IN VITRO ANTIFUNGAL, ANTI-INFLAMMATORY AND CYTOTOXIC ACTIVITIES OF RUMEX ABYSSINICUS RHIZOME EXTRACT AND BIOASSAY-GUIDED ISOLATION OF CYTOTOXIC COMPOUNDS FROM RUMEX ABYSSINICUS

Jibrel Abdulkadir Emami1,2*, Estifanos Ele Yaya1, Muhammad Iqbal Choudhary3, Sammer Yousuf3 and Tetemke Mehari Gebremedhin1

1Department of Chemistry, College of Natural and Computational Sciences, Addis Ababa University, Addis Ababa, Ethiopia
2Department of Chemistry, College of Natural and Computational Sciences, Dire Dawa University, Dire Dawa, Ethiopia
3International Center for Chemical and Biological Sciences, H. E. J. Research Institute of Chemistry, University of Karachi, Karachi-75270, Pakistan

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ABSTRACT. Rumex abyssinicus showed strong cytotoxicity against HeLa cells (IC50 = 22.25 µg/mL) and weak cytotoxicity against PC3 and BJ cells with percent inhibition of 58.6, 25.8 and 29.7% at 30.0 µg/mL. It showed moderate antifungal activity against Aspergillus niger with a percent growth inhibition of 55.5% at 3000 µg/mL. It also strongly inhibited oxidative burst with IC50 value of 24.8 µg/mL. DCM (100%) and DCM: EtOAc (1:1) fractions showed strong cytotoxicity against HeLa cells, whilst pet ether: DCM (1:1) fraction showed strong cytotoxicity against PC3 cells with IC50 values of 29.3, 26.3 and 24.3 µg/mL, respectively. Moreover, the DCM: EtOAc (1:1) fraction inhibited ROS production with IC50 value of 18.8 µg/mL. Cytotoxic fractions afforded chrysophanol (1), physcion (2), emodin (3), citreorosein (4) and β-sitosterol (5). Among the isolated compounds, emodin (3) showed strong cytotoxicity against HeLa cells, whilst chrysophanol (1) and physochin (2) showed strong cytotoxicity against PC3 cells with IC50 values of 8.94, 22.5, and 28.5 µM, respectively. In addition, emodin (3) and citreorosein (4) showed strong inhibition against ROS production with an IC50 value of 16.2 and 38.2 µg/mL. The findings of this study suggest R. abyssinicus as a good candidate for cancer and inflammation management.

KEY WORDS: Polygonaceae, Rumex abyssinicus Jacq., Cytotoxic, Antifungal, Anti-Inflammatory, Reactive oxygen species

INTRODUCTION

Rumex abyssinicus Jacq. (Polygonaceae), locally named in Amharic “mekmako”, is an indigenous perennial herb, up to 3 m tall, with thick and fleshy rhizome. It is a medicinal plant that grows in tropical Africa, including Madagascar, more commonly in cultivated lands. In many countries of tropical Africa, leaves and tender shoots are consumed as vegetable. The rhizomes are used to refine butter and give it a bright yellow color [1]. Furthermore, it is also used as a cosmetic in northern Ethiopia for dying the palms of the hands and feet [2]. Traditional healers use this plant for the treatment of various disorders such as amebiasis, hemorrhoid, hepatitis, common cold, wound, hypertension, toothache, headache, blood pressure, asthma, liver disease, abdominal pain, tuberculosis, lung diseases, leprosy, fever, tumor and cancer [3-6].

The phytochemical studies of R. abyssinicus extract revealed the presence of tannins, flavonoids, phenols, quinones, alkaloids, cartenoids, phlobatannins, terpenoids, naphththalenes, stilbenoids, steroid, glycosides, saponins, fats and oils [7-9]. Many compounds are reported from R. abyssinicus including chrysophanol (1), physcion (2), emodin (3), citreorosein (4), β-sitosterol (5), aloemodin, emodic acid, chrysophanol-8-β-D-glucoside, emodin-β-8-O-D-glucoside, physcion-β-8-D-glucoside, bianthron, emodin-chrysophanol, helminthosporin, epicatechin,

*Corresponding author. E-mail: jibrela56@gmail.com

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epicatechin-3-O-gallate, epicatechin-3-O-(4′-O-methyl)gallate, methyl gallate, oxalic acid, phenolic acid, rumexamide, oleanolic acid, lupeol, 3β,28-dihydroxy-20(22)-ene 3β-dihydroxy-20(29)-en-28-oic acid, stigmasterol, stigmasterol-3-O-β-D-glucoside stigmastane-3,6-dione and ergosta-6,22-diene-3,5,8-triol [2, 10-13].

Figure 1. Compounds isolated from cytotoxic fractions of *R. abyssinicus*.

*R. abyssinicus* reported to demonstrate wide range of bioactivities including antibacterial [14], antiviral [14], cholinesterase inhibitor [11], anthelmintic [15], wound healing [16], anti-inflammatory [14, 16], antioxidant [17], lytic activities against zoospores, *Trypanosomacidal* and anti-tumor [19]. In addition, the *R. abyssinicus* showed a chemopreventive potential against dimethylhydrazine-induced colon carcinogenesis in rats, and it also displayed anticancer activity in prostate, brain, breast, and leukemia cell cultures [6]. The current study was conducted to evaluate the antifungal, anti-inflammatory and cytotoxic activities of *R. abyssinicus* rhizome extract and to isolate lead compounds from cytotoxic fractions.

