

SYNTHESIS AND CHARACTERIZATION OF NOVEL SULFONAMIDES DERIVATIVES AND THEIR ANTIMICROBIAL, ANTIOXIDANT AND CYTOTOXICITY EVALUATION

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ABSTRACT. Five novel sulfonamides derivatives **HR5-HR8** and **HR14** were synthesized by sulfonylation of primary or secondary amine in the presence of base through nucleophilic substitution reaction. Structural elucidation was carried out through FT-IR, UV, ¹H NMR, MS and elemental analysis. Prepared compounds were evaluated against pathogenic strains of bacteria (*S. aureus* and *E. coli*) and fungi (*A. flavous* and *A. niger*). Results were compared against standard antifungal and bacterial drug already available in market (isiconazole and sulfamethoxazole). It was found that compound **HR14** showed good activity with MIC 1.5 µg/mL and 2.0 µg/mL for *S. aureus* and *E. coli*, respectively. While **HR5** showed best antifungal activity with zone of inhibition 27.2±0.12 mm (MIC: 5.25 µg/mL) and 18.1±0.12 mm (MIC: 12.5 µg/mL) against *A. flavous* and *A. niger*, respectively. Synthesized compounds were also tested for their *in vitro* antioxidant activity by using DPPH. Amongst all compounds **HR5** was found to have potential activity with 15.60% antioxidant activity at 6 mM concentration.

KEY WORDS: Sulfonamide, DPPH, AntiMICrobial activity

INTRODUCTION

Sulfonamides drugs have been used as preventive agents in chemotherapy against various diseases [1]. More than 30 drugs having sulfa drug as a functional group are in clinical use, such as antibacterial [2], antifungal [3], antiprotozoal [4], anti-inflammatory [5], and translational initiation inhibitors [6]. More recent use of sulfonamides are as an anticancer agent [7], antiviral HIV [8], and in Alzheimer's disease [9]. They are used effectively for the treatment of ulcerative colitis [10], urinary, intestinal and ophthalmic infections and also for obesity [11]. Beside their vital role in human medicine they are also showing their promising importance in field of veterinary and agricultural sciences. Due to presence of SO₂NH- group most important role of sulfonamide in medicinal field is as an antibacterial agent. Synthesis of bacterial DNA and RNA requires tetrahydrofolate as a co-factor, which is inhibited by sulfonamides, so production of new DNA and RNA dropped from lack of tetrahydrofolate which eventually decayed bacteria. Newer sulfonamides and their derivatives has obtained great attention in pharmaceutical field in order to compete life threatening issues caused by drug resistant strains of bacteria, i.e. *Methicillin* resistance as they have unusual ability of acclimatization against stress caused by antibiotics [12]. Disease causing organisms become much resistant when treated medically with routine antibiotic drug molecule, with appearance of additional species as per mutation, conjugation, transduction or transformation. So synthesis of new sulfonamides and

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their derivatives have got more attention from researchers for its application in the field of medicine sciences and medical chemistry. In the present study five sulfonamide derivatives have been synthesized by the reaction of *p*-toluene sulfonyl chloride with NH₂ group containing drugs such as ceftriaxone, cefepime, nicotinamide (vitamin B), cefadroxil, and nimsulide, respectively and their biological activities were evaluated by using bacterial and fungal strains such as *Escherichia coli*, *Aspergillum niger* and *Aspergillum flavus*.

