

ORIGINAL ARTICLE

# Long noncoding RNA metastasis-associated lung adenocarcinoma transcript 1 (MALAT1): A molecular predictor of poor survival in glioblastoma multiforme in Egyptian patients



Manal S. Fawzy<sup>a,\*</sup>, Eman A. Toraih<sup>b</sup>, Hoda Y. Abdallah<sup>b</sup>

<sup>a</sup> Department of Medical Biochemistry, Faculty of Medicine, Suez Canal University, P.O. 41522, Ismailia, Egypt

<sup>b</sup> Departments of Histology and Cell Biology (Genetics Unit), Faculty of Medicine, Suez Canal University, P.O. 41522, Ismailia, Egypt

Received 21 July 2016; accepted 12 August 2016

Available online 22 September 2016

## KEYWORDS

Glioblastoma multiforme;  
LncRNA;  
MALAT1;  
Real-time qPCR;  
Survival

**Abstract** *Background:* Long noncoding RNAs (lncRNAs) are a recently discovered class of transcribed RNA molecules with a length of more than 200 nucleotides. Recent studies have shown that lncRNA metastasis-associated lung adenocarcinoma transcript 1 (MALAT1) could play an important role in carcinogenesis and cancer progression in several types of malignancies.

*Objective:* As little is known about the role and clinical significance of lncRNA MALAT1 in glioblastoma multiforme (GBM) patients in Egyptian population, this study aimed to investigate the expressions of lncRNA-MALAT1 in human GBM samples and to correlate these expressions with the available clinicopathological features including patient survival data.

*Subjects and methods:* The relative expression of *MALAT1* was determined in 37 human glioblastoma formalin-fixed paraffin embedded (FFPE) tissue samples and 10 FFPE non-neoplastic brain tissues using quantitative reverse transcription polymerase chain reaction (qRT-PCR) technology.

*Results:* The current results revealed that lncRNA MALAT1 expression was down-regulated in all tumor specimens compared to normal tissues. A receiver operating characteristic (ROC) curve analysis showed high diagnostic performance; area under curve (AUC) =  $0.925 \pm 0.038$  ( $P < 0.001$ ), 95% CI = 0.850–1.00, with 94.6% sensitivity, and 72.7% specificity. Lower *MALAT1* expression was associated with poor prognosis; higher frequency of recurrence ( $P < 0.044$ ), lower overall survival ( $P < 0.005$ ), and shorter disease-free survival ( $P < 0.004$ ).

*Abbreviations:* DFS, disease-free survival; FFPE, formalin-fixed paraffin embedded; GBM, glioblastoma multiform; lncRNAs, long non-coding RNA; MALAT1, metastasis-associated lung adenocarcinoma transcript 1; NEAT2, nuclear-enriched abundant transcript 2; OS, overall survival; qRT-PCR, quantitative reverse transcription polymerase chain reaction; ROC, receiver operating characteristic; TBP, tata binding protein

\* Corresponding author.

E-mail address: [manal2\\_khashana@ymail.com](mailto:manal2_khashana@ymail.com) (M.S. Fawzy).

Peer review under responsibility of Ain Shams University.

<http://dx.doi.org/10.1016/j.ejmhg.2016.08.003>

1110-8630 © 2016 Ain Shams University. Production and hosting by Elsevier B.V.

This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

**Conclusion:** Taken together, we could postulate that MALAT1 might have a tumor-suppressive function in GBM in Egyptian population and this specific type of lncRNAs may be included in the lists of both potential prognostic biomarkers and the future therapeutic targets for glioblastomas. © 2016 Ain Shams University. Production and hosting by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## 1. Introduction

Glioma is by far the most common primary brain tumor associated with poor outcome and survival [1]. It has been classified into well-differentiated low grade astrocytomas (grade I–II), anaplastic astrocytomas (grade III), and glioblastoma multiforme (GBM; grade IV) according to World Health Organization (WHO) [2]. Despite standard treatment that typically includes neurosurgery, chemotherapy and, radiation, recent clinical trials have reported a median survival of only 14–16 months with a 26–33% 2-year survival rate [3]. It has been suggested that the WHO criteria to predict the patient clinical outcomes may not be sufficient alone to estimate patient prognosis [4]. More recently, improved understanding of glioma molecular genetics has led to identifying new potential biomarkers for early diagnosis, prognosis prediction, and novel therapeutic targets [5].

Long noncoding RNAs (lncRNAs) are a recently discovered class of transcribed RNA molecules with a length of more than 200 nucleotides. Similar to protein-coding genes, they have promoter structure, and are transcribed by RNA polymerase II, polyadenylated, and subjected to splicing. However, unlike mRNA, they do not encode proteins [6]. lncRNAs are thought to be important regulators of gene expression at transcription, translation and epigenetic levels [7,8]. An accumulating number of evidences suggested that lncRNAs may have critical roles in a wide range of biological processes [9]. Screening lncRNAs expression profile in glioma has revealed a significant contribution to pathogenesis [10], development and progression [11–13] by regulating cell growth and metastasis; indicating that lncRNAs play significant roles in glioma tumorigenesis [14].

The functional lncRNA-MALAT1 (metastasis-associated lung adenocarcinoma transcript 1); encoded by *MALAT1* gene, which is located at chromosome 11q13.1. [15], and also known as nuclear-enriched abundant transcript 2 (NEAT2), was one of the first lncRNAs found to have a pathogenic role [16] and it has been linked to various cancers besides lung adenocarcinoma [17]. Its expression profile has been found to be dysregulated and correlated with clinical parameters and prognosis in several types of human cancer, such as hepatocellular carcinoma [18], osteosarcoma [19], lung cancer [20], bladder cancer [21], and glioblastoma multiforme [22].

As there are no previous studies, up to the researchers' knowledge, on the expression of this type of lncRNAs in GBM patients among the Arab population, this study for the first time will aim to determine the expression levels of *MALAT1* in a sample of GBM patients and correlate these expressions with the available clinicopathological features including patient survival data in a sample of Egyptian population.