**RESULTS AND DISCUSSION**

**Cytotoxic activity**

The 80% EtOH extract of *R. abyssinicus* exhibited cytotoxicity against cervical cancer (HeLa) and prostate cancer (PC3) cell lines. *R. abyssinicus* inhibited the proliferation of HeLa and PC3 cancer cells by 58.6% and 25.8% at 30 µg/mL. However, the extract exerted less toxicity to normal cells (BJ) with a percent inhibition of 29.7% at 30 µg/mL. *R. abyssinicus* displayed potent cytotoxicity against HeLa cells with an IC<sub>50</sub> value of 22.25 ± 0.7 µg/mL, but lower compared to standard drug doxorubicin (IC<sub>50</sub> = 0.9 ± 0.14 µg/mL). In agreement with the current findings, Girma et al. (2013) reported that the 80% methanol in water rhizome extract of *R. abyssinicus* showed a chemopreventive potential against dimethylhydrazine-induced colon carcinogenesis in rats, and suggested COX-2 inhibition by the anthraquinones in the extract could be one mechanism for the observed chemopreventive effect [6].

The extract of *R. abyssinicus* was further partitioned sequentially into eight fractions using different polarities (Table 1), in order to determine the characteristics of cytotoxic phytochemicals present in them. Due to solubility problems in DMSO, cytotoxic activity of petroleum ether (100%) eluted fraction was not conducted. Among the fractions tested, 100% DCM (IC<sub>50</sub> = 24.3 ± 1.4 µg/mL) and 1:1 of DCM: EtOAc (IC<sub>50</sub> = 29.3 ± 5.5 µg/mL) eluted fractions displayed strong cytotoxicity against cervical cancer HeLa cells and suppressed the proliferation of HeLa cells by 65.2% and 56.2% at 30 µg/mL, respectively. Similarly, the pet ether: DCM (1:1) eluted fraction...
In vitro antifungal, anti-inflammatory and cytotoxic activities of *R. abyssinicus* rhizome extract showed significant cytotoxicity against PC3 cells with IC_{50} value 26.3 ± 1.2 µg/mL and inhibited the proliferation of PC3 cells by 64.9% at 30 µg/mL.

Table 1. Cytotoxic activity of *R. abyssinicus* extract, fractions and standard drugs.

<table>
<thead>
<tr>
<th>Crude extract, fractions and standard drug</th>
<th>% inhibition on HeLa 30 µg/mL</th>
<th>IC_{50} ± SD</th>
<th>% inhibition on PC3 50 µg/mL</th>
<th>IC_{50} ± SD</th>
<th>% inhibition on BJ 50 µg/mL</th>
<th>IC_{50} ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Crude extract</td>
<td>58.6</td>
<td>22.8 ± 0.3</td>
<td>25.8</td>
<td>29.7</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pet ether</td>
<td>INS</td>
<td>INS</td>
<td>INS</td>
<td>INS</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Pet ether: DCM (1:1)</td>
<td>9.14</td>
<td>64.9</td>
<td>26.3 ± 1.2</td>
<td>16.6</td>
<td></td>
<td></td>
</tr>
<tr>
<td>DCM</td>
<td>65.2</td>
<td>24.3 ± 1.4</td>
<td>33.3</td>
<td>29.59</td>
<td></td>
<td></td>
</tr>
<tr>
<td>DCM: EtOAc (1:1)</td>
<td>56.2</td>
<td>29.3 ± 5.5</td>
<td>38.2</td>
<td>54.7</td>
<td>44.4 ± 2.7</td>
<td></td>
</tr>
<tr>
<td>EtOAc</td>
<td>14.5</td>
<td>38.9</td>
<td>2.83</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>EtOAc : MeOH (1:1)</td>
<td>35.6</td>
<td>1.03</td>
<td>-0.7</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MeOH</td>
<td>-9.5</td>
<td>-50.3</td>
<td>9.6</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>MeOH : H_{2}O (1:1)</td>
<td>4.67</td>
<td>2.74</td>
<td>4.2</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Doxorubicin</td>
<td>101.2</td>
<td>0.9 ± 0.14</td>
<td>89.9</td>
<td>1.9 ± 0.08</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Cycloheximide</td>
<td>89.9</td>
<td></td>
<td>0.8 ± 0.1</td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

DCM = dichloromethane, EtOAc = ethyl acetate, MeOH = methanol, H_{2}O = water, Pet ether = petroleum ether.

The results in Table 1 indicated that the pet ether: DCM (1:1) and DCM (100%) fractions showed different cytotoxicity against HeLa and PC3 cells. Pet ether: DCM (1:1) fraction was found to be significantly cytotoxic to PC3 cells (IC_{50} = 26.3 ± 1.2 µg/mL, 64.9% inhibition at 30 µg/mL) but very weakly cytotoxic to HeLa cells (9.1% inhibition at 30 µg/mL). This finding indicated the responsible compounds/dose-response against the two cancer cells were different. On the other hand, DCM (100%) and DCM:EtOAc (1:1) eluted fractions were demonstrated to have comparable toxicity against HeLa and PC3 cancer cells. This may be due to the responsible compounds/dose-response of fractions being the same or exerting similar toxicity to HeLa and PC3 cancer cells. The presence of the same compounds in different fractions was also observed, but their quantity might be increased or decreased depending on the polarity of the compound and the eluent solvent used. An increment in percent inhibition of proliferation of cancer cells and a decrease in IC_{50} value of eluents indicated the cytotoxic activity of *R. abyssinicus* was related to responsible bioactive compounds and not to synergistic effect of chemical constituents found in crude extract. Furthermore, the secondary metabolites eluted from the column by less polar solvents were rich in steroids and anthraquinones, which are known to possess cytotoxic effects on cancer cells.

