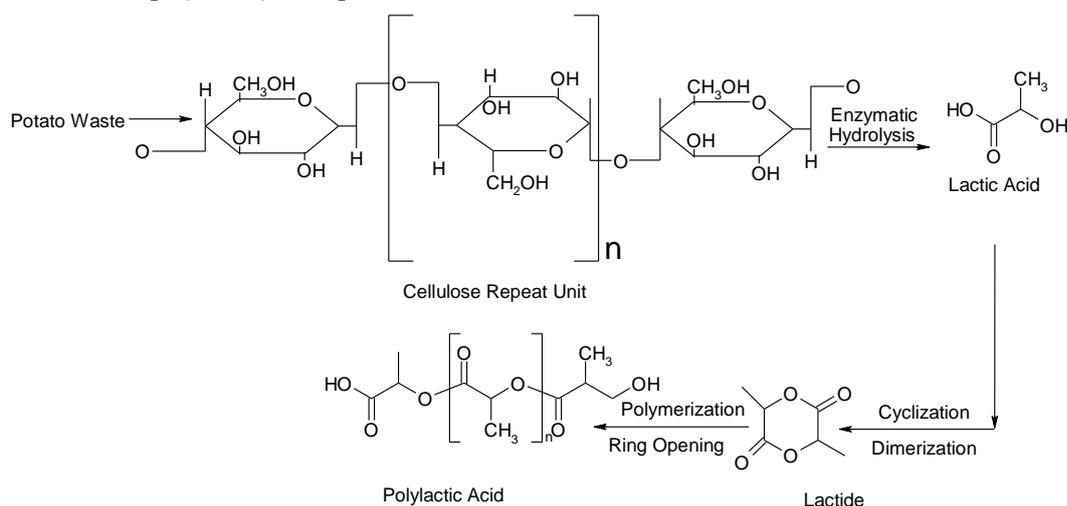
### Synthesis of Polylactic Acid

100g of the potato starch was hydrolyzed by the addition of 5% HCl. The mixture was heated on a hot plate at constant temperature of 60°C with constant stirring for 3 hours. The glucose was

then inoculated with *Lactobacillus bulgaricus* and incubated in an orbital shaker for 3-days to produce lactic acid (Akbar *et al.*, 2019) The lactic acid was then polymerized to form polylactic acid.



[Lin, *et al.*, 2012]



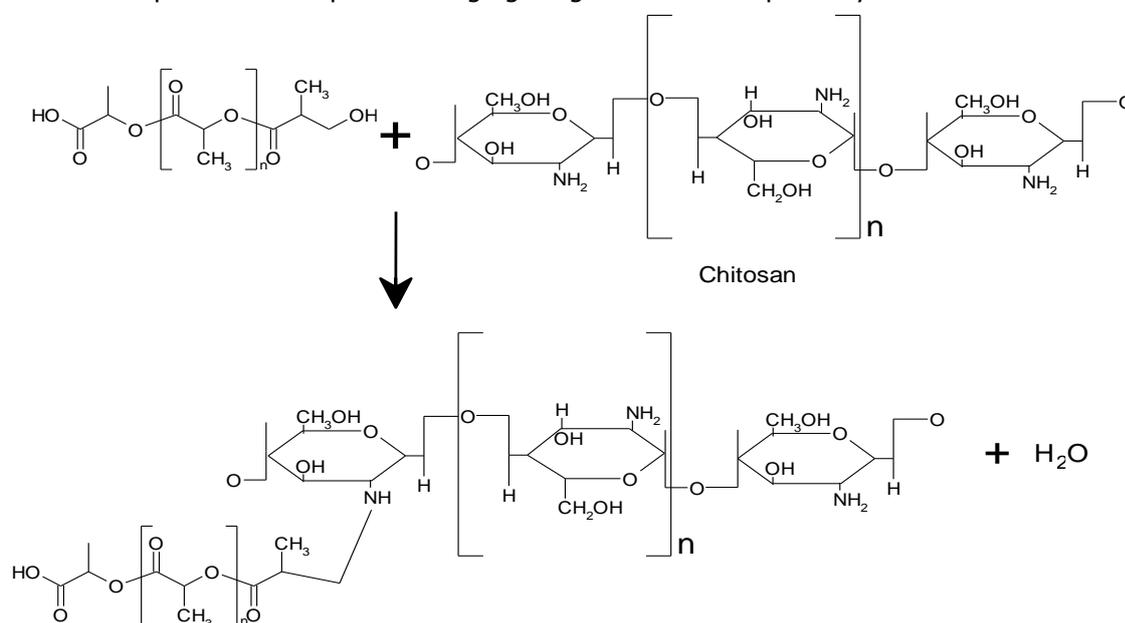
Scheme 2: Schematic representation of polylactic acid synthesis