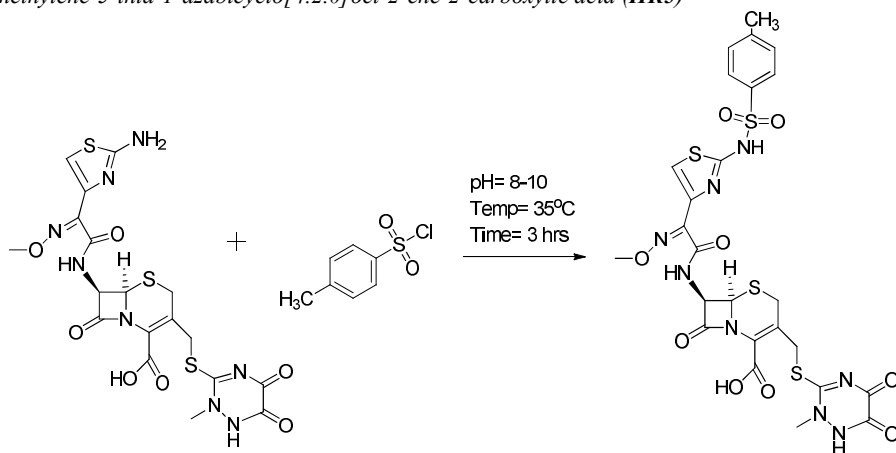
EXPERIMENTAL

¹H NMR spectra were conducted on Bruker 400 MHz spectrometer in DMSO-*d*₆ with tetramethylsilane as internal standard. MS data was recorded on Finnigan MAT 112 mass spectrometer. Elemental analysis was performed by using Perkin Elmer elemental analyzer. Melting points were taken on Gallenamp MP Apparatus MP70. Infrared spectra were recorded on Cary 630 Agilent FT-IR in the range between 4000-600 cm⁻¹. Absorption spectra were recorded by PGT90+ UV-Vis spectrophotometer.

General procedure

In this work efficient method based on Hinsberg test was used for preparation of sulphonamides, i.e. sulfonylation of primary or secondary amine in presence of base resulting in nucleophilic attack by amine. For sulfonylationtosyl chlorides were used [13] and base were used for neutralization of generated HCl, i.e. pyridine in synthesis of sulfonylmethylamide [14]. In the present work base sodium carbonate was used for neutralization of HCl. It was a one pot reaction, amine containing drug (0.001 M) in water was stirred and pH was noted, then equimolar sulfonyl chloride was added and mixture was allowed to stir for 2 hours and pH was monitored. Precipitates were separated by filtration and were purified by preparatory thin layer chromatography.

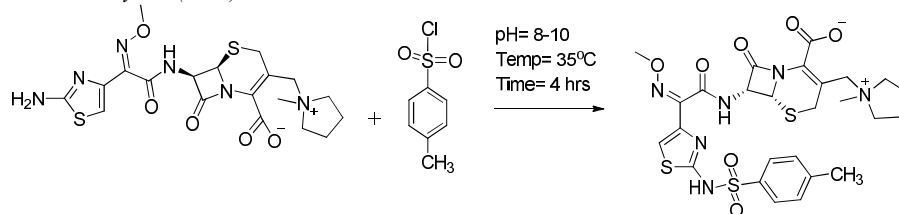
Synthesis of (6R,7R)-7-((Z)-2-(methoxyimino)-2-(2-(4-methylphenylsulfonamido)thiazol-4-yl)acetamido)-3-((2-methyl-5,6-dioxo-1,2,5,6-tetrahydro-1,2,4-triazin-3-yl)thio)methyl)-8-methylene-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylic acid (HR5)



Title compound **HR5** was prepared by above mentioned general procedure and was purified by using mobile phase DCM:EtOH, 50:50 with 83% yield. UV-Vis, λ_{max} : 270 nm. FT-IR (cm⁻¹):

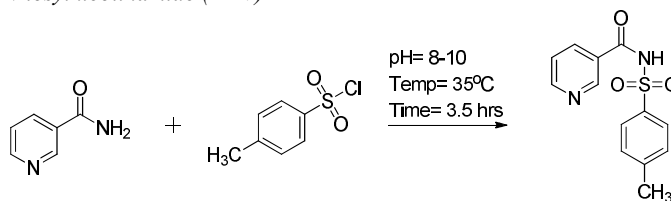
1178.19 (S=O str), 1037.18 (C–N str), 1633.80 (C=O), 846.97 (C–S str), 945.04 (S–N str), 3395.40 (–OH). $^1\text{H NMR}$ (400 MHz, DMSO- d_6 , δ): 11.22 (1H, s, –COOH), 8.36–8.47 (1H, d, J = 6.5, –CHNHCO), 8.11 (1H, s, –CONHNCH₃), 7.39–7.47 (2H, d, m, ArH), 7.22 (1H, s, –CHS–), 7.05–7.11 (2H, m, ArH), 5.68–5.76 (1H, d, J = 6.3, –CHNHCO–), 5.47–5.50 (1H, d, J = 1.2, –CHH=CH–), 5.12–5.23 (1H, d, J = 6.2, –NCHS–), 4.70–4.73 (1H, d, J = 1.1, –CHH=CH–), 4.56 (2H, s, –CH₂SC=N–), 4.15 (1H, s, –NHSO₂–), 3.92 (3H, s, –OCH₃), 3.47–3.56 (1H, d, J = 6.5, –CHHS–), 3.31 (3H, s, –NCH₃), 3.08–3.17 (1H, d, J = 6.3, –CHHS–), 2.49 (3H, s, –ArCH₃); MS (m/z , ESI): calcd. for C₂₆H₂₆N₈O₈S₄ [M+H]⁺ 706.0210312; found 706.0210615. Anal. calcd. For C₂₆H₂₆N₈O₈S₄: C, 42.36; H, 3.4; N, 15.81; O, 20.32; S, 18.10. Found: C, 42.16; H, 3.46; N, 15.84; O, 20.17; S, 18.13.