Anti-inflammatory activity

ROS are produced as by-products of normal biochemical reactions in the human body [20]. In inflammatory conditions, NADPH oxidases residing in the immune cells are activated and generate ROS in large quantities, creating an oxidative burst. Overproduction of ROS deregulates the cellular functions, which in turn enhances the inflammatory condition. Therefore, inhibition of ROS-induced oxidative burst is a potential therapeutic to prevent and manage inflammatory-mediated diseases. The plant extract that suppresses the production of ROS may be of paramount importance in regulating diseases that originate from immune cell disturbances. The effect of *R. abyssinicus* extract, fractions, or compounds on production of intracellular ROS from serum opsonized zymosan activated whole blood phagocytes was evaluated by luminol enhanced chemiluminescence technique. In this assay, luminol is used as a probe having a low molecular weight. It goes inside the cell and detects intracellular ROS.

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The extract of \textit{R. abyssinicus} significantly inhibited the production of whole blood ROS by 75.5\% at 50 μg/mL with IC\textsubscript{50} value of 24.8 ± 2.59 μg/mL. However, the inhibitory effect of extract on production of ROS from whole blood was lower compared to the standard drug ibuprofen with an IC\textsubscript{50} value of 11.2 ± 1.9 μg/mL. In agreement with the current findings, Mulisa \textit{et al.} [16] reported a dose-related anti-inflammatory activity of 80\% methanol extract of the rhizomes of \textit{R. abyssinicus} on carrageenan-induced mice paw edema following oral administration and attested the activity to secondary metabolites which have anti-inflammatory activity including tannins, flavonoids, steroids and anthraquinones [16].

Table 2. ROS production inhibition of \textit{R. abyssinicus} extract and fractions and standard drugs.

<table>
<thead>
<tr>
<th>Crude extract, fractions and standard drug</th>
<th>% inhibition of ROS</th>
<th>IC\textsubscript{50} ± SD (μg/mL)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Crude extract</td>
<td>75.5</td>
<td>24.8 ± 2.59</td>
</tr>
<tr>
<td>Pet ether</td>
<td>30.5</td>
<td></td>
</tr>
<tr>
<td>Pet ether:DCM (1:1)</td>
<td>30.5</td>
<td></td>
</tr>
<tr>
<td>DCM</td>
<td>58.8</td>
<td>39.8 ± 7.2</td>
</tr>
<tr>
<td>DCM:EtOAc (1:1)</td>
<td>89.8</td>
<td>18.8 ± 0.9</td>
</tr>
<tr>
<td>EtOAc</td>
<td>53.1</td>
<td>44.1 ± 2.3</td>
</tr>
<tr>
<td>EtOAc:MeOH (1:1)</td>
<td>79.4</td>
<td>26.3 ± 0.7</td>
</tr>
<tr>
<td>MeOH</td>
<td>22.7</td>
<td></td>
</tr>
<tr>
<td>MeOH:H\textsubscript{2}O (1:1)</td>
<td>15.1</td>
<td></td>
</tr>
<tr>
<td>Ibuprofen</td>
<td>73.2*</td>
<td>11.2 ± 1.9</td>
</tr>
</tbody>
</table>

DCM = dichloromethane, EtOAc = ethyl acetate, MeOH = methanol, H\textsubscript{2}O = water, Pet ether = petroleum ether, *= the % production of ROS inhibition was done at a concentration of 25 μg/mL.

This inhibition of whole blood ROS production by the crude extracts could be due to specific phytochemicals present in them. Therefore, the crude extract of \textit{R. abyssinicus} was further partitioned sequentially into eight solvents having different polarities (Table 2), and the capability of preventing the formation of ROS of each fraction was tested. Among the tested fractions, DCM: EtOAc (1:1), EtOAc:MeOH (1:1), DCM (100%), and EtOAc (100%) fractions inhibited more than 50\% production of intracellular ROS from zymosan activated whole blood phagocytes by 89.8\%, 79.4\%, 58.8\% and 53.1\% at a concentration of 50.0 μg/mL, respectively. The DCM: EtOAc (1:1), and EtOAc:MeOH (1:1) fractions with IC\textsubscript{50} values of 18.8 ± 0.9, and 26.3 ± 0.7 μg/mL were found to be strong inhibitors of ROS from human whole blood cells compared to DCM (100%) and EtOAc (100%) fractions with an IC\textsubscript{50} value of 39.8 ± 7.2 μg/mL, and 44.1 ± 2.3 μg/mL, respectively. However, the standard drug Ibuprofen (IC\textsubscript{50} = 11.2 ± 1.9 μg/mL) demonstrated stronger inhibitory potential on the production of ROS compared to all fractions.

\textbf{Antifungal activity}

The crude extract was evaluated for antifungal activity against six fungal strains; \textit{Trichophyton rubrum}, \textit{Candida albicans}, \textit{Aspergillus niger}, \textit{Microsporum canis}, \textit{Fusarium lini}, \textit{Candida glabarata}, and \textit{A. furrigatol} (Table 3). The disk diffusion method was used and the zones of growth inhibition of 80\% ethanolic \textit{R. abyssinicus} extract was measured in millimeters (mm) compared to the standard positive control, amphotericin B for \textit{Aspergillus niger} and miconazole for the rest of the tested fungi, and a negative control, DMSO. The zone of inhibition measured was presented in Table 3.

