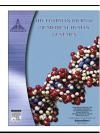


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ORIGINAL ARTICLE

Association between interleukin-4 (IL-4), gene polymorphisms (C-589T, T+2979G, and C-33T) and migraine susceptibility in Iranian population: A case—control study



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KEYWORDS

IL-4; Migraine; RFLP-PCR; Genotyping; Neuro-inflammation; Headache **Abstract** *Background:* Migraine is a chronic neurological disease characterized by recurrent moderate to severe headaches commonly in association with neuro-inflammation. Interleukin-4 (IL-4), an anti-inflammatory cytokine, plays an important role in modulating pain threshold and has an essential role in stimulation of pain receptors in the trigeminal nerve fibers.

Aim of the study: The current study aimed to investigate the possible associations between IL-4 single nucleotide polymorphisms (SNPs) and susceptibility to migraine in Iranian patients.

Patients and methods: In a prospective case—control study, we studied blood samples of 190 patients with migraine (migraineurs) and 200 healthy controls (HCs) for analysis of gene variants. Genotyping for the IL-4 SNPs: C-589T (rs2243250), T+2979G (rs2227284), and C-33T (rs2070874) were performed using PCR-RFLP. Statistical analysis was performed using the SPSS version 21.0 (SPSS, Chicago) and SNPStats version 1.14.0.

Results and conclusion: Among IL-4 SNPs, rs2243250 (TC genotype, OR = 0.25, 95% CI = 0.13–0.50, P = 0.001) and rs2227284 (TG and TT genotypes, OR = 0.44, 95% CI = 0.23–0.92, P = 0.029 and OR = 0.38, 95% CI = 0.18–0.79, P = 0.009 respectively) were significantly

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Abbreviations: IL-4, interleukin-4; IL-9, interleukin-9; TNF-α, tumor necrosis factor-α; IL-1β, interleukin-1β; CNS, central nervous system; CCR2, C-C chemokine receptor type 2; TGFB1, transforming growth factor-b1; NOS3, nitric oxide synthase 3; Th2, T helper type 2; SNPs, single nucleotide polymorphisms

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associated with migraine. No significant associations between IL-4 SNP rs2070874 (TC, TT and CC genotypes) and migraine were found. The most frequent genotypes in the migraineurs were CC in both SNPs rs2243250 (79%), and rs2070874 (71.5%), as well as GG for SNP rs2227284 (64%). There was no statistically significant relationship between these SNPs and different subclasses (common, classic and complicated) of migraine. Our findings revealed that in IL-4 rs2243250 and rs2227284 genotypes and allele frequencies have a role in susceptibility to migraine in our population. Therefore, it is suggested that in addition to other factors, IL-4 genetic variations also play a pivotal role in the progress of migraine.

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1. Introduction

Migraine is a rigorous and painful headache accompanying with sensory warning and is a public health problem of great impact on both the patient and society [1]. The two major subclasses of migraine are common migraine (without aura) and classic migraine (with aura or neurological symptoms) [2]. Since about half of migraineurs do not pursue medical attention and there is no economic, social or ethnic limitation, it is difficult to precisely determine disease prevalence in the community [3]. It seems that about 15 to 16% of women and 5 to 9% of men are affected with migraine and its prevalence is the highest among the ages of 30-49 worldwide [4]. Migraine etiology is multifactorial, involving both various genetic and environmental factors, but scientists consider three important mechanisms for its pathophysiology including: inflammatory, neurological and cardiovascular impairments [5,6]. Cell and molecular association studies may point to the novel molecules that mediate migraine disorder and enable its management. According to the theory of neuroinflammation, in the migraine, ion channels (Na2+, Ca2+, K2+) and inflammatory mediator (TNF-α and IL-1β) activation in the meninges sensory nerves, stimulates pain receptors in these area [7,8]. In addition to changes in the inflammatory cytokines, variations in the anti-inflammatory mediators in migraine patients have been reported but there are conflicting results on the mechanisms involved [9-14]. The interaction between immune cells is regulated by several mediators, including interleukins and cytokines, which play an essential role in pathobiological processes such as, inflammation, immunity and pain [15]. Widely, interleukins and their receptors are present in both peripheral neurovascular inflammation at meningeal/ganglia level, and essential sensitization processes in central nervous system (CNS) and likely to be involved in pain threshold modulation [16–21]. On the other hand, peripheral blood interleukins operate mainly at a level of areas innervated by trigeminal ganglion neurons related to neurovascular system [22]. The human IL-4 gene is located on chromosome 5q31 and consists of 25 kb. So far, numerous allelic variant polymorphisms (http://www.ncbi. nlm.nih.gov/SNP/) have been found in IL-4 gene, that the important ones are including -590C/T (rs2243250), -33C/T +3437C/G (rs2227282), (rs2070874), and 2979G/T (rs2227284) [23]. To understand the probable role of IL-4, as anti-inflammatory cytokine, in migraine headaches in the leading research we analyzed its imperative polymorphisms in migraineurs with three different subclass of disease and compared them to healthy controls.

