SQUARE WAVE ANODIC STRIPPING VOLTAMMETRIC DETERMINATION OF Hg(II) WITH N-p-CHLOROPHENYLCINNAMOHYDROXAMIC ACID MODIFIED CARBON PASTE ELECTRODE

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ABSTRACT. A new method has been developed for the electrochemical determination of Hg(II) with N-p-chlorophenylcinnamohydroxamic acid (CPCHA) modified carbon paste electrode by square wave anodic stripping voltammetry (SWASV). Hg(II) was accumulated on the electrode surface by the formation of the complex in an open circuit and the resulting surface was characterized by electrochemical reduction and stripping. The optimum voltammetric response was observed using a carbon paste composition of 7.5% (w/w) CPCHA and preconcentration time of 210 s in 0.1 M sodium acetate at pH 8 followed by electrochemical SWASV in 0.3 M NH4Cl solution at pH 4 at a reduction potential of -0.6 V. The voltammetric signals were linear in the range of 1-25 µM Hg(II) with a detection limit of 12.9 nM. The voltammetric response for six replicate measurements of 15 µM Hg(II) was reproduced with 3.8% relative standard deviation (RSD). Many coexisting metal ions had little or no effect on the determination of Hg(II). Five spiked samples of water were evaluated by using the developed method giving recoveries of Hg(II) in the range 98-105%.

KEY WORDS: Chemically modified carbon paste electrode, Square wave anodic stripping voltammetry, Hg(II) determination, N-p-Chlorophenylcinnamohydroxamic acid, Environmental water samples

INTRODUCTION

Mercury is one of the most toxic heavy metals that occur in the environment mainly due to industrial applications such as electrical equipment, batteries, paints and extraction of gold from mines and rivers [1]. It can have several effects on human health and living organisms even at very low concentrations, because of its high reactivity, extreme volatility and relative solubility in water and living tissues [2]. Therefore, the determination of mercury is necessary particularly at trace levels.

The analytical techniques which are frequently used for the determination of mercury include inductively coupled plasma-mass spectrometry (ICP-MS) [3], mercury analyzer [4-6], spectrofluorimetry [7], cold vapor-atomic absorption spectrometry (CVAAS) [8], inductively coupled plasma-optical emission spectrometry (ICP-OES) [9], X-ray fluorescence spectrometry (XRF) [10] and graphite furnace atomic absorption spectrometry (GFAAS) [11]. Although these techniques are very reliable and accurate, they have some significant limitations, such as long analysis time and the use of expensive instruments. Furthermore, several complex steps must be performed, and these require specially trained personnel [12]. Electrochemical methods, in particular anodic stripping voltammetry (ASV), is the most favorable technique for the determination of trace level of mercury due to its low cost, simplicity, speed, high sensitivity and the ability to carry out speciation analysis [2].

Chemically modified electrodes (CMEs) have received great attention due to their enhanced sensitivity and selectivity of electrochemical analytical techniques. Several studies showed that chemically modified carbon paste electrode (CMCPE) could be used successfully for the determination of heavy metal ions. For example, carbon paste electrodes modified with

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SiO$_2$ nanoparticles for Cu(II) and Cd(II) [13], coconut shell powder for Cd(II) [14], CrO$_3$ for simultaneous determination of Cu(II), Cd(II), Zn(II), Pb(II) and Ag(I) [15] and nitroso-S for Co(II) [16]. The determination of Hg(II) has been investigated using CPEs containing variety of modifiers by stripping voltammetry. Modifiers commonly used are EDTA [17], water hyacinth biomass [18], hydroxyapatite [19], multi-walled carbon nanotubes [20], hybrid mesostructured silica nanoparticles [21], Schiff base [22], silica nanoparticles with Schiff base [23], SnO$_2$ at MWCNTs [24] and organofunctionalised SBA-15 nanostructured silica [25]. Each of these methods has its own advantages and limitations. Mercury exists in most samples, especially in environmental water samples as Hg(II). Therefore, it is worthwhile to look for new chemically modified electrode with improved quality for the voltammetric determination of Hg(II) in environmental water samples.

N-p-Chlorophenylcinnamohydroxamic acid (CPCHA) (Scheme 1) has been used as an extracting agent for the spectrophotometric determination of Ce(IV) [26] and Nb(V) [27] and as a modifier in carbon paste electrode for the selective and sensitive voltammetric determination of Co(II) [28], Pb(II) [29] and Cd(II) [30] while its analogue N-phenylcinnamohydroxamic acid has been used for the selective and sensitive voltammetric determination of Cu(II) [31]. However, the complex formation reaction and voltammetric behavior of Hg(II) by using CPCHA modified carbon paste electrode have not been investigated. Hence the voltammetric behavior of Hg(II) at CPCHA modified carbon CPE was studied in the present investigation.