\textit{R. abyssinicus} extracts demonstrated moderate antifungal activity against \textit{Aspergillus niger} with growth inhibition of 55.5\% at a concentration of 3 mg/mL. It also showed low antifungal activity against \textit{Trichophyton rubrum}, \textit{Microsporum canis}, and \textit{Fusarium lini} pathogenic fungal.
In vitro antifungal, anti-inflammatory and cytotoxic activities of *R.* *abyssinicus* rhizome extract

Aspergillus niger is a fungal strain that causes a disease called "black mold" on certain fruits, produces potent mycotoxins; and is also a cause of pathogenic allergens generally associated with lung infections in individuals with a weak immune system [20]. The current findings are consistent with previous study which reported that MeOH, EtOAc and n-BuOH extracts of *R.* *abyssinicus* can be a potential source of antifungal agents [12].

Table 3. *In vitro* antifungal activity of *R.* *abyssinicus* extracts against pathogenic fungal strains.

<table>
<thead>
<tr>
<th>Fungal strain</th>
<th>% growth inhibition of (3mg/mL)</th>
<th>% growth inhibition of standard drug</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Amphotericin B</td>
<td>Miconazole</td>
</tr>
<tr>
<td><em>Trichophyton rubrum</em></td>
<td>20.4</td>
<td>70.0</td>
</tr>
<tr>
<td><em>Candida albicans</em></td>
<td>NI</td>
<td>110.0</td>
</tr>
<tr>
<td><em>Aspergillus niger</em></td>
<td>55.5</td>
<td>95.4</td>
</tr>
<tr>
<td><em>Microsporum canis</em></td>
<td>30.0</td>
<td>-</td>
</tr>
<tr>
<td><em>Fusarium lini</em></td>
<td>25.2</td>
<td>73.25</td>
</tr>
<tr>
<td><em>Candida glabrata</em></td>
<td>NI</td>
<td>110.8</td>
</tr>
<tr>
<td><em>A. furrigatol</em></td>
<td>NI</td>
<td>100.0</td>
</tr>
</tbody>
</table>

NI = no inhibition.

Isolation of compounds from cytotoxic fractions

Five compounds were isolated from the cytotoxic fractions of *R.* *abyssinicus*. The spectroscopic data of isolated compounds agree well with those reported for chrysophanol (1) [21], physcion (2) [22], emodin (3) [23], citreorosein (4) [24] and β-sitosterol (5) [25].

Cytotoxic activity of compounds

The isolated constituents were estimated for their cytotoxic effects against human cervical cancer (HeLa) and human prostate cancer (PC3) cells. Among the tested compounds, chrysophanol (1) and physcion (2) demonstrated strong cytotoxic activity against HeLa/PC3 cells with an IC50 values < 30 µM. According to the American National Cancer Institute (NCI), the compound is said to have cytotoxic activity if the IC50 value is < 30 µg/mL [26].

Emodin (3) inhibited the growth of HeLa and PC3 cancer cells by 76.7 and 36.6% at 30 µM. It showed strong cytotoxicity to HeLa cells with an IC50 value of 8.94 ± 1.12 µM. Deng et al. [26] reported that emodin (3) demonstrated a dos-dependent cytotoxic effect against PC3 cells [27]. Similarly, Yuenyongsawad et al. [27] reported that emodin (3) demonstrated cytotoxic effect against HeLa cells with an IC50 value of 0.86 µg/mL [27]. Emodin (3) is also known for its cytotoxic effect against many human cancer cells such as prostate cancer (PC3), liver cancer, lung adenocarcinoma, lung squamous carcinoma, promyelo leukemia, cervical cancer, non-small cell lung cancer, ovary cancer, melanoma cancer, colon cancer, hepatic carcinoma, breast cancer, chronic myeloid leukemia, and gastric cancer cells through its effects across multiple signaling pathways [26-30].

Chrysophanol (1) and physcion (2) demonstrated better cytotoxicity to PC3 cells with IC50 values of 22.5 ± 1.7 and 28.5 ± 2.2 µM. They exhibited weak cytotoxicity to HeLa cells. Chrysophanol (1) and physcion (2) reported to exhibit growth inhibitory activity against HeLa cells in a dose dependent manner [31]. According to Lee et al. [30], chrysophanol (1) and physcion (2) showed strong cytotoxic effects against several cancer cell lines, including non-small cell lung, ovary cancer, melanoma cancer, central nervous system, colon cancer, breast cancer, and human colorectal adenocarcinoma cells [27, 28].

