

https://dx.doi.org/10.4314/bestj.v20i3.1

BEST JOURNAL

4

أكمك

Nutritional Supplement and Pharmaceutical Potential of *Ipomea batata* L. Leaves: A Review

Ahmed, H^{*1}., Babura, S.R.¹ and Shaayau, S.²

¹Department of Plant Biology, Faculty of Natural and Pharmaceutical Sciences, Bayero University Kano ²Department of Biochemistry and Molecular Biology, Usmanu Danfodiyo University, Sokoto *hahmed.bot@buk.edu.ng

ABSTRACT

Sweet potatoes are an exceptionally essential crop in several parts of the world, being produced in more than 100 countries and positioned as sixth most major crop in the world. The world production of sweet potato have steady declined with the exception of America and Sub-saharan Africa in the last decade. The study was intended to provide latest information on the sweet potato leaves and its nutritional and medicinal potential. It was first introduced into Nigeria in the late 1694 through the early activities of Portuguese and Spanish explorers. Nigeria is one of the largest producers of sweet potato in sub-Saharan Africa with annual production estimated at 3.46 million tons per year. The sweet potato can, however, be harvested many times during the year considerably more than many other greens vegetables. It is one of the few vegetables that can be cultivated in both rainy and dry seasons. The sweet potato leaves are considered to be rich in protein, fibers, vitamin B, Vitamin C and minerals, such as iron, calcium, magnesium, zinc, dietary bioactive compounds, and contain more phenols than any other commercial vegetables. The pharmaceutical importance includes antiinflammatory, anti-cancer, antioxidant, anti-diabetes and anti-mutagenic among others. Sweet potato leaves could serve as an additional leafy green vegetable and promote food security for sustainable growth.

Keywords: Nutrition; Ipomea batata; antioxidant; Potato; Phenol; Vegetable

INTRODUCTION

Sweet potato (Ipomoea batatas L.) originated from Central America, Asia and Africa and widely grown as important staple food in most parts of the world. It is a herbaceous and creeping plant that originated from Central America and Northwestern part of South America in about 300 B.C and later introduced into Europe in the 16th century (Ejechi et al., 2020). The plant belongs to Convolvulacaea, and ranked the fifth most important food crop in the tropics and the seventh in the world food production after wheat, rice, maize, potato, barley, and cassava (Truong et al., 2018). It is an important food, feed and vegetable crop in most tropical developing countries. In Nigerian markets, three varieties are in popular demands: the purple, yellow and white potato (Aguoru et

al., 2015). It is mainly cultivated for its tubers and nutritional valuable source of carbohydrates, lipids, proteins, minerals and vitamin. The leaves have a good taste and much higher contents of calcium and vitamin K than spinach. About 200 wild potato species exist, in addition to thousands of primitive varieties.

Potatoes are the fourth most grown crop in the world, after the cereals rice, wheat, and maize and are the only major food crop that is a tuber (Navarre *et al.*, 2009). It was first introduced into Nigeria in the late 1694 through the early activities of Portuguese and Spanish explorers and was encouraged by the British colonials during the Second World War as their tubers were needed to feed their armed forces in West Africa (Udemezue, 2019).



Nigeria is one of the largest producers of sweet potato in sub-Saharan Africa with annual production estimated at 3.46 million tons per year with Egypt number one in Africa, followed by Malawi and Nigeria is the fourth biggest producer in Africa (Udemezue, 2019).

It is an important food security and early maturity crop that can be intercropped with some crops like yam and maize. More than 85% of the sweet potato production in Nigeria is done by peasant farmers (Udemezue, 2019). The crop has been identified to be the fourth most important root crop in Nigeria after cassava, yam and cocoyam. It is consumed and processed in most parts of the tropics and subtropical countries either eaten boiled, fried, roasted or dehydrated into chips, canned, creamed, cooked and frozen, dried and ground into flour to make biscuits, bread and other pastries (Udemezue, 2019). Sweet potato leaves (SPL) are also consumed in Taiwan and China, and are prepared in various ways (boiled, fried, baked, dehydrated, and fermented) (Leticia *et al.*, 2023). It can be found in dishes in the Philippines, Japan, South Pacific, Taiwan, Cameroon, Nigeria, Malawi, Ghana, Côte d'Ivoire, Burkina Faso and the United States.



Figure 1: An image of sweet potato leaves

Potato tubers are highly specialized organs evolved to improve a plant's chances of survival and to allow vegetative reproduction. Tubers are not derived from roots, but are modified stems, originating on stolons from axillary buds on the underground part of the stem (Navarre et al., 2009). Sweet potato is one of the most important food crops worldwide and its leaves (Figure 1) provide a dietary source of nutrients and various bioactive compounds. Sweet potato leaves (SPL) are considered to be a leafy vegetable consumed by human due to its dietary source of nutrients and various bioactive compounds with health benefits. It is one of the important food crops with high yield, drought tolerance, and the ability to grow in different climates and farming systems (Nguyen, et al, 2021).

Green leaves are nutrient-rich foods that have been an important part of the traditional human diet, and their potential contribution to protein intake is often overlooked (Balfany *et al.*, 2023). Sweet potato has a strong potential to contribute to better nutritional quality of our diets around the world. Therefore this review provides a contemporary information on nutritional quality and health benefit of sweet potato leaves.

Origin of sweet potato

The sweet potato is a herbaceous perennial that is grown as an annual by stem cuttings or plant sprouts from storage roots. Sweet potatoes originated somewhere in the region between the Yucatan Peninsula of Mexico and the mouth of the Orinoco river in north easternVenezuela (Austin, 1988).





Recent studies in chloroplast DNA and molecular phylogeny analyses confirmed the assertion (Roullier, *et al.*, 2013a,

b).Europeans in the 1500s spread the sweet potato to Africa and India, with it arriving in China prior to 1600 (Truong *et al.*, 2018).

