



STUDIES ON THE FOLIAR EPIDERMAL TISSUES OF THREE SPECIES OF *Digitaria* Haller IN JOS, PLATEAU STATE NIGERIA

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ABSTRACT

Foliar epidermal studies were carried out on Digitaria iburua Stapf, Digitaria exilis Stapf and Digitaria barbinodis Henr. with the aim of determining the patterns of variation in their epidermal characteristics and assessing the value of leaf epidermal characters in the identification of these culturally important species. Micro hairs, silica bodies, short cells, anticlinal wall pattern, epidermal cell shape and stomata index were diagnostic characters. While micro hairs were distributed on the surfaces of Digitaria exilis and Digitaria barbinodi, they were absent in Digitaria iburua. Short cells were present on the surfaces of Digitaria iburua and Digitaria barbinodis but absent on Digitaria exilis. Stomata index was smallest in Digitaria barbinodis distinguishing it from the other two species. The epidermal cell shape varied from rectangular to polygonal while the anticlinal wall pattern varied from straight to wavy in the three species. Other useful characters for distinguishing the species are epidermal cell size, stomata size and stomata number.

Keywords: *Digitaria iburua, microhairs, foliar, silica bodies, Digitaria exilis*

INTRODUCTION

The grass family, Poaceae, is noted for its wide diversity and complexity and so has posed many problems to the taxonomists using the traditional method based on gross morphology (Srivastava, 1978). The family is one of the largest and is economically the most important of all plants that cover the earth; being classified into about 50 tribes, 908 genera and more than 11,000 species (GPWG, 2001; Lowe, 1989). In West Africa, there are approximately 147 genera and 615 species (Gill, 1988).

The genus *Digitaria* Haller comprises annual and perennial grass species and is almost confined to the tropical and warm temperate regions of the world, frequently forming an important part of the savannah grasses in the tropics. According to Hutchinson and Dalziel (1972), there are 26 species of *Digitaria* in West Africa, 18 spp. of which also occur in Nigeria. *Digitaria exilis* Stapf (white fonio) is a staple food in various parts of West Africa and probably the oldest African cereal. It is the most culturally diverse and widely cultivated species in West Africa, locally called Acha, Ipouaga, Feningué, Findi, Kansambahon and Ova in Nigeria, Benin, Burkina Faso, Guinea, Mali and Togo respectively (Adoukonou-Sagbadja, 2010). *Digitaria iburua* Stapf (black fonio) is cultivated as a cereal in scattered localities from Côte d'Ivoire to northern Nigeria and southern Niger, and in Cameroon. It has also been reported to be grown in Guinea and DR Congo. Black fonio is only known from cultivation. Its origin is uncertain, but it may have been derived from *Digitaria ternata* (A.Rich.) Stapf (Portères, 1976). *Digitaria barbinodis* Hanr. is recorded only in Mali and Northern Nigeria (Burkill, 1994).

The black and white fonio require minimal processing due to grain size and location of

constituents. The starch of *Digitaria exilis* can be employed as an alternative binder to maize starch binding property in the formulation of paracetamol tablets (Musa *et al.*, 2008). Due to its low free sugar and low glycemic content, *Digitaria* species is recommended for lactating women, diabetic people and often used in diets of sick people (Kwon-Ndung *et al.*, 2003). The straw of *D. exilis* can be chopped and mixed with clay for building houses, walls burnt and the ash extracted with water to make potash and a source of heat for cooking (Hag and Ogbe, 1995). Black fonio is a staple food of the Birom people of the Jos Plateau in Northern Nigeria and an important supplementary food to people in the Atakora mountains of Togo and Benin. It is eaten as porridge or mixed with meal of other cereals. The grain is also eaten cooked like rice or in stews. In Benin and Nigeria black fonio is made into couscous types (wusu-wusu). In Togo it is used for brewing beer ('tchapalo') (Irvine, 1974).