2. Subjects and methods

2.1. Patients and samples

The study was approved by the ethics in medical research committee at Zahedan University of Medical Sciences, and was conducted with clinical samples from migraine patients $(N = 190, \text{ age: } 13 \text{ to } 66 \text{ years, age mean } \pm \text{SD: } 31.72$ \pm 10.17) who were treated at the Department of Neurology, Ali-ebn Abitaleb Hospital, Zahedan, Iran, from August 2013 to February 2014. Healthy controls (HCs) without any inflammatory, neurological diseases, migraine headache and specific systemic disease (N = 200, age: 15 to 75 years, age mean \pm SD: 35.1 \pm 12.2) from volunteer blood donors were selected during the same time. A diagnosis of migraine was made according to standardized criteria of international headache classification [24]. Patients were excluded if they had a history of inflammatory diseases or received antiinflammatory medicines. Patients adjusted in three definite groups including common (without aura, N = 96, 64 female and 32 male, age mean \pm SD: 31.0417 \pm 10.457) classic (with aura, N = 78, 56 female and 22 male, age mean \pm SD: 32.33 \pm 10.57) and complicated (N = 16, 4 male and 12 female, age mean \pm SD: 32.87 \pm 6.46) subtypes of migraine. All patients were informed of the study and participated voluntarily and written consents were taken. The work is carried out in accordance with the code of Ethics of the World Medical Association (Declaration of Helsinki) for experiments in humans.

2.2. Blood collection and DNA extraction

Whole blood (10 mL) samples were taken from all subjects and collected in separator tubes (contain EDTA, 0.5 M) and centrifuged for 15 min at 150 g (gravity) at 20 °C and then serum was stored at -20 °C in sterile plastic tubes for DNA extraction. Genomic DNA was extracted from the serum of 190 subjects with migraine headaches and 200 HCs using the DNA extraction kit (DIAtom DNA Prep., GORDIZ, Moscow, Russia) according to the manufacturer's instruction. DNA quality extracts were analyzed by electrophoresis. By Nano-Drop DNA concentrations about 60 ng/ μ l was obtained and ratio of 260/280 nm around 1.7 to 1.9 was accepted [25].

2.3. PCR analysis

Four IL-4 SNPs C-589T (rs2243250), T+2979G (rs2227284), and C-33T (rs2070874) were analyzed through restriction

fragment length polymorphism-polymerase chain reaction (RFLP-PCR) method [26]. PCR amplifications were performed in a final volume of 20 µl containing, 10 µl master mix (TAKARA, Tokyo, Japan), 0.7 µl (10 pmol) of each primer, 2 µL template DNA, and 6.8 µl DNase-free water was used. For rs2070874 and rs2227284, the amplification was performed with an initial denaturation step at 94 °C for 5 min; followed by 35 cycles at 94 °C for 30 s, 58 °C for 35 s, and 72 °C for 30 s with a final extension at 72 °C for 5 min. For SNP rs2243250, the cycling conditions were as follow: an initial denaturation step at 94 °C for 5 min, followed by 35 cycles at 94 °C for 30 s, 50 °C for 30 s, and 72 °C for 30 s with a final extension at 72 °C for 5 min. The PCR product was checked for size and purity by 1.5% agarose gel electrophoresis. The sizes of the fragments were 195bp, 220bp and 223bp for the C-589T, T+2979G and C-33T regions, respectively (Table 1).

