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**Review** 

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# Roles of *Trichoderma species* as a Potent Biochemical Agent for Sustainable Farming Practice: A Mini Review

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In the past few decades, fungi have attracted many scientists globally to develop an interest in using *Trichoderma species* and other related fungi specifically for organic farming purposes. *Trichoderma spp.* as an endophytic and saprophytic fungi found in diverse natural habitats such as soil, compost, plant roots, leaves, woods and so forth. They play a crucial role in different agricultural processes similar to the synthetic counter parts for the improvement of crop yield. In this mini review, numerous findings from old of 1989 to the recent of 2021 signifying the several researches as well as mechanisms in which *Trichoderma spp.* operate which are very crucial for the agricultural development and sustainability in providing cheaper, safer, healthier and nutritious produce as well as provision of environment free of contaminants. In this review it was gathered that these fungi could be one of the promising species that can burst farming practice when use as biofertilizer

**Keywords:** Ecosystem, Crop yield, Organic farming, Sustainability, *Trichoderma spp.* 

### 1. Introduction

Trichoderma species are endophytic and saprophytic in nature that is found to have been playing a vital and diverse role in the agricultural field and other essential fields of human endeavour (Woo et al., 2014). There are many species of Trichoderma but the notable ones amongst them include: Trichoderma harzianum, Trichoderma viride and Trichoderma atroviride; some of the strains are also available and still researches are underway to find more of them (Lee et al., 2016). These amazing fungi are available in diverse sources such as soil, compost, plant roots, leaves and can grow saprophytically on bark, wood and some other substrates (Zeilinger et al., 2016).

Agriculture, now the most challenging field in human endeavour in the global perspective that require more robust attention (Nchuchuwe & Adejuwon, 2012). One of the challenges is the crop production and yield which defend on the agricultural input of chemicals like synthetic fertilizers and pesticides to improve the soil fertility for more production outcome and eventually, these processes of applying chemicals were found to have more detrimental effect to the environment and to the final consumers (Massah & Azadegan, 2016). The chemicals such as fertilizers, insecticides and pesticides which eventually lead to the

environmental contamination in soil, air and water respectively.

In this new era of modern research, the use of microorganisms as a replacement of the chemical usage for more crops production is now in focus; with the priority focus on *Trichoderma species* that were known to have every potentiality in providing better, safer and qualitative feedback compared to chemical methods of application (Ajmal *et al.*, 2018). In this paper, we specifically focus on some important benefits associated with *Trichoderma species* in relation to agriculture and more reviews are needed for other benefits that can be derived from this amazing fungal species or other microbial species that can serve similar purpose.

### 2. Biocontrol and Cell wall degrading potential

Evidences in the literature have demonstrated the effectiveness of *Trichoderma species* on the enhancement of crop yield due to the pathogenicity combat of these fungal species which lead to their commercialization and now attracting the global attention toward the benefits. Although scientific research did not provide full details of mechanisms for which it operates, but it

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believes that some of the mechanisms of action of these Trichoderma species against a wide set of microbes like fundi, bacteria and virus to some certain cases indicate how effective are the Trichoderma spp. in biocontrol potentiality (Zeilinger et al., 2016). The author observed that the synthesis of bioactive compounds such as peptaibiotics in the cell is very crucial on the survival of both mycoparasites and saprotrophic species of Trichoderma for defence and colonization. Cuesta et al., (2012), described both species of Trichoderma and Streptomyces species produce antibiotics and clearly showed the antagonistic and chitinolytic activities against phytopathogens. In a related literature, where six Trichoderma species were tested against the soilborne pathogens Rhizoctonia solani, Sclerotium rolfsii, Sclerotinia sclerotiorum, the isolates exhibit the inhibitory effect upon dual culture with Trichoderma species where T. viride strains had more significant effect over *T. harzianum* strain (Amin et al., 2010), likewise when tested against Fusarium species (Küçük, 2017). The biocontrol potential of eleven some species of Trichoderma isolates were assessed against M. Phaseolina a soy bean pathogen of which two strains strains of T. Harzianum (T7 and T14) emerged the best for inhibiting pathogen growth (Khaledi & Taheri, Trichoderma 2016). Similarly, viride Trichoderma harzianum were described as the potent biocontrol agent and commercially available as biocides against plant pathogens (Ghahfarokhi & Goltapeh, 2010; Unarngam et al., 2020; Alvarado-Marchena & Rivera-Méndez, 2016). Also, T. asperillum demonstrates high antagonistic property towards Fusarium oxysporum f. sp. Lycoperci (FOL) (El-Komy et al., 2015). Other species known as Trichoderma gamsii isolated from a healthy plant root of Panax notoginseng, identified as nonpathogenic to its host plant, with ability of producing volatile organic compounds (Chen et al., 2016). Therefore, these features are of utmost importance for the Trichoderma species in successful combat against the fungal and bacterial related plant pathogens through antagonistic interactions (Degenkolb et al., 2015). Similarly, a combination of two beneficial fungi like Trichoderma spp. and Mycorrhizae produced the best growth and decrease the wilt disease in pigeon pea (Dehariya et al., 2015). Sometimes this depends on the plant type, density and probably the species of the microbes that colonized the roots of such plants. For instance, upon application of different concentrations of Trichoderma harzianum was experimentally reported to have increased seed germination, seedling survival and growth against Festuca arundinaceaschreb., Lespedeza cyrtobotryaMiq., Dianthus barbatus var. asiaticus Nakai, and Parthenocissus tricuspidata (Wang et al., 2016). Recently, the use of Trichoderma spp for pest

control and disease management were reported (Poveda, 2021; Mistra & Ansari, 2021).