Citreorosein (4) showed notable cytotoxicity to HeLa and PC3 cells and inhibited the growth by 48.3 and 34.5% at 30 µM. Lu et al. [32] reported that citreorosein (4) suppresses the gene expression of pro-inflammatory cytokines including tumor necrosis factor (TNF)-α, interleukin

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Table 4. Anti-inflammatory activity of compounds

<table>
<thead>
<tr>
<th>Compounds</th>
<th>% inhibition on HeLa</th>
<th>IC₅₀ ± SD</th>
<th>% inhibition on PC3</th>
<th>% inhibition of ROS IC₅₀ ± SD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chrysophanol (1)</td>
<td>33.8 %</td>
<td>-</td>
<td>64.5</td>
<td>22.5 ± 1.7</td>
</tr>
<tr>
<td>Physcion (2)</td>
<td>29.5</td>
<td>-</td>
<td>56</td>
<td>28.5 ± 1.2</td>
</tr>
<tr>
<td>Emodin (3)</td>
<td>86.7</td>
<td>8.94 ± 1.12</td>
<td>36.6</td>
<td>-</td>
</tr>
<tr>
<td>Citreorosein (4)</td>
<td>48.3</td>
<td>-</td>
<td>34.5</td>
<td>-</td>
</tr>
<tr>
<td>β-sitosterol (5)</td>
<td>8.4</td>
<td>-</td>
<td>16.5</td>
<td>-</td>
</tr>
</tbody>
</table>

Extracts, fractions, or compounds that have a potential to inhibit ROS-induced oxidative burst can serve as effective anti-inflammatory agents. Therefore, these results provided evidence for the anti-inflammatory potential of *R. abyssinicus* extract, fractions, and isolated active...
In vitro antifungal, anti-inflammatory and cytotoxic activities of *R. abyssinicus* rhizome extract. Furthermore, it indicates the potential application of *R. abyssinicus* in the prevention and management of ROS-induced inflammatory conditions as a natural source of anti-inflammatory drug candidates.

**EXPERIMENTAL**

**Materials**

All solvents were ACS grade (Carlo Erba reagents S.A.S, Val de Reuil, France); Dulbecco's Modified Eagle Medium (Gibco, ThermoFisher Scientific, USA); fetal bovine serum (ThermoScientific and ScienCell, USA); penicillin-streptomycin (Invitrogen, ThermoFisher Scientific, USA); 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (Invitrogen, ThermoFisher Scientific, ScienCell, USA); Sabouraud dextrose agar (Thermo-Scientific and ScienCell, USA); amphotericin B (Formepharma, Pakistan); Miconazole (Formepharma, Pakistan); doxorubicin (ICN, USA); Hanks Balanced Salt Solution (HBSS™) (Sigma, St. Louis, USA); zymosan (Fluka, Buchs, Switzerland); Luminol (Research Organics, Cleveland, USA); 96-well white half area plates (Costar, NY, USA). A human cervical cancer (HeLa), human prostate cancer (PC3), and normal human foreskin fibroblasts (BJ) cells were obtained from the cell bank at Dr. Panjwani Center for Molecular Medicine and Drug Research (PCMD), International center for chemical and biological sciences (ICCBS), University of Karachi, Karachi, Pakistan.

**Apparatus and instrument**

The compounds reported in this work were isolated using vacuum liquid chromatography (VLC) which carries 1 kg silica gel 60HF, and three sizes (big, medium, and small) column chromatography (CC) which can carry 200.0, 80.0, and 50.0 g silica gel 60HF, respectively. Fractions collected from VLC were purified using column (50mm x50cm) which can carry 200 g silica gel 60HF. TLC was performed on pre-coated plates (Silica gel 60 F254, 230-400 mesh, Merck) and Al₂O₃ plates. Spots were detected by observation under UV light (Vilber Lourmat). Spraying agents used were 10% cerium (IV) sulphate (Ce(SO₄)₂) and 5% KOH. Melting point (mp) was determined in capillary tube with a digital electrothermal melting point apparatus. UV-Vis spectral measurements were done on a Shimadzu UV-VIS recording spectrophotometer, UV-160, spectronic genesys spectrophotometer. The IR (KBr) spectral measurements were done on a Perkin Elmer 1600 and Pye Unicam Infrared spectrophotometer SP3-300. Electron ionized mass spectrometry (EI-MS) was carried out using JEOL-600H-1. The nuclear magnetic resonance (NMR) spectra were recorded in deuterated solvents (CDCl₃ or (CD₃)₂CO) on a Bruker Avance III 400 and Avance Neo 600 MHz NMR spectrometers. All chemical shifts (δ) are reported in parts per million (ppm) with the solvent signal as a reference relative to TMS (δ = 0) as internal standard, while the coupling constants (J) are given in Hertz (Hz).

**Plant collection**

The rhizome of *R. abyssinicus* was collected in October, 2018, inside Arat Kilo Campus, Addis Ababa University, Addis Ababa, Ethiopia. This plant was identified and authenticated by a taxonomist and voucher specimen (JA-04-2019) was deposited at the National Herbarium, Department of Biology, College of Natural and Computational Sciences, Addis Ababa University, Addis Ababa, Ethiopia. Botanical names have been transcribed according to the nomenclature system used by The World Flora Online (http://www.theplantlist.org).
Extraction and fractionation

The air-dried and pulverized rhizome of *R. abyssinicus* (1.0 kg) was soaked in EtOH:H₂O (8:2) (5 L) for two weeks at room temperature (20-25 °C). The combined extracts were filtered with Whatman filter paper (type 2) and concentrated under pressure using rotary evaporator (Heidolph Hei-VAP, Germany) preset at 40 °C yielding a dark red powder (41.0 g). The extract (40.0 g) was pre-adsorbed on silica gel 60HF (60.0 g) and introduced to vacuum liquid chromatography (VLC) packed with silica gel 60HF (500 g) as stationary phase. The elution was carried out with different solvent gradient systems using (2000 mL each): pet ether (100%), pet ether: DCN (1:1), DCN (100%), DCN: EtOAc (1:1), EtOAc (100%), EtOAc: MeOH (1:1), MeOH (100%), and MeOH: H₂O (1:1) successively. The eluents were separately collected and evaporated to dryness to afford 3.5, 7.6, 9.6, 0.9, 4.5, 2.6 g each, respectively.