2.4. RFLP analysis

Final volume of 20 μ L including 2 μ L of 10 \times Buffer, 0.5 μ L of enzyme, 7 µL of PCR product, 10.5 µL of double distilled water was used for all amplification products overnight at 37 °C, and 10 μL sample loaded for electrophoresis. AvaII (Thermo Scientific) endonuclease digested pattern for rs2243250 amplification product were 177bp and 18bp for the CC, 195bp, 177bp and 18bp for the TC and 195bp for the TT genotypes (Table 1). AluI (Thermo Scientific) endonuclease digested pattern for rs2227284 amplification product were 122bp and 98bp for the GG, 122bp, 98bp, 53bp, and 45bp for the TC and 122bp, 53bp, and 45bp for the TT genotypes (Table 1). BsmAI (Thermo Scientific) endonuclease digested pattern for rs2070874 amplification product were 178bp and 45bp for the CC, 178bp, 140bp, 45bp, and 38bp for the TC and 140bp, 45bp, and 38bp for the TT genotypes (Table 1).

2.5. Statistical analysis

SPSS version 21.0 (SPSS, Chicago) and SNPStats version 1.14.0 were used for all the statistical analyses. The association between genotypes and IL-4 was estimated using the odds ratio (OR) and 95% confidence intervals (95% CI) from logistic regression analyses. The Hardy–Weinberg equilibrium (HWE) was tested with the X2 test for any of the SNPs under

consideration. The significance level was set at $P \le 0.05$ for all the tests.

3. Results

3.1. Association of IL-4 SNP (rs2070874 C/T) and migraine

The C/C, T/C and T/T genotypes of -33 C/T were found in 74%, 19% and 7% in HCs, in comparison with 79%, 15% and 6% in migraineurs, respectively. The allele frequency of IL-4 rs2070874 (C/T) were 83.5% (C), 16.5% (T) in HCs and 86% (C), 14% (T) in migraineurs, respectively. Distributions of IL-4 polymorphisms in rs2070874 (C/T) were not significantly different between patients and controls for TC (OR = 0.727, P = 0.412), and TT (OR = 0.847, P = 0.773) genotypes and also C (OR = 1.246, P = 0.438) and T (OR = 0.8, P = 0.434) alleles (Table 2). Similarly, there were no associations with migraine classic, common and complicated subtypes and IL-4 rs2070874 (G/T) SNP in this population (Table 3).

3.2. Association of IL-4 SNP (rs2243250 C/T) and migraine

The C/C, T/C and T/T genotypes of -589 C/T were found in 45%, 44% and 11% in HCs, in comparison with 71.5%, 18% and 10.5% in migraineurs, respectively. The allele frequency of IL-4 rs2243250 (C/T) were 67% (C), 33% (T) in HCs and 80.5% (C), 19.5% (T) in migraineurs, respectively. There were significant associations between TC (OR = 0.256, P = 0.001) and CC (OR = 0.386, P = 0.00) genotype and also C (OR = 2.036, P = 0.002) and T (OR = 2.036, P = 0.000) alleles of IL-4 rs2243250 SNP and migraine (Table 2). TC genotype could be considered as protective and CC genotype could be considered as risk factor in migraine headaches. There were no significant differences between patients and controls for the TT (OR = 2.0602, P = 0.287) genotype (Table 2). There were no associations with migraine subtypes and IL-4 SNP (rs2243250 C/T) in this population (Table 3).