## 2. Subjects and methods

### 2.1. Patients and tissue samples

The present study included 37 formalin-fixed paraffin embedded (FFPE) glioblastoma samples, fulfilling the WHO criteria of GBM and 10 FFPE non-neoplastic brain tissue specimens. GBM patient samples (9 females and 28 males, aged 35 to 60 years old) have been assessed retrospectively from the archive of the Pathology Departments, Mansoura University Hospitals and Suez Canal University Hospitals, Egypt, from 2010 to 2013. Detailed patients' data were retrieved from their medical follow up records. All patients had GBM (i.e. grade IV), undergone surgical removal and post-operative irradiation, and followed for more than 3 years. The work has been carried out in accordance with the code of ethics of the world medical association (Declaration of Helsinki) for experiments on humans. All patients gave written informed consent, except for deceased individuals or patients who provided archived tissue samples and can't be traced.

### 2.2. Total RNA extraction

Following deparaffinization in xylene and washing with alcohol, the total RNA was extracted from tumor and control FFPE tissue sections (4–5  $\mu$ m) collected in sterile eppendorf tubes. Qiagen RNeasy FFPE Kit (Cat. No. 73504, Qiagen, Hilden, Germany) has been used following the protocol supplied by the manufacturer. Extracted total RNA concentration and purity at the absorbance ratio 260/280 nm were determined by NanoDrop ND-1000 spectrophotometer (NanoDrop Tech., Inc. Wilmington, DE, USA). In addition, RNA degradation and contamination were assessed by 1.5% agarose gel electrophoresis.

### 2.3. Reverse transcription

High Capacity cDNA Reverse Transcription Kit (Applied Biosystems, P/N 4368814) was used for reverse transcription (RT) reaction. For each 20  $\mu$ l RT reaction, 10  $\mu$ l (10 ng) RNA sample was combined with 10  $\mu$ l of 2 $\times$  RT reaction mix containing 2  $\mu$ l of 10 $\times$  RT Buffer, 0.8  $\mu$ l of 25 $\times$  dNTP Mix (100 mM), 2  $\mu$ l of 10 $\times$  RT random primers, 1  $\mu$ l of MultiScribe™ Reverse Transcriptase, 1  $\mu$ l of RNase inhibitor, and 3.2  $\mu$ l of nuclease-free water. RT was carried out in a T-Professional Basic, Biometra PCR System (Biometra, Goettingen, Germany) at 25 °C for 10 min, followed by 37 °C for 120 min, and finally 85 °C for 5 min, then held at 4 °C. Appropriate negative controls were included in each experiment.

#### 2.4. Gene expression analysis

Real-time polymerase chain reaction (PCR) was performed in accordance with the Minimum Information for Publication of Quantitative Real-Time PCR Experiments (MIQE) guidelines. Expression of *MALAT1* and the endogenous control *TBP* (Tata binding protein) were quantified using TaqMan® assays (Applied Biosystems, assay ID Hs00273907\_s1, and Hs00427620\_m1, respectively) and Taqman® Universal PCR master mix II, No UNG (2×) (Applied Biosystems, P/N 4440043). Apart for *TBP*, none of the other conventionally used housekeeping genes in GBM studies, were found to be suitable as they showed variation in RNA expression [23]. Hence, *TBP* expression was used for normalization of qPCR data in the current work. The PCR reactions were carried out in a final volume of 20 µl, including 1.33 µl RT product, 10 µl 2 × TaqMan® Universal PCR Master Mix, 1 µl TaqMan® assays. All reactions included a non-template control (with water instead of cDNA), and a no-reverse transcriptase control. The PCR was performed on StepOne™ Real-Time PCR System (Applied Biosystems) as follows: 95 °C for 10 min followed by 40 cycles of 92 °C for 15 s and 60 °C for 1 min.

#### 2.5. Statistical analysis

Statistical Package for the Social Sciences (SPSS) for Windows software (version 20.0) was used for statistical graphics and analyses. The following parameters were considered besides the expression values of the *MALAT1* analyzed: age at diagnosis, gender and tumor site. Categorical variables were compared using the chi-square ( $\chi^2$ ) or Fisher's exact tests where appropriate, while Student's *t* test or One way ANOVA were used to compare continuous variables between two groups or more than two groups, respectively in case the data distribution was concordant with normal distribution (Shapiro–Wilk test) and after checking variance homogeneity (Levene's test). If the data did not meet the criteria mentioned above, the non-

parametric Mann–Whitney–*U* test or Kruskal–Wallis tests were used for comparison between groups with subsequent Bonferroni's post hoc test for multiple testing. The receiver operating characteristic (ROC) curves were derived and area-under-the curve (AUC) analysis performed to get the best cut-off value of *MALAT1* for discriminating GBM patients from controls. *P*-values < 0.05 were considered statistically significant. Disease-free survival is defined as the time between diagnosis of disease and recurrence or distant metastasis. Overall survival is defined as time from diagnosis of disease to death of patients with brain cancer [24]. Median follow-up time was computed by the Kaplan–Meier method. For analysis of disease-free and overall survival, frontal and fronto-temporal tumors were combined for comparison with those with parietal and temporo-parietal. Log rank (Mantel–Cox) was used for survival curves comparison. The fold change of *MALAT1* expression was calculated using Livak's method based on the threshold cycle ( $C_T$ ) value with the following equation: relative quantity =  $2^{-\Delta\Delta CT}$  [25].

### 3. Results

#### 3.1. Baseline characteristics of the study population

The main patient characteristics have been summarized in Table 1. In addition, the mean ± SD of the GBM patients overall survival (OS) and disease-free survival (DFS) were 15.4 ± 5.17 months and 14.7 ± 5.34 months with a range of (8–27) and (6–27) months, respectively. Twenty three patients (62.2%) showed short (≤1 year) disease free survival (DFS) and only 12 (32.4%) showed a higher overall survival rate (> 1 year). As shown in Table 1, male gender was significantly associated with shorter DFS than females (*P* < 0.04).

