Bull. Chem. Soc. Ethiop. **2021**, 35(2), 243-255. © 2021 Chemical Society of Ethiopia and The Authors DOI: https://dx.doi.org/10.4314/bcse.v35i2.3 ISSN 1011-3924 Printed in Ethiopia Online ISSN 1726-801X

LEVELS OF MAJOR AND TRACE METALS IN EGGPLANT AND SOIL

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(Received May 9, 2021; Revised October 13, 2021; Accepted October 18, 2021)

ABSTRACT. Eggplant is one of the widely used edible vegetables in the world including Ethiopia. The aim of this study was to determine the metal contents of eggplants and their corresponding soils collected from five different areas of Ethiopia. Levels of nine selected metals (Na, Ca, Mn, Fe, Cu, Ni, Zn, Pb and Cd) were determined using flame atomic absorption spectrometry. The mean concentration ranges (mg/kg) of the metals in eggplant were Na (1,384-1,917) > Ca (110-158) > Fe (55.9-94.8) > Mn (34.5-44.2) > Zn (17.9-29.2) > Cu (4.3-10.0) > Pb (1.8-4.5) and in soil Fe (4,714-5,508) > Na (843-1,120) > Ca (237-788), Mn (617-763) > Zn (50.7-99.4) > Cu (44.8-74.0) > Pb (13.8-15.8), respectively. Ni and Cd were below the detection limit. The results indicated that eggplant is a good source of essential macro and micro metals. It is also free from toxic metal Cd. However, it contains higher level of Pb, well above the WHO/FAO permissible level. The bioavailability of the metals in eggplant was investigated by analysis of soil pH, electrical conductivity, total dissolved solid and salinity. Transfer factor exhibited a higher accumulation of Na from the soil to the eggplant but for the other metals it was < 1.

KEY WORDS: Eggplant, Minerals, Soil properties, Transfer factor, Ethiopia

INTRODUCTION

Vegetable is any edible part of a plant, fungus or several algae, which is not sugary to taste [1]. Vegetables belonging to the *Solanaceae* family have great importance, because they have around 90 genera and 3000–4000 species. Plants such as potato, tomato, eggplant, bell pepper and pepper from *Solanaceae* family are widely used by humans mainly for food.

The fruit of the eggplant (*Solanum melangema*) is botanically classified as a berry and contains many edible soft seeds that are bitter because they contain nicotinoid alkaloids. The name of eggplant differs from country to country. It is known as brinjal in England, baigan in India, deberjan in Ethiopia and many more names all over the world. The varieties of eggplant lead to different shapes, sizes and colors of its fruit. The skin of the eggplant has many colors but the purple, green and white are more common [2]. The shape of fruit varies from oblong, ovoid, to long cylindrical. Bitterness in eggplant is due to the presence of glycol alkaloids [3].

Eggplant is generally cultivated as fruit vegetable in sub-tropical and tropical regions of the world. Eggplant is a high-yielding crop and is well-adapted to hot and wet environments. The highest production of eggplant is coming from China and India. Egypt, Turkey and Indonesia have equal rate in the production of eggplant [4].

Vegetables are appreciated because they contribute with many nutrients to the human diet, such as macro minerals Na, K, Ca, Mg, P, and also contain trace minerals such as Cu, Cr, Fe, Mn, Zn, Co and Ni [5]. Eggplant is a good source of minerals and vitamins and in total nutritional value. Eggplant is a popular vegetable that is consumed globally on a daily basis, which is rich in dietary fiber, low in calories and protein, phenolic constituents, water soluble sugars, free reducing sugars and folate [6]. Eggplant contains vitamins such as B6, K and C. Eggplant is a good source of essential minerals such as K, Mg, Na, P, Cu, Cr, Fe, Mn, Ni and Zn [7]. Eggplant is an important source of nutrients in the diets of low income consumers [8].

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Eggplant contains many bioactive constituents that have healthy effects, for example flavonoids, minerals, vitamins, and other products which possess several medicinal properties [9]. The phenolic compounds reduce risk of developing chronic diseases such as cardiovascular disease, cancer, diabetes and delayed vital functions disorders with age [10]. The American Diabetes Association recommends an eggplant-based diet as a choice for the controlling of type-2 diabetes because of the high-insoluble fiber and low soluble carbohydrate contents. Especially white eggplant is used to treat type-2 diabetes [11].

Eggplant has also been used in traditional medicine to treat many diseases. Eggplant is also used as a purgative, to ease urination, and to increase sex drive or sex initiative [12]. Eggplant is used for the treatment of several diseases, including diabetes, arthritis, asthma, cancer and bronchitis. Eggplant is ranked among the top ten vegetables for the free radical scavenging activity [9].