Scheme 1. Structure of N-p-chlorophenylcinnamohydroxamic acid (CPCHA).

The aim of this work was to develop a new CPE modified with CPCHA for the selective preconcentration and determination of Hg(II) by SWASV. The method has been applied for the determination of traces of Hg(II) in potable and environmental waters.

**EXPERIMENTAL**

*Reagents and chemicals*

All the chemicals used were of analytical grade. Graphite powder (spectroscopic grade RBW) was from SGL Carbon (Ringsdorff, Germany) and paraffin oil was supplied from (Uvasol, Merck). Anhydrous sodium acetate, acetic acid, sodium hydroxide, ammonium chloride, ammonia solution and hydrochloric acid were obtained from (BDH, England). Standard stock solution of Hg (1000 mg/L, atomic absorption standard) was purchased from Sigma-Aldrich (USA). Reagent grade nitrate, sulfate and chloride salts of other cations (Merck and Sigma-Aldrich) were used as received. Sodium acetate and ammonium chloride solutions of various concentrations were prepared in distilled water. The pH of sodium acetate and ammonium chloride solutions were adjusted to the desired values by adding acetic acid (0.1 M) or sodium hydroxide (0.1 M) and ammonia solution (0.1 M) or hydrochloric acid (0.1 M), respectively. The working standard solutions, for optimization studies, were prepared daily from the standard stock solution by suitable dilution. N-p-chlorophenylcinnamohydroxamic acid was prepared by the condensation of N-p-chlorophenylhydroxylamine with cinnamoyl chloride at low temperature in diethyl ether medium made alkaline with a saturated solution of sodium bicarbonate [32].
Apparatus and instruments

All the voltammetric experiments for the determination of mercury(II) were performed by using CHI 840C electrochemical analyzer (CH Instruments, USA) connected to IBM Personal computer 130100DX4 for recording of voltammograms and processing of data. All the experiments were carried out using a conventional three electrode system with a UMCPE or CMCPE as a working electrode, a platinum wire as an auxiliary electrode, and a silver/silver chloride (Ag/AgCl) as a reference electrode. Three different cells (10 mL) supplied with an electrical spiral stirrers were used for the preconcentration and stripping step. The pH measurements were carried out using a pH meter (sensION, SHA Snilu Instruments CO. LTD, China). A magnetic stirrer (Stuart Scientific, Germany) with a Teflon coated stirring bar was used for stirring the solutions. A stop clock (Harris Digitimer, Japan) was employed for time measurement.

Electrode preparation

Unmodified carbon paste was prepared by adding 0.36 mL paraffin oil to 1 g graphite powder. Modified carbon pastes were prepared by substituting corresponding amounts of the graphite powder (5%, 7.5%, 10%, and 12.5% weight-to-weight ratio of the modifier (CPCHA) relative to the graphite powder) and adding the paraffin oil (0.36 mL) and manually homogenizing the mixture by using mortar and pestle. Both unmodified and modified active pastes were finally pressed into a cavity of the assembly made from 1 mL plastic syringe of 3 mm outer diameter with copper wire inserted at one end forming the external electric contact. The electrode surface was polished on clean paper until it had a shiny surface. The fresh electrode was pretreated by using cyclic voltammetry (4–5 runs) in the supporting electrolyte solution to get reproducible results. Renewal of the electrode was done by removing a thin layer of the surface and replacing with a fresh paste.

Procedure

The CVs were performed at unmodified and modified carbon paste electrodes in the potential range of -1.0 to 1.0 V (at 100 mV/s scan rate). All the quantitative measurements were carried out by SWASV. The CMCPE was immersed in a preconcentration cell containing 0.1 M sodium acetate solution (pH 8) and known amount of Hg(II) solution with constant stirring (400 rpm) under open circuit for a preselected period of time. The electrode was then taken out from the preconcentration cell, rinsed with distilled water and placed into a stripping cell containing 0.1 M NH₄Cl solution (pH 4) as supporting electrolyte. Following the application of a deposition potential of -0.6 V (verses Ag/AgCl) for 60 s without stirring and the electrode was scanned anodically from -0.6 V to 0.3 V after 10 s equilibration time with pulse amplitude 0.035 V; frequency 15 Hz and step potential of 0.004 V. After each measurement, the electrode was renewed and pretreated by scanning (4–5 runs) in 0.1 M NH₄Cl solution. Fresh supporting electrolyte solution was used in each set of experiments to avoid potential build up of Hg(II) in the electrochemical cell. All electrochemical measurements were made at room temperature (22 ± 2 °C) under atmospheric conditions (without inert gas bubbling).