Synthesis of (6R,7R)-7-((Z)-2-(methoxyimino)-2-(2-(4-methylphenylsulfonamido)thiazol-4-yl)acetamido)-3-((1-methylpyrrolidin-1-ium-1-yl)methyl)-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-2-ene-2-carboxylate (HR6)



Designated compound **HR6** was prepared by following the general procedure mentioned earlier and was purified by using mobile phase DCM:EtOH, 40:60. Compound was obtained with a good yield, 85%. λ_{max} 280 nm. FT-IR (cm^{-1}): 1158.15 (S=O str), 1045.15 (C–Nstr), 1649.55 (–N–C=O), 825.38 (C–S str), 930.60 (S–N str), 3378.15 (–OH). $^1\text{H NMR}$ (400 MHz, DMSO- d_6 , δ): 8.01–8.03 (1H, d, J = 6.1, –CHNHCO–), 7.62–7.65 (2H, d, J = 8.3, –ArH), 7.44 (1H, s, –C=CHS–), 7.32–7.35 (2H, d, J = 8.2, –ArH), 5.52–5.58 (1H, d, J = 6.3, –NHCHCO–), 5.08–5.10 (1H, d, J = 5.9, –SCHNCO–), 4.28 (1H, s, –NHSO₂–), 4.01 (3H, s, –OCH₃), 3.81 (2H, s, –CH₂NCH₃), 3.61 (1H, s, –CH₂NCH₃), 3.37–3.48 (4H, t, J = 7.5, CH₃NCH₂CH₂–), 3.20–3.29 (1H, d, J = 6.5, –CHHS–), 3.07–3.14 (1H, d, J = 6.4, –CHHS–), 2.49 (3H, s, –ArCH₃), 1.44–1.60 (4H, q, J = 6.9, CH₃NCH₂CH₂CH₂–); MS (m/z , ESI): calcd. for C₂₆H₃₀N₆O₇S₃ [M+H]⁺ 634.1094327; found 634.0923741. Anal. calcd. for C₂₆H₃₀N₆O₇S₃: C, 49.20; H, 4.76; N, 13.24; O, 17.64; S, 15.5. Found: C, 49.35; H, 4.75; N, 13.23; O, 17.63; S, 15.16.

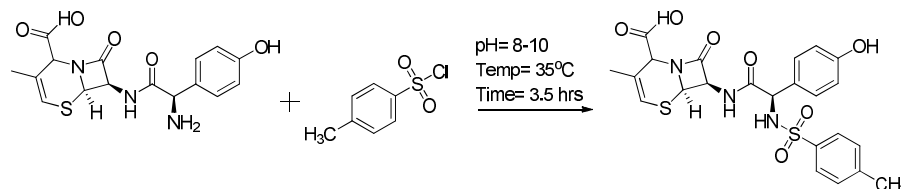
Synthesis of N-tosylnicotinamide (HR7)