➤ **Modification of polylactic acid with chitosan**

Poly (lactic acid) modification was carried out by amidation reaction between the amino groups, in the glucosamine unit of chitosan and the terminal carboxyl group of polylactic acids chains (Suryani *et al.*, 2017). 2g of the chitosan powder was added to the polylactic acid and the mixture was dissolved in 200ml of THF.

The two solutions were mixed and stirred at constant rate of 350rpm at 30°C for 3hrs (Formin *et al.*, 2017) The modified polylactic acids was purified by dialysis in deionized water for 48hours with water changes at 6hrs interval, after which the polylactic acids were recovered and freeze dried for 48hrs under vacuum atmosphere of 0.5 mBar at -46°C in the collector (Layla *et al.*, 2018).

The process was repeated using 4g & 6g of chitosan respectively



Scheme 3: Schematic representation of modification of polylactic acid with chitosan

**Synthesis of films of polyethylene and chitosan modified polylactic acid nano composites.**

The film of polyethylene and chitosan modified polylactic acid was prepared using 5%, 10% and 15% polyethylene (PE).

The polymer blends each for polyethylene with Ch-PLA were subjected to mechanical agitation for 30 minutes until a homogeneous mixture was obtained. The films were then poured in mould and oven dried at 50°C for 72hrs.

The films were characterized using FTIR, SEM and mechanical properties using tensile test equipment, universal testing machine (model exceed E. 43, Japan) and Enzymatic degradation to assess biodegradability (Karim, 2018).

**RESULTS AND DISCUSSION**

**FTIR SPECTRUM**

The FTIR spectrum of Chitosan, chitosan modified polylactic acid displayed the following absorption characteristics shown table 1:

Sample	Absorption band (cm <sup>-1</sup> )		Functional group
Chitosan	3 0 0	- 3 6 0 0	OH stretching & bending vibration
	3 4	5 7	NH stretching and bending
	1 6 5 9	- 1 7 5 9	C = O stretching
	1 5	4 7	NH <sub>2</sub> stretching
P L A	2 9 1 8 ,	2 8 7 3	CH <sub>2</sub> stretching
	3 3	6 5	OH stretching vibration
	1 7	6 0	C = O stretching and bending vibration in the amide
P L A - C h	2 8 0 0	- 3 0 0 0	CH - Stretching & bending vibration in the glucopyranose ring.
	3 4	5 0	CH Stretching and wagging
	1 5	5 4	OH stretching
	2 8	4 0	NH - Stretching in the 2 <sup>o</sup> amide confirming, Amidation reaction
	2 8	4 0	CH - S t r e t c h i n g