Analysis of real samples

In order to test the practical application of the proposed method, the Hg(II) content was determined in five different types of water samples (tap water, bottled water, well water, stream water, river water) by spiking method. The well water, stream water, river water samples were filtered before the analysis to remove suspended particles. It was not necessary to filter the tap water and bottled water. Each water sample was spiked with 15 µM Hg(II) and placed in 10 mL of chemical cell, preconcentrated for 3.5 min and followed by voltammetric measurements.
RESULTS AND DISCUSSION

Voltammetric behavior of Hg(II) at CPCHA modified carbon paste electrode

The cyclic voltammetry of unmodified and modified carbon paste electrodes without and with Hg(II) preconcentration on the electrode surface at open circuit were run in 0.3 M NH₄Cl at pH 5. Figure 1 shows the CVs of CPCHA modified electrode with and without Hg(II) preconcentration. The voltammograms of the unmodified electrode with and without Hg(II) preconcentration are the same (overlapped curve a) and the CPCHA modified electrode (curve b) without Hg(II) preconcentration did not show any significant peak in the potential range -1 V to 1 V, whereas the CV of the CPCHA modified electrode with Hg(II) preconcentration (curve c) shows a well-defined anodic peak at 0.11 V and a broad cathodic peak at about -0.64 V. The anodic peak is due to the oxidation of Hg(0) to Hg(II) which formed complex with the CPCHA modifier in the preconcentrating solution while the cathodic peak is due to the reduction of Hg(II) to Hg(0). The modified carbon paste electrode also shows a small anodic peak at about 0.69 V due to the CPCHA modifier. The anodic peak current of Hg(II) at the CPCHA modified electrode is substantially larger and sharper than the corresponding cathodic peak current. Hence the anodic peak of Hg(II) was systematically studied by square wave anodic stripping voltammetry for analytical applications.

![Figure 1](image1.png)

Figure 1. Cyclic voltammograms: (a) bare CPE with and without Hg(II) preconcentration, (b) CPCHA modified CPE without Hg(II) preconcentration and (c) CPCHA modified CPE with Hg(II) preconcentration from 0.1 M sodium acetate solution pH 10; preconcentration time: 5 min; Hg(II) concentration: 100 µM; supporting electrolyte: 0.3 M NH₄Cl pH 5; scanned from -1 to 1 V and back at scan rate: 0.1 V/s.

The square wave voltammograms of the unmodified and modified carbon paste electrodes were run in the presence of 80 µM Hg(II). As observed in the voltammogram shown in Figure 2, the anodic peak current of Hg(II) at the CPCHA modified electrode is much larger than that of the unmodified electrode. Furthermore, there is a slight shift in the peak potential of square wave voltammogram of CPCHA modified electrode towards negative potential, which indicates that Hg(II) is easily oxidized at the CPCHA modified electrode compared to that at unmodified electrode. Therefore, the use of the CPCHA made it possible to significantly improve the analytical signal of the electrode.

Square wave anodic stripping voltammetric determination of Hg(II)

Even though the modifier, CPCHA, is a water insoluble compound [33], the possibility of its leaching during the voltammetric measurements was tested by its color reaction with V(V) [34] which clearly indicated the absence of CPCHA in the electrolyte solution. Therefore, there was no leaching of the compound during the voltammetric measurements. Leaching was also not observed in the previous voltammetric studies using CPCHA modified electrode [28–30].

**Electrochemical processes**

Based on the similar studies reported in the literature [28, 30, 31], the possible electrochemical processes that occur at the modified electrode can be described by the following steps (where HL represents CPCHA):

1. **Preconcentration step** (open circuit cell, 0.1 M NaAc, pH 8):
   \[
   \text{Hg}^{2+} + 2\text{HL} \rightarrow \text{HgL}_2 + 2\text{H}^+ \\
   \text{Solution} \quad \text{CPE surface} \quad \text{CPE surface} \quad \text{Solution}
   \]

2. **Reduction step** (closed circuit cell, 0.3 M NH₄Cl, pH 4, -0.6 V):
   \[
   \text{HgL}_2 + 2\text{H}^+ + 2e \rightarrow \text{Hg}^0 + 2\text{HL} \\
   \text{CPE surface} \quad \text{Solution} \quad \text{CPE surface} \quad \text{CPE surface}
   \]