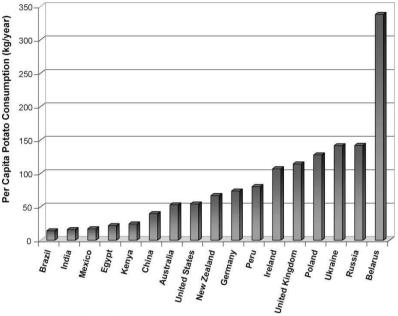


Figure 2: Consumption of potatoes in 17 countries in 2005 (Naverra et al., 2009)

Dietary Importance of Potatoes

Consumption of sweet potato leaves affects serum lipid profiles in both humans and animals, and thus shows potential for reducing the risks associated with the development of cardiovascular disease. With most crops, including potatoes, nutrient profiles are available only for a few varieties. Thus, surprisingly little is known about what vitamins and nutrients are in potatoes. In most of the developed world, potatoes are by far the most eaten vegetable (Figure 2) due to its valuable source of dietary vitamins. minerals. and phytonutrients while in the developing world, potato consumption is increasing at about 5% a year (Navarre et al., 2009). When incorporated into the diets of animals, potato leaves have sweet potential mechanisms to improve dietary protein and

amino acid intake, as well as improve growth performance (Johnson and Ralphenia, 2010). Hotz et al. (2012) reported that increase in consumption of orange-fleshed sweet potatoes improved the vitamin A status of children, pregnant women, and lactating mothers. Sweet potato leaves are rich in crude protein, crude fat, crude fiber, carbohydrate, polyphenols, vitamin C, carotenoids, minerals, and have higher levels of polyphenols than the root, and flesh root tissue (Sun et al., 2017; Makori et al., 2022). The amounts of polyphenols in sweet potato leaves are higher than in most commonly consumed commercial vegetables (Figure 3) such as spinach, kale, amaranth, eggplant, cabbage, cauliflower, green peas, and lettuce (Tan et al., 2023).

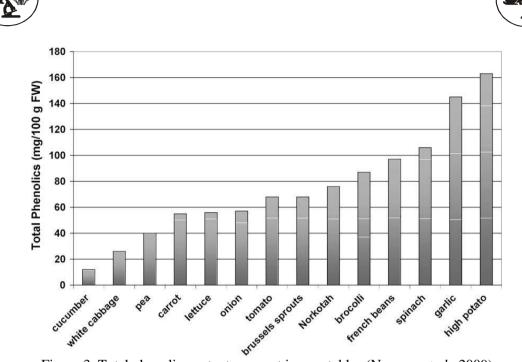


Figure 3. Total phenolic contents present in vegetables (Naverre et al., 2009)

Mineral Elements

All the sweet potato parts (roots, vines, and young leaves) are used as foods, animal feeds and traditional medicine around the world. Potatoes are an important source of different dietary minerals. A wide range of mineral elements occurs in fruits and vegetables, which are a primary dietary source. Fresh sweet potato provides about 50% more calories than Irish potatoes. Its tuberous roots contain about 27% carbohydrate and high concentrations of vitamin A, C, calcium and iron (Coleman and Morre, 2003). The sweet potato leaves (SPL) contains essential mineral elements such as Ca, K, Mg, Na, P, Fe, Cu, Fe, Zn, Niacin, Vitamin B₁, B₂, B₆, Vitamin C, Vitamin E, biotin and β -carotene (Table 1).

Its leaves, stems and roots are a valuable source of bioactive carbohydrates, lipids, proteins, carotenoids, anthocyanins, phenolic acids and flavonoids. These bioactive metabolites possess many biological activities, such as antioxidant, antidiabetic, anticancer, hepatoprotective, antimicrobial, and immunostimulant, antiulcer antimutagenic, and anti-bacterial activity which be helpful for maintaining and may promoting human health (Yoshimoto et al., 2006). Promoting sweet potato leaves consumption in Africa is necessary because it can be a good alternative to reducing malnutrition and health problems caused by nutrients deficiency or poor eating habit.



Elements	Quantity (mg/100g DW)
Water	86.81
Energy (kcal)	42
Carbohydrate (g)	8.82
Protein (g)	2.49
Total fibre (g)	5.3
Total lipid (g)	0.51
Na	8.06 - 832.31
Mg	220.2-910.5
Р	131.1 – 2639.8
Ca	229.7-1958.1
Κ	479.3 - 4280.6
Cu	0.7 - 1.9
Zn	1.2 - 3.2
Mn	1.7 - 10.9
Fe	1.9 - 21.8
Vitamin B3	0.856 - 1.498
Vitamin B6	0.12 - 0.329
Vitamin B2	0.248 - 0.254
Vitamin B1	0.053 - 0.128
Vitamin C	0.0627 - 0.081
Vitamin E	0.00139 - 0.00284
Vitamin B5	0.32 - 0.66
b-carotene	0.273 - 0.4
Biotin	0.003 - 0.008

 Table 1: Mineral and vitamins composition of sweet potato leaves

Source: Ayeleso et al., 2016; Nguyen et al., 2021

Phenolics

Plant phenolics possesses great potential health-promoting compounds and are the most abundant antioxidant compounds in the diet that can have effects on cardiovascular disease, eye health, longevity and mental acuity and (Manach et al., 2004; Scalbert et al., 2005). These has been illustrated by many of the reports in the popular press about positive health benefit coffee, green tea, and wine due to phenolic content. Sweet potato leaves contains at least 6 biologically active polyphenolic compounds and 15 anthocyanin with remarkable multifaceted activity including antioxidation, antimutagenicity, anti-inflammation and anticarcinogenesis (Islam et al., 2014).

Vitamin B₆ (pyroxidine)

Vitamin B_6 is water soluble nutrient and may involve in many metabolic functions than any other nutrient (Tambasco-Studart et al., 2005), it also serve as a cofactor for many enzymes, especially those involved in protein and folate metabolism. It has anticancer and strong antioxidant activities involved in hemoglobin biosynthesis, lipid and glucose metabolism and immune and nervous system function (Denslow et al., 2005; Theodoratou et al., 2008). Very little research has been conducted on this vitamin in potato, thus little is known about how much its concentrations vary among genotypes; ranges of 0.26-0.82 mg/200 g FW have been reported (Rogan et al., 2000).

Recently much has been learned about vitamin B_6 synthesis in plants including identification of the key genes *PDX1* and *PDX2* (Tambasco-Studart *et al.*, 2005). Such information should enable new approaches to further enhance vitamin B_6 concentrations in potatoes.

Vitamin B₉ (Folate)

Potatoes were the third most important overall source of folate in the Dutch diet (Table 2), providing 7% of the total folate intake (Konings *et al.*, 2001), provided 9–12% intake in a Norway (Brevik *et al.*, 2005) and ~10% in Finland (Alfthan *et al.*, 2003).