Evaluation of micronutrients levels of vegetables is a growing trend in nutritional studies throughout the world as different micronutrients are necessary for different body functions [13]. Heavy metals are the major contaminants of food, because significant level of toxic chemical elements are frequently released into the seas, rivers, lakes and irrigation channels from industrial wastes and other anthropogenic sources [14]. Plants accumulate heavy metals (toxicants) like Cd and Pb from contaminated soil and water. Human and animals get affected or contaminated by accumulating these metals through the consumption of contaminated plants or vegetables cultivated in contaminated soils and water [15]. The heavy metals have high impact on the contamination of soil and water and may affect vegetables, plants and human health because they are persistent [16].

Environmental factors like temperature, soil, water and genetic factors have their own effects in the chemical composition as well as the physical characteristics of eggplant. Therefore, it is important to determine the metal content of eggplant. Soil is essential for growth of vegetables so it is important to determine the metal contents as well as the pH, salinity, total dissolved solid and electrical conductivity of the soil. Soil serves as a natural medium for growth of plants [17]. Soil organic carbon, nitrogen, phosphorus, and potassium are the major indicators of soil quality and fertility as they are strongly connected to plant growth and productivity [18].

Heavy metals occupy a special position in soil chemistry because they play very important physiological roles in nature. The solubility of heavy metal ions in soil is mainly influence by many factors such as pH, conductivity, and moisture content. Heavy metals may enter the food chain as a result of their uptake by edible plants [17]. Measuring the pH and electrical conductivity (EC) parameters provides valuable information for assessing soil condition for growth of plant, nutrient cycling and biological activity [19].

Some studies have been done on the levels of metals in the eggplant in some countries, South America [2], Turkey [20, 21] and Bangladesh [22, 23]. Recently, some studies have been done on the levels of metals in selected vegetables in Addis Ababa, Ethiopia [24], onion around Meki Town and Lake Ziway, Ethiopia [25], Swiss chard, Addis Ababa, Ethiopia [26] and lettuce from five farms in Ethiopia [27]. However, there is no any report in the literature on the levels of metals in eggplant cultivated in Ethiopia.

The main objective of this study was to determine the metal contents in eggplants and their corresponding soil collected from five different areas of Ethiopia by flame atomic absorption spectrometry (FAAS). Specifically, the scope of the study was (i) to determine metal contents of eggplants and corresponding soil by FAAS, (ii) to determine EC, pH, salinity and total dissolved solid (TDS) content of soil to assess the bioavailability of metals in the eggplant, (iii) to study the correlation of the metals in the eggplants and their corresponding soil, and (iv) to compare the results of this study with results reported in literature.

EXPERIMENTAL

Equipment and apparatus

Plastic knife was used to separate the skin of the eggplant from the fruit. Electronic balance (OPTECH, A205EC, Italy) with precision of ± 0.0001 g was used for weighing both eggplant and soil sample. Ceramic mortar and pestle were used to grind eggplant and soil samples. 250 mL round bottom flasks fitted with reflux condenser were used with Kjeldahl digestion block (UK) to digest the dried and powdered eggplant and soil sample. A refrigerator (Hitachi, Tokyo, Japan) was used to keep the digested sample until the analysis. Volumetric flasks (50 mL) were used during dilution and preservation of samples and preparation of metals standard solutions. Micro pipettes (0-100 µL and 100-1000 µL) were used for measuring volumes. pH/ion meter (WTW Inolab pH/ION Level 2, Germany) using unfilled pH glass electrode was used to measure the pH of soil sample. The EC, salinity and TDS of soil samples were measured using Thermo Orion EC meter (Orion 162A) (USA). Magnetic stirrer was used to stir the mixture of soil sample with distilled water. ZEEnit 700p (Germany) flame atomic absorption spectrometry (FAAS) was used for the determination of various metals viz. Na, Ca, Mn, Fe, Cu, Zn, Ni, Pb and Cd.

Chemicals and reagents

All chemicals and reagents used in the study were of analytical grade. HNO₃ (69.5% reagent grade, ACS, ISO, European Union), HClO₄ (70%, Research-lab Fine Chem Industries, India) and HCl (37%, Riedel-de Haen, AG, Germany) were used for digestion of eggplant and soil samples. Stock standard solution containing 1000 mg/L, in 2% HNO₃, of the metal Na, Ca, Mn, Fe, Cu, Zn, Ni, Pb, and Cd (Buck Scientific Puro-Graphictm, USA) were used for the preparation of calibration standard and spiking experiments.