3.3. Association of IL-4 SNP (rs2227284 T/G) and migraine

The G/G, T/G and T/T genotypes of +2979 T/G were found in 27%, 38% and 35% in HCs, in comparison with 46.5%,

Table 1 Primer sequences and restriction enzymes used for detection of IL-4 gene polymorphisms.						
SNPs (rs number)	Sequence $(5' \rightarrow 3')$ F: Forward R: Reverse	Digestion pieces (restriction enzyme)	Amplicon size			
C-33T (rs2070874)	F: CAA GTT ACT GAC AAT CTG GTG T R: CGG CAC ATG CTA GCA GGA A	Allele C: 178, 45 (BsmAI) Allele T: 140, 45, 38 (BsmAI)	223 bp			
C-589T (rs2243250)	F: TAA ACT TGG GAG AAC ATG GT R: TGG GGA AAG ATA GAG TAA TA	Allele C: 177, 18 (AvaII) Allele T:195 (AvaII)	195 bp			
T + 2979G (rs2227284)	F: CTA CTC TTG GCA GTT GCT GGA A R: GGA ACT CTC TGT AGA ATT ATG AAC TTT AGG TC	Allele T: 122, 53, 45 (AluI) Allele G: 122, 98 (AluI)	220 bp			

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Table 2	Genotype and	allelic	frequencies	of IL-4	SNPs in	patients and	control subjects.

SNP (rs number)	Genotypes and Alleles	Patient n (%)	Control n (%)	^a OR (95% CI)	P-value
C-33T (rs2070874)	CC	150(79%)	148(74%)	1.00	-
	TC	28(15%)	38(19%)	0.727 (0.340–1.557)	0.412
	TT	12(6%)	14(7%)	0.846 (0.271–2.636)	0.773
	TC + TT	40(21%)	52(26%)	0.759(0.390-1.477)	0.417
	C	164(86%)	167(83.5%)	1.246 (0.712–2.176)	0.438
	T	26(14%)	33(16.5%)	0.8 (0.4–1.4)	0.434
C-589T (rs2243250)	CC	136(71.5%)	90(45%)	1.00	_
	TC	34(18%)	88(44%)	0.256 (0.13-0.502)	0.001***
	TT	20(10.5%)	22(11%)	0.602 (0.236–1.533)	0.287
	TC + TT	54(28.5%)	110(55%)	0.325(0.179-0.589)	0.000^{***}
	C	153(80.5%)	134(67%)	2.036 (1.279-3.241)	0.002^{**}
	T	37(19.5%)	66(33%)	0.428 (0.263-0.696)	0.000***
T + 2979G (rs2227284)	GG	88(46%)	54(27%)	1.00	-
	TG	58(30.5%)	76(38%)	0.448 (0.237-0.925)	0.029^{*}
	TT	44(23.5%)	70(35%)	0.386 (0.188-0.790)	0.009^{**}
	GG + TG	102(54%)	146(73%)	0.429(0.236-0.780)	0.005**
	G	117(62%)	92(46%)	1.881 (1.25–2.816)	0.002^{**}
	T	73(38%)	108(54%)	0.581 (0.386-0.873)	0.008**

^{*} p < 0.05.

30.5% and 23.5% in migraineurs, respectively. The allele frequency of IL-4 rs2243250 (T/G) were 46% (G), 54% (T) in HCs and 61% (C), 39% (T) in migraineurs, respectively. There were significant associations between TG (OR = 0.448, P = 0.029) and TT (OR = 0.386, P = 0.009) genotypes and also G (OR = 1.881, P = 0.002) and T (OR = 0.581, P = 0.008) alleles of IL-4 rs2227284 SNP and migraine (Table 2). T allele could be considered as protective and C allele could be considered as risk factor in migraine headaches. There were no associations with migraine subtypes and IL-4 rs2227284 SNP in this population (Table 3).