#### 3.2. Expression of *MALAT1* in glioblastoma multiform

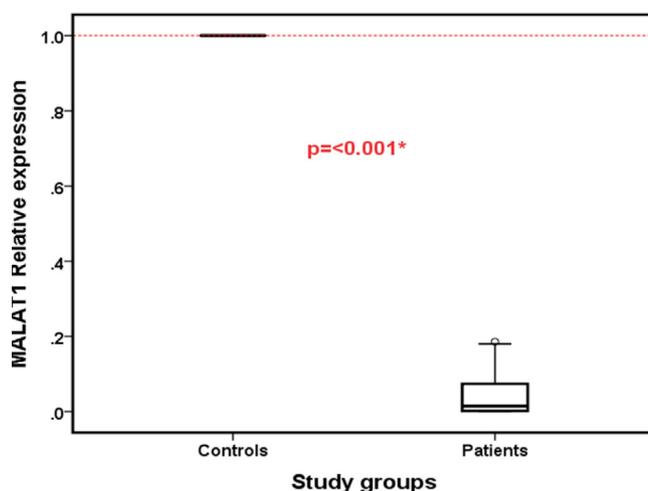
Compared with normal brain tissues, all GBM patients showed low expression levels of lncRNA MALAT1

**Table 1** Baseline patient characteristics (*n* = 37) and survival.

	Mean ± SD or Number (%)	OS (mo)	<i>P</i> value	DFS (mo)	<i>P</i> value
Age (years)	51.5 ± 5.9				
<i>Age, categories</i>					
35-	16 (43.2)	17.3 ± 7.08	0.241	17.0 ± 6.89	0.123
50-	21 (56.8)	13.9 ± 2.25		13.09 ± 2.98	
<i>Gender</i>					
Female	9 (24.3)	18.5 ± 6.9	0.086	18.5 ± 6.9	<b>0.040*</b>
Male	28 (75.7)	14.3 ± 4.13		13.5 ± 4.20	
<i>Tumor site</i>					
Frontal	18 (48.6)	14.4 ± 5.09	0.179	13.5 ± 5.14	0.133
Fronto-temporal	4 (10.8)	19.0 ± 3.46		19.0 ± 3.46	
Parietal	1 (2.7)	11.0		11.0	
Temporo-parietal	14 (37.8)	19.5 ± 5.52		15.4 ± 5.72	
<i>Recurrence</i>					
Non-recurrent	30 (81.1)	15.5 ± 5.25	0.719	15.3 ± 5.26	0.259
Recurrent	7 (18.9)	15.0 ± 5.22		12.4 ± 5.47	

OS, overall survival; DFS, disease-free survival; mo, months. Student's *t* and ANOVA tests were used.

\* Statistically significant, *P* < 0.05.



**Figure 1** LncRNA-MALAT1 expression levels in glioblastoma. Data are represented as medians. The box defines upper and lower quartiles (25% and 75%, respectively) and the error bars indicate upper and lower adjacent limits. Mann–Whitney  $U$  test was used; \*Statistically significant.

( $P < 0.001$ , Fig. 1) with a median expression value (quartile) = 0.015 (0.001–0.086).

### 3.3. ROC curve analysis

Relative expression levels were used to determine *MALAT1* sensitivity and specificity at the cutoff fold change value for discriminating GBM patients from controls. The curve showed high diagnostic performance with an AUC =  $0.925 \pm 0.038$  ( $P < 0.001$ ), 95% CI = 0.850–1.00, with 94.6% sensitivity and 72.7% specificity (Fig. 2).

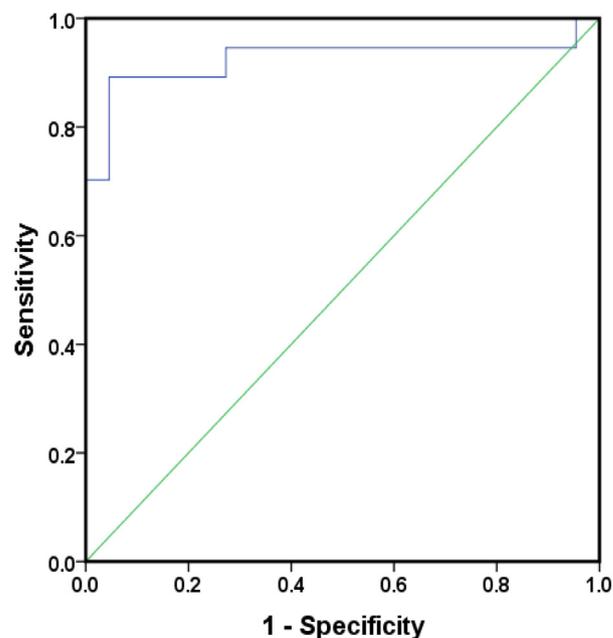
### 3.4. LncRNA *MALAT1* relative expression and clinicopathological features in GBM patients

Median *MALAT1* relative expression was not associated significantly with age (< 50 vs.  $\geq 50$ ,  $P = 0.387$ ) or gender (females vs. males,  $P = 0.196$ ), however fronto-temporal brain tissue samples showed a significant lower expression in comparison to other sites, ( $P = 0.025$ ); Fig. 3. Lower *MALAT1* expression, in addition, was associated with poor prognosis; higher frequency of recurrence, lower overall survival, and shorter disease-free survival (Fig. 3).

As summarized in Table 2, *MALAT1* expression was directly correlated with shorted overall survival ( $r = 0.359$ ,  $P = 0.029$ ) and disease-free survival ( $r = 0.361$ ,  $P = 0.028$ ) and was inversely correlated with recurrence ( $r = -0.334$ ,  $P = 0.044$ ).