Digitaria iburua Stapf, *Digitaria exilis* Stapf and *Digitaria barbinodis* Hanr. are very similar morphologically, often confused (Hutchinson and Dalziel, 1972). Their identification depends principally on their seed morphology which has seasonal occurrence. This often takes some time to develop, therefore studies have to wait for a considerable length of time for the plant to mature and the organ to be available. The leaf epidermis is described as an important character after cytology for solving this taxonomic problem (Stace, 1984; Srivastava, 1978).

Metcalfe (1960) studied abaxial epidermal anatomy of *Digitaria borbonica* Desv., *D. brazzae* (Franch.) Stapf, *D. horizontalis* Willd., *D. nzilanjana* (Rendle) Stapf and *D. wallichiana* (Wight & Am.) Stapf. Webster (1983) revised the genus *Digitaria* of Australia.

He studied morphological as well anatomical characters of 38 species of *Digitaria*. While studying the leaf epidermal anatomy of the genus, he examined the abaxial leaf surface only. Several studies have also been conducted on other genera and tribes belonging to the family Poaceae using leaf epidermal and anatomical features to aid their identification and classification (Sharma and Salam, 1984; Sharma and Mittal, 1985; Gill and Mensah, 2001; Kharazian, 2007; Folorusno and Olaniyan, 2009).

The present investigation aimed to explore the possible use of microscopic characters of the leaf to identify samples of these very similar but economically important and culturally diverse grass species. The study will be useful in the determination of the patterns of variation in epidermal characteristics, assessing their value in species identification and classification, and also in establishing the taxonomic relationships among the species studied.

MATERIALS AND METHODS

The study was carried out in the Botany Laboratory, Department of Plant Science & Technology, University of Jos. Fresh leaf samples of three *Digitaria* species (*Digitaria exilis*, *Digitaria barbinodis* and *Digitaria iburua*) were collected from the botanical garden, University of Jos (9.933°N 8.883°E) and Riyom Local Government Area of Plateau state (9.567°N 8.667°E). Preparation of leaf samples for permanent slides to enhance epidermal morphology follows the method of Wilkinson (1979), with slight modifications. The leaf samples were soaked in concentrated nitric acid for three to five hours in order to remove the colouring pigments and surface debris followed by washing in several changes of water to remove excess reagents. Using the fine grade carmel hair brush, epidermal peels were carefully removed from the leaf sample surfaces and cleared properly. The cleared epidermal peels were stained in aqueous solution of Safranin O for 4-8 minutes, rinsed carefully in water to remove excess stain and mounted in Canada balsam.

The slides of both abaxial and adaxial surfaces of the leaves were prepared, labelled, viewed for micromorphological characters with the digital Olympus BX 51 light microscope and photomicrographs were captured on the computer. Where measurements were taken, in the cases of stomata length and width and the epidermal cells, the range corresponds to two major levels of discontinuity (smallest and highest values). Specimens were observed at X400 objective magnification.

The stomata index was determined according to Metcalfe and Chalk (1979) using the formula:

$$\frac{S}{E+S} \times 100 \text{ stomata index (S.I)}$$

Where S = Number of stomata per unit area and E = number of epidermal cells in the same area

RESULTS

The results obtained from the study of the surface features of *Digitaria* species are shown in Plates 1a-3b as photomicrographs and tabulated in Tables 1 and 2. Figure 1 is the dendrogram of possible phyletic relationships among the three species. Anatomical description of the foliar epidermal characters with

specific references to stomata complex, epidermal cell complex, silica bodies, short cells and micro hairs, for each species, are described below.

***Digitaria iburua* Stapf**

The distribution of stomata was epiamphistomatic (stomata abundant on the adaxial surface and scanty on the abaxial epidermis). Large numerous paracytic stomata type with dome shaped subsidiary cells were observed on both surfaces (Plate 1a). The stomata size on the abaxial surface measured 30-37.4µm long and 20.4-23.8µm wide while stomata size measuring 34.0-37.4µm long and 20.4µm wide was found on the adaxial surface (Table2). The stomata index on the abaxial and adaxial surfaces was 19.34% and 29.3% respectively.