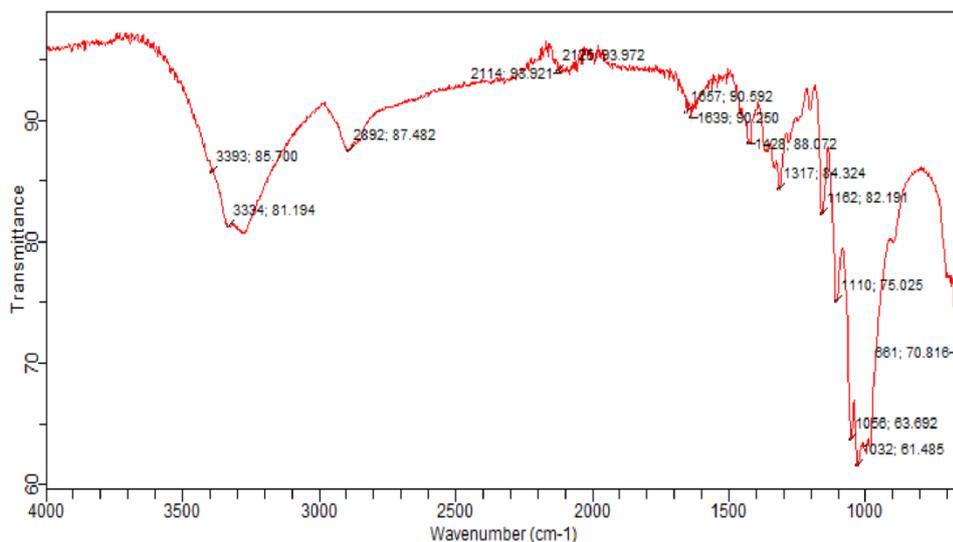


Figure 1a: FTIR spectrum of poly(lactic acid)