4. Discussion

Migraine is a severe neurological disorder that causes a strong throbbing or pulsating pain in one area of the head and can be accompanied by nausea, vomiting and extreme photophobia [2]. Several studies used a candidate gene approach to elucidate genetic contribution to neuropathic pain phenotypes; however, the data are limited and inconsistent [27]. The genetic background of migraine consists of common or overlapped pathways and the responsible genes may provide insight regarding the pathophysiological mechanisms that can explain their comorbidity with migraine [28,29]. Cytokines, small protein molecules secreted in response to immune stimuli, are involved in signaling that activates CNS glial cells and this activation is part of a poorly understood interaction between immune challenge and host that can lead to the development or facilitation of pathologic pain [30]. Data from the large cohort of Caucasian women (n = 25,713) in 77 different SNPs suggested that there is an association between variants in some inflammatory mediators including TNF- α rs673 (OR = 0.52, CI = 0.30-0.89, p = 0.017), CCR2rs1799864

(OR = 1.12, 95% CI = 1.03-1.21, p = 0.007), TGFB1rs1800469 (OR = 0.93, 95% CI = 0.89–0.89, p = 0.009), rs3918226 (OR = 1.13, 95% CI = 1.01–1.27, NOS3 p = 0.04), and IL-9 rs2069885 (OR = 1.12, 95% CI = 1.02– 1.24, p = 0.02) with migraine [31]. In this study, for the first time we focused on anti-inflammatory cytokine, IL-4 in patients with migraine headaches to examine the hypothesis that say migraine headaches could be caused by change in immune system [32]. It has been characterized that IL-4 is required for the generation of the Th2-derived cytokines and several reports indicate a reduced level of IL-4 during migraine attack [33-36]. Perini et al. have found no significant differences between IL-4 plasma levels in migraine patients with and without aura (p = 0.07); as well as patients outside and during the attacks (p = 0.06) and also between HCs and patients (p = 0.06) [16]. Conversely, Sarchielli et al. reported that levels of IL-4 were reduced at the time of migraine attack (p < 0.004) and its levels at the end of the attack returned to those detected at attack onset [35]. Moreover, Munno et al. suggested that IL-4 plasma levels were decreased during attacks and undetectable in 62.5% of patients with migraine headaches without aura [37]. In other study, although no significant fluctuations of the IL-4 plasma levels during the headache-free period in children with migraine and tensiontype headaches have been found, immune dysfunction (abnormal changes in pro- and anti-inflammatory cytokines) in migraineurs could not be totally excluded [38]. Such controversies may be due to different population studies, long medical history of migraineurs and frequent intake of prophylactic drugs or analgesic treatments. In relation to adult glioma, there were close association between IL4 (rs2243248, T-1098G), IL6 (rs1800795, G-174C) and overall risk of glioma [39]. Another study demonstrated no association between IL-4 rs2227284 SNP and rheumatoid arthritis (RA) that is the

^{**} p < 0.01.

^{***} p < 0.001: significant p-value.

^a Adjusted for sex and age.

SNP (rs number)	Genotypesand Alleles	Common n (%)	Classic n (%)	Complicated n (%)	^a OR (95% CI)	P-value
C-33T	CC	74(77%)	67(85.5%)	8(50%)	1.00	-
(rs2070874)	TC	12(13%)	11(14.5%)	4(25%)	1.397(0.567-3.44)	0.467
	TT	10(10%)	0	4(25%)	1.035(0.271-3.954)	0.959
	TC + TT	22(23%)	11(14.5%)	8(50%)	1.279(0.584-2.801)	0.538
	C	80(83%)	73(93%)	10(62.5%)	0.706(0.307-1.623)	0.411
	T	16(17%)	5(7%)	6(37.5%)	1.41(0.616–3.251)	0.411
C-589T (rs2243250)	CC	68(71%)	60(77%)	8(50%)	1.00	-
	TC	13(14%)	15(19%)	4(25%)	1.298(0.559-3.014)	0.544
	TT	15(15%)	3(4%)	4(25%)	0.849(0.285-2.528)	0.768
	TC + TT	28(29%)	18(23%)	8(50%)	1.114(0.546-2.271)	0.767
	C	84(87%)	67(86%)	10(62.5%)	0.58(0.309-1.093)	0.090
	T	12(13%)	11(14%)	6(37.5%)	1.82(0.95-3.459)	0.064
T + 2979G (rs2227284)	GG	44(46%)	37(48%)	6(37%)	1.00	-
	TG	25(26%)	28(35%)	4(26%)	1.181(0.558-2.498)	0.664
	TT	27(28%)	13(17%)	6(37%)	0.942(0.411-2.160)	0.888
	TG + TT	52(54%)	41(52%)	10(63%)	1.073(0.561-2.050)	0.832
	G	57(59%)	51(66%)	8(50%)	0.526(0.274-1.006)	0.050
	T	39(41%)	27(34%)	8(50%)	1.901(0.993-3.638)	0.050