### 3.5. *MALAT1* expression and survival of patients with different clinical variables

To explore the prognostic value of the lncRNA-MALAT1 expression in GBM patients, Kaplan–Meier analysis with the log-rank test was used in comparing survival of patients with different clinical variables (Fig. 4). Poor survival has been



Area	Std. Error	Asymptotic Sig.	Asymptotic 95% Confidence Interval	
			Lower Bound	Upper Bound
0.925	0.038	.000	0.850	1.000

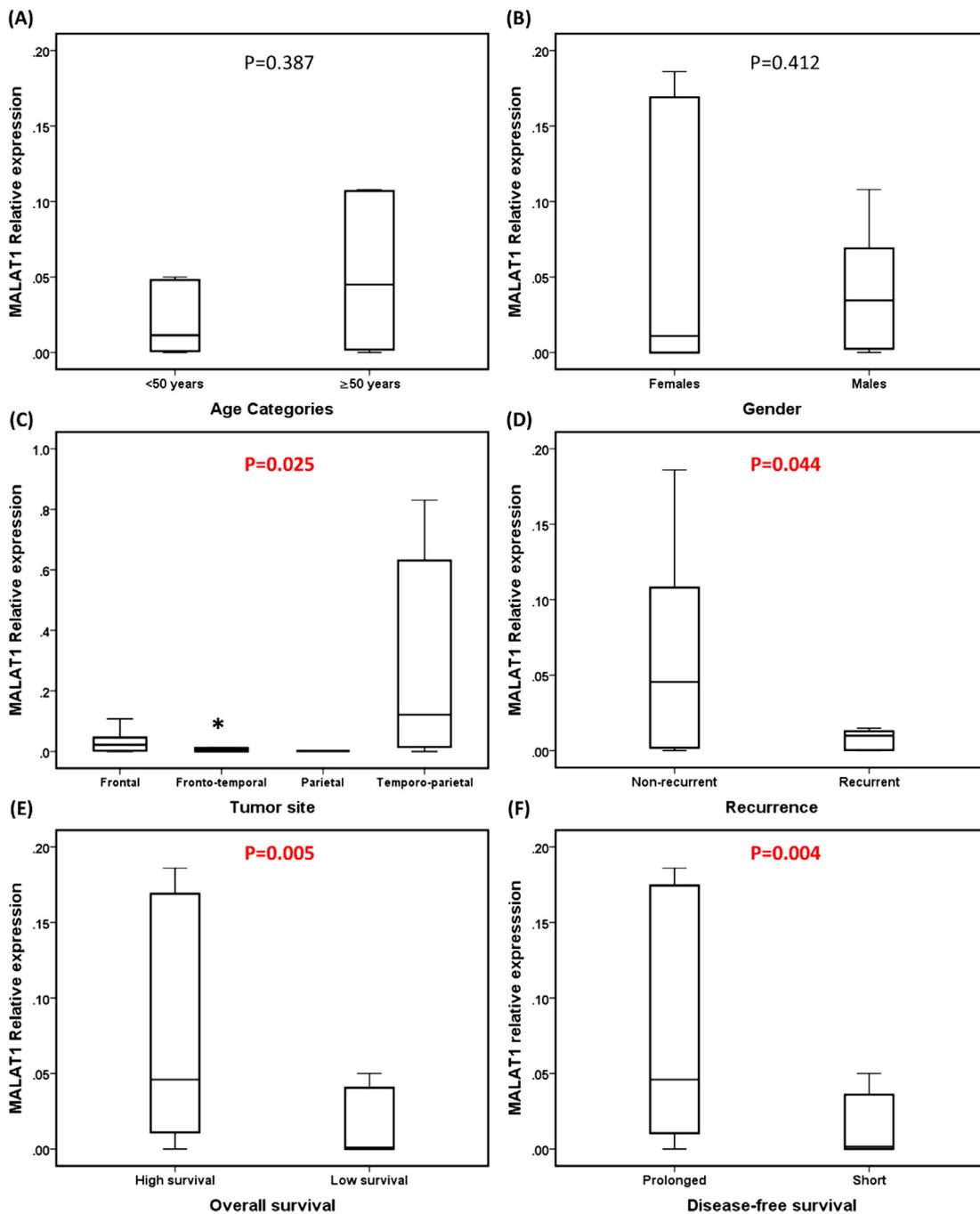
**Figure 2** Receivers operating characteristic curve of MALAT1. AUC: area under curve; CI: confidence interval; SE, standard error; SEN, sensitivity; SP, specificity.

shown in older patients [median (95% CI); 14 (12.5–15.4) and 16 (8.81–23.1) in patients  $\geq 50$  year-old and < 50 year-old, respectively] and among males [median (95% CI); 14 (12.2–15.7) and 22 (13.6–30.3) in males and females, respectively]. Otherwise, there were no significant differences of lncRNA *MALAT1* expressions and survival of patients regarding the tumor location or recurrence rate (Fig. 4).

## 4. Discussion

Human transcriptome analysis has been revealed that the majority of human genome transcripts are non-coding RNAs, including lncRNAs that previously were considered as transcriptional noises [26]. However, an accumulating number of evidences have revealed that lncRNAs could have important functions and may interact with a broad range of RNA molecules through competitively binding with miRNAs, suggesting their vital roles in a wide range of biological processes and human diseases [27]. More specifically, they have been found to be involved in brain development and in the pathogenesis of gliomas, and could serve as novel biomarkers for early diagnosis, prediction of prognosis, and therapeutic targets in gliomas [28,29].