Crop	Folate content	Reference	
	(µg 100/g FW)		
Rice	6-9	USDA,	
Sweet potato	11	USDA	
Onions	10-19	Konings et al., 2001; USDA	
Tomato	8-30	Konings et al., 2001; USDA	
Banana	13-20	Vahteristo et al., 1997; USDA	
Carrot	16-19	Vahteristo et al., 1997; USDA	
Corn (yellow)	19	USDA	
Orange	18-30	Konings et al., 2001; USDA	
Cassava	27	USDA	
Peas (green)	25-65	Han and Tayler, 2003; USDA	
Strawberry	13 -96	Tulipani et al., 2008	
Snap bean	37	USDA	
Wheat	38	USDA	
Lettuce	38-43	Konings et al., 2001; USDA	
Broccoli	63-114	Vahteristo et al., 1997; USDA	
Spinach	100- 194	Konings et al., 2001; USDA	
Peanut	110-240	Holland et al., 1996; USDA	
Lentis	151-479	Han and Tayler, 2003; USDA	
Beans	143-525	Han and Tayler, 2003; USDA	

Table 2: Folate (Vit. B₆) content in various plant foods.

Source: (Naverra et al., 2009)

Vitamin C

Potatoes are a good source of many vitamins and minerals. The primary storage proteins in tubers are patatins, which account for 40% of the soluble protein content (Prat, 1990). The best known symptom of vitamin C deficiency is scurvy, which in severe cases causes loss of teeth, liver spots, and bleeding. Vitamin C has a major role in detoxifying reactive oxygen species in plants, which are the primary source of vitamin C in the human diet. Leafs and chloroplasts can contain 5 to 25 mM L-

ascorbate, respectively (Wheeler et al., 1998). Plants may have multiple vitamin C biosynthetic pathways, with all of the enzymes of the L-galactose pathway recently characterized (Laing et al., 2007; Wolucka and Montagu, 2007). Vitamin C is а cofactor for numerous enzymes, functioning as an electron donor. Numerous studies have shown that vitamin C levels decrease rapidly during cold storage of potatoes and losses can approach a 60% decrease (Keijbets and Ebbenhorst-Seller, 1990).



Fried potatoes has a 10% decreased in vitamin C after 6 months of storage at -18° C while pre-freezing operations caused 51% loss (Duray *et al.*, 2009), however this sounds cautionary about the importance of how potatos are handled during processing.

Anti-bacterial

Islam (2008)reported the potent antibacterial activity of the leaf water extract of three sweet potato cultivars against Staphylococcus aureus, Bacillus cereus, and E. coli. In contrast, ethanol leaf extract of Brazilian sweet potato did not show any anti-bacterial activities against S. aureus, S. mutans, and S. mitis (Pochapski, et al., 2011), and this could be attributed to methodology differences in the of antimicrobial assay and inherent difference in phytochemical compositions in the SPL due to difference in location. Acetone and ethanol extracts of sweet potato leaves showed antimicrobial activity against Salmonella typhimurium and Pseudomonas aeruginosa (Adsul et al., 2012), peptone, water and ethanol extracts of sweet potato leaves against E. coli, Salmonella typhi, S. Aspergillus niger, Penicillium aureus, aeroginosa and K. pneumonia (Ayeleso et al., 2016). There are still limited studies on the antimicrobial activity of SPL, therefore further studies are needed to elucidate the mechanism of activity.

Anti-cancer

Methanolic leaf extract of SPL inhibit proliferation of all human prostate cancer cells (PC-3, C4-2B, C4-2, DU145, and LNCaP), induce apoptosis, and reductions of chlorogenic survival. Leaves of sweet potato have been recognized as a potent anti-cancer food source against various cancer cells, including HCT-116 colon cancer, HeLa cancer, MCF-7 breast cancer, prostate cancer, colorectal cancer, and lung cancer (Ezekiel *et al.*, 2013; Gundala *et al.*, 2013; Lim *et al.*, 2013; Vishnu *et al.*, 2019). Chlorogenic acid one of the major phenolics compound in potato was reported to be a



strong and selective inhibitor of angiogenic enzymes (matrix metalloproteinase-2, and matrix metalloproteinase-9), A549 human lung cancer cells responsible for tumor metastasis and invasion and (Feng *et al.*, 2005; Jin *et al.*, 2005; Chen *et al.*, 2011).

Anti-diabetes

Phenolic such as caffeoylquinic acid derivatives, and anthocyanins were found to key among the hypoglycemic be contributors in SPL (Table 3). Chlorogenic acid reduces the release of glucose into the blood-stream, lowering the glycemic index, thereby benefitting diabetic patients and reducing the risk of type II diabetes (Bassoli et al., 2008). Polyphenol contents in leaves of 116 sweet potato cultivars grown in China showed anti-diabetic activity (Sun et al., 2014).

Antimutagenic

Sweet potato leaves are a good supplementary resource of antioxidants and antimutagenic compounds (Table 3). An investigation was conducted to examine the effects of 82 kinds of vegetable juice and plant components on the division and multiplication of cancer cells, and it was found that sweet potato has especially high cancer checking rates (Islam, 2014).Sweet potato leaves contain a high content of polyphenolics including, 3-mono-Ocaffeoylquinic acid (chlorogenic acid, ChA), 3,4-di-O-caffeoylquinic acid (3,4-diCOA), 3,5-di-O-caffeoylquinic acid (3,5-diCQA), 4,5-di-O-caffeoylquinic acid (4,5-diCQA), 3,4,5-tri-*O*-caffeoylquinic and acid (Yoshimoto et al., 2006).

Antioxidant activity

Sweet potato leaf contain various antioxidant compounds which contribute to the physiological defense against oxidative stress (Figure 4) and free radical-mediated reactions, leading to an increase in antioxidant defense, suppression of lowdensity lipoprotein (LDL) oxidation and DNA damage in human lymphocytes (Nagai et al., 2011; Chao et al., 2013).



Polyphenol antioxidants, especially caffeoylquinic acid derivatives(3,4,5-tri-O-caffeoylquinic acid, 3,5-di-O-caffeoylquinic acid, and 4,5-di-O-caffeoylquinic acid), exhibit strong antioxidant capacity (Ghasemzadeh *et al*, 2012; Xi *et al.*, 2015; Zhang *et al.*, 2015).

Polyphenols in SPL causes an increase in glutathione by facilitating the expression of –glutamylcysteine synthetase and inhibiting glutathione reductase (Zhange *et al.*, 1997; Moskaug *et al.*, 2005).

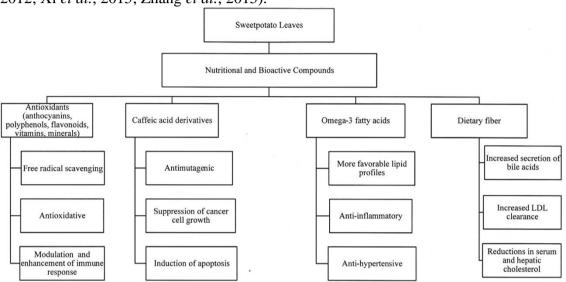


Figure 4: Synergy effects of the major nutritional and bioactive compounds within sweet potato leaves in disease prevention and health promotion (John and Ralphania 2010).