One of the main reasons of the Trichoderma species serving as biocontrol agent is due its ability to secrete enzymes of carbohydrates, lipids and proteins degradation. It was reported that Trichoderma species produce lytic enzymes that serve as an antifungal against the pathogenic fungi (Rajeswari, 2014). The secretion of antifungal enzymes and chitinolytic action of Trichoderma isolates of T. viride, T. harzianum and T. hamatum were screened and tested; with the *T. viride* identified to be the most potent fungal antagonist against Fusarium oxysporum, Aspergillus niger and Sclerotum rolfsii as well as secretion of the main cell wall degrading enzymes protease, chitinase and  $\beta$ -glucanase (Goltapeh & Danesh, 2006). Therefore, the ability of Trichoderma species as a bio fertilizer producing power is mainly due to the secretion of degradative enzymes. It was believed that the agricultural waste decomposition also improves the soil fertility which is solely due to the action of microbial degradative enzymes. Some of the activities that Trichoderma exhibit are as a result of these secretions. For instance, the use of Trichoderma species as biological control agents were due to their ability to secrete hydrolytic which include chitinases enzymes glucanases which are among the cell wall degrading enzymes of phytopathogenic fungi against nematodes and insects (González et al., 2012). Report shows that, upon isolation of fifty fungal species from fouled soil in oil refinery which was screen for its ability to secrete xylanase enzyme from one of the strain identified as Hypocrea lixii and was tested to have the potentiality in utilizing sludge of sunflower oil as the sole source of carbon and eventually high content of xylanase was traced (Sakthiselvan, Naveera & Partha, 2014). It was demonstrated that the alginate pellets formulated from three isolates of Trichoderma species and the comparison were made for the in vitro chitinase and also β-1.3-glucanase synthesis in which the addition of the dried fungal mycelium increases the fold of chitinase and  $\beta$ -1,3-glucanase (El-Katatny et al., 2003). In a related finding, assessment for the induction of chitinases and glucanases enzymes was demonstrated using different strains of Trichoderma upon cultured in liquid media containing different inducers, the highest level of enzymes activities of both chitinase and glucanase were reported (Prasetyawan & Sulistyowati, 2018). As a result, Trichoderma spp. are used in the current industrial trends for enzymes production. antibiotics, biofuels and other metabolites; therefore, more researches are required to enhance their efficiency and more safety for the application (Błaszczyk et al., 2014). Application of

T. viride on the solid state fermentation process on grape marc and wine lees; after 10 days, the important enzymes for the cell wall degradation (chitinase, pectinase and β-glucanase) of varying concentrations were produced and are also predicted for their involvement in protecting the plants from disease and are eco-friendly (Bai et al., 2008). Also, Trichoderma reesei was described as the major source of hemicelluloses and cellulases used in various industries for the production of biofuels due to the presence of biosynthetic pathways leading to the synthesis of secondary metabolites but mechanisms for their secretion still not clear (Martinez et al., 2008). The secretion of the enzymes seems not to affect the secretion of other beneficial metabolites. Because, when an in vitro study of Trichoderma against Rhizoctonia solani species conducted it revealed that enzymatic activities of Trichoderma species which include chitinase, cellulase, lipase, protease and phosphate solubilisation capability had shown a positive response for inhibition of the pathogen's growth as well as synthesis of growth hormone (Rahmansyah, 2014).

## 3. Bio fertilizer and phosphate solubility

The major limiting factor that affects the crop yield among the farmers in developing countries globally is the issue of soil fertility and thus, maintaining the soil quality could subside the problems concerning land degradation, soil infertility as well as a rapid decline in crop production for better farming practice (Mohammadi & Sohrabi, 2012). In a similar review as reported previously, the author stated that; bio fertilizers are a product of cells of certain different types of beneficial microorganisms that consist of important components of nutrients which play a key role in soil protection, crop productivity and of ecosystem. sustainability the Some microorganisms that are often used as biofertilizers are nitrogen fixers, phosphorus and potassium solubilize (Mohammadi & Sohrabi, 2012). Augmentation of some certain beneficial fungi contribute immensely in achieving a desired goal such as application of Mycorrhizae and Trichoderma species on dry land increased the growth and weight of both fresh and dry bulb of red onion significantly as previously reported (Sakthiselvan, Naveena & Partha, Improving the fertility and more conducive soil, some of the microbes were identified as thermotolerant and capable of solubilizing phosphate compounds such as the bacteria and fungi which were isolated from varying compost plants and bio fertilizers; with the exception of Streptomyces thermophiles J57 that lacked pectinase, all of the

isolates possessed; chitinase, CMcase, pectinase, protease, nitrogenase and lipase activities and all were able to solubilize phosphate containing compounds such as calcium phosphate, Israel rock phosphate among others (Chang & Yang, 2009).

The study was conducted to assess the effect of the application of different doses of chemical fertilizer (potassium fertilizer) K2O along with Pochonia chlymydosporia and Trichoderma harzianum for the management of disease caused by nematodes (*Meloidogyne incognita*); upon addition of double doses of K<sub>2</sub>O fertilizer along with the two fungi (P. chlymydosporia and T. harzianum) that served as bio fertilizer enhanced the plant growth and reduced the population of galls in a roots system (Hisamuddin, 2014). Therefore, the ability of Trichoderma species as a bio fertilizer producing power is mainly due to the secretion of the degradative enzymes. In another finding, the Trichoderma enhances cell wall degradative enzymes as well availability of phosphorus in the soil (Saravanakumar et al., 2016). It was believed that the agricultural waste upon decomposition and decay can also improve the soil fertility which is solely due to the action of microbial degradative enzymes.