Isolation of compounds

Pet ether: DCN (1:1), DCN (100%), and DCN: EtOAc (1:1) eluted fractions were separately pre-adsorbed on silica gel 60HF and fractionated on different size column packed with silica gel 60HF. A pet. ether: DCN (1:1) fraction (7.0 g) was pre-adsorbed on silica gel 60HF (10.0 g), packed with silica gel 60HF (180 g) on CC (50 mm x 50 cm), and eluted with pet ether: acetone (99.75:0.25, 99.5:0.5, 99:1, 49:1, 24:1(2), 9.5:0.5, 9:1, 8:2, 7:3, 1:1 400 mL each), successively. A total of 40 fractions (100 mL each) were collected (F1–F40). Fraction F9–F14 (1.0 g) was adsorbed on silica gel 60HF (1.0 g), packed with silica gel 60HF (50.0 g) on CC (25 mm x 50 cm), and eluted with pet ether: acetone (99.5: 0.5, 99:1, 49:1(2), 24:1(2), 9:1, 8:2, 200 mL each). Out of 40 fractions (50 mL each) collected (Fa1–Fa40), fraction Fa21–Fa23 afforded compound 1 (250 mg) as a clear orange solid after solvent removal under reduced pressure. In addition, fraction F24–F30 (1.9 g) was also pre-adsorbed on silica gel 60HF (2.0 g), packed with silica gel 60HF (50.0 g) on CC (25 mm x 50 cm), and eluted with pet ether: acetone (99:1, 49:1, 24:1, 9:1, 8:2, 1:1 200 mL each) to give Fb1–Fb56 a fractions (25 mL each). Compound 2 (27.5 mg) was obtained from Fb18–Fb21 fraction as an orange solid.

DCM (100%) fraction (9.0 g) was adsorbed on silica gel 60HF (9.0 g), packed with silica gel 60HF (200.0 g) on CC (50 mm x 50 cm), and eluted with pet ether: acetone (99.75:0.25, 99.5:0.5, 99:1, 49:1, 24:1, 9.5:0.5, 9:1, 8:2, 7:3, 1:1 400 mL each) successively. A total of 40 fractions (100 mL each) were collected (F1–F40). Fraction F12–F18 (1.0 g) pre-adsorbed on silica gel 60HF (2.0 g), packed with silica gel 60HF (50.0 g) on CC (25 mm x 50 cm) and eluted with pet ether: acetone (99.75:0.25, 99:1, 49:1, 24:1, 9:1, 8:2, 200 mL each). A total of 30 fractions (50 mL each) were collected (Fc1–Fc30), and fraction Fe10–Fe14 and Fe18–Fe22 afforded compounds 1 (12.2 mg) and compound 2 (15.6 mg), respectively, as orange solids. In addition, fraction F22–F29 (1.7 g) was pre-adsorbed on silica gel 60HF (2.0 g) and packed with silica gel 60HF (50.0 g) on CC (25 mm x 50 cm) and eluted with pet ether: acetone (99.75:0.25, 99:0.5, 99:1, 49:1, 24:1, 9.5:0:5, 9:1, 8:2, 7:3, and 1:1 200 mL each). A total of 80 fraction (25 mL each) were collected (Fd1–Fd80). Fraction Fd32–Fd37 and Fd59–Fd65 afforded compound 24 (725 mg) and compound 4 (140 mg) as white and red solids, respectively.

DCM: EtOAc (1:1) fraction (0.7 g) was adsorbed on silica gel 60F(1.0 g), packed with silica gel 60F (20.0 g) on CC (50 mm x 50 cm), and eluted with pet ether: acetone (49:1, 24:1, 9.5:0.5, 9:1, 2:8, 2:7, 3:1, and 0:1 200 mL each) successively. A total of 40 fractions (25.0 mL each) were collected (Fe1–Fe80). Compound 4 (17.1 mg) and compound 5 (27.8 mg) were obtained as a red and orange solids from fraction Fe32–Fe34 and Fe44–Fe56, which were eluted with 9:1 and 8:2 pet ether in acetone.

Characterization and identification of compounds

Chrysophanol (1). An orange-red powder; mp 194-197 °C; UV-Vis (MeOH) λmax nm: 259, 290 and 433; IR (KBr): νmax cm⁻¹: 3417, 1676, and 1631; EI MS: [M]+ peak at m/z 254.0 corresponding Bull. Chem. Soc. Ethiop. 2022, 36(4)
C_{13}H_{29}O_{8} (calc. mass, 254.0579); 1H-NMR (500 MHz, CDCl$_3$), chemical shift $\delta$ in ppm, coupling constant $J$ in Hz: $\delta$H 1.43 12.11 (1H, s, OH-8), 12.00 (1H, s, OH-1), 7.81 (1H, dd, $J$ = 7.55, 0.9, H-5), 7.65 (1H, $d$, $J$ = 8.45, H-6), 6.74 (1H, $d$, $J$ = 0.9 Hz, H-4), 7.27 (1H, dd, $J$ = 8.45, 0.9, H-7), 7.09 (1H, brx, H-2), 2.45 (3H, s, H-11); 13C NMR (125 MHz, CDCl$_3$): $\delta$C 162.8 (C-1), 124.4 (C-2), 149.4 (C-3), 121.4 (C-4), 119.9 (C-5), 137.0 (C-6), 124.6 (C-7), 162.5 (C-8), 192.6 (C-9), 182.0 (C-12), 22.3 (C-11), 133.3 (4a), 115.9 (8a), 113.8 (9a), 133.7 (10a).