Following above mentioned procedure labeled compound **HR7** was purified by using mobile phase DCM:EtOH, 60:40 having 76% yield. λ_{max} 285 nm. FT-IR (cm^{-1}): 1150.35 (S=O str), 1055.12 (C–N str), 1641.45 (C=O), 815.32 (C–S str), 930.16 (S–N str), 3388.12 (–OH). $^1\text{H NMR}$ (400 MHz, DMSO- d_6 , δ): 9.24 (1H, s, –ArH), 8.84–8.88 (1H, d, J = 8.3, –ArH), 8.51–8.54 (1H, d, J = 8.2, –ArH), 8.15 (1H, s, –NHSO₂–), 7.95–7.97 (2H, d, J = 8.1, –ArH), 7.68–7.71 (1H, t, J = 6.4, –ArH), 7.40–7.42 (2H, d, J = 8.2, –ArH), 2.33 (3H, s, –ArCH₃). MS (m/z , ESI): calcd. For C₁₃H₁₂N₂O₃S [M+H]⁺ 276.5352716; found 276.573781. Anal. calcd. For

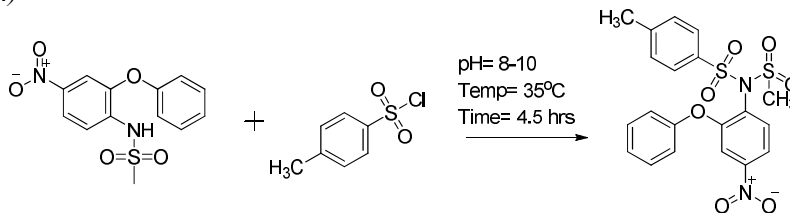
$C_{13}H_{12}N_2O_3S$: C, 56.51; H, 4.38; N, 10.14; O, 17.37; S, 11.60. Found: C, 56.35; H, 4.41; N, 10.8; O, 17.43; S, 11.56.

Synthesis of (6R,7R)-7-((R)-2-(4-hydroxyphenyl)-2-(4-methylphenylsulfonamido)acetamido)-3-methyl-8-oxo-5-thia-1-azabicyclo[4.2.0]oct-3-ene-2-carboxylic acid (HR8)



By following general procedure labeled compound **HR8** was purified by using mobile phase DCM:EtOH, 80:20 having 87% yield. λ_{max} 275 nm. FT-IR (cm^{-1}): 1160.35 (S=O str), 1053.12 (C-N str), 1647.45 (C=O), 816.32 (C-S str), 940.16 (S-N str), 3368.12 (-OH). 1H NMR (400 MHz, DMSO- d_6 , δ): 9.48 (1H, s, -COOH), 8.01-8.03 (1H, d, $J = 6.1$, -CHNHCO), 7.25-7.29 (1H, d, $J = 6.2$, -CHNHCO), 6.98-7.17 (4H, m, -ArH), 6.72-6.85 (4H, m, -ArH), 6.27 (1H, s, -C=CHS-), 6.03 (1H, s, -ArOH), 5.57-5.72 (1H, t, $J = 7.1$, -CHNHCO-), 5.06 (1H, s, -NCHCOOH), 4.84-4.86 (1H, d, $J = 6.4$, -CHN(S)), 4.78-4.82 (1H, d, $J = 6.3$, -NHCHAr), 3.31 (3H, s, -ArCH₃), 2.49 (3H, s, CH₃CH=CH-); MS (m/z , ESI): calcd. For $C_{23}H_{23}N_3O_7S_2$ [M+H]⁺ 517.5342617; found 517.5783681. Anal. calcd. For $C_{23}H_{23}N_3O_7S_2$: C, 53.37; H, 4.48; N, 8.12; O, 21.64; S, 12.39. Found: C, 53.35; H, 4.46; N, 8.14; O, 21.63; S, 12.36.

Synthesis of 4-methyl-N-(methylsulfonyl)-N-(4-nitro-2-phenoxyphenyl)benzenesulfonamide (HR14)



Labeled compound **HR14** was synthesized by adopting above procedure in yield 56% and was purified by using mobile phase DCM: EtOH, 40:60. λ_{max} 290. FT-IR (cm^{-1}): 1154.28 (S=O str), 1079.40 (C-N str), 1595.70 (C=O), 805.16 (C-S str), 975.07 (S-N str), 3761.15 (-OH). 1H NMR (400 MHz, DMSO- d_6 , δ): 7.69-7.72 (4H, m, -ArH), 7.39-7.43 (4H, m, -ArH), 7.12-7.14 (3H, d, $J = 8.1$, -ArH), 6.72-6.74 (2H, d, $J = 8.2$, -ArH), 2.94 (3H, s, CH₃SO₂-), 2.35 (3H, s, -ArCH₃); MS (m/z , ESI): calcd. for $C_{20}H_{18}N_2O_7S_2$ [M+H]⁺ 462.0362647; found 462.753910. Anal. calcd. for $C_{20}H_{18}N_2O_7S_2$: C, 51.94; H, 3.92; N, 6.06; O, 24.22; S, 13.87. Found: C, 51.65; H, 3.86; N, 6.10; O, 24.23; S, 13.76.