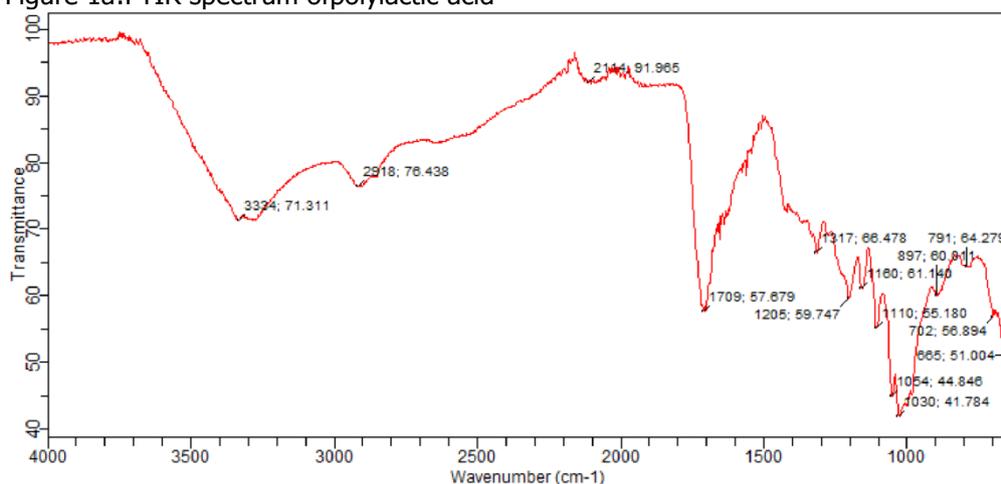


Figure 1b: FTIR Spectrum of Chitosan

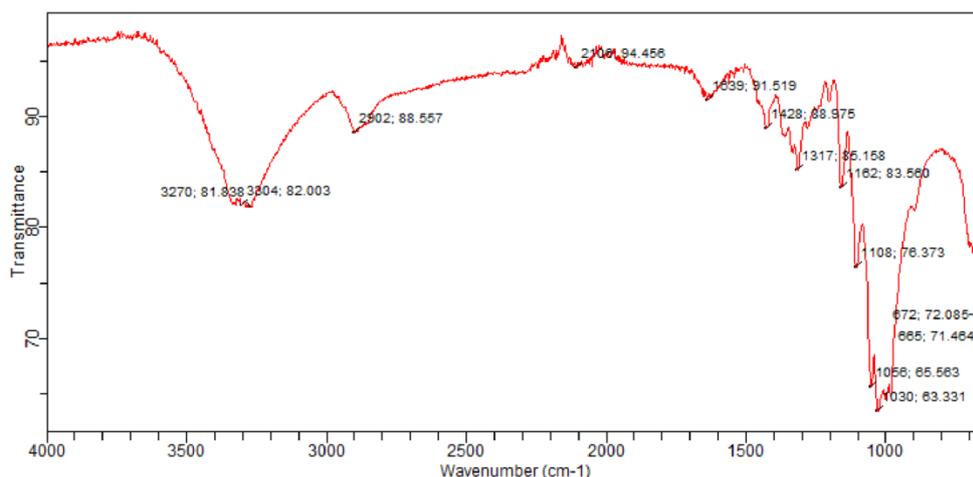


Figure 1c: FTIR spectrum of chitosan modified poly(lactic acid)

From table 1 and Figure (1a, 1b, and 1c) It was established that absorption bands in the region of 3000-2800cm<sup>-1</sup> was due OH stretching and bending vibration in both chitosan, PLA and PLA-Chit nanocomposites. While absorption peaks at 2800-3000cm<sup>-1</sup> were due CH stretching and bending vibration in the glucose pyranose rings of both chitosan, PLA & Ch-PLA nano composites. Whereas absorption bands at 1750-1760C was due to C=O stretching vibrations in both chitosan, PLA and the Ch-PL matrix. Another peak at 1428 – 1448 was due to CH-Bending vibration. An absorption at 1554cm<sup>-1</sup> was due to NH stretching vibration in the 2<sup>o</sup> amide confirming amidation reaction.

**Mechanical properties of polyethylene/PLA-Ch nanocomposites**

Analysis of tensile strength and elongation at break of the nano composites films yield a very good results. The results are depicted in table 2.

Table 2: Tensile testing values for Ch-PL/polyethylene nano composites

P.E (wt%)	S a m p l e	v o l u m e s	1 0 0 m l	&	2 0 0 m l
	Ch-PLA (wt%)	Tensile strength (PMA)	Elongation (%)		
5	95 in 100 ml	1 4 0 . 9 6 0 3 2 . 3 8			
1	0 90 in 100 ml	1 8 6 . 2 0 0 3 1 . 9 6			
1	5 85 in 100 ml	2 0 0 . 6 1 0 3 0 . 1 9			
5	95 in 200 ml	2 4 0 . 7 2 0 2 9 . 6 5			
1	0 90 in 200 ml	2 8 0 . 0 0 6 2 9 . 0 2			
1	5 85 in 200 ml	2 8 5 . 7 3 0 2 8 . 7 3			

From table 2; it was observed that the tensile strength of the composites increases as a factor of polylactic-chitosan ratio, which indicates that the higher the increases in PLA-Ch concentration (%) the higher the tensile strength and stiffer the composites films is.

Conversely, the elongation of the bioplastics to break decreases with increase in the concentration of Ch-PLA.

**Scanning Electron Microscopy of the Bioplastics**

The morphology of the Ch-PLA/polyethylene bioplastics showed a homogeneous surface independent of concentration of Ch-PLA or P.E in the composites.

The results may be attributed to the biocompatibility of the Ch-PLA to the polyethylene in the composites. The results is in conformity with Nurshahida and Zainoha (2016). The micrographs of Ch-PLA, PE/Ch-PLA and chitosan were shown in Figure 1.