3. **Stripping step** (closed circuit cell, 0.3 M NH₄Cl, pH 4, positive scan: -0.6 V to 0.3 V):
   \[
   \text{Hg}^0 \rightarrow \text{Hg}^{2+} + 2e \\
   \text{CPE surface} \quad \text{Solution}
   \]

Briefly the accumulation of Hg(II) on the electrode surface takes place by the complexation of Hg(II) with the modifier CPCHA in the open circuit cell at pH 8. The reduction of Hg(II) in

**Figure 2.** Square wave voltammograms of 80 µM Hg(II) in 0.3 M NH₄Cl pH 4 at (a) unmodified carbon paste electrode and (b) CPCHA modified carbon paste electrode: preconcentration time 5 min in 0.1 M NaAc; deposition potential -0.6 V; deposition time; 60 s and potential scan from -0.6 V to 0.3 V after 10 s equilibration time with pulse amplitude 0.025 V, frequency 15 Hz and step potential 0.004 V.
the complex \( \text{HgL}_2 \) to \( \text{Hg}^0 \) occurs at an applied potential of \(-0.6 \) \( \text{V} \) in closed circuit cell in \( 0.3 \) \( \text{M NH}_4\text{Cl} \) at \( \text{pH 4} \). The stripping of \( \text{Hg}^0 \) occurs after oxidation to \( \text{Hg}^{2+} \) on positive scan from \(-0.6 \) \( \text{V} \) to \( 0.3 \) \( \text{V} \) in the closed circuit cell in \( 0.3 \) \( \text{M NH}_4\text{Cl} \), at \( \text{pH 4} \).

Effect of amount of CPCHA in the carbon paste

The accumulation of \( \text{Hg(II)} \) at the CPCHA modified CPE was based on the complex formation reaction between the metal ion, \( \text{Hg(II)} \) and the modifier, CPCHA. Therefore, the amount of CPCHA in the carbon paste had significant influence on the voltammetric response of the modified electrode. The effect of the amount of CPCHA in the modified carbon paste electrode was investigated by preparing six different modified carbon paste electrodes (2.5%, 5%, 7.5%, 10%, 12.5% and 15% weight-to-weight ratio of CPCHA to graphite powder in the carbon paste) under identical conditions. The respective peak currents were 1.05, 2.37, 5.34, 4.25, 2.49 and 2.00 \( \mu\text{A} \). This result clearly indicates that the maximum peak current was obtained with 7.5% CPCHA in the CPE. Higher amounts of CPCHA containing electrodes gave lower peak currents. This could be related to a decrease in the carbon content of the paste and, consequently, reduction of the conductive electrode area (carbon particles) hampering the electron transfer at the electrode surface. Hence an electrode containing 7.5% CPCHA was applied for all subsequent experiments.

Effect of composition and concentration of preconcentration solution

The complex formation reaction of \( \text{Hg(II)} \) with CPCHA at the electrode surface and the anodic stripping voltammetric properties of \( \text{Hg(II)} \) preconcentration in \( \text{NH}_4\text{Cl}, \text{NaAc and Na}_2\text{CO}_3 \) buffer solutions (each one \( 0.1 \) \( \text{M, pH 8} \)) were investigated. The peak current of \( \text{Hg(II)} \) preconcentrated in \( \text{Na}_2\text{CO}_3 \) (3.61 \( \mu\text{A} \)) was slightly smaller than \( \text{NaAc} \) (9.01 \( \mu\text{A} \)). This could be due to the precipitate formation of \( \text{Hg(II)} \) as \( \text{HgCO}_3 \) at \( \text{pH 8} \). Preconcentration of \( \text{Hg(II)} \) in \( \text{NH}_4\text{Cl} \) solution showed a much smaller peak current (1.99 \( \mu\text{A} \)). This is due to complex formation \( \text{Hg(II)} \) with ammonia and chloride at high concentration that can hinder the accumulation of \( \text{Hg(II)} \) on the electrode surface. Among these, best voltammetric result of \( \text{Hg(II)} \) was obtained from preconcentration in \( \text{NaAc} \) solution. The effect of \( \text{NaAc} \) concentration was also tested in the range \( 0.1–0.5 \) \( \text{M} \) at \( \text{pH 8} \). The respective peak currents were 9.01, 8.70, 5.71, 2.00 and 1.00 \( \mu\text{A} \). This result shows that the maximum peak current of \( \text{Hg(II)} \) was obtained from preconcentration in \( 0.1 \) \( \text{M NaAc} \) solution. It was observed that increasing the concentration of \( \text{NaAc} \) decreased the peak current. This is due to the formation of a weak complex between acetate ions and \( \text{Hg(II)} \). Hence, increasing the concentration of acetate ions decreases the accumulation of \( \text{Hg(II)} \) that complexes with CPCHA at the electrode surface. Thus \( 0.1 \) \( \text{M sodium acetate solution was used for preconcentration of} \text{Hg(II)} \) throughout the study.