The antioxidant capacity of SPL was found to be much higher than that of other leafy vegetable, and the potato leaf antioxidants tend to vary with the color. Antioxidant capacity of purple SPL was significantly higher than other colored SPL red, yellow, and white (Ji *et al.*, 2015). For instance, purple SPL exhibited higher antioxidants capacity than *Celosia argente*, *Gynura bicolor*, *Perilla frutescens*, *Amaranthus tricolor* and *Houttuynia cordata*, and other

commercial leafy vegetables, due to higher antioxidant (Chu et al., 2000; Lako et al., 2007; Tang et al., 2013). The purple SPL extracts depress neuro-inflammatory responses in lipopolysaccharide-activated BV-2 microglia cells by inhibiting production of pro-inflammatory mediators, such as inducible nitric oxide synthase (iNOS), cyclooxygenase 2 (COX-2), nitric oxide (NO), and TNF (Nguyen et al., 2012).



Phytochemical class	Compound	Function	Reference
Phenolics	Caffeic acid	Antioxidant	Xi et al., 2015;
	derivatives		Jung et al., 2006
		Anti-mutagenic	Yoshimoto et al., 2002
		Antidiabettic	Jung et al., 2006
		Anticancer	Gundala <i>et al.</i> , 2013;
		1 milliounicor	Tang <i>et al.</i> , 2013
		Anti-inflammatory	Ye <i>et al.</i> , 2009;
		rinti initiatinitatory	Zieli <i>et al.</i> , 2021
	Caffeoylquinic	Antioxidant	Zhang <i>et al.</i> , 2016
	acid derivatives	7 milloAldulit	Zhang et u., 2010
		Anticancer	Kurata <i>et al.</i> , 2007;
			Ezekiel <i>et al.</i> , 2013;
			Taira <i>et al.</i> , 2014
		Anti-hypertension	Ishiguro <i>et al.</i> , 2007
		Antidiabetic	Zhang <i>et al.</i> , 2016
		Heart protection	Ezekiel <i>et al.</i> , 2013;
		riean protection	Chao <i>et al.</i> , 2013,
	Chlorogenic acid	Antidiabetic	Bassoli <i>et al.</i> , 2008
	Chilorogenie acid	Anticancer	Ezekiel <i>et al.</i> , 2003;
		7 milleaneer	Gundala <i>et al.</i> , 2013;
			Chen <i>et al.</i> , 2013,
	Quinic acid	Anticancer	Ezekiel <i>et al.</i> , 2013;
	Quinic aciu	Anticalicei	Gundala <i>et al.</i> , 2013,
Flavonoids	Anthocyanins	Antioxidant activity	Chao <i>et al.</i> , 2003;
1 lavonolas	7 minoe yamns	Antioxidant activity	Reddivari <i>et al.</i> , 2003,
		Anti-mutagenic	Islam <i>et al.</i> , 2007
		Anticancer	Konczak <i>et al.</i> , 2004;
		7 milleaneer	Reddivari <i>et al.</i> , 2007,
		Hyppglycemic	Nagamine <i>et al.</i> , 2004
		Hepato-protection	Wang <i>et al.</i> , 2014;
		neputo protection	Olowu <i>et al.</i> , 2011
		Anti-inflammatory	Karlsen <i>et al.</i> , 2007;
		7 mili-milanimator y	Lee <i>et al.</i> , 2015
	Quercetin	Antioxidant	Krochmal-Marczak <i>et al.</i> ,
	Quereeum	mitomunt	2020; Chao <i>et al.</i> , 2003
		Anticancer	Ezekiel <i>et al.</i> , 2013
		Anti-inflammatory	Lee <i>et al.</i> , 2015
	Apigenin	Anticancer	Liu <i>et al.</i> , 2005
	Kaempferol	Anticancer	Ezekiel <i>et al.</i> , 2013
	Myricetin	Anticancer	Knekt <i>et al.</i> , 2002
		Antidiabetic	Jung <i>et al.</i> , 2006
	Fisetin	Anticancer	Lu <i>et al.</i> , 2005
		Anti-inflammatory	Geraets <i>et al.</i> , 2009
	Morin	Anticancer	Kawabata <i>et al.</i> , 1999
		Anti-inflammatory	Galvez <i>et al.</i> , 2001
	Isorhamnetin	Cardioprotection	Sanchez <i>et al.</i> , 2007
	Luteolin	Anticancer	Lim <i>et al.</i> , 2007
	Lucom	Anti-inflammatory	Jang <i>et al.</i> , 2007
		- interminiation y	······

Table 3: phytochemical compounds present in potato leaf and their health benefit

Source: Nguyen, et al, 2021



CONCLUSION

Evidence has been provided on the nutritional benefits of sweet potato leave and its potential to improve human health including the reduction of oxidative damage and the prevention of some diseases. These

REFERENCE

- Adsul VB, Khatiwora E, Torane R, Deshpande NR. (2012) Antimicrobial activities of Ipomoea carnea leaves. *Journal of Natural Product and Plant Resources.***2** (5): 597-600
- Aguoru, C., Uhia, P. and Olasan, J. (2015) Varietal Characterization and Taxonomic Evaluation of Sweet Potato (*Ipomoea batatas*) Using Macro- and Micromorphological Evidence. *Open Access Library Journal*, 2, 1-7. doi: 10.4236/oalib.1101757.
- Austin DF. (1988) The taxonomy, evolution, and genetic diversity of sweet potato and related wild species. In: *Exploration, Maintenance, and Utilization of the* Sweet potato *Genetic Resources*. Lima, Peru: International Potato Center, pp. 27–60.
- Ayeleso, T.B., Ramachela, K. and Mukwevho, E (2016) A review of therapeutic potentials of sweet potato: Pharmacological activities and influence of the cultivar. *Tropical Journal of Pharmaceutical Research*, 15 (12): 2751-2761

http://dx.doi.org/10.4314/tjpr.v15i12.31

- Balfany, C., Gutierrez, J., Moncada, M. and Komarnytsky, S. (2023) Current Status and Nutritional Value of Green Leaf Protein. Nutrients, 15, 1327. https://doi.org/10.3390/nu15061327
- Bassoli, B.K., Cassolla, P., Borba-Murad, G.R., Constantin, J., Salgueiro-Pagadigorria, C.L., Bazotte, R.B., da Silva, R.S.D.S.F. and de Souza, H.M. (2008) Chlorogenic acid reduces the plasma glucose peak in the oral glucose

health promoting benefits are attributed to the presence of various constituents having strong bioactivity. Therefore, SPL can be an alternative natural dietary source providing additional applications in the food supplement and nutraceutical industries.

tolerance test: Effects on hepatic glucose release and glycaemia. *Cell Biochemistry and Function*, 26, 320–328.