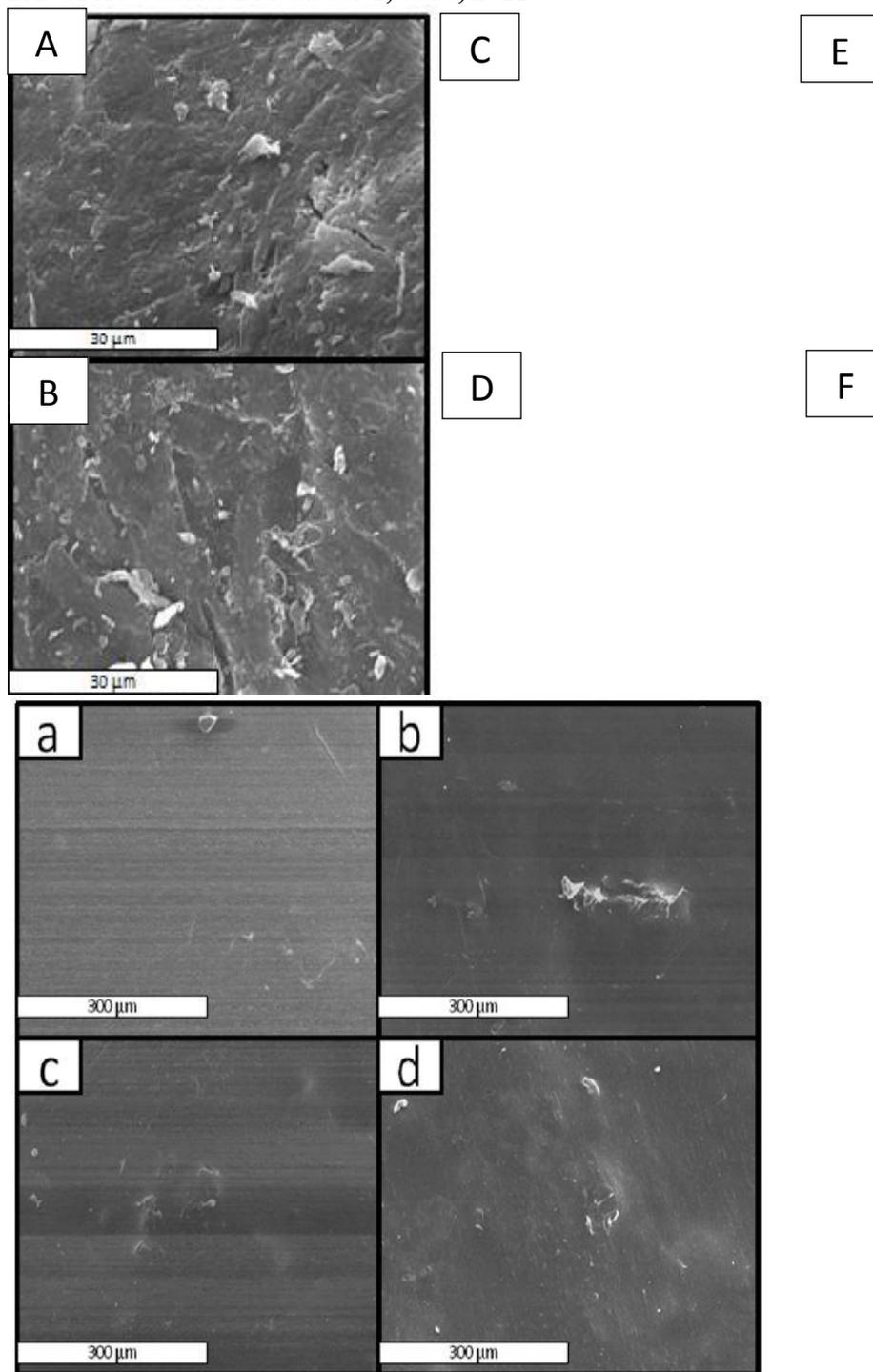


Figure 2: (a) Chitason (b) PLA (c) Ch-PLA (d) 5% PE/Ch-PLA (e)10% PE/Ch-PLA (f) 15% PE/Ch-PLA

**Biodegradability of the bioplastics**

Biodegradation of the bioplastics were observed to be a factor of Ch-PLA concentrations. The results establish that biodegradation of the bioplastics occur effectively at concentration dependent manner of Ch-PLA content in both the samples as show in Figure 1. The results in agreement with Akbar et al., (2019).