#### 4. Macro/Micro Nutrients

Nutrients are required by every living matter be it plant, animal or microbes. All nutrients are either classified as macro or micronutrients depending on the quantity requirement. In the plant Carbon, Nitrogen, Sulphur and Phosphorus are referred to as macro elements for macronutrient synthesis while Zinc, Copper, Molybdenum, Manganese are called microelements for micronutrients supplement. In the plant, all the macronutrients are automatically made in the process called photosynthesis which are referred autotrophs. But in the micronutrients/elements, plants compete around their niche. Such microelements are crucial to plant growth and health. Related work had been published where some Trichoderma species serve as nutrients supplement booster on plants. Ülker et al., (2011), reported that certain elements such as Ca2+ and Mn2+ have the potential of enhancing lipases activities. Experimental prove shows that; application of Trichoderma species on citrus orchard trees with deficiency in some certain nutrients had shown a significant increase in nitrogen (N) and manganese (Mn) supplement (Haris et al., 2016). Another evidence of cyanobacteria-based inoculant interacts with the rice plant (Oryza sativum L.) leads to the supplementation of micronutrients such as Zn, Mn, Cu and Fe (Adak et al., 2016). Li et al., (2015), prove the nutritional supplementation was

noticed while using Trichoderma species against tomato plant in a copper (Cu) deficient soil but nutrients uptake was enhanced on the soil with phosphate due to their phosphate solubilizing potentials. Similarly, composted kitchen waste using Trichoderma harzianum was observed to have improved the growth, yield and the nutritional quality of the tomato (Lycopersicon esculentus Mill.) (Molla et al., 2012). Thus, improvement of the nutrients synthesis in addition to that of the photosynthesis process and its uptake will lead to the early harvest of the product within a limited period. Another impressive habit of this fungus that contributes to the better yield is the exhibition of resistivity to certain abiotic factors.

### 5. Trichoderma and Systemic Resistance

The induction of systemic resistivity to a plant is an interesting and added advantage for a plant to possess such an opportunity. In agriculture, one of the selection criteria that is required for most of the farming practices, especially in a warm temperate region, is the plant with resistance capability to abiotic stresses such as disease and drought. This is a natural phenomenon. But in some plants, the resistivity is intrinsic and therefore, external factors are needed to make it active. Some microorganisms were known to possess such potentialities as such, they act as elicitors and hence are called bio elicitors. Trichoderma species are among the successful bio stimulants that enhance the plant growth, induce disease resistance in plants and longlasting use in horticulture due to the activation of mitogen (López-Bucio, Pelagio-Flores & Herrera-Estrella, 2015). Even though other fungi can perform a similar role like Rhizosphere fungi. It was revealed that: Rhizosphere of fungi native possesses the potentiality for growth promotion as well as induction of resistance in sunflower when tested against Plasmoparahalstedii thereby reducing the severity of the mildew disease (Nagaraju et al., 2012). Some of the microbes were believed to induce such resistivity either directly or indirectly due to the secretion of certain secondary metabolites. Many root-associated mutualists such as Trichoderma, Bacillus, Pseudomonas as well as Mycorrhiza species are able to sensitize the immune system of the plant and renders defence indirectly (Pieterse et al., 2014).Carmona-Hernandez et al., (2019), made the assertion that; the defence response experienced by some plants is as a result of overproduction of certain enzymes and other metabolites. Trichoderma harzianum interaction with maize plant; it was noticed that over 300 different proteins were up-regulated and notably among them include the enzymes involved in carbohydrate metabolisms like

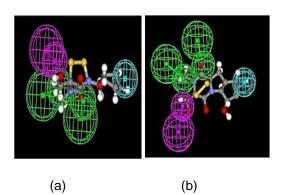
glucanases and chitinases and other proteins that are associated with the disease resistance and stress (Harman & Shoresh, 2007). Another evidence emerged that; Trichoderma species induce resistance over Fusarium oxysporum f. spp. on greenhouse crops which subsequently substantiated the incidence of the disease more especially when applied a week prior to the pathogen's inoculation (Ei-Khallal, Elsharkawy et al., 2015a). A recent study plant-microbes' involving the interactions; revealed that plants can be able to host microbes which help in modifying and reshaping the rhizosphere microbiomes in order to prevent the plants from pathogens and insects attack thereby recruiting microorganisms that suppress pathogen due to the synthesis of secondary metabolites (Berendsen, Pieterse & Bakker, 2012). In the same vein, Phoma species which are a plant growth promoting fungi as well, that induce systemic resistivity on Colletotrichumorbiculare; hence suppress the disease in cucumber (Elsharkawy et al., 2015b). Similarly, Trichoderma spp. was reported to have induced disease resistance on tomato plant caused by Fusarium oxysporum f. sp. (FORL) as it significantly reduces the disease severity (Hibar et al., 2007; Shanmugam, Chugh, & Sharma, 2015). Also in cucumber plant (Koike et al., 2001)

### 6. Growth Hormone and bio elicitors

Growth promoting efficiency is one of the unique features associated with Trichoderma spp. Many findings were published up to the molecular level proving the secretion of growth factor or hormone that is a structural and functional similarity with a popular plant growth hormone known as auxin. The plant hormones are responsible for both roots and shoot elongations. Some are synthetic and others like indole acetic acids (IAA) are naturally produced by the plant. For example, Trichoderma atroviride serves as bio elicitor for stimulation of growth and biosynthesis of secondary metabolites around plant roots (Ming et al., 2013). Therefore, secretion of growth hormone by *Trichoderma spp.*and other related microbes will enhance the actions of both synthesizing hormones and burst the growth due to the synergistic effect of the two expected hormones induced in a particular plant. Moreover, when some isolates of Trichoderma species were used for the assessment of the germination and seeds vigour in rice (Oryza sativa L.) the isolates were discovered to speed up the germination capacity as well as vigour of the rice seeds across the parameters used for such assessment (Doni et al., 2014). For these reasons, the microorganisms like Trichoderma definately possess some mechanisms or

biosynthetic pathways responsible for the secretion of such important biochemical substances. Trichoderma harzianum Rifaii involved in promoting plant growth; in which three possible mechanisms were suggested and these include: medium acidification, synthesis of chelating metabolites as well as redox activity accompanied by solubilisation of the insoluble and sparingly soluble minerals that play a vital role in plant growth (Bezuidenhout, Rensburg & Rensburg, 2012). Trichoderma promote plant growth on maize seedlings more than when treated with 10µL of indole acetic acid (López-Coria, Hernandez-Mendea & Sanchez-Nieto, 2016).