Antioxidant activity

DPPH radical scavenging assay. Using DDPH *in vitro* antioxidant activity of synthesized compounds was evaluated by a reported method [15]. All compounds were run in triplicate in order to produce precision of results. Trolox was used for standard curve. Using R^2 value relative concentrations of compounds were determined, and scavenging %, directly representing antioxidant activity, was determined using formula: Inhibition % = $(1 - \text{sample}_{530} / \text{blank}_{530}) \times 100$. The results are given in Table 1.

Table 1. Antioxidant activity of sulfonamide derivatives **HR5-HR8** and **HR14**.

Compound	% antioxidant activity	
	4 mM	6 mM
Trolox	1.53	12.87
HR5	1.52	15.60
HR6	1.47	15.19
HR7	1.48	15.06
HR8	1.26	14.17
HR14	1.22	13.07

Biological activity

Antibacterial activity. Growth media used was Luria-Bertain broth as it is highly efficient in bacterial growth [16]. Media was prepared using 4.0 g of tryptone, 2.0 g of yeast extract and 4.0 g of sodium chloride in 400 mL distilled water. Value of pH of media was maintained at 7.0. Above mentioned media was autoclaved at 125 °C for 30 min. Sample solutions were prepared in 5-50 µg concentration range. Three test tubes were labeled for each bacterial strain, i.e. *S. aureus* and *E. coli*. 2 mL of LB Broth and 20 µL of bacterial strain were added in above sterilized tubes. After that stocks of 5, 10, and 20 µL containing 5, 12.5, and 50 µg were added in them. Then these tubes were incubated at 37 °C for 72 hours. After this OD of each medium and control medium were taken at 600 nm. Graph was plotted between concentration and OD of compounds showing a comparative study for synthesized compounds (Table 2).

Table 2. Determination of MIC µg/mL synthesized products against bacterial strains.

S. No.	Name of compound	<i>S. aureus</i>	<i>E. coli</i>
1	HR5	2	5
2	HR6	6.5	8.5
3	HR7	3	4.0
4	HR8	2.5	3.0
5	HR14	1.5	2.0
6	Sulfmethoxazole	0.2	0.04

Antifungal activity. Antifungal activity of compounds was evaluated by performing well diffusion test [17-21] using PDA (potato dextrose agar). A 24 h yeast culture of PDA was used to prepare inoculum. Sterile saline solution (0.85%) was used for making suspension. Spectrophotometer was used to adjust turbidity of above suspension at 600 nm for getting final concentration matching with 0.5 McFarland standard. Agar medium was autoclaved for 30 min at 120 °C then cooled at 50 °C and inoculated with 1ml of above suspension having absorbance 0.5 McFarland. This inoculated medium were then poured into all assay plates 9cm in diameter and were allowed to cool down until solidified. Upon solidification, equidistance four wells 6mm in diameter were cut out of agar 6 µL of medium was added into these wells having synthesized compounds. These plates were then incubated at 27 °C for 48 h. MIC values in µg/mL and zone of inhibition in mm were calculated for each compound, comparing it with standard antifungal isoconazol (ISC) in concentration 1.0 µg/mL in each plate as +ve control. Results are given in Table 3 and 4.

Cytotoxicity test. *In vitro*, cytotoxicity test was performed using Vero cell line. Assay was based on protocol described by Borenfreund and Puerner (1984). 10% FBS (Fetal Bovine Serum) containing Trypsin enzymes were used for cell growth in 96-well plates for 24 hours. After that 100 µL of each sample and standard was loaded in above plate. MIC values were determined by comparison to doxorubicin hydrochloride as a reference drug. Two fold dilutions of test

compounds and doxorubicin were prepared in ethanol (1 mL). Each dilute was finally added to media at room temperature giving a final concentration of 100, 50, 12.5 $\mu\text{g mL}^{-1}$. This loaded plate was incubated for 48 hours at 37 °C, then natural red dye 10 μL (40%) was introduced in all wells and incubated at same temperature for 4 hours. Then plate was washed two times with PBS and finally one time with acidified ethanol. Absorbance was recorded at 540 nm in a MICrotitre plate reader spectrophotometer. Activity of each well was found using given formula and is presented in Table 5. Cytotoxicity of sample = (1-Experimental well abs/ abs of negative control) x100.