The lncRNA *MALAT1* gene produces a highly abundant and ubiquitously expressed transcript of > 8000 nucleotides that is retained in the nucleus where it is thought to form molecular scaffolds for ribonucleoprotein complexes [30]. *MALAT1* was found to act as decoys that bind to and interfere with the function of other RNAs or proteins (e.g. miRNAs, transcription factors, or RNA-binding proteins),



**Figure 3** LncRNA MALAT1 relative expression and clinicopathological features in GBM patients. Data are represented as medians. The box defines upper and lower quartiles (25% and 75%, respectively) and the error bars indicate upper and lower adjacent limits. \*N.B. In posthoc test parietal tumor group was merged with temporo-parietal one as test could not be performed by single sample.

especially those involved in cell cycle regulation, cell migration, and cancer metastasis [31].

Initially, *MALAT1* was identified by Ji and co-workers in (2003) [32], through subtractive hybridization in a screen for genes associated with metastasis in non-small cell lung cancer. Subsequently, several studies have associated *MALAT1* expression with various cancers and metastasis [33–35]. In the current study, *MALAT1* showed significant lower expression levels in all GBM tissues compared with normal brain tissues. This is consistent with a recent study finding conducted

by Han et al. [22] who found that lncRNA MALAT1 could have a tumor-suppressive function in glioma. This inhibitory effect could be due to suppression of both growth and cell invasion. At the molecular level, they suggested that this could be mediated via regulation of the ERK/MAPK (extracellular signal-regulated kinase/mitogen-activated protein kinase) pathway and expression of MMP2 (matrix metalloproteinase 2). The ERK/MAPK pathway is one of the most important signal transduction pathways, and *MALAT1* upregulation in glioma cells inhibits the growth and invasion of tumor by

**Table 2** Spearman's rho correlation analysis between brain tissue MALAT1 relative expression and the clinicopathological features in GBM patients ( $n = 37$ ).

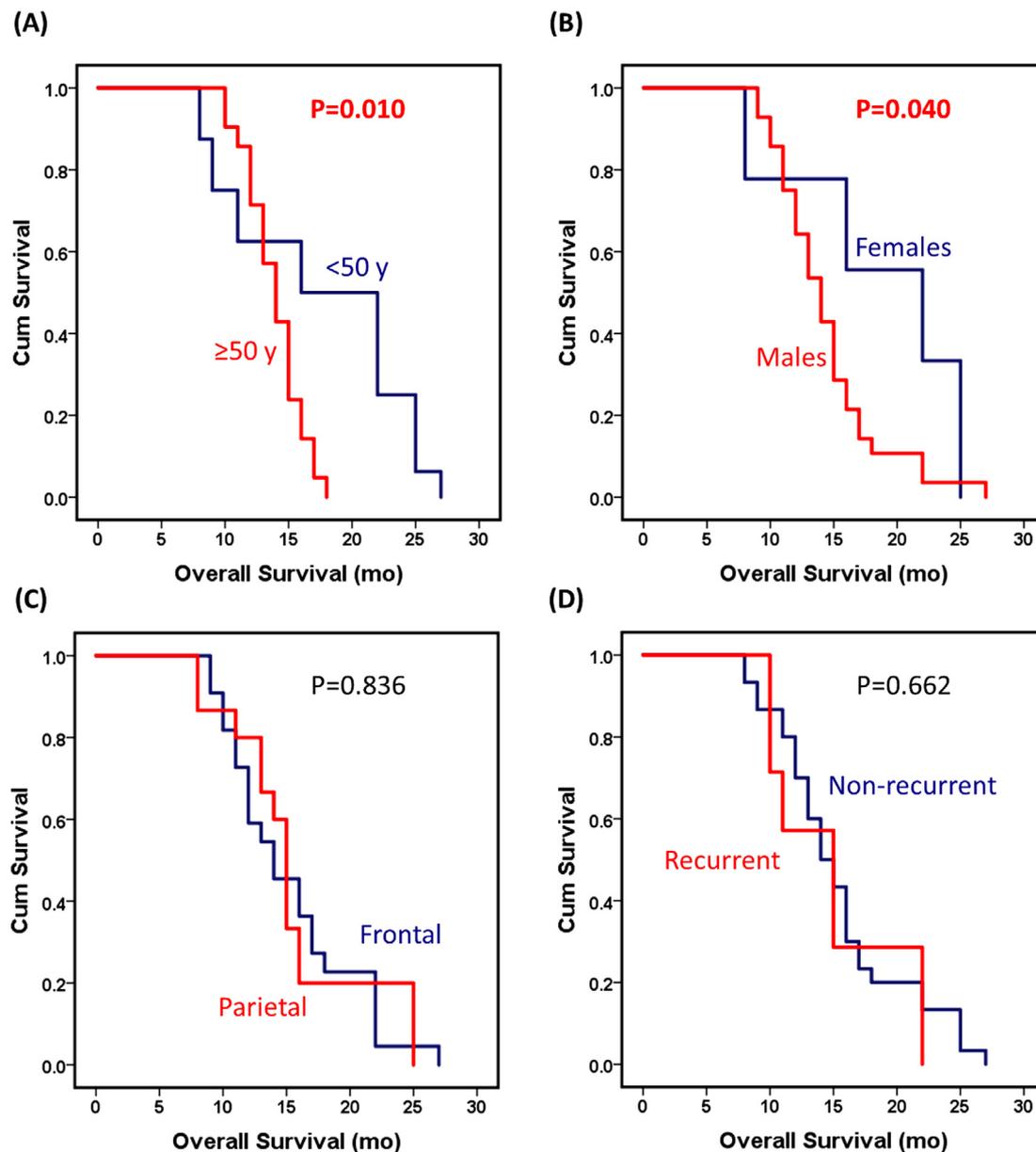
	Correlation coefficient	<i>P</i> value
Age	0.104	0.539
Gender	0.142	0.402
Tumor site	0.050	0.768
OS	0.359	<b>0.029*</b>
DFS	0.361	<b>0.028*</b>
Recurrence	-0.334	<b>0.044*</b>

OS, overall survival; DFS, disease-free survival.