Martínez-Medina et al., (2014), confirmed the elevation of auxin and decrease in abscisic acid and cytokinins content induced by the isolates that promote plant growth and defence-related metabolites. The molecular docking simulations revealed that *T. harzianum* synthesizes gliotoxin which has a structural identity with the growth hormone gibberellic acids through its receptor (GIDI) and this mimics the structure of gibberellic acid (GA<sub>3</sub>) in particular with similar response (Bezuidenhout, Rensburg & Rensburg, 2012). Also, the growth of Arabidopsis seedlings tested in vivo upon inoculated with T. virens and T. atroviride and the result indicated the auxinrelated characteristics including the lateral roots stimulation as well as biomass production (Contreras-Cornejo et al., 2009). In a related finding, Trichoderma viride and Trichoderma harzianum tested against M. javanica on tomato plants, the results show that all the densities used for the two fungal species involved in the suppression of nematodes, root galling and also enhanced the growth of the tomato plants (Al-Hazmi & TariqJaveed, 2016).



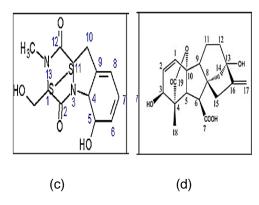


Fig.1: Molecular similarity between (a) Gibberellic Acid (b) Gliotoxin,

Numbered structures for (c) gibberellic acid 3 and (d) gliotoxin (Bezuidenhout, Rensburg & Rensburg, 2012).

### 7. Trichoderma spp. and yield enhancement potentials

Trichoderma inoculants combination with other beneficial microorganisms enhance the growth of plants which resulted in providing the appreciable yield (Baker, 1989; Harman et al., 2004; Szczech et al., 2017). For instance, combined Trichoderma spp. with Mycorrhizae and reported the increase in onion growth and the weight of both dry and wet onion bulbs by 63.95% and 55.10% respectively (Made Sudantha, Astiko & Author, 2017). Another report indicated the growth enhancement of red beet and cabbage with huge yield increment (Topolovec-Pintaric, Zutic & Dermic, 2013), tomato yield increment was also documented (Molla et al., 2012; Konappa et al., 2018) and sorghum (López-Bucio, Pelagio-Flores Herrera-Estrella, 2015) upon application of Trichoderma species. However, the notice in the growth begin with the seed germination (Amooaghaie, Saeri & Azizi, 2015; Doni et al., 2014: Mastouri, Biorkman & Harman, 2010: Srivastava et al., 2010; Toghueo et al., 2016; You et al., 2016), length of the roots (Samolski et al., 2012; Saravanakumar et al., 2016), number of the leaves (López-Coria, Hernandez-Mendea & Sanchez-Nieto, 2016; Made Sudantha, Astiko & Author, 2017; Molla et al., 2012), lateral branches (Srivastava et al., 2010), chlorophyll content (Shoresh, Harman & Mastouri, 2010; López-Bucio, Pelagio-Flores & Herre-Estrella, 2015; John et al., 2010; Kumar, Manigundan & Amaresan, 2017) to the crop yields. The yield of grape fruit was again increased by two Trichoderma strains (Pascale et al., 2017). Inoculating soil with Trichoderma improved the rice yield cultivated under continues flooding condition (Khadka et al., 2019). T. harzianum and

*T. hamatum* were applied on field study, where *T. harzianum* increased the crop yield of chickpea from 12% to 28% while *T. hamatum* from 12% to 24% respectively.

### 8. Conclusion

The role of biological agents such as *Trichoderma* species for improvement of agricultural yield cannot be overemphasized considering the above-outlined contributions made by these amazing fungi compiled from several findings which could feasibly represent a viable as well as environmentally benign to human with efficiency similar to that of the synthetic chemicals used for the successful faming processes and more safer to the entire ecosystem, these could save yield from declining, ameliorate the contaminated soil and improve its fertility with time. Therefore, future finding should be focused on health implication upon prolong consumption of the produce obtain on application of these microbes when used as biofertilizers. If found safe for consumption, this could be our future organic fertilizer to replace the chemically synthetic form.

#### Conflict of interest

The authors declare that there is no conflict of interests regarding the publication of this article.

#### References

- Adak, A., Prasanna, R., Babu, S., Bidyarani, N., Verma, S., Pal, M., ... Nain, L. (2016). Micronutrient enrichment mediated by plant-microbe interactions and rice cultivation practices. *Journal of Plant Nutrition*, 39(9), 1216–1232. https://doi.org/10.1080/01904167.2016.114
- Ajmal, M., Ali, H.I., Saeed, R., Akhtar, A., Tahir, M., Mehboob, M.Z., & Ayub, A. (2018). Biofertilizer as an alternative for chemical fertilizers. *Journal of Agriculture and Applied Sciences*, 7(1), 1-7
- Al-Hazmi, A. S., & TariqJaveed, M. (2016). Effects of different inoculum densities of *Trichoderma harzianum* and *Trichoderma viride* against *Meloidogyne javanica* on tomato. *Saudi Journal of Biological Sciences*, 23(2), 288–292. https://doi.org/10.1016/j.sjbs.2015.04.007
- Alvarado-Marchena, L., & Rivera-Méndez, W. (2016). Molecular identification of *Trichoderma spp.* in garlic and onion fields and In Vitro antagonism trials on Sclerotium cepivorum. *Revista Brasileira de Ciencia Do Solo*, 40, 1–9. https://doi.org/10.1590/18069657rbcs2015 0454
- Amin, F., Razdan, V. K., Mohiddin, F. A., Bhat, K. A., & Banday, S. (2010). Potential of