- Chao, P.Y., Huang, W.Y., Hu, S.P., Lo, H.F., Lin, K.H., Huang, M.Y., Chang, T.R. and Yang, C.M. (2013) Indigenous purple vegetable extracts protect against hydrogen peroxide-induced DNA damage in human lymphocytes. *Food Nutrition Science*, 4, 62–70.
- Chen, C.M., Li, S.C., Chen, C.Y.O., Au, H.K., Shih, C.K., Hsu, C.Y., Liu, and J.F. (2011) Constituents in purple sweet potato leaves inhibit in vitro angiogenesis with opposite effects ex vivo. *Nutrition*, 27, 1177–1182.
- Chu, Y.H., Chang, C.L. and Hsu, H.F. (2000) Flavonoid content of several vegetables and their antioxidant activity. *Journal of the Science of Food and Agriculture*, 80, 561–566.
- Coleman SW and Moore JE (2003) Feed quality and animal performance. *Field Crops Research*, 84: 17-29.
- Denslow, S. A., Walls, A. A., and Daub, M. E. (2005). Regulation of biosynthetic genes and antioxidant properties of vitamin B6 vitamers during plant defense responses. *Physiological and Molecular Plant Pathology*, 66(6), 244– 255.
- Roullier, C., Benoit, L., Doyle, B. M. and Lebot, V. (2023) Historical collections reveal patterns of diffusion of sweet potato in Oceania obscured by modern plant movements and recombination. *PNAS* 110: 2205–2210.



- Ejechi, M.E., Ode, I.O. and Sugh, E.T (2020) Empirical analyses of production behaviour among small-scale sweet potato farmers in Ebonyi State Nigeria .51 (1): 17-21 http://www.ajol.info/index.php/naj
- Ezekiel, R.; Singh, N.; Sharma, S. and Kaur,
 A. (2013) Beneficial phytochemicals in potato—A review. *Food Research International*, 50, 487–496.
- FAOSTAT. (2020). Statistics division of food and agriculture Organization of the United Nations. http://www.fao.org/faostat/en/#data.
- Feng, R.; Lu, Y.Bowman, L.L.; Qian, Y.; Castranova, V. and Ding, M. (2005) Inhibition of activator protein-1, NF-_B, and MAPKs and induction of phase 2 detoxifying enzyme activity by chlorogenic acid. *Journal of Biological Chemistry*, 280, 27888–27895.
- Galvez, J.; Coelho, G.; Crespo, M.; Cruz, T.; Rodríguez-Cabezas, M.; Concha, A.; Gonzalez, M.; and Zarzuelo, A. (2001) Intestinal anti-inflammatory activity of morin on chronic experimental colitis in the rat. Aliment. *Pharmacology.*, 15, 2027–2039.
- Geraets, L.; Haegens, A.; Brauers, K.; Haydock, J.A.; Vernooy, J.H.;Wouters, E.F.; Bast, A. and Hageman, G.J. (2009) Inhibition of LPS-induced pulmonary inflammation by specific flavonoids. *Biochemical and Biophysical Research Communication*, 382, 598–603.
- Ghasemzadeh, A.; Omidvar, V.; Jaafar, H.Z. (2012) Polyphenolic content and their antioxidant activity in leaf extract of sweet potato (*Ipomoea batatas*). *Journal* of Medicinal Plants Research, 6, 2971– 2976.
- Goyer, A. and Navarre, D. A. (2007). Determination of folate concentrations in diverse potato germplasm using a trienzyme extraction and a microbiological assay. *Journal of*

Agricultural Food Chemistry, 55, 3523–3528

- Gundala, S.R.; Yang, C.; Lakshminarayana, N.; Asif, G.; Gupta, M.V.; Shamsi, S. and Aneja, R. (2013) Polar biophenolics in sweet potato greens extract synergize to inhibit prostate cancer cell proliferation and in vivo tumor growth. *Carcinogenesis*, 34, 2039–2049.
- Han, J. Y., and Tyler, R. T. (2003). Determination of folate concentrations in pulses by a microbiological method employing trienzyme extraction. *Journal of Agricultural Food Chemistry*, *51*, 5315–5318.
- Holland, B., Unwin, I. D. and Buss, D. H. (1996). Vegetables, Herbs and Spices. Fruits and Nuts. Royal Society of Chemistry, Cambridge, UK.
- Hotz, C., Loechl, C., de Brauw, A., Eozenou, P., Gilligan, D., Moursi, M., Munhaua, B., van Jaarsveld, P., Carriquiry, A. and Meenakshi, J.V., (2012). A large scale intervention to introduce orange sweet potato in rural Mozambique increases vitamin A intakes among children and women. *British Journal of Nutrition*. 108:163–176.
- Ishida, H.; Suzuno, H.; Sugiyama, N.; Innami, S.; Tadokoro, T. and Maekawa, A. (2000) Nutritive evaluation on chemical components of leaves, stalks and stems of sweet potatoes (*Ipomoea batatas* poir). *Food Chemistry*. 68, 359– 367
- Ishiguro, K.; Yoshimoto, M.; Tsubata, M. and Takagaki, K. (2007) Hypotensive effect of sweet potato [*Ipomoea batatas*] tops. *Journal of Japanese Society of Food Science Technology*, 54, 45–49.
- Islam, M.S.; Yoshimoto, M.; Terahara, N. and Yamakawa, O. (2002) Anthocyanin compositions in sweet potato (*Ipomoea* batatas L.) leaves. Bioscience. Biotechnology and Biochemistry, 66, 2483–2486.