Effect of pH of preconcentration solution

The \( \text{pH} \) of the preconcentration solution (as shown in the above mechanism) has a direct effect on the complex formation reaction of \( \text{Hg(II)} \) with CPCHA. Hence the effect of \( \text{pH} \) of the preconcentration solution (0.1 \( \text{M NaAc} \) on the accumulation of \( \text{Hg(II)} \)) was investigated in the \( \text{pH} \) range 5–11. The peak current of CPCHA modified electrode as a function of \( \text{pH} \) is shown in Figure 3. The square wave voltammograms at different \( \text{pH} \) are shown as inset in Figure 3. It was observed that the peak current increased sharply with increasing \( \text{pH} \) from 5 to 8 and then decreased from 9–11. This is due to the increased complex formation of \( \text{Hg(II)} \) with CPCHA at the electrode surface with increasing \( \text{pH} \) and due to the completion of complex formation \( \text{Hg(II)} \) with acetate at higher (\( \text{pH} > 8 \)). Hence 0.1 \( \text{M sodium acetate pH 8} \) was used for all subsequent experiments.
Figure 3. Effect of pH of preconcentration solution on the square wave anodic stripping voltammetric peak current. Each bar represents mean ± 3 standard deviation. Hg(II) concentration: 80 µM. Other conditions are as in Figure 2. Square wave voltammograms at different pH as inset (a, pH 5; b, pH 6; c, pH 11; d, pH 10; e, pH 7; f, pH 9; g, pH 8).

One would expect the precipitation of Hg(II) as Hg(OH)$_2$ at such higher pHs. However, this was not observed in this investigation due to the fact that the Hg(II) solution was spiked into the preconcentration cell just before dipping the electrode in the cell and subsequent stable complex formation of Hg(II) with CPCHA at the electrode surface. It should also be noted that the acetate ions form weak complexes with Hg(II) that also prevent the precipitation of Hg(OH)$_2$.

**Effects of composition and concentration of supporting electrolyte**

The effect of composition of supporting electrolytes including H$_2$SO$_4$ + KCl, NH$_3$.OH.HCl, NaH$_2$PO$_4$ and NH$_4$Cl buffer solution (each 0.3 M, pH 4) were examined. The peak currents were very low in H$_2$SO$_4$ + KCl (2.19 µA) and NaH$_2$PO$_4$ (2.87 µA). This might be due to the inability of the ions of the two salts to supply hydrogen ions required during the stripping steps (as shown in the above mechanism). While the peak current for NH$_3$.OH.HCl (5.94 µA) was comparable with that of NH$_4$Cl (6.60 µA), it has unstable background current. Among these, the electrochemical response in NH$_4$Cl solution was the best peak shape, highest stripping peak current, reproducibility and background stability. This might be due to the differences in the extent of providing hydrogen ions required for the reduction step (as shown in the above mechanism) and consequently favoring the proceeding stripping step. Hence, NH$_4$Cl buffer solution was selected throughout this study.

The effect of concentration of NH$_4$Cl on the peak current was also studied in the range 0.1–0.5 M at pH 8. The respective peak currents obtained were 2.00, 3.10, 6.61, 2.40 and 1.11 µA. This result shows that the concentration of the supporting electrolyte have significant effect on the current response, maximum and reproducible peak current was obtained in 0.3 M NH$_4$Cl solution and hence it was selected for the subsequent experiments.

Effect of pH of supporting electrolyte

The effect of pH of the supporting electrolyte (0.3 M NH$_4$Cl) was also studied by varying the pH in the range 3–8. The results are shown in Figure 4. Peak current increased with increasing pH up to pH 4. This is due to reduction of Hg(II) at the CME surface involves hydrogen ions (as shown in the above mechanism). The peak current decreased with further increase in the pH beyond 4. The increasing pH hinders the reduction reaction. Maximum peak current was observed at pH 4.