\* Statistically significant,  $P < 0.05$ .

reducing the phosphorylated ERK1/2 expression rather than affecting the total ERK1/2, leading to inactivation of this signaling cascade [22]. In earlier studies, it has been proved that knockout of *MALAT1* in lung cancer cells could significantly reduce the expression of several metastasis-related genes, including Glypican 6 (*GPC6*) [36] and C-X-C motif chemokine 5 (*CXCL5*) [37], leading to MAPK pathway inactivation. Hence *GPC6* or *CXCL5* expression down-regulation could be one of the underlying mechanisms by which *MALAT1* inactivate ERK/MAPK pathway in glioma which require further future validation.

In addition, Bernard et al. [38] have found that *MALAT1* expression in B-cell malignancies was decreased compared with solid tumors. Another study demonstrated significantly lower *MALAT1* expression levels in the plasma of multiple



**Figure 4** Kaplan–Meier curves and the log-rank (Mantel–Cox) test for comparison of the patients' survival with different clinical variables. Cum. Cumulative; mo, months.

myeloma patients [39]. The results of the present study, however, were a little different from others. It has been found that *MALAT1* expression was increased in glioma tissues compared with paired adjacent brain normal tissues by Ma et al. [14]. *MALAT1* was shown, in addition, to act as oncogene in several types of human cancer, such as lung cancer [40], breast cancer [41], cervical cancer [42], bladder urothelial carcinoma [21], gastric cancer [43], colorectal cancer (CRC) [44], hepatocellular carcinoma [34], pancreatic duct adenocarcinoma [45] and melanoma [46]. However, *MALAT1* was downregulated in the cell culture, with the cells exhibiting high metastatic potential for ovarian cancer metastasis [47]. This discrepancy of results among some studies and ours could be due to the fact that *MALAT1* function may vary with various cancer types and context; the same gene may play opposite roles in different cancer types or in different stages of cancer progression [48]. Additionally, emerging evidence has suggested that lncRNAs can potentially interact with other classes of non-coding RNAs including miRNAs [49], or are regulated by transcription factors [50], indicating that lncRNAs may have regulatory roles in a wide range of cellular processes at various levels. In addition, the recent molecular sub-classification of GBM includes *IDH1* (isocitrate dehydrogenase 1) mutation, *p53* mutation, *EGFR* (epidermal growth factor receptor) amplification and *MGMT* (O-6-methylguanine-DNA methyltransferase) promoter methylation status, all of which have proved to be partly associated and are useful in clinical practice [13]. Their aberration may result in significant epigenetic changes, including DNA methylation level, mRNA expression level [51], and lncRNA expression level [52]. Therefore, it is recommended to investigate whether there are some associations between *MALAT1* signature and these well-recognized genetic biomarkers by an intriguing multi-dimension analysis that can give useful explanations for the potential result discrepancies in GBM molecular research.

In the present study, we further found that lower *MALAT1* expression was associated with poor prognosis (i.e. higher frequency of recurrence, lower overall survival and shorter disease-free survival). Several reports indicate that *MALAT1* contributes to the complex molecular mechanisms involved in the control of cell growth, differentiation and motility that could predict the overall prognosis and survival [21]. For example, the expression of *MALAT1* was associated with prostate cancer progression and prognosis [53], GBM survival [13], tumor progression and survival in lung, liver and breast cancer [54], invasion and metastasis of CRC cells [55], proliferation and metastasis of gallbladder cancer cells [56] and tumor recurrence of hepatocellular carcinoma after liver transplantation [57]. All these findings indicate that lncRNA MALAT-1 could be considered as an important novel candidate for future therapeutic intervention. However, the applicability and epigenetic regulation of MALAT-1 targeted strategies for the clinical treatment of GBM requires additional studies.

## 5. Conclusion

In summary, the current study does confirm the lncRNA MALAT1 association with the glioblastoma and its potential role as prognostic biomarker. The study was limited by the relatively small sample size, and the fact that all patients were

grade IV gliomas. We recommend further studies of a larger scale with different glioma grades, to explore the correlation of *MALAT1* expression with different WHO grades and to confirm its possible role as prognostic biomarker. Further functional investigations of this type of lncRNAs on tumor cell lines and xenograft models may increase our understanding of its detailed molecular roles in GBM pathogenesis and progress. Hence, this may be an interesting target for future therapeutic interventions in GBM patients.

## Declaration of conflict of interest

The authors declare that they have no conflict of interest.

## Acknowledgements

The authors would like to thank the Oncology Diagnostic Unit, Suez Canal University, Egypt, for providing the facilities for performing the current work.