- *Trichoderma Species* as Biocontrol, 2(10), 38–41.
- Amooaghaie, R., Saeri, M. R., & Azizi, M. (2015).
  Synthesis, characterization and biocompatibility of silver nanoparticles synthesized from *Nigella sativa* leaf extract in comparison with chemical silver nanoparticles. *Ecotoxicology and Environmental Safety*, 120. https://doi.org/10.1016/j.ecoenv.2015.06.0
- Bai, Z., Jin, B., Li, Y., Chen, J., & Li, Z. (2008). Utilization of winery wastes for *Trichoderma viride* biocontrol agent production by solid-state fermentation. *Journal of Environmental Sciences*, *20*(3), 353–358. https://doi.org/10.1016/S1001-0742(08)60055-8
- Baker, R. (1989). Improved *Trichoderma spp.* for promoting crop productivity. *Trends in Biotechnology*, 7(2), 34–38. https://doi.org/10.1016/0167-7799(89)90055-3
- Berendsen, R. L., Pieterse, C. M. J., & Bakker, P. A. H. M. (2012). The rhizosphere microbiome and plant health. *Trends in Plant Science*, 17(8), 478–486. https://doi.org/10.1016/j.tplants.2012.04.00
- Bezuidenhout, J., Rensburg, L. Van, & Rensburg, P. J. Van. (2012). Molecular Similarity between Gibberellic Acid and Gliotoxin: Unravelling the Mechanism of Action for Plant Growth Promotion by, 2, 703–712.
- Błaszczyk, L., Siwulski, M., Sobieralski, K., Lisiecka, J., & Jędryczka, M. (2014). *Trichoderma spp.* Application and prospects for use in organic farming and industry. *Journal of Plant Protection Research*, 54(4), 309–317. https://doi.org/10.2478/jppr-2014-0047
- Carmona-Hernandez, S., Reyes-Pérez, J., Chiquito-Contreras, R., Rincon-Enriquez, G., Cerdan-Cabrera, C., & Hernandez-Montiel, L. (2019). Biocontrol of Postharvest Fruit Fungal Diseases by Bacterial Antagonists: A Review. *Agronomy*, *9*(3), 121.
  - https://doi.org/10.3390/agronomy9030121
- Chagas, L. F. B., Chagas, A. F., Fidelis, R. R., De Carvalho Filho, M. R., & De Oliveira Miller, L. (2017). *Trichoderma asperellum* efficiency in soybean yield components. *Comunicata Scientiae*, 8(1), 165–169. https://doi.org/10.14295/CS.v8i1.1754
- Chang, C. H., & Yang, S. S. (2009). Thermotolerant phosphate-solubilizing microbes for multi-functional biofertilizer preparation. *Bioresource Technology*, 100(4), 1648–1658.

https://doi.org/10.1016/j.biortech.2008.09.0

- Chen, J. L., Sun, S. Z., Miao, C. P., Wu, K., Chen, Y. W., Xu, L. H., ... Zhao, L. X. (2016). Endophytic *Trichoderma gamsii* YIM PH30019: A promising biocontrol agent with hyperosmolar, mycoparasitism, and antagonistic activities of induced volatile organic compounds on root-rot pathogenic fungi of Panax notoginseng. *Journal of Ginseng Research*, *40*(4), 315–324. https://doi.org/10.1016/j.jgr.2015.09.006
- Contreras-Cornejo, H. A., Macías-Rodríguez, L., Cortés-Penagos, C., & López-Bucio, J. (2009). Trichoderma virens, a plant beneficial fungus, enhances biomass production and promotes lateral root growth through an auxin-dependent mechanism in Arabidopsis. *Plant physiology*, *149*(March), 1579–1592 https://doi.org/10.1104/pp.108.130369
- Cuesta, G., García-de-la-Fuente, R., Abad, M., & Fornes, F. (2012). Isolation and identification of actinomycetes from a compost-amended soil with potential as biocontrol agents. *Journal of Environmental Management*, *95*(SUPPL.), S280–S284. https://doi.org/10.1016/j.jenvman.2010.11. 023
- Degenkolb, T., Fog Nielsen, K., Dieckmann, R., Branco-Rocha, F., Chaverri, P., Samuels, G. J., Brückner, H. (2015). Peptaibol, Secondary-Metabolite, and Hydrophobin Pattern of Commercial Biocontrol Agents Formulated with Species of the *Trichoderma harzianum* Complex. *Chemistry & Biodiversity*, 12(4), 662–684. https://doi.org/10.1002/cbdv.201400300
- Dehariya, K., Shukla, A., Sheikh, I. A., & Vyas, D. (2015). *Trichoderma* and *Arbuscular mycorrhizal* fungi based biocontrol of *Fusarium udum* butler and their growth promotion effects on pigeon pea. *Journal of Agricultural Science and Technology*, 17(2), 505–517.
- Doni, F., Anizan, I., Che Radziah, C. M. Z., Salman, A. H., Rodzihan, M. H., & Yusoff, W. M. W. (2014). Enhancement of rice seed germination and vigour by *Trichoderma* spp. Research Journal of Applied Sciences, Engineering and Technology, 7(21), 4547– 4552. https://doi.org/10.19026/rjaset.7.832
- Ei-Khallal, S. (2007). Induction and modulation of resistance in tomato plants against Fusarium wilt disease by bioagent fungi (arbuscular mycorrhiza) and/or hormonal elicitors (Jasmonic acid & Salicylic acid): 2-Changes in the antioxidant enzymes, phenolic compounds and. Australian Journal of Basic and Applied Sciences, 1(4), 717–732.