Table 3. Determination of MIC $\mu\text{g/mL}$ products against fungal strains.

S. No.	Name of compound	<i>A. flavous</i>	<i>A. nyger</i>
1	HR5	5.25	12.5
2	HR6	8.50	14.5
3	HR7	12.5	30.0
4	HR8	50.0	65.5
5	HR14	7.50	13.0
6	Isoconazole	0.50	0.76

Table 4. Diameter of zone of inhibition (mm \pm SD).

S. No.	Name of compound	<i>A. flavous</i>	<i>A. nyger</i>
1	HR5	27.2 \pm 0.12	18.1 \pm 0.12
2	HR6	26.5 \pm 0.30	16.3 \pm 0.33
3	HR7	18.9 \pm 0.11	14.9 \pm 0.22
4	HR8	12.4 \pm 0.22	10.6 \pm 0.10
5	HR14	25.5 \pm 0.10	17.1 \pm 0.11
6	Isoconazole	30	29.5

Table 5. Cytotoxicity values of sulfonamide derivatives.

Compound	Absorbance			% Activity		
	100	50	12.5	100	50	12.5
HR5	0.89	0.90	0.987	44	40	5.2
HR6	0.90	0.94	0.96	40	24	16
HR7	0.94	0.98	0.99	24	8.0	4.0
HR8	0.94	0.97	0.99	24	12	4.0
HR14	0.90	0.94	0.99	40	24	4.0
Doxorubicin hydrochloride	0.76	0.89	0.93	96	44	28

RESULT AND DISCUSSION

A series of five sulfonamides were synthesized in aqueous basic media by simple reaction of five amino group containing drugs; ceftriaxone, cefepime, nicotinamide (vitamin B), cefadroxil and nimsulide with paratoluenesulphonyl chloride with continuous stirring and details of reaction conditions are explained in experimental section and synthetic pathway of sulfonamides is explained in general procedure. The compounds were obtained in good to excellent yield (55–87%). Elemental analysis was performed for the conformation of all the compounds and measurement of absorption maximum (λ_{max}) provided the justification. The synthesized compounds were characterized by FT-IR; the characteristics band at 1148–1155.5 cm^{-1} of S=O stretching and 1048-1055 cm^{-1} for (C-N) and 813-814 cm^{-1} (C-S) and 930-958.9 cm^{-1} (S-N) for all compounds reveals the formation of sulfonamides. Mass spectral data of all synthesized compounds was obtained by ESI-MS. The molar mass for compound **HR5**, $\text{C}_{25}\text{H}_{24}\text{N}_8\text{O}_9\text{S}_4$, was originated as 708.77(calcd. 708.05). Correspondingly, observed mass for compound **HR6**,

$C_{26}H_{30}N_6O_7S_3$, was found as 634.13 (calcd. 634.75) and it proved the formation of the desired product. The prominent peaks for compounds, **HR7** and **HR8**, were recorded at m/z 122.05 and 363.09 for stable fragments $[C_6H_6N_2O]^+$, $[C_{16}H_{17}N_3O_5S]^+$, respectively. Observed and calcd molar masses of compounds **HR7** and **HR8** were found as 276.07 (calcd. 276.31), and 517.10 (calcd. 517.57), respectively. The major peak for compound **HR14** was noticed at m/z 462.06 and showed good agreement with calculated molecular masses of concerned compound. The structures of all the compounds were also confirmed by 1H NMR by dissolving in DMSO. 1H NMR spectra of compounds **HR5-HR8**, all $ArCH_3$ showed their chemical shift values from 21.39 to 21.84 ppm, and found to be very much similar to the literature values. In **HR5** and **HR6** two methyl signals, i.e. $-NCH_3$ and $-OCH_3$ were also recorded at 38.65, 60.21 ppm and 50.39, 63.99, respectively. The chemical shift value of $-OCH_3$ was observed on downfield side than $-NCH_3$ due to strong electron withdrawing influence of oxygen than nitrogen. In **HR7** a prominent peak of methyl (C1) was noticed at 21.39 ppm. In this molecule there are two rings, one of them is attached with methyl and second ring contains nitrogen. In second ring chemical shift values of C10, C12 were found on downfield side due to electron withdrawing influence of nitrogen and peaks of C2, C4 were appeared on high field side due to the electron donating effect of $-CH_3$ group. In **HR8** a small peak was noticed at 174.17 ppm due to carbon atom of $-COOH$ group.