The epidermal cell shape was rectangular on both surfaces while the anticlinal wall pattern was highly wavy in the abaxial surface but slightly wavy in the adaxial surface. The epidermal size on the abaxial measured 53.0-158.6µm long and 20.4-34.0µm wide while epidermal size measuring 68.0-142.8µm long and 20.4-27.2µm wide was found on the adaxial surface (Table 2). Micro hairs were absent on both surfaces. Numerous short cells and dumb-bell shaped silica bodies were seen in the intercoastal zones on both surfaces.

***Digitaria exilis* Stapf**

The stomata distribution was hypoamphistomatic (stomata abundant on the adaxial surface and scanty on the abaxial epidermis). Large numerous paracytic stomata type with dome shaped subsidiary cells present (Plate 2a). The stomata size on the abaxial measured 30.6-37.4µm long and 23.8-30.6µm wide while stomata size measuring 30.6-40.8µm long and 23.8-30.6µm wide was found on the adaxial surface (Table 2). The stomata index on the abaxial and adaxial surfaces was 24.2% and 17.8% respectively. The epidermal cell shape on both surfaces was long and polygonal and the anticlinal wall pattern was straight. The epidermal size on the abaxial surface measured 74.8-136.0µm long and 20.4-34.0µm wide while epidermal size measuring 74.8-129.2µm long and 34.0-44.2µm wide was seen on the adaxial surface (Table 2). Micro hairs and dumb-bell shaped silica bodies were present on both surfaces while short cells were absent.

***Digitaria barbinodis* Henrard**

Stomata distribution was epiamphistomatic. Paracytic stomata type, numerous on the adaxial than on the abaxial surface was observed. The stomata type observed on both surfaces was paracytic with triangular dome shaped subsidiary cells (Plate 3a). The stomata size on the abaxial measured 27.2-34.0µm long and 23.8µm wide while stomata size measuring 23.8-30.6µm long and 20.4-23.8µm wide was found on the adaxial surface (Table2). The stomata index on the abaxial and adaxial surfaces was 4.36% and 8.5% respectively. On both surfaces, the epidermal cell shape was rectangular while the anticlinal wall pattern was straight. The epidermal size on the abaxial measured 23.8-30.6µm long and 20.4-23.8µm wide while epidermal size measuring 54.4-129.2µm long and 34.0-37.4µm wide was seen on the adaxial surface (Table2).

Short cells were present on the adaxial surface but absent on the abaxial surface while dumb bell silica bodies were present on the intercoastal zones on both surfaces. Micro hairs were also observed on both surfaces.

TABLE 1: QUALITATIVE EPIDERMAL CHARACTERS EXTRACTED FROM THE MICROGRAPHS OF THE STUDIED SPECIES

SPECIES	LEAF SURFACE	STOMATA TYPE	EPIDERMAL CELL SHAPE	ANTICLINAL WALL PATTERN	SHORT CELL	SILIA BODIES	MICRO HAIRS
					+/-	+/-	+/-
<i>D. iburua</i>	AB	Paracytic	Long rectangular	Highly wavy	+	+	-
	AD	Paracytic	Long rectangular	Slightly wavy	+	+	-
<i>D. exilis</i>	AB	Paracytic	Polygonal	Straight	-	+	+
	AD	Paracytic	Polygonal	Straight	-	+	+
<i>D. barbinodis</i>	AB	Paracytic	Short rectangular	Straight	-	+	+
	AD	Paracytic	Short rectangular	Straight	+	+	+