It is evident from the above discussion that the preconcentration and the voltammetric measurement steps require NaAc and NH$_4$Cl solutions of different concentrations and different pH. Hence it was necessary to carry out the preconcentration and the voltammetric measurement separately. Therefore, 0.1 M NaAc solution of pH 8 and 0.3 M NH$_4$Cl solution of pH 4 were used for an open-circuit preconcentration and voltammetric measurements, respectively.

![Figure 4. Effect of pH of supporting electrolyte solution on the square wave anodic stripping voltammetric peak current. Each bar represents mean ± 3 standard deviation. Hg(II) concentration: 80 µM and other conditions are as in Figure 2.](image)

Effect of preconcentration time

The other important parameter that affects the voltammetric response is the time allowed for the accumulation of the Hg(II) from the preconcentration solution at the electrode surface. The effect of accumulation time on the peak current was studied from 30 s to 300 s. The effect of time on the anodic peak current of the modified electrode is shown in Figure 5. The peak current increases with increasing preconcentration time, indicating an enhancement of Hg(II) uptake at the electrode surface. Normally, the increase in the response continues until a maximum peak current, presumably corresponding to either saturation or an equilibrium surface coverage, is attained. The results obtained indicated attainment of steady-state accumulation levels of Hg(II) at the electrode surface at exposure times of 210 s. After that there was no further increase in the peak current up to 300 s. Thus, to obtain a linear relationship between Hg(II) concentration and the peak current, a relatively short preconcentration time must be employed to avoid saturation effect. Therefore, 210 s preconcentration time was selected throughout this study.
Square wave anodic stripping voltammetric determination of Hg(II)

Effect of preconcentration time on the square wave anodic stripping voltammetric peak current. Each bar represents mean ±3 standard deviation. Hg(II) concentration: 80 µM. Other conditions are as in Figure 2.

Effect of reduction potential and reduction time

The effect of the reduction potential on the anodic peak current of Hg(II) was investigated in the range from -0.2 V to -0.8 V. The effect of the reduction potential on the anodic peak current of the modified electrode is shown in Figure 6. At more negative reduction potentials, Hg(II) is reduced more completely, thus increasing the peak current. The peak current increased as the reduction potential became more negative up to -0.6 V and then decreased. At more positive potentials a decrease in the anodic peak current was caused by an inefficient reduction of Hg(II) to Hg(0) at the electrode surface. The maximum peak current was obtained at a potential of -0.6 V, providing maximum analytical signal for Hg(II) determination.

Figure 5. Effect of preconcentration time on the square wave anodic stripping voltammetric peak current. Each bar represents mean ±3 standard deviation. Hg(II) concentration: 80 µM. Other conditions are as in Figure 2.

Figure 6. Effect of reduction potential on the SWASV peak current. Each bar represents mean ±3 standard deviation. Hg(II) concentration 80 µM. Other conditions as in Figure 2.
Reduction time is also the most important factor for determination of the detection limit in stripping voltammetric analysis. At the same reduction potential, a longer reduction time will cause Hg(II) to be reduced completely and consequently lead to a higher peak current. However, when the reduction time is too long the reduced Hg(0) covers the entire effective electrode surface, and the peak current does not change with increasing reduction time. The effect of the reduction time on the anodic peak current of Hg(II) was performed at constant reduction potential with different reduction times from 20-80 s. The peak current was found to increase linearly with increasing reduction time up to 60 s and then remained constant. Therefore 60 s reduction time was used for all subsequent measurements.

**Effect of frequency and amplitude**

In SWASV amplitude and frequency are the most important parameters affecting the shape and sensitivity of the peak current. Effect of amplitude was studied by varying the value from 20 to 50 mV. The peak current increased and became broader with increasing amplitude, maximum amplitude of 35 mV was chosen for the study. A similar pattern of the peak current was obtained with increasing the frequency (from 15 to 35 Hz). Frequency of 25 Hz was selected for this work.

**Figure 7.** Anodic stripping voltammograms of Hg(II) at different concentrations (a) 1; (b) 5; (c) 10; (d) 15; (e) 20 and (f) 25 µM in 0.3 M NH₄Cl solution pH 4, preconcentration time 3.5 min with pulse amplitude 0.055 V, frequency 25 Hz and step potential 0.004 V. Other conditions as in Figure 2.