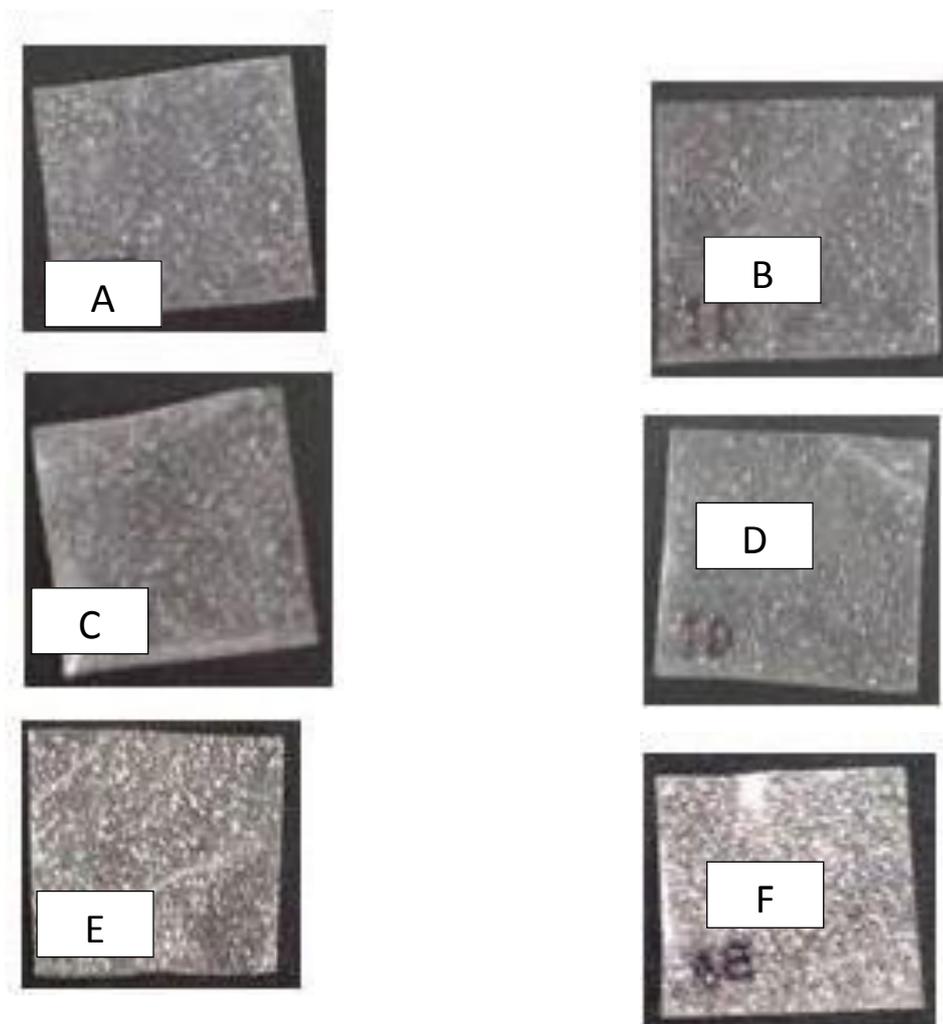


Figure 3: Biodegradable samples (a,c and e)Ch-PLA in 100ml (b,d, and f) Ch-PLA in 200ml vol.

### CONCLUSION

Biodegradable polyethylene/biorenewable polymers composites were successfully prepared using grasshopper chitosan and potato waste (peel) based polylactic acid matrix. Concentration of Ch-PLA affect both physical and mechanical properties of the bioplastics composites by increasing the tensile strength in Ch-PLA concentration dependent manner. While the elongation to break of the nanocomposite was found to decrease in Ch-PLA content. The biodegradability of the composites was enhanced by enzymatic degradation in Ch-PLA dependent manner. The results indicated that it

is very possible to manufacture films of synthetic polyethylene (P.E) with natural polymers Ch-PLA that is biodegradable as a means of utilization of potato waste and plastic waste in the industrial preparation of biodegradable plastics for food packaging.

### Acknowledgements

The authors acknowledge the intellectual contribution and untiring efforts given by Prof. Haruna Musa Danbatta during the conduct of this research and when preparing this manuscript. Equally acknowledged the parental courage and support by Prof. A.A Salis that lead to the success of this research.

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