Description of sampling sites

The sampling sites at Bishoftu, Koka, Meki, Ziway and Alemtena, where eggplant and soil samples were collected are located in Oromia region of Ethiopia (Figure 1). These sampling sites were selected due to their maximum production of eggplant in Ethiopia. The Oromia area is under continuous cultivation throughout the year and has been supplying wide variety of vegetables like eggplant, tomato, onion, cabbage, green pepper, potato, etc. to the capital city and for local consumption for a long period of time. Modern farming practices such as mechanized farming, application of agro-chemicals and selected seeds are significant agricultural inputs for getting better yield in the area.

Collection and preparation of eggplant and soil samples

About one-kilogram eggplants were collected from each of the five different sample sites in polyethylene bags. The eggplant samples kept in polyethylene bags were transported to the laboratory of the Chemistry Department of Addis Ababa University. The skin of the eggplant was separated from the fruit and the fruit was chopped into small pieces using plastic knife in order to facilitate drying. The samples were air-dried for one week. The dried samples were ground into powder using mortar and pestle and sieved to 0.425 mm mesh size. The sieved samples were stored in polyethylene bags and placed in a clean and dry place until digestion.



Figure 1. Map of Ethiopia showing sites from where samples were collected.

The soil samples were collected from five sample sites from the base of the uprooted eggplant using auger and placed in polyethylene bags. The soil samples were dried at ambient temperature for 10 days and crushed into powder using mortar and pestle and sieved to 0.425 mm mesh size. The sieved samples were stored in polyethylene bags and placed in a clean and dry place until the time of digestion.

Sample decomposition

A wet digestion method was used for sample decomposition in this study. Wet acid digestion is based on changing different digestion parameters like volume ratio of reagents added, digestion temperature and digestion time. The optimization of digestion procedures described in next sections was carried out based on the usage of smaller reagent volume, shorter digestion time and reasonable mild temperature for obtaining clear and colorless solutions of the resulting digestates.

Optimization of digestion procedure for eggplants

0.5 g of powder and homogenized sample was transferred to a 250 mL round bottom flask. Different volumes of HNO₃ and HClO₄ at different proportions (v/v) were added and digested at different temperatures (150, 180, 210, 240, 270 and 300 °C) for different periods of time (105,

120, 135, 165 and 180 min). The optimization procedure was determined based on the formation of a clear colorless solution. The digested solution was allowed to cool down and 5 mL of distilled water was added to dissolve residue formed on cooling and gradually swirled and filtered into 50 mL volumetric flask through Whatman 93 filter paper. The clear solution was diluted up to 50 mL in a volumetric flask with distilled water and kept in a refrigerator until analysis using FAAS. The optimization procedures for the sample preparation are shown in Table 1.

ent	Trial	Reagents used	Reagent	Temperature	Digestion	Observation
80 90		-	volume ratio	(°C)	time (h)	
re			(mL)			
for	1	HNO3:HClO4	3:1	300	3:00	Cloudy colorless
E	2	HNO3:HClO4	2:2	300	3:00	Clear light yellow
tio	3	HNO ₃ :HClO ₄	4:1	300	3:00	Clear and colorless*
iza es	4	HNO3:HClO4	3:2	300	3:00	Clear light yellow
um um	5	HNO3:HClO4	5:1	300	3:00	Clear and colorless
0p vol	6	HNO3:HClO4	4:2	300	3:00	Deep yellow
for	1	HNO3:HClO4	4:1	300	1:45	Clear and light yellow
E	2	HNO3:HClO4	4:1	300	2:00	Clear and light yellow
tio	3	HNO3:HClO4	4:1	300	2:15	Clear and light yellow
iza	4	HNO ₃ :HClO ₄	4:1	300	2:30	Clear and colorless*
e tim	5	HNO3:HClO4	4:1	300	2:45	Clear and colorless
tim Op	6	HNO3:HClO4	4:1	300	3:00	Clear and colorless
	1	HNO3:HClO4	4:1	150	2:30	Yellow solution
unc	2	HNO3:HClO4	4:1	180	2:30	Yellow solution
uizatioi nperat	3	HNO3:HClO4	4:1	210	2:30	Clear and light yellow
	4	HNO3:HClO4	4:1	240	2:30	Cloudy and colorless
ter	5	HNO ₃ :HClO ₄	4:1	270	2:30	Clear and colorless*
Op for	6	HNO3:HClO4	4:1	300	2:30	Clear and colorless

Table 1. Different conditions tested for optimization of digestion procedure for 0.5 g of eggplant samples.