Synthesized compounds were screened for their antibacterial and antifungal activities using sulfamethoxazole and isoconazol as reference antibacterial and antifungal agents. All developed compounds showed moderate to good activity for both bacterial and fungal strains but compound **HR14** exhibited excellent activity against the *E. coli* and *S. aureus* (MIC 1.5 and 2.0) and compound **HR5** showed good activity against *A. Flavous* and *A. Nyger* (MIC 5.25 and 12.5). Synthesized compounds were also screened for their antioxidant activity. Compound **HR5** showed excellent and pronounced activity at 4 mM concentration. The MIC values and zone of inhibitions are presented in Table 1-4. Cytotoxicity evaluation clearly shows that developed sulfonamides exhibited poor activity than standard drug, i.e. doxorubicin. Cytotoxicity values are presented in Table 5.

CONCLUSION

Five novel sulfonamides derivatives **HR5-HR8** and **HR14** were synthesized and evaluated for their antimicrobial, antioxidant and cytotoxicity test. Most of the synthesized compounds showed promising antimicrobial and antioxidant activity, suggesting a possible clinical significance of novel compounds. Compound **HR14** showed remarkable antimicrobial results, but compound **HR5** was found to have potential antioxidant activity. However their cytotoxic effects are not so pronounced.

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REFERENCES

1. Hansch, C.; Sammes, P.G.; Taylor, J.B. *Comprehensive Medicinal Chemistry*, Vol. 2, Pergamon Press: Oxford; **1990**; chap. 7.1.
2. Kanda, Y.; Kawanishi, Y.; Oda, K.; Sakata, T.; Mihara, S.; Asakura, K.; Kanemasa, T.; Ninomiya, M.; Fujimoto, M.; Kanoike, T. Synthesis and structure-activity relationships of potent and orally active sulfonamide ETB selective antagonists. *Bioorg. Med. Chem.* **2001**, *9*, 897-907.
3. Stokes, S.S.; Albert, R.; Buurman, Ed.T.; Andrews, B.; Shapiro, A.B.; Green, O.M.; McKenzie, A.R.; Otterbein, L.R. Inhibitors of the acetyltransferase domain of N-