TABLE 2: QUANTITATIVE EPIDERMAL CHARACTERS EXTRACTED FROM THE MICROGRAPHS OF THE STUDIED SPECIES

SPECIES	LEAF SURFACE	STOMATA LENGTH (µm)	STOMATA WIDTH (µm)	EPIDERMAL LENGTH (µm)	EPIDERMAL WIDTH (µm)	STOMATA NUMBER	EPIDERMAL NUMBER	STOMATA INDEX (%)
<i>D. iburua</i>	AB	30.0(34.04±1.81)37.4	20.4(21.70±1.29)23.8	53.0(124.26±11.12)158.6	20.4(24.82±2.08)34.0	4.0(7.0±1.57)11.0	23.0(29.0±2.09)34.0	19.34
	AD	34.0(35.36±1.24)37.4	20.4(20.4±0.00)20.4	68.0(108.12±4.89)142.8	20.4(24.8±1.86)27.2	10.0(13.0±1.36)15.0	29.0(31.0±1.31)34.0	29.3
<i>D. exilis</i>	AB	30.6(35.7±1.66)37.4	23.8(26.86±1.54)30.6	74.8(110.76±4.29)136.0	20.4(25.5±2.07)34.0	6.0(10.0±1.55)13.0	26.0(29.0±1.5)33.0	24.62
	AD	30.6(36.38±1.85)40.8	23.8(26.52±1.43)30.6	74.8(102.0±4.11)129.2	34.0(30.06±2.8)44.2	5.0(6.0±1.21)9.0	22.0(29.0±2.0)35.0	17.8
<i>D. barbinodis</i>	AB	27.2(32.20±1.51)34.0	23.8(23.8±0.00)23.8	23.8(25.84±1.65)30.6	20.4(22.44±1.29)23.8	1.0(3.0±1.16)5.0	64.0(75.0±2.6)84.0	4.36
	AD	23.8(25.84±1.65)30.6	20.4(22.44±1.29)23.8	54.4(92.86±4.58)129.2	34.0(34.68±1.17)37.4	4.0(7.0±1.31)9.0	64.0(73.0±2.3)81.0	8.5

Key: Min. (mean ± standard error) max.

AB: Abaxial

AD: Adaxial

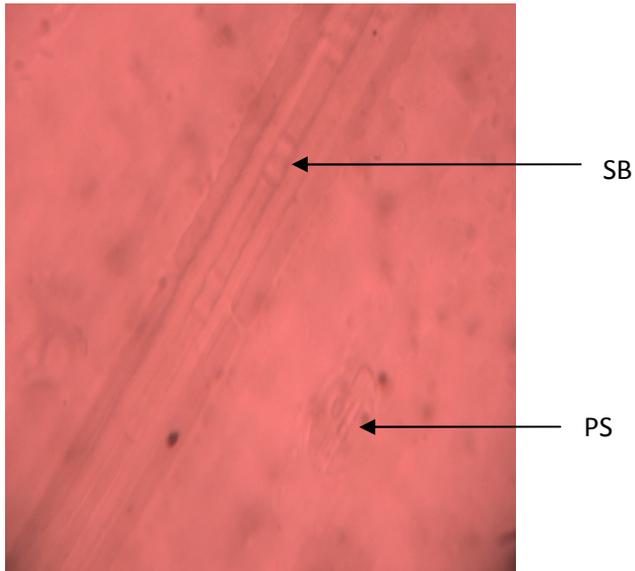


Plate 1a

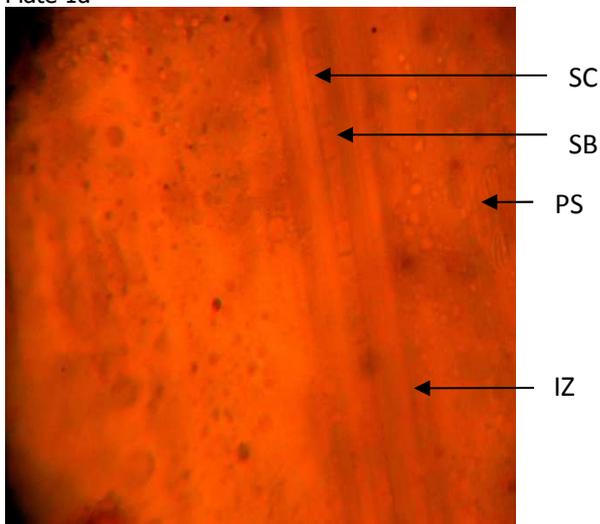


Plate 1b

Plates 1a and 1b: Abaxial and adaxial surfaces of *D. iburua*

Abbreviations: Paracytic stoma (PS), silica body (SB) and intercoastal Zone (IZ), short cell (SC)