*Shows the optimized volume ratio, time and temperature.

Optimization of digestion procedure for soil sample

0.5 g of powder and homogenized soil sample was transferred to a 250 mL round bottom flask. Different volumes of HNO₃, HClO₄ and HCl at quantified proportions (v/v) were added and digested at different temperatures (150, 180, 210, 240, 270 and 300 °C) for different period of time (105, 120, 135, 165 and 180 min). The optimization procedure was determined based on the formation of a clear colorless solution. The digested solution was allowed to cool and 5 mL of distilled water was added to dissolve residue formed on cooling and gradually swirled and filtered into 50 mL volumetric flask through Whatman 93 filter paper. The clear solution was diluted up to 50 mL in a volumetric flask with distilled water and kept in a refrigerator until analysis using FAAS. The optimization procedures for the soil sample preparation are shown in Table 2.

Digestion of eggplant samples

Using the optimized conditions (Table 1), 0.5 g eggplant powdered sample was transferred to a 250 mL round bottom flask and 4 mL 69.5% HNO₃ and 1 mL 70% HClO₄ were added. The mixture was digested on a Kjeldhal digestion block for the optimized time of 2:30 h at the optimized temperature 270 °C. After 2:30 h of the digestion time, the digested solution was allowed to cool to room temperature for about 30 min without disassembling the condenser.

After disassembling the condenser, 10 mL of distilled water was added to the solution. This was done by rinsing the neck of the round bottom flask and tip of condenser that was in contact to dissolve the residue formed due to cooling and minimizing dilution of filter paper by digest residue while filtering with Whatman 110 mm filter paper into 50 mL volumetric flask. Finally, the cooled solution was filled up to the mark with distilled water. This procedure was done three times for each eggplant samples. The digestion of the blank reagent was done simultaneously with the samples. All the solutions were kept in refrigerator until the analysis.

Table 2. Different conditions tested for optimization of digestion procedure for 0.5 g of soil samples.

reagent	Trial	Reagents used	Reagent volume ratio (mL)	Temperat ure (°C)	Digestion time (h)	Observation
for	1	HNO3:HC1:HClO4	1:1:1	300	3:00	Deep yellow
u	2	HNO3:HCl: HClO4	2:1:1	300	3:00	Deep yellow
tio	3	HNO3:HC1:HClO4	3:2:1	300	3:00	Light yellow
uiza les	4	HNO3:HCI:HClO4	3:1:1	300	3:00	Clear and colorless*
um	5	HNO3:HC1:HClO4	6:2:2	300	3:00	Clear and colorless
0p vol	6	HNO3:HC1:HClO4	6:1:1	300	3:00	Clear light yellow
	1	HNO3:HC1:HClO4	3:1:1	150	3:00	Deep yellow
n ure	2	HNO3:HC1:HClO4	3:1:1	180	3:00	Clear light yellow
tio	3	HNO3:HC1:HClO4	3:1:1	210	3:00	Clear light yellow
uiza npe	4	HNO3:HC1:HClO4	3:1:1	240	3:00	Clear and light yellow
tim ter	5	HNO3:HCI:HClO4	3:1 :1	270*	3:00	Clear and colorless*
Op for	6	HNO3:HC1:HClO4	3:1:1	300	3:00	Clear and colorless
for	1	HNO3:HC1:HClO4	3:1:1	270	1:45	Deep yellow
u	2	HNO3:HC1:HClO4	3:1:1	270	2:00	Clear and light yellow
izatio	3	HNO3:HC1:HClO4	3:1:1	270	2:15	Clear and light yellow
	4	HNO3:HC1:HClO4	3:1:1	270	2:30	Clear and light yellow
tin Ie	5	HNO3:HCl	3:1:1	270	2:45	Clear and colorless*
Op tim	6	HNO ₃ :HCl	3:1:1	270	3:00	Clear and colorless

*Shows the optimized volume ratio volume ratio, time and temperature.

Digestion of soil samples

Using the optimized conditions (Table 2), 0.5 g soil powder samples were weighed and transferred to a 250 mL round bottom flask and 5 mL of 3:1:1 ratio of HNO₃, HCl and HClO₄ were added. The mixture was digested on a Kjeldhal digestion block for the optimized time of 2:30 h at the optimized temperature of 270 °C. After 2:30 h of the digestion time the digested solution was allowed to cool to room temperature for about 30 min without disassembling the condenser. After disassembling the condenser, 20 mL distilled water was added to the digest to dissolve the residue formed on cooling and vigorously swirled and filtered into 50 mL volumetric flask through Whatman 110 mm filter paper. Finally, the clear solution was diluted with distilled water. This procedure was done three times for each soil samples. The digestion of the blank reagent was done simultaneously with the samples. All the solutions were kept in refrigerator until the analysis.