El-Katatny, M. H., Hetta, A. M., Shaban, G. M., & El-Komy, H. M. (2003). Improvement of Cell Wall Degrading Enzymes Production by Alginate Encapsulated *Trichoderma spp. Food Technology and Biotechnology*, *41*(3), 219–225.

- El-Komy, M. H., Saleh, A. A., Eranthodi, A., & Molan, Y. Y. (2015). Characterization of novel *Trichoderma asperellum* isolates to select effective biocontrol agents against tomato fusarium wilt. *Plant Pathology Journal*, 31(1), 50–60. https://doi.org/10.5423/PPJ.OA.09.2014.00 87
- Elsharkawy, M. M., Hase, T., Yagi, Y., Shimizu, M., & Hyakumachi, M. (2015a). Induction of systemic resistance against fusarium crown and root rot disease by blast processing. *Journal of Plant Interactions*, 10(1), 262–269. https://doi.org/10.1080/17429145.2015.106
  - https://doi.org/10.1080/17429145.2015.106 6038
- Elsharkawy, M. M., Shivanna, M. B., Meera, M. S., & Hyakumachi, M. (2015b). Mechanism of induced systemic resistance against anthracnose disease in cucumber by plant growth-promoting fungi. Acta Agriculturae Scandinavica Section B: Soil and Plant Science, 65(4), 287–299. https://doi.org/10.1080/09064710.2014.100 3248
- Ghahfarokhi, R. M., & Goltapeh, M. E. (2010). Potential of the root endophytic fungus Piriformospora indica; Sebacina vermifera and Trichoderma species in biocontrol of take-all disease of wheat Gaeumannomyces graminis var. tritici in vitro. Journal of Agricultural Technology, 6(1), 11–18.
- Goltapeh, E. M., & Danesh, Y. R. (2006). Pathogenic interactions between *Trichoderma species* and *Agaricus bisporus*. *Mycopathologia*, 29–37.
- González, I., D., I., Martínez, B., Arias, Y., González, N., Miranda, L., & Peteira, B. (2012). Induction of chitinases and glucanases in *Trichoderma spp.* strains intended for biological control. *Biotecnología Aplicada*, 29(March), 12–16.
- Haris, M., Ali, R., Arshad Khan, M., Shahzad, H., Rashid, A., Waheed, M., & Bibi, H. (2016). Micronutrients Availability in Citrus with *Trichoderma* Application. *International Journal of Science and Engineering Investigations*, *5*(58), 15–19.
- Harman, G. E., Howell, C. R., Viterbo, A., Chet,
  I., & Lorito, M. (2004). *Trichoderma species*Opportunistic, avirulent plant symbionts. *Nature Reviews Microbiology*, 2(1), 43–56.
  https://doi.org/10.1038/nrmicro797

- Harman, G. E., & Shoresh, M. (2007). The mechanisms and applications of symbiotic opportunistic plant symbionts. NATO Security through Science Series A: Chemistry and Biology, 131–155. https://doi.org/10.1007/978-1-4020-5799-1-7
- Hibar, K., Phytopathologie, L. De, De, I. S. A., Daami-remadi, M., Mahjoub, M. El, Phytopathologie, D., & Chott-mariem, I. S. A. De. (2007). Induction of Resistance in Tomato Plants against Fusarium oxysporum f. sp. radicis-lycopersici by Trichoderma spp. Tunisian Journal of Plant Protection, 2(1), 47–58.
- Hisamuddin, R. S. (2014). Management of Root-Knot Disease in Phaseolus vulgaris Using Potassium Fertilizer and Biocontrol Agents. *Journal of Plant Pathology & Microbiology*, 5(5). https://doi.org/10.4172/2157-7471.1000242
- John, R. P., Tyagi, R. D., Prévost, D., Brar, S. K., Pouleur, S., & Surampalli, R. Y. (2010). Mycoparasitic *Trichoderma viride* as a biocontrol agent against *Fusarium oxysporum f. sp. adzuki* and Pythium arrhenomanes and as a growth promoter of soybean. *Crop Protection*, *29*(12), 1452–1459.
  - https://doi.org/10.1016/j.cropro.2010.08.00
- Khadka, R. B., & Uphoff, N. (2019). Effects of Trichoderma sedling treatment with system of Rice intensification management and with conventional management of transplanted rice. *Peer J*, 7, e5877.
- Khan, M. R., Ashraf, S., Rasool, F., salati, K. M., Mohiddin, F. A., & Haque, Z. (2014). Field performance of Trichoderma species against wilt disease complexof Chickpea caused by Fusarium oxysporum f. sp. ciceri and Rhizoctonia solani. Turkish Journal of Agriculture and Forestry, 38(4), 447-454.
- Khaledi, N., & Taheri, P. (2016). Biocontrol mechanisms of Trichoderma harzianum against soybean charcoal rot caused by Macrophomina phaseolina. *Journal of Plant Protection Research*, *56*(1), 21–31. https://doi.org/10.1515/jppr-2016-0004
- Koike, N., Hyakumachi, M., Kageyama, K., Tsuyumu, S., & Doke, N. (2001). Induction of systemic resistance in cucumber against several diseases by plant growth-promoting fungi: Lignification and superoxide generation. *European Journal of Plant Pathology*, 107(5), 523–533. https://doi.org/10.1023/A:1011203826805
- Küçük, Ç. (2017). In vitro Antagonistic Activity against *Fusarium Species* of Local *Trichoderma spp.* Isolates. *J. Biol. Environ. Sci*, 11(32), 67–74. Retrieved from