_Emodin_ (3). An orange-yellow powder; mp 204-208 °C; UV-Vis (MeOH) $\lambda_{max}$ nm: 265, 289 and 439; IR (KBr) $\nu_{max}$ cm$^{-1}$: 3468, 1672, and 1629; EI MS: [M$^+$] peak at m/z = 284.1 corresponding to molecular formula (calc. mass, 284.0685); 1H-NMR (400 MHz, CDCl$_3$), chemical shift $\delta$ in ppm, coupling constant $J$ in Hz: $\delta$H 12.35 (1H, s, OH-8), 12.15 (1H, s, OH-1), 7.65 (1H, s, H-4), 7.39 (1H, $d$, $J$ = 2.5, H-5), 7.10 (1H, s, H-2), 3.76 (3H, s, OCH$_3$), 2.47 (3H, s, H-11); 13C NMR (100 MHz, CDCl$_3$): $\delta$C 165.2 (C-1), 124.5 (C-2), 148.5 (C-3), 121.3 (C-4), 106.8 (C-5), 166.6 (C-6), 108.2 (C-7), 162.5 (C-8), 190.8 (C-9), 182.1 (C-10), 22.2 (C-11), 133.2 (4a), 110.3 (8a), 113.7 (9a), 135.3 (10a), 56.1 (OCH$_3$).

_Citroerosein_ (4). An orange amorphous needles; mp 254-256 °C; EI MS: a [M$^+$] peak at m/z = 270.1 corresponding to molecular formula C$_{13}$H$_{12}$O$_{5}$ (calc. mass, 270.0528); UV-Vis (MeOH) $\lambda_{max}$ nm: 265, 289 and 437; IR (KBr) $\nu_{max}$ cm$^{-1}$: 3388, 2923,1687, 1633; 1H-NMR (400 MHz, CDCl$_3$), chemical shift $\delta$ in ppm, coupling constant $J$ in Hz: $\delta$H 7.56 (1H, s, H-4), 7.13 (1H, s, H-2), 7.24 (1H, $d$, $J$ = 2.24, H-5), 6.65 (1H, $d$, $J$ = 2.28, H-7) and $\delta$ 2.46 (3H, s, CH$_3$); 13C NMR (100 MHz, CDCl$_3$): $\delta$C 163.2 (C-1), 124.9 (C-2), 149.5 (C-3), 121.5 (C-4), 109.7 (C-5), 166.5 (C-6), 108.8 (C-7), 166.2 (C-8), 191.7 (C-9), 182.2 (C-10), 134.2 (4a), 110.4 (8a), 114.4 (9a), 136.6 (10a), 21.9 (CH$_3$).

_β-Sitosterol_ (5). White solid; mp 134-136 °C; UV-Vis (CHCl$_3$) $\lambda_{max}$ nm: no absorption 600-200; IR (KBr) $\nu_{max}$ cm$^{-1}$: 3415, 2937, 2843, 1659, 1447, 1381 and 1048; EI MS: [M$^+$] peak at m/z 414.4 corresponding to molecular formula C$_{28}$H$_{48}$O$_{5}$ (calc. mass, 414.3862); 1H-NMR (500 MHz, CDCl$_3$), chemical shift $\delta$ in ppm, coupling constant $J$ in Hz: $\delta$H 5.33 (1H, m, H-6), 5.30 (1H, m, H-3), 0.99 (3H, s, H-19), 0.90 (3H, s, H-18), 0.83 (3H, t, $J$ = 7.45, H-29), 0.81 (3H, d, $J$ = 6.85, H-26), 0.79 (3H, d, $J$ = 6.8, H-27), 0.66 (3H, s, H-18); 13C NMR (100 MHz, CDCl$_3$): $\delta$C 37.3 (C-1), 31.7 (C-2), 71.8 (C-3), 42.3 (C-4), 140.8 (C-5), 121.7 (C-6), 31.9 (C-7), 31.9(C-8), 50.1 (C-9), 36.5 (C-10), 21.1(C-11), 39.8 (C-12), 42.3 (C-13), 56.8(C-14), 26.1(C-15), 28.2 (C-16), 56.1 (C-17), 119.1 (C-18), 19.4 (C-19), 36.1 (C-20), 18.8 (C-21), 34.0 (C-22), 26.1 (C-23), 45.9 (C-24), 29.2(C-25), 19.8 (C-26), 19.0 (C-27), 23.1 (C-28), 12.0 (C-29).

_Biological activities_

_Cell lines and culture conditions_

The human cervical cancer (HeLa), prostate cancer (PC3), human fibroblast (BJ) normal cells were separately cultured in Dulbecco’s Modified Eagle Medium (DMEA), supplemented with 5% for HeLa and PC3 cells and 10% for BJ cells of fetal bovine serum (FBS). 100 IU/mL of penicillin, 100 µg/mL of streptomycin in 75 cm$^2$ flasks, and kept in 5% CO$_2$ incubator at 37 °C.