- Islam, S. (2014). Nutritional and medicinal qualities of sweet potato tops and leaves. Cooperative extension service (p. FSA6135). Fayetteville, Arkansas: University of Arkansas.
- Islam, S. (2008) Antimicrobial activities of Ipomoea batatas (L.) leaf. Journal of Food Agriculture and Environment, 6, 14.
- Jang, S.; Kelley, K.W. and Johnson, R.W. (2008) Luteolin reduces IL-6 production in microglia by inhibiting JNK phosphorylation and activation of AP-1. *Proceedings of the Nattional Academic of Science*. USA, 105, 7534–7539.
- John, M and Ralpania, D.P (2010) Sweet potato leaves: properties and synergistic interactions that promote health and prevent disease. *Nutrition Reviews*. 68(10):604–615doi:10.1111/j.1753-4887.2010.00320.x
- Ji, H.; Zhang, H. Li, H. and Li, Y. (2015) Analysis on the nutrition composition and antioxidant activity of different types of sweet potato cultivars. *Food Nutrition Science*, 6, 161–167.
- Jin, U.H., Lee, J.Y.; Kang, S.K.; Kim, J.K.; Park, W.H.; Kim, J.G.; Moon, S.K. and Kim, C.H. (2005) A phenolic compound, 5-caffeoylquinic acid (chlorogenic acid), is a new type and strong matrix metalloproteinase-9 inhibitor: Isolation and identification from methanol extract of *Euonymus alatus*. *Life Science*, 77, 2760–2769
- Jung, U.J.; Lee, M.K.; Park, Y.B.; Jeon, S.M. and Choi, M.S. (2006). Antihyperglycemic and antioxidant properties of caffeic acid in db/db mice. *Journal of Pharmacological Experiment*, 318, 476–483
- Karlsen, A.; Retterstøl, L.; Laake, P.; Paur,
 I.; Kjølsrud-Bøhn, S.; Sandvik, L. and
 Blomhoff, R. (2007) Anthocyanins
 inhibit nuclear factor-_ B activation in
 monocytes and reduce plasma
 concentrations of pro-inflammatory

mediators in healthy adults. *Journal of Nutrition*, 137, 1951–1954.

- Kawabata, K.; Tanaka, T.; Honjo, S.; Kakumoto, M.; Hara, A.; Makita, H.; Tatematsu, N.; Ushida, J.; Tsuda, H. and Mori, H. (1999) Chemopreventive effect of dietary flavonoid morin on chemically induced rat tongue carcinogenesis. *International Journal of Cancer*, 83, 381–386.
- Keijbets, M. J. H. and Ebbenhorst-Seller, G. (1990). Loss of vitamin C (L-ascorbic acid) during long-term cold storage of Dutch table potatoes. *Potato Research*, *33*, 125–130.
- Knekt, P.; Kumpulainen, J.; Järvinen, R.; Rissanen, H.; Heliövaara, M.; Reunanen, A.; Hakulinen, T. and Aromaa, A. (2002) Flavonoid intake and risk of chronic diseases. *American Journal of Clinical and Nutrition*, 76, 560–568.
- Konczak, I. and Zhang, W. (2004) Anthocyanins—More than nature's colours. *Journal of Biomedical and Biotechnology*, 239–240.
- Konings, E. J., Roomans, H. H., Dorant, E., Goldbohm, R. A., Saris, W. H., and van den Brandt, P. A. (2001). Folate intake of the Dutch population according to newly established liquid chromatography data for foods. *American Journal of Clinical and Nutrition*, 73, 765–776.
- Krochmal-Marczak, B.; Cebulak, T.; Kapusta, I.; Oszmian´ski, J.; Kaszuba, J. and Z[·] urek, N. (2020) The content of phenolic acids and flavonols in the leaves of nine varieties of sweet potatoes (*Ipomoea batatas* L.) depending on their development, grown in Central Europe. *Molecules*, 25, 3473.

Kurata, R.; Adachi, M.; Yamakawa, O. and Yoshimoto, M. (2007) Growth suppression of human cancer cells by polyphenolics from sweet potato (*Ipomoea batatas* L.) leaves. *Journal of Agriculture and Food Chemistry.*, 55, 185–190.



- Laing, W. A., Wright, M. A., Cooney, J. and Bulley, S. M. (2007). The missing step of galactose pathway of ascorbate biosynthesis in plants, an L-galactose gse, of biosynthesis in plants, an Lgalactose guanyltransferase, increases leaf ascorbate content. *Proceedings of the National Academy of Sciences of the United States of America*, 104 (22), 9534–9539.
- J.; Trenerry, V.C.;Wahlqvist, Lako, M.;Wattanapenpaiboon, N.: Sotheeswaran, S.; Premier, R. (2007) Phytochemical flavonols, carotenoids and the antioxidant properties of a selection of Fijian wide fruit. vegetables and other readily available foods. Food Chemistry. 101, 1727-1741
- Lebot, V (2013a). Disentangling the Origins of Cultivated Sweet potato (*Ipomoea batatas* (L.) Lam.). *PLoS ONE* 8(5): e62707.
- Lee, S.L.; Chin, T.Y.; Tu, S.C.; Wang, Y.J.; Hsu, Y.T.; Kao, M.C. and Wu, Y.C. (2015) Purple sweet potato leaf extract induces apoptosis and reduces inflammatory adipokine expression in 3T3-L1 differentiated adipocytes. Evid. Based Complement. *Alternative Medicine*, 126302.
- Lenti, M.; Gentili, C.; Pianezzi, A.; Marcolongo, G.; Lalli, A.; Cancedda, R. and Cancedda, F.D. (2009) Monogalactosyldiacylglycerol antiinflammatory activity on adult articular cartilage. *Natural Product Research*, 23, 754–762
- Leticia X. López-Martínez, Alejandra A. López-Pérez, Aarón F. González-Córdova, Gustavo Adolfo González-Aguilar, Citali Colin Chávez, Lilia M. Beltrán-Barrientos, Julieta Domínguez-Soberanes, and Manuel Vargas-Ortiz (2023) Technologies for the Use and Consumption of Sweet Potato Leaves and Their Bioactive Compounds, *Food*

Science and Technology. 3 (3), 379-393DOI: 10.1021/acsfoodscitech.2c00405

- Lim, D.Y. Jeong, Y.; Tyner, A.L. and Park, J.H. (2007) Induction of cell cycle arrest and apoptosis in HT-29 human colon cancer cells by the dietary compound luteolin. *American Journal* of Physiology and Gastrointestine Liver Physiology, 292, G66–G75.
- Lim, S.; Xu, J.; Kim, J.; Chen, T.Y.; Su, X.; Standard, J.; Carey, E.; Griffin, J.; Herndon, B. and Katz, B. (2013) Role of anthocyanin-enriched purple-fleshed sweet potato p40 in colorectal cancer prevention. *Molecule and Nutrition Food Research*, 57, 1908–1917.
- Liu, L.Z.; Fang, J.; Zhou, Q.; Hu, X.; Shi, X and Jiang, B.H. (2005) Apigenin inhibits expression of vascular endothelial growth factor and angiogenesis in human lung cancer cells: Implication of chemoprevention of lung cancer. Molecule and Pharmacology, 68, 635–643.
- Lu, X.; Jung, J.I.; Cho, H.J.; Lim, D.Y.; Lee, H.S.; Chun, H.S.; Kwon, D.Y.; Park, J.H. (2005) Fisetin inhibits the activities of cyclin-dependent kinases leading to cell cycle arrest in HT-29 human colon cancer cells. *Journal of Nutrition*, 135, 2884–2890.
- Manach, C., Scalbert, A., Morand, C., Remesy, C., and Jimenez, L. (2004). Polyphenols: food sources and bioavailability. American Journal of clinical Nutrition, 79, 727-747 McKillop, D. J., Pentieva, K., Daly, D., McPartlin, J. M., Hughes, J., Strain, J. J., Scott, J. M., and McNulty, H. (2002). The effect of different cooking methods on folate retention in various foods that are amongst the major contributors to folate intake in the UK diet. British Journal of Nutrition, 88, 681–688.