- http://jbes.uludag.edu.tr/PDFDOSYALAR/3 2/mak02.pdf
- Kumar, K., Manigundan, K., & Amaresan, N. (2017). Influence of salt tolerant *Trichoderma spp.* on the growth of maize (*Zea mays*) under different salinity conditions. *Journal of Basic Microbiology*, 57(2), 141–150. https://doi.org/10.1002/jobm.201600369
- Kumar, S., Thakur, M., & Rani, A. (2014). trichoderma: Mass production, formulation, quality control, delivery and its scope in commercialization in india for the management of plant diseases. *African Journal of Agricultural Research*, 9(53), 3838 3852.
- Konappa, N., Krishnamurthy, S., Siddaiah, C.N.,Ramachandrappa, N.S., & Chodappa, S. (2018). Evaluation of biological efficacy of Trichoderma asperellum against tomato bacterial wiltcaused by Ralstonia solanaccearum. Egyptian Journal of Biological Pest Control, 28(1), 1-11.
- Lee, S., Yap, M., Behringer, G., Hung, R.,& Bennett, J.W. (2016). Volatile organic compounds emitted by Trichoderma species mediated plant growth. *Fungal Biology and Biotechnology*, 3(1), 1-14
- Li, R. X., Cai, F., Pang, G., Shen, Q. R., Li, R., & Chen, W. (2015). Solubilisation of phosphate and micronutrients by *Trichoderma harzianum* and its relationship with the promotion of tomato plant growth. *PLoS ONE*, 10(6), 1–16. https://doi.org/10.1371/journal.pone.01300 81
- López-Bucio, J., Pelagio-Flores, R., & Herrera-Estrella, A. (2015). Trichoderma as biostimulant: Exploiting the multilevel properties of a plant beneficial fungus. *Scientia Horticulturae*, 196, 109–123. https://doi.org/10.1016/j.scienta.2015.08.0 43
- López-Coria, M., Hernández-Mendoza, J. L & Sánchez-Nieto, S. (2016). *Trichoderma asperellum* Induces Maize Seedling Growth by Activating the Plasma Membrane H(+)-ATPase. *Molecular Plant-Microbe Interactions: MPMI*, 29(10), MPMI07160138R. https://doi.org/10.1094/MPMI-07-16-0138-P
- Made Sudantha, I., Astiko, W., & Author, C. (2017). The Effort of Increasing Growth And Harvest of Local Variety Red Onion With Applications of Some Dose of Indigenous *Mycorrhizal* and Bioactivator *Trichoderma spp.* in Dry Land, 10(9), 42–49. https://doi.org/10.9790/2380-1009014249
- Massah, J., & Azadegan, B. (2016). Effect of chemical fertilizers on soil compaction and

degration. Agricultural Mechanization in Asia, Africa, and latin America, 47(1), 44-50

- Martínez-Medina, A., Del Mar Alguacil, M., Pascual, J. A., & Van Wees, S. C. M. (2014). Phytohormone Profiles Induced by *Trichoderma* Isolates Correspond with Their Biocontrol and Plant Growth-Promoting Activity on Melon Plants. *Journal of Chemical Ecology*, 40(7), 804–815. https://doi.org/10.1007/s10886-014-0478-1
- Martinez, D., Berka, R. M., Henrissat, B., Saloheimo, M., Arvas, M., Baker, S. E., ... Brettin, T. S. (2008). Genome sequencing and analysis of the biomass-degrading fungus Trichoderma reesei (syn. Hypocrea jecorina). *Nature Biotechnology*, 26(5), 553–560. https://doi.org/10.1038/nbt1403
- Mastouri, F., Björkman, T., & Harman, G. E. (2010). Seed Treatment with *Trichoderma harzianum* Alleviates Biotic, Abiotic, and Physiological Stresses in Germinating Seeds and Seedlings. *Phytopathology*, 100(11), 1213–1221. https://doi.org/10.1094/PHYTO-03-10-0091
- Ming, Q., Su, C., Zheng, C., Jia, M., Zhang, Q., Zhang, H., ... Qin, L. (2013). Elicitors from the endophytic fungus *Trichoderma atroviride* promote Salvia miltiorrhiza hairy root growth and tanshinone biosynthesis. *Journal of Experimental Botany*, *64*(18), 5687–5694.
  - https://doi.org/10.1093/jxb/ert342
- Misra, V., & Ansari, M. I. (2021). Role of Trichoderma in Agriculture and Disease Management. *Plant- Promoting Microbes* for Sustainable Biotic and Abiotic Stress Management, 425 - 440.
- Mohammadi, K., & Sohrabi, Y. (2012). Bacterial Biofertilizers for Sustainable Crop Production: a Review. *Journal of Agricultural and Biological Science*, 7(5), 307–316.
- Molla, A. H., Manjurul Haque, M., Amdadul Haque, M., & Ilias, G. N. M. (2012). Trichoderma-Enriched Biofertilizer Enhances Production and Nutritional Quality of Tomato (Lycopersicon esculentum Mill.) and Minimizes NPK Fertilizer Use. Agricultural Research, 1(3), 265–272. https://doi.org/10.1007/s40003-012-0025-7
- Nagaraju, A., Murali, M., Sudisha, J., Amruthesh, K. N., & Murthy, S. M. (2012). Beneficial microbes promote plant growth and induce systemic resistance in sunflower against downy mildew disease caused by Plasmopara halstedii, 3(5), 12–18.
- Nchuchuwe, F. F., & Adejuwon, K.D.(2012). The challenges of Agricultural development in Africa: the case of Nigeria. *International Journal of academic Reseach in*

Progressive Education and Development, 1(3), 45-61.