## References

- [1] Siegel R, Ma J, Zou Z, Jemal A. Cancer statistics, 2014. *CA Cancer J Clin* 2014;64:9–29.
- [2] Louis DN, Ohgaki H, Wiestler OD, Cavenee WK, Burger PC, Jouvet A, et al. The 2007 WHO classification of tumors of the central nervous system. *Acta Neuropathol* 2007;2007(114):97–109.
- [3] Weathers S-P, Gilbert MR. Advances in treating glioblastoma. *F1000Prime Rep* 2014;6:46.
- [4] Johnson DR, Galanis E. Incorporation of prognostic and predictive factors into glioma clinical trials. *Curr Oncol Rep* 2013;15:56–63.
- [5] Appin CL, Brat DJ. Molecular pathways in gliomagenesis and their relevance to neuropathologic diagnosis. *Adv Anat Pathol* 2015;22:50–8.
- [6] Quinn JJ, Chang HY. Unique features of long non-coding RNA biogenesis and function. *Nat Rev Genet* 2016;17:47–62.
- [7] Wu P, Zuo X, Deng H, Liu X, Liu L, Ji A. Roles of long noncoding RNAs in brain development, functional diversification and neurodegenerative diseases. *Brain Res Bull* 2013;97:69–80.
- [8] Yan B, Yao J, Liu J-Y, Li XM, Wang XQ, Li YJ, et al. LncRNA-MIAT regulates microvascular dysfunction by functioning as a competing endogenous RNA novelty and significance. *Circ Res* 2015;116:1143–56.
- [9] Rinn JL, Chang HY. Genome regulation by long noncoding RNAs. *Annu Rev Biochem* 2012;81:145–66.
- [10] Han L, Zhang K, Shi Z, Zhang J, Zhu J, Zhu S, et al. LncRNA profile of glioblastoma reveals the potential role of lncRNAs in contributing to glioblastoma pathogenesis. *Int J Oncol* 2012;40:2004–12.
- [11] Zhang X, Sun S, Pu JK, Tsang AC, Lee D, Man VO, et al. Long noncoding RNA expression profiles predict clinical phenotypes in glioma. *Neurobiol Dis* 2012;48:1–8.
- [12] Zhang JX, Han L, Bao ZS, Wang YY, Chen LY, Yan W, et al. HOTAIR, a cell cycle associated long noncoding RNA and a strong predictor of survival, is preferentially expressed in classical and mesenchymal glioma. *Neuro-Oncology* 2013;15:1595–603.
- [13] Zhang XQ, Sun S, Lam KF, Kiang KM, Pu JK, Ho AS, et al. A long non-coding RNA signature in glioblastoma multiforme predicts survival. *Neurobiol Dis* 2013;58:123–31.
- [14] Ma KX, Wang HJ, Li XR, Li T, Su G, Yang P, et al. Long noncoding RNA MALAT1 associates with the malignant status and poor prognosis in glioma. *Tumour Biol* 2015;36:3355–9.
- [15] Hutchinson JN, Ensminger AW, Clemson CM, Lynch CR, Lawrence JB, Chess A. A screen for nuclear transcripts identifies