- Moskaug, J.Ø.; Carlsen, H.; Myhrstad, M.C. and Blomhoff, R. (2005) Polyphenols and glutathione synthesis regulation. *American Journal of Clinical Nutrition*, 81, 277S–283S.
- Nagai, M.; Tani, M.; Kishimoto, Y.; Iizuka, M.; Saita, E.; Toyozaki, M.; Kamiya, T.; Ikeguchi, M. and Kondo, K. (2011)
 Sweet potato (*Ipomoea batatas* L.) leaves suppressed oxidation of low density lipoprotein (LDL) in vitro and in human subjects. *Journal of Clinical Biochemistry and Nutrition*, 48, 203–208
- Nagamine, R.; Ueno, S.; Tsubata, M.; Yamaguchi, K.; Takagaki, K.; Hira, T.; Hara, H.; Tsuda, T. (2014) Dietary sweet potato (*Ipomoea batatas* L.) leaf extract attenuates hyperglycaemia by enhancing the secretion of glucagonlike peptide-1 (GLP-1). *Food Function*, 5, 2309–2316.
- Nagamine, R.; Ueno, S.; Tsubata, M.; Yamaguchi, K.; Takagaki, K.; Hira, T.; Hara, H.; Tsuda, T. (2014) Dietary sweet potato (*Ipomoea batatas* L.) leaf extract attenuates hyperglycaemia by enhancing the secretion of glucagonlike peptide-1 (GLP-1). *Food Function*, 5, 2309–2316.
- Navarre, A. D., Goyer, A and Shakya, R. (2009) Nutritional Value of Potatoes: Vitamin, Phytonutrient, and Mineral Content, Chapter 14. USDA-ARS, Washington State University, 395 - 425 USA
- Nguyen, H.C.; Chen, C.-C.; Lin, K.-H.; Chao, P.-Y.; Lin, H.-H. and Huang, M.-Y. (2021) Bioactive Compounds, Antioxidants, and Health Benefits of Sweet Potato Leaves. *Molecules*, 26, 1820. <u>https://doi.org/</u> 10.3390/molecules26071820
- Olowu, A.O.; Adeneye, A.A.; Adeyemi, O.O. (2011) Hypoglycaemic effect of *Ipomoea batatas* aqueous leaf and stem extract in normal and streptozotocin-

induced hyperglycaemic rats. Jundishapur Journal of Natural Pharmaceutical, 2, 56–61.

- Pochapski, M.T.; Fosquiera, E.C.; Esmerino, L.A.; Dos Santos, E.B.; Farago, P.V.; Santos, F.A. and Groppo, F.C. (2011)
 Phytochemical screening, antioxidant, and antimicrobial activities of the crude leaves' extract from *Ipomoea batatas* (L.) Lam. *Pharmacognosy Magazine*, 7, 165.
- Prat, S., Frommer,W. B., Hofgen, R., Keil, M., Kobmann, J., Koster-Topfer, M., Liu, X. J., Muller, B., Pena-Cortes, H., Rocha-Sosa, M., Sanchez-Serrano, J. J., Sonnewald, U., and Willmitzer, L. (1990). Gene expression during tuber development in potato plants. *FEBS Letters*. 268, 334–338.
- Reddivari, L.; Vanamala, J.; Chintharlapalli, S.; Safe, S.H.; and Miller, J.C., Jr. (2007) Anthocyanin fraction from potato extracts is cytotoxic to prostate cancer cells through activation of caspase-dependent and caspaseindependent pathways. *Carcinogenesis*, 28, 2227–2235.
- Rogan, G. J., Bookout, J. T., Duncan, D. R., Fuchs, R. L., Lavrik, P. B., Love, S. L., Mueth, M., Olson, T., Owens, E. D., Raymond, P. J., and Zalewski, J. (2000). Compositional analysis of tubers from insect and virus resistant potato plants. *Journal of Agricultural and Food Chemistry*, 48, 5936–5945.
- Roullier C, Duputié A, Wennekes P, Benoit L, Bringas VMF, Rossel G, Tay D, McKey, D and Lebot, V (2013a). Disentangling the Origins of Cultivated Sweet potato (*Ipomoea batatas* (L.) Lam.). *PLoS ONE* 8(5): e62707
- Roullier C, Benoit L, McKey DB, and Lebot V. (2013b). Historical collections reveal patterns of diffusion of sweet potato in Oceania obscured by modern plant movements and recombination. *PNAS* 110: 2205–2210



- Sanchez, M.; Lodi, F.; Vera, R.; Villar, I.C.; Cogolludo, A.; Jimenez, R.; Moreno, L.; Romero, M.; Tamargo, J. and Perez-Vizcaino, F.(2007) Quercetin and isorhamnetin prevent endothelial dysfunction, superoxide production, and overexpression of p47phox induced by angiotensin II in rat aorta. *Journal of Nutrition*, 137, 910–915.
- Xi, L. and Song, Z. (2014) Effects of domestic cooking methods on polyphenols and antioxidant activity of sweet potato leaves. *Journal of Agriculture and Food Chemistry*, 62, 8982–8989.
- Sun, H.; Mu, T.; Xi, L.; Zhang, M.; and Chen, J. (2014) Sweet potato (*Ipomoea batatas* L.) leaves as nutritional and functional foods. *Food Chemistry*, 156, 380–389.
- Taira, J.; Uehara, M.; Tsuchida, E. and Ohmine,W. (2014) Inhibition of thecatenin/Tcf signaling by caffeoylquinic acids in sweet potato leaf through down regulation of the Tcf-4 transcription. *Journal of Agriculture and Food Chemistry*, 62, 167–172.
- Tambasco-Studart, M., Titiz, O., Raschle, T., Forster, G., Amrhein, N., and Fitzpatrick, T. B. (2005). Vitamin B6 biosynthesis in higher plants. *Proceedings of the National Academy* of Sciences of the United States of America, 102(38), 13687–13692
- Tan, W., Xin-bo, G., Zhang-ying, W., Rong, Z., Chao-chen, T., Bing-zhi, J., Rui-xue, J., Yuan-yuan, D., Shao-hai, Y. and Jing-yi, C. (2023) Metabolic profile and morphological characteristics of leaf tips among different sweet potato (*Ipomoea batatas* Lam.) varieties, *Journal of*

Integrative Agriculture. Accepted 10 March, 2023.