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Cytotoxic activity

The cytotoxicity assays (cell viability test) on HeLa, PC3, and BJ was performed according to microculture MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide) method as mentioned by Mosmann [42]. Briefly, 100 μL per well of cell solutions (6 × 10^4 cells per mL HeLa cells, 1 × 10^5 cells per mL PC3 cells, or 6× 10^4 cells per mL BJ cells), were added into 96-well plate and incubated for 24 h at 37 °C. After overnight incubation, medium was removed and 200 μL of fresh medium was added with extract (30 μg/mL)/fraction (30 μg/mL)/compound (30 μM)/standard (30 μM) for 48 h. After this, 200 μL MTT (0.5 µg/mL) was added to each well and incubated further for 4 h at 37 °C. Then 100 μL DMSO was added to each well to dissolve the formazan crystal. The extent of MTT reduction to formazan within cells was calculated by measuring the absorbance at 570 nm (PC and HeLa) or 550 nm (BJ) using a microplate reader (Spectra Max Plus, Molecular Devices, CA, USA). Standard drug doxorubicin as a positive control and DMSO as a negative control was used to find the percent growth inhibition or decrease in viable cells. The IC_{50} value of active compounds (inhibition > 50% at 30 μg/mL) was calculated using the EZ-Fit software.

\[
\text{% inhibition} = \frac{100 - \left( \frac{\text{Mean of O.D. of test substance} - \text{Mean of O.D. of NC}}{\text{Mean of O.D. of PC} - \text{Mean of O.D. of NC}} \right)}{100} 
\]

where O.D. is optical density, NC is negative control and PC is positive control.

Selectivity index

High SI value (> 2) of a compound suggests selective toxicity against cancer cells, while a compound with SI value < 2 is considered to give general toxicity which can also cause cytotoxicity in normal cells [43]. Each SI value is calculated using the formula:

\[
\text{SI} = \frac{\text{IC}_{50} \text{ normal cell}}{\text{IC}_{50} \text{ cancer cell}} 
\]

Anti-inflammatory activity test

Luminol-enhanced chemiluminescence assay was performed, as described by Helfand et. al. [44] with slight modifications. Briefly, 25 μL of 1:20 diluted whole blood in HBSS++ was incubated with 25 μL of test sample (50.0 µg/mL for screening and three different concentrations, 1.0, 10.0, and 100.0 µg/mL for active samples), each in triplicate. Control wells received HBSS++ and cells, but no test sample. Test was performed in white half area 96 well plates which were incubated at 37 °C for 15 min in the thermostat chamber of luminometer (Lab systems, Helsinki, Finland). After incubation, 25 μL of 0.30% serum opsonized zymosan (SOZ) and 25 μL of 7 x 10^5 M of intracellular ROS detecting probe, luminol was added into each well, except blank wells (containing only HBSS++). The results were monitored as relative light units (RLU) reading, with peak and total integral values set with repeated scans at 50 s intervals, and 1 s point of measuring time. Standard used for the assay was Ibuprofen. The inhibition percentage (%) for each extract was calculated using the following formula:

\[
\text{% Inhibition of ROS} = \frac{(\text{RLU}_{\text{control}} - \text{RLU}_{\text{sample}})}{\text{RLU}_{\text{control}}} \times 100\% 
\]

Antifungal activity

The antifungal activity of extract to pathogenic fungi, i.e. trichophyton rubrum, candida albicans, aspergillus niger, microsporum canis, and candida glabarata, was determined using agar tube dilution method [45]. The fungal strains were grown in Sabouraud dextrose agar (SDA) which

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contained 2% maltose (pH 5.5-5.6), prepared by mixing 32.5 g of SDA in 500.0 mL water and steamed to dissolve the contents. Into each screw caps tubes 4 mL media was dispensed. The tubes were autoclaved at 121 °C for 15 min. Tubes were allowed to cool to 50 °C and non-solidified SDA was loaded with 66.6 µL of extracts pipetted from the stock solution (180.0 µg/mL) to give the final concentration of 3000 µg/mL. Then the tubes were allowed to solidify in slanting position at room temperature. Each tube was inoculated with a 4mm diameter piece of fungus removed from a mycelial plugs cut from the edge of seven day old cultures. For non-mycelial growth, an agar surface streak was employed. Media supplemented with DMSO was used as negative control and reference antifungal drug Amphotericin B for *Aspergillus niger* and Miconazole for the remaining tested fungal species were used as a positive control. The tubes that were incubated at 27 °C for 7 days and cultures were examined twice-weekly during incubation. Growth in the extract amended media was determined by measuring linear growth (mm) and growth inhibition calculated with reference to the negative control. The % inhibition from 0-39% considered as low, 40-59% as moderate, and 60-69% as good and above 70 as significantly active. The % inhibition of fungal growth for each extract was calculated using the following formula:

\[
\text{% Fungal growth inhibition} = 100\% - \frac{\text{linear growth in test (mm)}}{\text{linear growth in control (mm)}} \times 100\%
\]

**CONCLUSION**

The bioassay-guided fractionation and isolation of compounds from *R. abyssinicus* against cancer cells resulted in isolation of cytotoxic compounds chrysphanol (1), physicon (2) and emodin (3) that may be a potential therapeutic agent in treatment of various cancers. The strong oxidative burst inhibitory potential of *R. abyssinicus* extract, fractions and isolated compounds emodin (3) and citreorosein (4) proved the effectiveness of *R. abyssinicus* and its isolates as natural alternatives to regulate various forms of pro-oxidative, immune disorders and inflammation. The present finding supports the folk medicinal value of *R. abyssinicus* in treatment of different ailments and provides scientific backing for utilizing them in such herbal preparations.

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