**Calibration curve, detection limit and reproducibility**

Under the above optimized conditions, the relationship between the anodic peak current and the Hg(II) concentration was studied. Well-defined peaks were observed following a 210 s preconcentration time and reduction potential of -0.6 V. Stripping voltammograms and calibration plot for Hg(II) concentration in the range 1.0 µM to 25 µM are shown in Figure 7. The peak current increased with increasing Hg(II) concentration and a linear calibration plot was obtained with calibration equation: 

\[ I_p = 0.334 [\text{Hg}] (\mu M) + 0.845 \]

and a correlation coefficient of 0.998. The detection limit was calculated by 3 x SD of the lowest concentration of
the linear range (1.0 µM Hg(II)). The detection limit was found to be 12.9 nM. The repeatability and consistency of the proposed electrode was evaluated by six replicate determination of 15 µM Hg(II). A relative standard deviation (RSD) of 3.83% was obtained for six repeated measurements of 15 µM Hg(II) solution which demonstrate the good precision of the proposed method.

Comparison of the performance of CPCHA modified carbon paste electrode with other electrodes

The analytical performance of the proposed method was compared with several reported methods in the literature and the results are summarized in Table 1. The results show that the limit of detection obtained in this study was lower or comparable to most of the reported methods in the literature. Some reported methods achieved lower detection limits of Hg(II), having preconcentration times which are several times longer than that of the present study (210 s). A shorter accumulation time enables faster measurements, therefore in the same duration time more analyses can be made. The proposed method also has good linear range. Thus the proposed method is able to detect the presence of Hg(II) at trace levels.

Table 1. Comparison of the analytical performance obtained at CPCHA/CPE with different electrodes reported in the literature for the electrochemical determination of Hg(II).

<table>
<thead>
<tr>
<th>Electrode type</th>
<th>Technique</th>
<th>Preconcentration time (min)</th>
<th>Liner range (µM)</th>
<th>LOD (nM)</th>
<th>Ref.</th>
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<tr>
<td>HA-NP/GCE</td>
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<td>0.20–210</td>
<td>141</td>
<td>[19]</td>
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<tr>
<td>NG/GCE</td>
<td>DPASV</td>
<td></td>
<td>0.99–8.90</td>
<td>50</td>
<td>[35]</td>
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<tr>
<td>PdO-NP/CPE</td>
<td>DPASV</td>
<td>3</td>
<td>0.25–100</td>
<td>19.3</td>
<td>[36]</td>
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<td>WBM/CPE</td>
<td>DPASV</td>
<td>10</td>
<td>1.98–3.96</td>
<td>965</td>
<td>[18]</td>
</tr>
<tr>
<td>TCA/GCE</td>
<td>SWASV</td>
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<td>0.008–3</td>
<td>5.0</td>
<td>[2]</td>
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<tr>
<td>MTZ-MUS-2/CPE</td>
<td>DPASV</td>
<td>5</td>
<td>0.1–1</td>
<td>99</td>
<td>[21]</td>
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<td>EDTA/CPE</td>
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<td>50–350</td>
<td>8.6</td>
<td>[17]</td>
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<td>0.005–5</td>
<td>0.38</td>
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<td>1–25</td>
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</tbody>
</table>

1Hydroxyapatite nanoparticle; 2N-doped graphene; 3palladium oxide supported onto natural phosphate; 44-tertbutyl-1-(ethoxycarbonylmethoxy)thiacalix[4]arene; 5mesostructured silica nanoparticles functionalized with a 5-mercapto-1-methyltetrazole derivative; 6ethylene diamine tetra acetic acid; 7multi-wall carbon nanotubes; 8silica nanoparticles modified by a newly synthesized Schiff base ligand.

Effects of other ions

CMCPE has significant capability to enhance selectivity because of the modifier-analyte interaction. The selectivity of the CMCPE was evaluated by adding different amounts of other ions (which are commonly present together with Hg(II) in different samples) into known concentration of Hg(II) solutions during the preconcentration step, and the results are shown in Table 2. Alkali and alkaline earth metal ions, NH₄⁺, Cl⁻ and NO₃⁻ (up to 100-fold molar excess); Ni(II) and SO₄²⁻ (up to 10-fold molar excess); Cd(II) and Co(II) (up to 5-fold molar excess) and Cu(II) (up to 2 fold) have no significant effect on the determination of Hg(II). The lower tolerance limit of Co(II) [28], Cd(II) [30] and Cu(II) [31] are due to their complex formation reactions with the CPCHA at the electrode surface. The results show that the presence of Pb(II) and Fe(III) interferes significantly by decreasing the
Hg(II) signal, because both Pb(II) [29] and Fe(III) [37] form stable complexes with CPCHA and prevent the complex formation and accumulation of Hg(II) at the electrode surface.