- acetylglucosamine-1-phosphate-uridylyltransferase/glucosamine-1-phosphate acetyltransferase (GlmU). Part 2: Optimization of physical properties leading to antibacterial aryl sulfonamides. *Bioorg. Med. Chem. Lett.* **2012**, *22*, 7019-7023.
4. Chibale, K.; Haupt, H.; Kendrick, H.; Yardley, V.; Saravanamuthu, A.; Fairlamb, A.H.; Croft, S.L. Antiprotozoal and cytotoxicity evaluation of sulfonamide and urea analogues of quinacrine. *Bioorg. Med. Chem. Lett.* **2001**, *11*, 2655-2657.
 5. Rahavi Ezabadi, I.; Camoutsis, C.; Zoumpoulakis, P.; Geronikaki, A.; Soković, M.; Glamočilija, J.; Čirič, A. Sulfonamide-1,2,4-triazole derivatives as antifungal and antibacterial agents: Synthesis, biological evaluation, lipophilicity, and conformational studies. *Bioorg. Med. Chem.* **2008**, *16*, 1150-1161.
 6. Kennedy, J.F.; Thorley, M. *Pharmaceutical Substances*, 3rd ed., Kleeman, A.; Engel, J.; Kutscher, B.; Reichert, D. (Eds.), Thieme: Stuttgart; **1999**.
 7. Serradeil-Le Gal, C. An overview of SR121463, a selective non-peptide vasopressin V2 receptor antagonist. *Cardiovascular Drug Rev.* **2001**, *19*, 201-214.
 8. Natarajan, A.; Guo, Y.; Harbinski, F.; Fan, Y.-H.; Chen, H.; Luus, L.; Diercks, J.; Aktas, H.; Chorev, M.; Halperin, J.A. Novel arylsulfonamide-oxindole hybrid as an anticancer agent that inhibits translation initiation. *J. Med. Chem.* **2004**, *47*, 4979-4982.
 9. Vullo, D.; De Luca, V.; Scozzafava, A.; Carginale, V.; Rossi, M.; Supuran, C.T.; Capasso, C. The extremo- α -carbonic anhydrase from the *Thermophilic bacterium* Sulfurihydrogenibiumazorense is highly inhibited by sulfonamides. *Bioorg. Med. Chem.* **2013**, *21*, 4521-4525.
 10. Wilson, C.O.; Gisvold, O.; Block, J.H. *Wilson and Gisvold's Textbook of Organic Medicinal and Pharmaceutical Chemistry*, 11th ed., Block, J.; Beale, J.M. (Eds.), Lippincott Williams and Wilkins: Philadelphia; **2004**.
 11. Levin, J.I.; Chen, J.M.; Du, M.T.; Nelson, F.C.; Killar, L.M.; Skala, S.; Sung, A.; Jin, G.; Cowling, R.; Barone, D.; March, C.J.; Mohler, K.M.; Black, R.A.; Skotnicki, J.S. Anthranilate sulfonamide hydroxamate TACE inhibitors. Part 2: SAR of the acetylenic P1' group. *Bioorg. Med. Chem. Lett.* **2002**, *12*, 1199-1202.
 12. Livermore, D.M. Antibiotics resistance in staphylococci. *Int. J. Antimicrobiol. Agents* **2000**, *16*, 3-10.
 13. Whitaker, D.T.; Whitaker, K.S.; Johnson, C.R.; Haas, J. *p-Toluenesulfonyl Chloride. Encyclopedia of Reagents for Organic Synthesis*. John Wiley and Sons: New York; **2006**; DOI: 10.1002/047084289X.rt136.pub2.
 14. Online Publication, Working with hazardous chemicals. *Organic Syntheses*, Coll. Vol. 4, p. 943 (**1963**); Vol. 34, p. 96 (**1954**). DOI:10.15227/orgsyn.034.0096.
 15. Mohammed, H. Natural and synthetic flavonoid derivatives with potential antioxidant and anticancer activities. *PhD Dissertation*, Chemie, Pharmazie, Bio- und Werkstoffwissenschaften der Universität des Saarlandes, Saarbrücken, Germany, **2009**.
 16. Sezonov, G.; Joseleau-Petit, D.; D'Ari, R. *Escherichia coli* physiology in Luria-Bertani broth. *J. Bacteriology* **2007**, *189*, 8746-8749.
 17. Magalia, S.; Camero, T. Susceptibilidad de *Candida albicans* 'invitro' mediante los pozos de difusión. *Boletín Venezolano de Infectología* **1997**, *7*, 5-8.
 18. Magaldi, S.; Camero, T.; Mata, S.; Ortigoza-Medrano, E.; Arroyo-Espinosa, D.I. Pruebas de sensibilidad de *Candida albicans* frente a los de uso comercial. *Boletín Sociedad Venezolana de Microbiología* **1998**, *18*, 16-20.
 19. Magaldi, S.; Mata, S.; Camero, T.; Marcano, C.; Hartung, C. Determinación de la sensibilidad antifúngica en agentes de cromosistemas mediante técnica de los pozos de difusión. *Antibióticos e Infección* **1999**, *7*, 17-20.
 20. Magaldi, S.; Mata, S.; Hartung, C.; Verde, G.; Deibis, L.; Roldán, Marcano, Y. 'In vitro' susceptibility of 137 *Candida* sp. isolates from HIV positive patients to several antifungal drugs. *Mycopathologia* **2000**, *149*, 63-68.
 21. Magaldi, S.; Rios, A.; Hartung, C.; Verde, G.; Spencer, L.; Mata, S. In vitro susceptibility to fluconazole of *Candida* spp. isolates comparing three different methods. *J. Mycol. Med.* **2001**, *11*, 123-126.