- Pieterse, C. M. J., Zamioudis, C., Berendsen, R. L., Weller, D. M., Van Wees, S. C. M., & Bakker, P. A. H. M. (2014). Induced Systemic Resistance by Beneficial Microbes. *Annual Review of Phytopathology*, 52(1), 347–375. https://doi.org/10.1146/annurev-phyto-082712-102340
- Poveda, J. (2021). Trichoderma biocontrol agent against pest: new uses for a mycoparasite". Biological Control, 104634
- Rahmansyah, M. (2014). Antagonism Competence of *Trichoderma Spp.* Isolates against *Rhizoctonia solani* KUHN, *3*(4), 171–179.
- Rajeswari, P. (2014). Inhibition of Pectinolytic and Cellulolytic Enzymes of Fusarium oxysporum by Trichoderma spp and Pseudomonas fluorescens on Arachishypogaea . L, 3(3), 817–822.
- Sakthiselvan, P., Naveena, B., & Partha, N. (2014). Molecular characterization of a xylanase-producing fungus isolated from fouled soil. *Brazilian Journal of Microbiology*, 45(4), 1293–1302. https://doi.org/10.1590/S1517-83822014000400020
- Samolski, I., Rincón, A. M., Pinzón, L. M., Viterbo, A., & Monte, E. (2012). The qid74 gene from *Trichoderma harzianum* has a role in root architecture and plant biofertilization. *Microbiology*, 158(1), 129– 138. https://doi.org/10.1099/mic.0.053140-0
- Saravanakumar, K., Yu, C., Dou, K., Wang, M., Li, Y., & Chen, J. (2016). Synergistic effect of Trichoderma-derived antifungal metabolites and cell wall degrading enzymes on enhanced biocontrol of Fusarium oxysporum f. sp. cucumerinum. Biological Control, 94, 37–46. https://doi.org/10.1016/j.biocontrol.2015.12.001
- Shanmugam, V., Chugh, P., & Sharma, P. (2015). Cold-tolerant *Trichoderma species* for the management of Fusarium wilt of tomato plants. *Annals of Microbiology*, 65(1), 543–551. https://doi.org/10.1007/s13213-014-0890-3
- Shoresh, M., Harman, G. E., & Mastouri, F. (2010). Induced Systemic Resistance and Plant Responses to Fungal Biocontrol Agents. *Annual Review of Phytopathology*, 48(1), 21–43. https://doi.org/10.1146/annurev-phyto-073009-114450
- Srivastava, R., Khalid, A., Singh, U. S., & Sharma, A. K. (2010). Evaluation of

- Arbuscular mycorrhizal fungus, Pseudomonas fluorescent and Trichoderma harzianum formulation against Fusarium oxysporum f. sp. lycopersici for the management of tomato wilt. Biological Control, 53(1), 24–31. https://doi.org/10.1016/j.biocontrol.2009.11.012
- Szczech, M., Nawrocka, J., Felczyński, K., Małolepsza, U., Sobolewski, J., Kowalska, B., ... Kancelista, A. (2017). *Trichoderma atroviride* TRS25 isolate reduces downy mildew and induces systemic defence responses in cucumber in field conditions. *Scientia Horticulturae*, 224(June), 17–26. https://doi.org/10.1016/j.scienta.2017.05.0 35
- Toghueo, R. M. K., Eke, P., Zabalgogeazcoa, Í., de Aldana, B. R. V., Nana, L. W., & Boyom, F. F. (2016). Biocontrol and growth enhancement potential of two endophytic *Trichoderma spp.* from *Terminalia catappa* against the causative agent of Common Bean Root Rot (Fusarium solani). *Biological Control*, 96, 8–20. https://doi.org/10.1016/j.biocontrol.2016.01.008
- Topolovec-Pintaric, S., Zutic, I., & Dermic, E. (2013). Enhanced growth of cabbage and red beet by *Trichoderma viride*. *Acta Agriculturae Slovenica*, 101(1), 87–92. https://doi.org/10.2478/acas-2013-0010
- Pascale, A., Vinale, F., Manganello, G., Nigro, M., Lanzuice, S., Ruocco, M., ...& Lorito, M. (2017). Trichoderma and its secondary metabolites improve yield and quality of grapes. *Crop Protection*, 92, 176-181.
- Prasetyawan, S., & Sulistyowati, L. (2018, January). Glucanase and Chitinase from Some Isolates of Endophytic Fungus Trichoderma spp. In IOP Conference Series: Materials Science and Engineering (Vol. 299, No. 1, p. 012026). IOP Publishing. https://doi.org/10.1088/1757-899X/299/1/012026
- Ülker, S., Özel, A., Çolak, A., & Karaoğlu, Ş. A. (2011). Isolation, production, and characterization of an extracellular lipase from *Trichoderma harzianum* isolated from soil. *Turk J Biol*, *35*, 543–550. https://doi.org/10.3906/biy-1004-107
- Unartngam, J., Srithongkum, B., Intanoo, W., Charoenrak, P., & Chamsuarng, C. (2020).

  "Morphological and Molecular based identification of Trichoderma CB-Pin-01 biological control agent of plant pathogenic fungi in Thailand". IJAT 16 (16), 175 188
- Wang, F., Wang, Z., Kou, C., Ma, Z., & Zhao, D. (2016). Responses of wheat yield, macroand micro-nutrients, and heavy metals in soil and wheat following the application of

- manure compost on the North China Plain. *PLoS ONE*, 11(1), 1–18. https://doi.org/10.1371/journal.pone.01464
- Woo, S.L., Ruocco, M., Vinale, F., Nigro, M., Marra, R., Lombardi, N., ... & Lorito, M. (2014). Trichoderma- based products and their widespread used in agriculture. the open Mycology Journal, 8(1).
- You, J., Zhang, J., Wu, M., Yang, L., Chen, W., & Li, G. (2016). Multiple criteria-based screening of *Trichoderma* isolates for biological control of Botrytis cinerea on tomato. *Biological Control*, 101, 31–38. https://doi.org/10.1016/j.biocontrol.2016.06 .006
- Zeilinger, S., Gruber, S., Bansal, R., & Mukherjee, P. K. (2016). Secondary metabolism in *Trichoderma* Chemistry meets genomics. *Fungal Biology Reviews*, 30(2), 74–90. https://doi.org/10.1016/j.fbr.2016.05.001