In general, several ions commonly present in water samples do not interfere in the determination of Hg(II) and therefore, the sensor is quite selective and can be applied for determination of Hg(II) in the absence of Pb(II).

Table 2. Change in SWASV peak current of 15 x 10^{-6} M Hg(II) in presence of other ions.

<table>
<thead>
<tr>
<th>Interfering ion</th>
<th>Concentration (M)</th>
<th>Change in peak current (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Alkali and alkaline earth metal ions</td>
<td>15 x 10^{-4}</td>
<td>&lt; 5</td>
</tr>
<tr>
<td>NH_4^+</td>
<td>15 x 10^{-4}</td>
<td>-1.4</td>
</tr>
<tr>
<td>Cd(II)</td>
<td>7.5 x 10^{-5}</td>
<td>1.6</td>
</tr>
<tr>
<td>Co(II)</td>
<td>7.5 x 10^{-5}</td>
<td>3.1</td>
</tr>
<tr>
<td>Cu(II)</td>
<td>3.0 x 10^{-4}</td>
<td>-4.2</td>
</tr>
<tr>
<td>Ni(II)</td>
<td>1.5 x 10^{-3}</td>
<td>2.2</td>
</tr>
<tr>
<td>Pb(II)</td>
<td>15 x 10^{-6}</td>
<td>-42</td>
</tr>
<tr>
<td>Fe(III)</td>
<td>15 x 10^{-6}</td>
<td>-7.3</td>
</tr>
<tr>
<td>Cl^-</td>
<td>1.5 x 10^{-4}</td>
<td>-3.5</td>
</tr>
<tr>
<td>SO_4^{2-}</td>
<td>1.5 x 10^{-4}</td>
<td>1.2</td>
</tr>
<tr>
<td>NO_3^-</td>
<td>15 x 10^{-4}</td>
<td>&lt; 5</td>
</tr>
</tbody>
</table>

Analytical applications

The applicability and feasibility of the proposed method for the analysis of real samples was assessed by its application to the determination of Hg(II) in five types of water samples. Tap water, bottled water, well water, stream water and river water samples were employed. The water samples do not require the pretreatment, but they still need to adjust the pH to 8 using 0.1 M sodium acetate solution. Hg(II) was not detected in the tap water, bottled water, well water, and stream water while appreciable concentration (1.8 µM) of Hg(II) was detected and determined in the river water. Recovery of Hg(II) in water samples were determined by the spiking method. The recovery results are summarized in Table 3. The percentage recoveries were found in the range 98–105% which are within the acceptable range. Thus the proposed method can be used for the precise and reliable determination of Hg(II) in environmental water samples.

Table 3. Percentage recovery of Hg(II) in water samples (N = 3).

<table>
<thead>
<tr>
<th>Sample</th>
<th>Original (µM)</th>
<th>Added (µM)</th>
<th>Found (µM)</th>
<th>Recovery (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tap water</td>
<td>ND</td>
<td>15</td>
<td>14.7 ± 0.17</td>
<td>98.0</td>
</tr>
<tr>
<td>Bottled water</td>
<td>ND</td>
<td>15</td>
<td>14.9 ± 0.12</td>
<td>99.5</td>
</tr>
<tr>
<td>Well water</td>
<td>ND</td>
<td>15</td>
<td>14.8 ± 0.48</td>
<td>98.6</td>
</tr>
<tr>
<td>Stream water</td>
<td>ND</td>
<td>15</td>
<td>15.3 ± 0.22</td>
<td>102</td>
</tr>
<tr>
<td>River water</td>
<td>1.8</td>
<td>15</td>
<td>17.6 ± 0.86</td>
<td>105</td>
</tr>
</tbody>
</table>

ND = Not detected.

CONCLUSION

A new method has been developed for the square wave anodic stripping voltammetric determination of Hg(II) using N-p-chlorophenylcinnamohydroxamic acid modified carbon paste electrode. The method is relatively cheap and easy, and has proved to be a simple and selective for the determination of Hg(II) at trace level in water samples. The method offers attractive...
properties compared to the previous studies such as simplicity of electrode preparation, high stability, short preconcentration time (3.5 min), and the use of non-deaerated solution. The proposed method has good linear range: 1–25 μM Hg(II) and a lower detection limit: 12.9 nM Hg(II) than most of the reported methods and not interfered strongly by other ions, which are normally associated with mercury in environmental waters, alloys and complex materials. The developed method has been applied successfully to the determination Hg(II) in environmental water samples without any prior sample treatment except filtration.

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REFERENCES