Determination of pH, conductivity, salinity and TDS of soil samples

Soil pH, EC, salinity and TDS of the soil samples were determined [19] by mixing 10 g of the soil with 25 mL of distilled water in 50 mL beaker. The mixture was stirred in a magnetic stirrer for 30 min. The pH was measured after calibrating the pH meter with technical pH buffers of 4.01 (for acidic solutions) and 10.00 (for alkaline solutions). The other three parameters (EC,

salinity and TDS) were determined after direct calibration with standard solution according to the procedure given in the instruction manual of the manufacturer [28].

Calibration of instrument for the determination of selected metals by FAAS

For the analysis of all eggplant and soil samples, FAAS instrument was calibrated with known concentration of standard solutions for each target metal. Calibration metal standard solutions were prepared for each of the metals from a standard solution containing 10 mg/L which was prepared from standard stock solutions that contained 1000 mg/L. These standards were diluted with deionized water to obtain four working standards for each metal of interest. Three replicate runs were carried out for each sample. The concentration of Na, K, Ca, Mn, Fe, Ni, Cu, Zn, Pb and Cd were determined by FAAS in both the eggplant and soil samples. Calibration curves were prepared to determine the concentration of metals in the sample solutions. Calibration curve exhibited good linearity with R^2 values ranged > 0.99, which showed that the two variable concentration and absorbance had a good positive correlation and linearity.

Validation of optimized procedure

The efficiency of the optimized procedure was verified by adding the known concentration of each metal to 0.5 g sample of eggplant and soil. The spiked and non-spiked samples were digested and analyzed in the same way using optimized procedure of sample analysis. The results obtained from recovery analysis are shown in Table 3. The percentage of recovery was found to vary from 93 to 109% and 94 to 103% for eggplant and soil samples, respectively, which were within the acceptable range for all the metals.

Metals		Soil						
	Na	Ca	Fe	Zn	Na	Ca	Fe	Zn
Unspiked sample (mg/kg)	1,451±1	110±1	94.8±0.6	17.9±0.3	1,016±1	266±1	5,508±1	44.8±0.2
% spiked	20	50	50	50	20	50	20	50
Amount added (mg/kg)	290	55	47	9	203	133	1102	22
Spiked sample (mg/kg)	1721±1	170±1	140±1	27±0.1	1226±1	391±1	6,588±1	70±0.1
% Recovery	93	109	96	101	103	94	98	100

Table 3. Recovery test results for the optimized procedure of eggplant and soil samples.

RESULTS AND DISCUSSION

Levels of metals in eggplant samples

The results of the analysis (Table 4) showed that metal contents of eggplant varied from the area to area where the eggplant was harvested. This variation could be attributed to the mineral composition of the soil in which the plant had been grown and also to the environmental factors influencing the mineral content of the plant. The concentration of metals in eggplant samples collected from Bishoftu was found as Na > Ca > Fe > Mn > Zn > Cu > Pb. The pattern of concentration of the metal in eggplant collected from Meki, Alemtena and Ziway were the same as that of Bishoftu. But the distribution of the concentration of metals in eggplant samples collected from Koka was Na > Ca > Fe > Mn > Zn > Pb > Cu.

The higher amounts of Na and Pb were found to occur in the sample collected from Koka as compared other sample sites. The concentration of Ca, Cu and Fe were higher in Alemtena, Meki and Bishoftu sample sites, as compared to other sample sites. The concentrations of Mn

and Zn were higher in the sample site of Ziway as compared to other sample sites. The concentrations Na were highest than other metals concentrations in each sample site and the concentrations of Pb were lowest than other metals in each sample site except Koka sample site. The concentrations of Ni and Cd in eggplant samples collected from five different sample sites were below the detection limit.