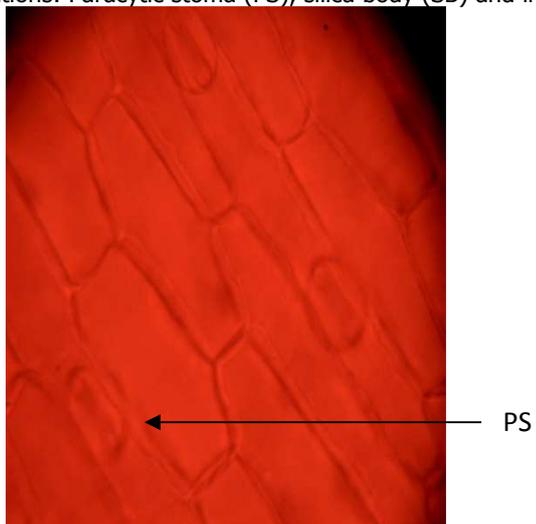


Plate 2a

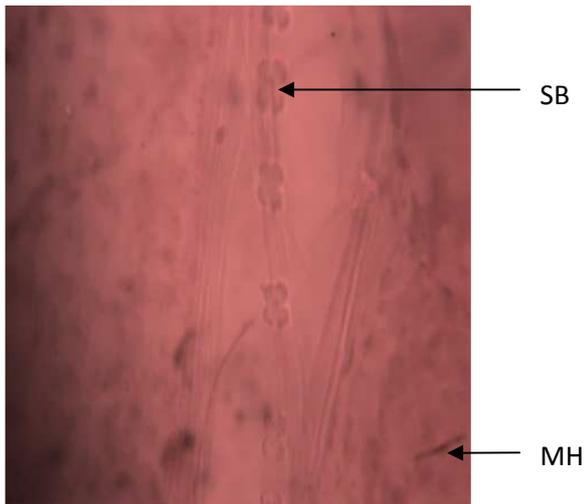


Plate 2b

Plates 2a and 2b: Adaxial and abaxial surfaces *D. exilis*

Abbreviations: Paracytic Stoma (PS), micro hairs (MH), silica bodies (SB)