- two linked noncoding RNAs associated with SC35 splicing domains. *BMC Genomics* 2007;8:39.
- [16] Wu Y, Huang C, Meng X, Li J. Long noncoding RNA MALAT1: insights into its biogenesis and implications in human disease. *Curr Pharm Des* 2015;21:5017–28.
- [17] Yoshimoto R, Mayeda A, Yoshida M, Nakagawa S. MALAT1 long non-coding RNA in cancer. *Biochim Biophys Acta* 2016;1859:192–9.
- [18] Lin R, Maeda S, Liu C, Karin M, Edgington TS. A large noncoding RNA is a marker for murine hepatocellular carcinomas and a spectrum of human carcinomas. *Oncogene* 2007;26:851–8.
- [19] Fellenberg J, Bernd L, Delling G, Witte D, Zahlten-Hinguranage A. Prognostic significance of drug-regulated genes in high-grade osteosarcoma. *Mod Pathol: Off J United States Can Acad Pathol* 2007;20:1085–94.
- [20] Schmidt LH, Spieker T, Koschmieder S, Schaffers S, Humberg J, Jungen D, et al. The long noncoding malat-1 RNA indicates a poor prognosis in non-small cell lung cancer and induces migration and tumor growth. *J Thorac Oncol: Off Pub Int Assoc Study Lung Cancer* 2011;6:1984–92.
- [21] Han Y, Liu Y, Nie L, Gui Y, Cai Z. Inducing cell proliferation inhibition, apoptosis, and motility reduction by silencing long noncoding ribonucleic acid metastasis-associated lung adenocarcinoma transcript 1 in urothelial carcinoma of the bladder. *Urology* 2013;81, 209.e1-7.
- [22] Han Y, Wu Z, Wu T, Huang Y, Cheng Z, Li X, et al. Tumor-suppressive function of long noncoding RNA MALAT1 in glioma cells by downregulation of MMP2 and inactivation of ERK/MAPK signaling. *Cell Death Dis* 2016;7:e2123.
- [23] Aithal MGS, Rajeswari N. Validation of housekeeping genes for gene expression analysis in glioblastoma using quantitative real-time polymerase chain reaction. *Brain Tumor Res Treat* 2015;3:24–9.
- [24] Tong D, Heinze G, Pils D, Wolf A, Singer CF, Concini N, et al. Gene expression of PMP22 is an independent prognostic factor for disease-free and overall survival in breast cancer patients. *BMC Cancer* 2010;10:682.
- [25] Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta CT) Method. *Methods* 2001;25:402–8.
- [26] Ponting CP, Oliver PL, Reik W. Evolution and functions of long noncoding RNAs. *Cell* 2009;136:629–41.
- [27] Harries LW. Long non-coding RNAs and human disease. *Biochem Soc Trans* 2012;40:902–6.
- [28] Qureshi IA, Mehler MF. Emerging roles of non-coding RNAs in brain evolution, development, plasticity and disease. *Nat Rev Neurosci* 2012;13:528–41.
- [29] Sun Y, Wang Z, Zhou D. Long non-coding RNAs as potential biomarkers and therapeutic targets for gliomas. *Med Hypotheses* 2013;81:319–21.
- [30] Ren D, Li H, Li R, Sun J, Guo P, Han H, et al. Novel insight into MALAT-1 in cancer: therapeutic targets and clinical applications. *Oncol Lett* 2016;11:1621–30.
- [31] Hrdlickova B, de Almeida RC, Borek Z, Withoff S. Genetic variation in the non-coding genome: involvement of micro-RNAs and long non-coding RNAs in disease. *Biochim Biophys Acta* 2014;1842:1910–22.
- [32] Ji P, Diederichs S, Wang W, Böing S, Metzger R, Schneider PM, et al. MALAT-1, a novel noncoding RNA, and thymosin beta4 predict metastasis and survival in early-stage nonsmall cell lung cancer. *Oncogene* 2003;22:8031–41.
- [33] Cheetham SW, Gruhl F, Mattick JS, Dinger ME. Long noncoding RNAs and the genetics of cancer. *Br J Cancer* 2013;108:2419–25.
- [34] Li G, Zhang H, Wan X, Yang X, Zhu C, Wang A, et al. Long noncoding RNA plays a key role in metastasis and prognosis of hepatocellular carcinoma. *Biomed Res Int* 2014;2014:780521.
- [35] Vassallo I, Zinn P, Lai M, Rajakannu P, Hamou M-F, Hegi ME. WIF1 re-expression in glioblastoma inhibits migration through attenuation of non-canonical WNT signaling by downregulating the lncRNA MALAT1. *Oncogene* 2016;35:12–21.
- [36] Yiu GK, Kaunisto A, Chin YR, Toker A. NFAT promotes carcinoma invasive migration through glypican-6. *Biochem J* 2011;440:157–66.
- [37] Merabova N, Kaminski R, Krynska B, Amini S, Khalili K, Darbinyan A. JCV agnoprotein-induced reduction in CXCL5/LIX secretion by oligodendrocytes is associated with activation of apoptotic signaling in neurons. *J Cell Physiol* 2012;227:3119–27.
- [38] Bernard D, Prasanth KV, Tripathi V, Colasse S, Nakamura T, Xuan Z, et al. A long nuclear-retained non-coding RNA regulates synaptogenesis by modulating gene expression. *EMBO J* 2010;29:3082–93.
- [39] Isin M, Ozgur E, Cetin G, Erten N, Aktan M, Gezer U, et al. Investigation of circulating lncRNAs in B-cell neoplasms. *Clin Chim Acta* 2014;431:255–9.
- [40] Gutschner T, Hämmerle M, Eissmann M, Hsu J, Kim Y, Hung G, et al. The noncoding RNA MALAT1 is a critical regulator of the metastasis phenotype of lung cancer cells. *Cancer Res* 2013;73:1180–9.
- [41] Qi P, Du X. The long non-coding RNAs, a new cancer diagnostic and therapeutic gold mine. *Mod Pathol* 2013;26:155–65.
- [42] Tripathi V, Shen Z, Chakraborty A, Giri S, Freier SM, Wu X, et al. Long noncoding RNA MALAT1 controls cell cycle progression by regulating the expression of oncogenic transcription factor B-MYB. *PLoS Genet* 2013;9:e1003368.
- [43] Okugawa Y, Toiyama Y, Hur K, Toden S, Saigusa S, Tanaka K, et al. Metastasis-associated long non-coding RNA drives gastric cancer development and promotes peritoneal metastasis. *Carcinogenesis* 2014;35:2731–9.
- [44] Ji Q, Zhang L, Liu X, Zhou L, Wang W, Han Z, et al. Long non-coding RNA MALAT1 promotes tumour growth and metastasis in colorectal cancer through binding to SFPQ and releasing oncogene PTBP2 from SFPQ/PTBP2 complex. *Br J Cancer* 2014;111:736–48.
- [45] Liu JH, Chen G, Dang YW, Li CJ, Luo DZ. Expression and prognostic significance of lncRNA MALAT1 in pancreatic cancer tissues. *Asian Pac J Cancer Prev* 2014;15:2971–7.
- [46] Tian Y, Zhang X, Hao Y, Fang Z, He Y. Potential roles of abnormally expressed long noncoding RNA UCA1 and Malat-1 in metastasis of melanoma. *Melanoma Res* 2014;24:335–41.
- [47] Liu SP, Yang JX, Cao DY, Shen K. Identification of differentially expressed long non-coding RNAs in human ovarian cancer cells with different metastatic potentials. *Cancer Biol Med* 2013;10:138–41.
- [48] Vogelstein B, Papadopoulos N, Velculescu VE, Zhou S, Diaz LA, Kinzler KW. Cancer genome landscapes. *Science (New York, NY)* 2013;339:1546–58.
- [49] Salmena L, Poliseno L, Tay Y, Kats L, Pandolfi PP. A ceRNA hypothesis: the Rosetta Stone of a hidden RNA language? *Cell* 2011;146:353–8.
- [50] Yang JH, Li JH, Jiang S, Zhou H, Qu LH. ChIPBase: a database for decoding the transcriptional regulation of long non-coding RNA and microRNA genes from ChIP-Seq data. *Nucleic Acids Res* 2013;41:D177–87.
- [51] Turcan S, Rohle D, Goenka A, Walsh LA, Fang F, Yilmaz E, et al. IDH1 mutation is sufficient to establish the glioma hypermethylator phenotype. *Nature* 2012;483:479–83.
- [52] Ellis BC, Molloy PL, Graham LD. CRNDE: a long non-coding RNA involved in cancer, neurobiology, and development. *Front Genet* 2012;3:270.
- [53] Sowalsky AG, Xia Z, Wang L, Zhao H, Chen S, Bublely GJ, et al. Whole transcriptome sequencing reveals extensive unspliced mRNA in metastatic castration-resistant prostate cancer. *Mol Cancer Res* 2015;13:98–106.

- [54] Diederichs S. Non-coding RNA in malignant tumors. A new world of tumor biomarkers and target structures in cancer cells. *Pathologie* 2010;31:258–62 [Abstract].
- [55] Xu C, Yang M, Tian J, Wang X, Li Z. MALAT-1: a long non-coding RNA and its important 3' end functional motif in colorectal cancer metastasis. *Int J Oncol* 2011;39:169–75.
- [56] Wu XS, Wang XA, Wu WG, Hu YP, Li ML, Ding Q, et al. MALAT1 promotes the proliferation and metastasis of gallbladder cancer cells by activating the ERK/MAPK pathway. *Cancer Biol Ther* 2014;15:806–14.
- [57] Lai MC, Yang Z, Zhou L, Zhu QQ, Xie HY, Zhang F, et al. Long non-coding RNA MALAT-1 overexpression predicts tumor recurrence of hepatocellular carcinoma after liver transplantation. *Med Oncol* 2012;29:1810–6.