Metals	Concentration of metals (mean \pm SD, mg/kg, n = 3)								
	Bishoftu	Koka	Alemtena	Meki	Ziway				
Na	$1,451 \pm 1$	$1,917 \pm 1$	$1,858 \pm 1$	$1,384 \pm 1$	$1,600 \pm 1$				
Ca	110 ± 1	114 ± 1	158 ± 1	120 ± 1	135 ± 1				
Mn	39.5 ± 0.1	34.5 ± 0.2	41.8 ± 0.4	43.3 ± 0.2	44.2 ± 0.3				
Fe	94.8 ± 0.6	85.3 ± 0.3	55.9 ± 0.3	72.2 ± 0.1	85.0 ± 0.2				
Cu	6.40 ± 0.02	4.3 ± 0.02	6.5 ± 0.01	10.0 ± 0.3	6.7 ± 0.01				
Zn	17.9 ± 0.1	18.2 ± 0.2	26.3 ± 0.5	21.6 ± 0.4	29.2 ± 0.2				
Pb	4.20 ± 0.01	4.5 ± 0.01	1.8 ± 0.01	2.9 ± 0.01	3.6 ± 0.03				
Ni	BDL	BDL	BDL	BDL	BDL				
Cd	BDL	BDL	BDL	BDL	BDL				

Table 4. Concentration of metals (mg/kg) in eggplant samples.

BDL is below detection limit.

Comparison of the heavy metal contents in eggplant sample with other reported values

As shown in Table 5, the concentrations of Mn and Pb in eggplant of this study were higher than the concentration of Mn and Pb in other reported values. The concentration of Cd in eggplant is reported in some studies [2, 20, 23] but it was not detected in the eggplants in this study. The concentration of Cu in the eggplant of this study is lower than those reported by researchers [20, 21, 23] and but comparable with the study [2]. At the same time, it is much higher than that reported in the study of Rashid et al. [22]. The concentration of Fe in the eggplant of this study is comparable with the other studies [2, 22] while it is lower than in the study of Uddin *et al.* [23]. The concentration of Zn in the eggplant is higher in the study of Demirezen and Aksoy [21] while the results of other studies are comparable with the results of present study. The concentration of Ni in the eggplant is reported in some studies [20, 21], but it is not detected in present study. The concentrations of Zn and Cu in the eggplant of this study are well within the permissible levels of WHO/FAO [29] in the fruits and vegetables. Ni and Cd are not detected in the present study. However, the concentration of Pb in the eggplant of this study is well above the permissible levels of WHO/FAO [29] in the fruits and vegetables.

Table 5. Comparison of heavy metals concentration (mg/kg) of the eggplant with values (concentration) reported in the literature.

Zn	Cu	Mn	Fe	Ni	Cd	Pb	Country origin	Reference
21.4	8.4	13.4	63.4	NR	2.40	3.24	South America	[2]
35.8	0.13	28.8	96.8	NR	NR	NR	Bangladesh	[22]
53.9	22.2	NR	NR	2.18	0.30	3.3	Turkey	[21]
16.4	11.7	NR	285	NR	NR	0.24	Bangladesh	[23]
39.7	14.4	8.2	NR	1.5	1.1	NR	Turkey	[20]
50	40	-	-	1.0	0.2	0.3	WHO/FAO*	[29]
22.6	6.8	40.7	78.6	ND	ND	3.4	Ethiopia	This study

NR = not reported. ND = not detected. *WHO/FAO permissible levels in fruits and vegetables.

The concentration of metals in soil samples

Table 6 shows that the metal contents of soil samples varied between the areas in which the plant was cultivated. This variation can be attributed to the mineral composition of soil in which the plant has been grown and the environmental factors influence the mineral content of the plant. The distribution of the concentration of metals in soil samples that were collected from Koka, Alemtena and Meki were Fe > Na > Mn > Ca > Zn > Cu > Pb. The distribution of the concentration of metals in eggplant samples that collected from Bishoftu was Fe > Na > Mn > Ca > Cu > Zn > Pb. The distribution of the concentration of metals in eggplant samples that collected from Ziway was Fe > Na > Mn > Ca > Zn > Cu > Pb. The results showed that the higher amounts of Na, Ca, Zn and Pb were obtained in the soil collected from Ziway as compared to other sample sites. The higher concentration of Mn, Cu and Fe were obtained in the sample collected from Bishoftu as compared to other sample sites. The concentrations of Pb were lowest than other metals concentrations in each sample site. The concentrations of Ni and Cd in soil samples were below the detection limit.