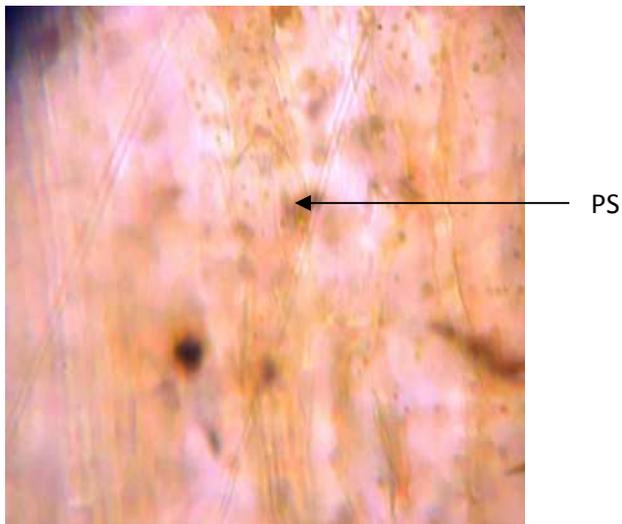


Plate 3a

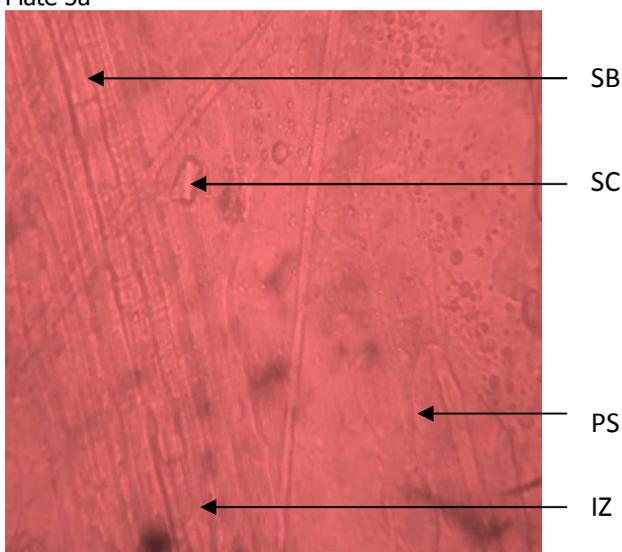


Plate 3b

Plates 3a and 3b: Abaxial and adaxial surfaces of *D. barbinodis*

Abbreviations: Paracytic stoma (PS), intercoastal Zone (IZ), silica bodies (SB)

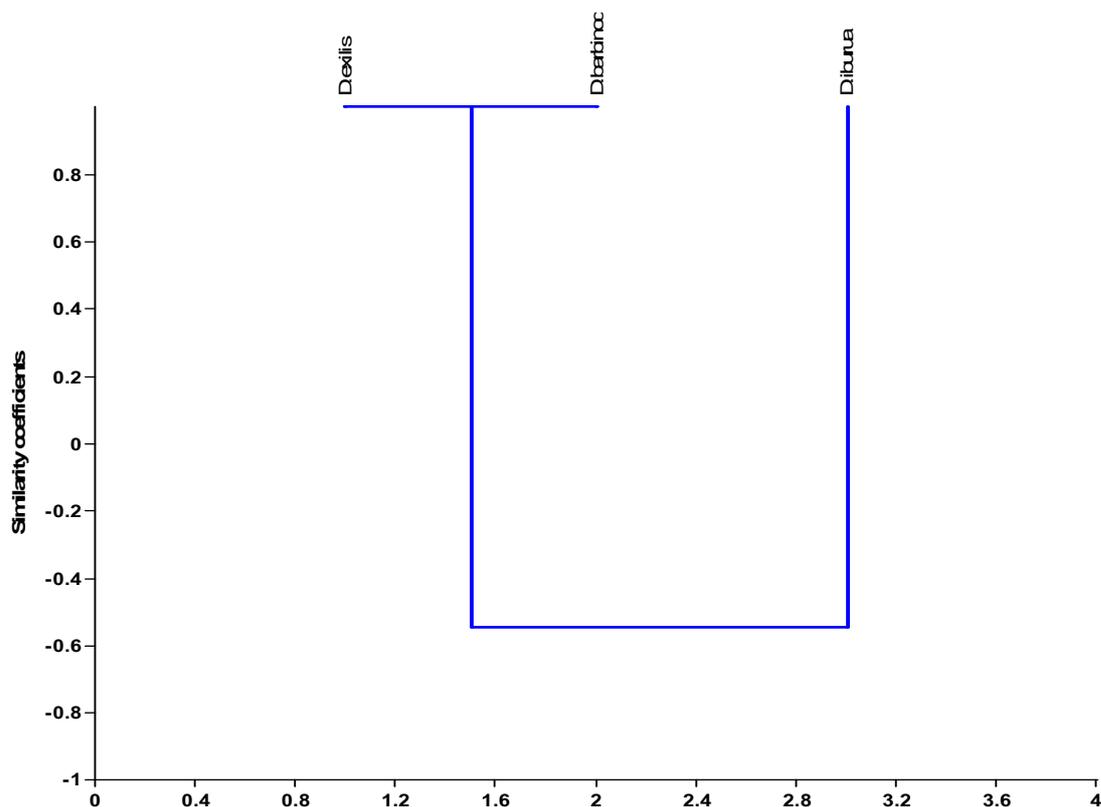


Fig. 1: Dendrogram of relationships among the *Digitaria* species studied based on epidermal foliar characters.