- Tang, S.C.; Lo, H.F.; Lin, K.H.; Chang, T.J.; Yang, C.M. and Chao, P.Y. (2013) The antioxidant capacity of extracts from Taiwan indigenous purple-leaved vegetables. *Journal of Taiwan Society of Horticulture and Science*, 59, 43–57.
- Tang, S.C.; Lo, H.F.; Lin, K.H.; Chang, T.J.; Yang, C.M.; and Chao, P.Y. (2013) The antioxidant capacity of extracts from Taiwan indigenous purple-leaved vegetables. *Journal of Taiwan Society of Horticulture and Science*, 59, 43–57.
- Theodoratou, E., Farrington, S. M., Tenesa, A., McNeill, G., Cetnarskyj, R., Barnetson, R. A., Porteous, M. E., Dunlop, M. G., and Campbell, H. (2008). Dietary vitamin B6 intake and the risk of colorectal cancer. *Cancer Epidemiology and Biomarkers Prevention*, 17, 171–182.
- Truong, V. D., Avula, R.Y., Pecota, K.V and Yencho, G.C (2018) Sweet potato Production, Processing, and Nutritional Quality, *Handbook of Vegetables and Vegetable Processing*, 2, Second Edition. Ed: Muhammad Siddiq and Mark A. Uebersax. John Wiley & Sons Ltd. Pp.811-838
- Tulipani, S., Mezzetti, B., Capocasa, F., Bompadre, S., Beekwilder, J., de Vos, C. H., Capanoglu, E., Bovy, A., and Battino, M. (2008). Antioxidants, phenolic compounds, and nutritional quality of different strawberry genotypes. *Journal of Agriculture and Food Chemistry*, 56, 696–704. Udemezue J.C. (2019) Profitabilities and Constraints to Sweet Potato Production in Nigeria. *Current Trends*

in Biomedical Engineering Bioscience; 19 (2):556007.

and

Ahmed et al. (2023)

Biological and Environmental Sciences Journal for the Tropics 20(3) December, 2023 ISSN 0794 – 9057; eISSN 2645 - 3142

ber, 2023



USDA National Nutrient Database for Standard Reference (SR20) a.gov/main/site_main.htm?modecode=12 354500

- Vahteristo, L., Lehikoinen, K., Ollilainen, V., and Varo, P. (1997). Application of an HPLC assay for the determination of folate derivatives in some vegetables, fruits and berries consumed in Finland, *Food Chemistry*, 59(4), 589–597.
- Vishnu, V.R.; Renjith, R.S.; Mukherjee, A.; Anil, S.R.; Sreekumar, J.; Jyothi, A.N. (2019)Comparative study on the chemical structure and in vitro antiproliferative activity of anthocyanins in purple root tubers and leaves of sweet potato (Ipomea batatas). Journal of Agriculture and Food Chemistry, 67:2467-2475.
- Wang, W.; Li, J.; Wang, Z.; Gao, H.; Su, L.; Xie, J.; Chen, X.; Liang, H.; Wang, C. and Han, Y. (2014) Oral hepatoprotective ability evaluation of purple sweet potato anthocyanins on acute and chronic chemical liver injuries. *Cell Biochemistry* and Biophysical Research, 69, 539–548.
- Wheeler, G. L., Jones, M. A., and Smirnoff, N. (1998). The biosynthetic pathway of vitamin C in higher plants. *Nature* (*London, England*) *Nature*, 393, 365– 369.
- Wolucka, B. A., and Montagu, M. v. (2007). The VTC2 cycle and the de novo biosynthesis pathways for vitamin C in plants: An opinion. *Phytochemistry*, 68(21), 2602–2613.
- Xi, L.; Mu, T. and Sun, H. (2015) Preparative purification of polyphenols from sweet potato (*Ipomoea batatas* L.) leaves by AB-8 macroporous resins. *Food Chemistry*, 172, 166–174.
- Xi, L.; Mu, T.; Sun, H. (2015) Preparative purification of polyphenols from sweet potato (*Ipomoea batatas* L.) leaves by AB-8 macroporous resins. *Food Chemistry*, 172, 166–174

- Ye, Z.; Liu, Z.; Henderson, A.; Lee, K.; Hostetter, J.; Wannemuehler, M. and Hendrich, S. (2009) Increased CYP4B1 mRNA is associated with the inhibition of dextran sulfate sodium–induced colitis by caffeic acid in mice. *Experimental Biology and Medicine*, 234, 605–616.
- Yoshimoto, M., Kurata, R., Okuno, S., Ishiguro, K., Yamakawa, O., Tsubata, M., Mori, S. and Takagaki, K. (2006). Nutritional value and physiological functions of sweet potato leaves. *Acta Horticulture*. 703, 107-116 DOI: 10.17660/ActaHortic.2006.703.11
- Yoshimoto, M.; Yahara, S.; Okuno, S.; Islam, M.S.; Ishiguro, K. and Yamakawa, O. (2002) Antimutagenicity of mono-, di-, and tricaffeoylquinic acid derivatives isolated from sweet potato (*Ipomoea batatas* L.) leaf. *Bioscience Biotechnology and Biochemistry*, 66, 2336–2341.
- Zhang, K.; Yang, E.B.; Tang,W.Y.;Wong, K.P.; Mack, P. (1997) Inhibition of glutathione reductase by plant polyphenols. *Biochemical Pharmacology*, 54, 1047–1053.
- Zhang, L.; Tu, Z.C.; Yuan, T.; Wang, H.; Xie, X.; Fu, Z.F. (2016) Antioxidants and glucosidase inhibitors from *Ipomoea batatas* leaves identified by bioassayguided approach and structure-activity relationships. Food Chem., 208, 61–67.
- Zhang, L.; Tu, Z.C.; Wang, H.; Fu, Z.F.; Wen, Q.H.; Chang, H.X.; Huang, X.Q. (2015)
 Comparison of different methods for extracting polyphenols from *Ipomoea batatas* leaves, and identification of antioxidant constituents by HPLC-QTOF-MS2. *Food Research International*, 70, 101–109
- Zieli´nska, D.; Zieli´nski, H.; Laparra-Llopis, J.M.; Szawara-Nowak, D.; Honke, J.; Giménez-Bastida, J.A. (2021) Caffeic acid modulates processes associated with intestinal inflammation. *Nutrients*, 13, 554.