Metals	Concentration of metals (mean \pm SD, mg/kg, n = 3)								
	Bishoftu	Koka	Alemtena	Meki	Ziway				
Na	$1,016 \pm 1$	$1,083 \pm 1$	$1,320 \pm 2$	843 ± 1	$1,120 \pm 2$				
Ca	266 ± 1	248 ± 2	396 ± 1	237 ± 1	788 ± 1				
Mn	763 ± 1	649 ± 2	653 ± 1	617 ± 1	628 ± 1				
Fe	$5,508 \pm 1$	$4,714 \pm 1$	$5,010 \pm 2$	$4,970 \pm 1$	5,355 ±1				
Cu	74.0 ± 0.2	50.7 ± 0.2	58.3 ± 0.2	51.3 ± 0.2	74.0 ± 0.2				
Zn	44.8 ± 0.2	55.5 ± 0.2	73.9 ± 0.4	51.5 ± 0.5	99.4 ± 0.2				
Pb	15.0 ± 0.1	14.9 ± 0.1	13.8 ± 0.1	14.8 ± 0.2	15.8 ± 0.2				
Ni	BDL	BDL	BDL	BDL	BDL				
Cd	BDL	BDL	BDL	BDL	BDL				

Table 6. Concentration of metals (mg/kg) in soil.

Physicochemical properties of soil

The physicochemical properties of soil of the eggplant farmlands are given in Table 7. The pH of the farmland soil lied between 6.81 and 8.13. The highest pH (8.13) was found in Koka farmland soil and the lowest pH (6.81) was found in Bishoftu farmland soil. Thus, the pH of all the farmland soil samples was near to neutral and in the normal pH range of soil 6.5-8.4 recommended for the cultivation of vegetables. The highest EC of 303 μ s and TDS of 176 mg/L were found in Ziway farmland soil, and the lowest conductivity of 208 μ s and TDS of 124 mg/L were found in Bishoftu farmland soil. The salinity of all the farmland soil was in the range 0.1-0.3%.

Table 7. Mean concentration (mean \pm SD), pH, conductivity, salinity and total dissolved solid of soil sample.

Sample site	pН	EC (µs)	Salinity (%)	TDS (mg/L)
Bishoftu farmland	6.81 ± 0.02	208 ± 4	0.1	124 ± 3
Koka farmland	8.13 ± 0.03	282 ± 2	0.2	134 ± 2
Meki farmland	7.60 ± 0.04	293 ± 5	0.3	133 ± 3
Alemtena farmland	7.80 ± 0.03	287 ± 3	0.2	143 ± 2
Ziway farmland	8.10 ± 0.02	303 ± 3	0.2	176 ± 3

Transfer factor of metals from soil to plants

Transmission of metals from soil to plant is studied using an index called transfer factor (TF). It is calculated as a ratio of the concentration of a metal in the plant to the concentration of metal in soil both represented in the same units. If TF is higher or equals to one, the absorption of metal from soil by the plant is higher. On the other hand, lower value of TF indicates the poor response of plants towards metal absorption and the plant can be used for human consumption. Loading and accumulation of metals in the soil depend on different factors such as the chemical form of elements, pH, and organic matter content as well as electric conductivity [30].

Metal transfer factor from soil to plant is a key component of human exposure to metals via the food chain. Table 8 summarizes the TF of metals for eggplants collected from five farmlands. The TF of metals in eggplant was in the order Na > Ca > Zn > Pb > Cu > Mn > Fe. The TF value of Na is higher than the other metals in all the sample sites with TF > 1. This indicates that the Na may get accumulated in the eggplant from other sources (water, fertilizers and air). The TF values of the other metals (Ca, Mn, Fe, Cu, Zn and Pb) were less than one in all the sample sites. This indicates lower uptake of metals by eggplant from the soil. The lower TF values indicate the poor response of plants toward metal absorption and the plants can be used for human consumption.

Metals	Sample sites								
	Bishoftu	Koka	Alemtena	Meki	Ziway				
Na	1.43	1.77	1.41	1.64	1.43				
Ca	0.486	0.460	0.399	0.506	0.171				
Mn	0.052	0.053	0.046	0.071	0.070				
Fe	0.017	0.018	0.011	0.015	0.016				
Cu	0.086	0.085	0.111	0.195	0.091				
Zn	0.399	0.328	0.356	0.419	0.293				
Pb	0.282	0.302	0.130	0.196	0.229				

Table 8. Transfer factor (TF) values of eggplant samples.