A study of the dendrogram in Figure 1 reveals that *D. exilis* and *D. barbinodis* are very similar, almost inseparable while *D. iburua* appears distant from both.

DISCUSSION

Broadly, the leaf epidermal attributes of the three species are quite similar regarding the length and width of the stomata, while the stomata index is quite different. In this study, the stomata index is helpful in species delimitation; thus separating the species into two groups. *Digitaria iburua* and *Digitaria exilis* with stomata index greater than 10% and *Digitaria barbinodis* having stomata index less than 10%. Paracytic stomata type was observed which had dome shaped or triangular subsidiary cells. Triangular subsidiary cells were observed in *Digitaria barbinodis* while dome shaped subsidiary cells were observed in *D. iburua* and *D. exilis*. This is in agreement with the study of Syed *et al.* (2002) which reported the presence of paracytic stomata type with dome shaped, parallel or triangular subsidiary cells in the *Digitaria* species studied. Metcalfe and Chalk (1954) pointed out that certain characters of the epidermis of Poaceae such as shape of the subsidiary cells of the stomata, micro hairs, short cells and silica bodies are important systematically. Metcalfe and Chalk (1979) and Aworinde *et al.* (2009) reported that stomata complex is highly constant for a given species and can be used in species identification.

Furthermore, the shape of the epidermal cells and the anticlinal wall pattern vary between the species studied and can be used as distinguishing tools with *Digitaria iburua* possessing rectangular

epidermal cell shape and wavy anticlinal wall pattern, *Digitaria exilis* having long polygonal cell shape and straight anticlinal walls pattern while rectangular cell shape and straight anticlinal wall pattern were observed in *Digitaria barbinodis*. This is partly in agreement with the study of Farooq (2009) which reported polygonal cell shape and sinuous anticlinal wall pattern in the *Digitaria* species studied.

The closeness observed in the dendrogram between *Digitaria exilis* and *Digitaria barbinodis* is in consonance with previous studies. The presence of micro hairs on the surfaces of *Digitaria exilis* and *Digitaria barbinodis* distinguished them from *Digitaria iburua* that lack micro hairs, in agreement with the findings of Farooq (2009) which reported the presence of micro hairs in some *Digitaria* species. Ogei-odia *et al.* (2010) stressed the importance of micro hairs as diagnostic character. In this study, the presence or absence of micro hairs can be useful in distinguishing the species. Other characteristic features of the leaf epidermis were short cells which were present on both the adaxial and abaxial surfaces of *Digitaria iburua*, but only observed on the adaxial surface of *Digitaria barbinodis* while absent on the surfaces of *Digitaria exilis*, thereby providing a diagnostic tool to distinguish between these species. This is partly in agreement with the findings of Syed *et al.* (2002), which reported the presence of short cells in the three *Digitaria* species studied.

The presence of dumb bell-shaped silica bodies observed between the intercoastal zones on the three species studied appears to be synapomorphic for the species which thus supports the reported similarities (Hutchinson and Dalziel, 1972). This is in agreement with Farooq (2009) and Freire *et al.* (2005) who also observed presence of dumb bell-shaped silica bodies in the species studied.

CONCLUSION

Leaf epidermal studies have proved to be very important in providing information of taxonomic importance. Diagnostic characters for distinguishing the species include presence or absence of micro hairs, epidermal cell shape, anticlinal wall pattern, stomata index and presence or absence of silica bodies. Thus, micromorphological characters

presented in the current study sustain the difference between the three species on the basis of a key given below:

Silica bodies present on both surfaces in the three species

1. micro hairs present..... 2
- 2 short cells only on abaxial surface *Digitaria barbinodis*
- 2 short cells on both adaxial and abaxial surfaces *Digitaria exilis*

1. micro hairs absent.....*Digitaria iburua*

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