Analysis of variance (ANOVA)

Analysis of variance (ANOVA) uses the F statistic to compare whether the difference between sample means of two or more sets of data is significant or not. If $F_{calculated} > F_{critical}$, it means sample means are significantly different. If $F_{calculated} < F_{critical}$, sample means are not significantly different. During the processes of sample preparation and analysis, a number of random errors may be introduced in each aliquot and in each replicate measurement. The variation in sample mean of the analyte was tested by using ANOVA, whether the source of variation was from experimental procedure or heterogeneity among the samples (i.e. difference in mineral contents of soil, water, atmosphere; variation in application of agrochemicals like fertilizers, pesticides, herbicides etc or other variations in cultivation procedures). The ANOVA results (Table 9) showed that there were significant differences in the mean values of Na, Ca, Mn, Zn and Pb between eggplant samples but there was no significant difference in the mean values of Fe and Cu between the samples. The ANOVA results (Table 9) showed that there were significant differences in the mean values of Na, Ca, Mn, Fe, Cu and Zn between the soil of eggplant but there was no significant difference in the mean values of Pb between the soil samples.

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Parameters	Na	Ca	Mn	Fe	Cu	Zn	Pb			
Eggplant										
$F_{Calculated}$	55.4	38.5	32.4	0.445	2.00	6.60	29.0			
F _{Critical}	3.48	3.48	3.48	3.48	3.48	3.48	3.48			
p-value	8.7×10^{-7}	4.7×10^{-7}	1.1×10^{-5}	0.774	0.171	0.007	1.8×10^{-5}			
Soil										
$F_{Calculated}$	4.74	12.8	9.40	13.3	21.4	9.41	0.548			
F _{Critical}	3.48	3.48	3.48	3.48	3.48	3.48	3.48			
p-value	7.6×10^{-7}	0.001	0.002	0.001	6.9×10^{-5}	0.002	0.705			

Table 9. Analysis of variance (ANOVA) in eggplant and soil samples at 95% confidence level.

Pearson correlation of metals

The Pearson correlation coefficient (r) is used to measure the strength of linear association between two variables. A correlation coefficient of +1 indicates that there is a strong (perfect positive) correlation between the two variables. A correlation coefficient of -1 indicates that there is a strong (negative) correlation relationship between the two variables. The correlation values are categorized as no correlation (r = 0.00-0.19), low correlation (r = 0.20-0.39), medium correlation (r = 0.40-0.59), higher correlation (r = 0.60-0.79) and strong correlation (r = 0.8-1.00) [31].

The correlation coefficients between metals in eggplant shows strong correlation between Zn/Mn and Cu/Na. Medium positive correlation between Mn /Ca, Zn/Ca, Cu/Ca and Ca/Na. Pb/Fe and Cu/Mn have low positive correlation. Zn/Cu has no correlation coefficient between them. All the correlation between Ca/Na, Mn/Na, Fe/Na, Zn/Na, Pb/Na, Fe/Mn, Cu/Fe, Zn/Fe, Zn/Na, Pb/Ca, Pb/Cu, Fe/Ca and Pb/Zn show negative correlation.

The correlation coefficients between metals in soil shows strong correlation between Cu/Fe, Pb/Na and Pb/Ca. Medium positive correlation between Zn/Na, Zn/Ca, Fe/Mn, Cu/Mn, Zn/Cu, Fe/Ca and Pb/Mn. The correlation coefficients between metals Ca/Na, Mn/Na, Cu/Na and Pb/Zn in the soil sample have low positive correlation. The correlation coefficients between metal Zn/Fe and Pb/Cu in the soil have no correlation coefficient between them. All the correlations between Fe/Na, Mn/Ca, Cu/Ca, Zn/Mn and Pb/Fe have negative correlation.

CONCLUSION

The present study determined the levels of metals Na, Ca, Mn, Fe, Ni, Cu, Zn, Pb and Cd in eggplants and their corresponding soils collected from five different sample sites in Ethiopia by FAAS. The concentration of Ni and Cd were below detection limit. The levels of metals (except Fe and Cu) in the eggplants from five different sites were significantly different at p = 0.05. Pearson correlation coefficients showed strong positive correlation between most of the metals except Na which showed strong negative correlation with other metals. The investigation of the metal levels revealed that for most of the metals determined, there was a direct relationship between the levels in the eggplant and the soil in which the plant was grown. The study also showed that the metals were found with some of the values reported in the literature. The concentrations of Pb slightly exceeded the permissible levels by WHO, which could be attributed to the agricultural practices employed such as the use of fertilizer and herbicides [32, 33]. The study showed that Na transfer factor from the soil to the plant was > 1, which indicated

that the eggplant absorbs Na from other sources than soil such as water and air while Fe transfer factor was lowest indicating that it was strongly retained in the soil. Other metals showed transfer factor < 1.

ACKNOWLEDGMENTS

The authors express their gratitude to the Department of Chemistry, Addis Ababa University, Ethiopia for providing the laboratory facilities.

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