commonest autoimmune disease [40]. It has been confirmed that IL-4 gene polymorphisms may influence the function of mononuclear cells to produce not only IL-4 but also other cytokines [41]. In the Pakistani cohort study, it has been reported that the most frequent genotypes of IL-4 in the asthma and allergic rhinitis groups were TT for SNP rs2243250, and GG for SNP rs2227284 [42]. As well as, IL-4 rs2070874 was not found to be associated with either asthma or allergic rhinitis in the Pakistani cohort [42]. Recently Shang et al. have demonstrated that IL-4 rs2243250 SNP may be associated with high levels of serum IL-4, which may increase the risk of atopic dermatitis in children [43]. Lu et al. findings suggest that SNP in IL-4 rs2070874 (OR: 3.438, 95% CI: 1.032-11.458, P = 0.044) may be a risk factor for hepatocellular carcinoma in Chinese males [44]. It is notable that, there are not available any data from previous epidemiological studies exploring theses IL-4 SNPs in migraineurs population all over the word. Therefore, our results for the first time provided evidence that enhance our understanding of how migraine may relate to, an anti-inflammatory cytokine, IL-4 gene variation. Mentioned reports are relatively conflicting with our recent leading study (but in different population and disease) which revealed that the IL-4 rs2243250 SNP in the genotype TC and also rs2227284 SNP in the genotypes TG and TT playa protective role. These polymorphisms could resulting to changing in Th2 release IL-4, or affecting IL-4 affinity to their cell targets and consequently unbalance between Th1/Th2 cytokines may possibly influence the spreading of pain producing processes in migraine. IL-4 appears to be a prospective target for future development of migraine-specific preventive therapies. The data presented here must be viewed with caution due to the relatively small number of patients enrolled and therefore these results should be taken as preliminary. Similar studies enrolling greater sample sizes and composed of other ethnic groups from different countries may contribute to confirming our findings.

5. Conclusion

In conclusion, our results implicate that the IL-4 rs2243250 (TC) and rs2227284 (TG and TT) SNPs have a protective role in susceptibility to migraine disease in Iranian patients. No significant associations between IL-4 SNP rs2070874 (TC, TT and CC genotypes) and migraine were found. There was no statistically significant relationship between these SNPs and different subclasses (common, classic and complicated) of migraine.

Conflict of interest

All the authors declare that they do not have financial disclosure or conflicts of interest.

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References

- [1] Goadsby PJ. Recent advances in the diagnosis and management of migraine. BMJ 2006;332(7532):25-9.
- [2] Lipton RB, Bigal ME, Steiner J, Silberstein SD, Olesen J. Classification of primary headaches. Neurology 2004;63 (3):427-35.
- [3] Victor TW, Hu X, Campbell JC, Buse DC, Lipton RB. Migraine prevalence by age and sex in the United States: a life-span study. Cephalalgia 2010;30(9):1065-72.
- [4] Stovner LJ, Hagen K. Prevalence burden and cost of headache disorders. Curr Opin Neurol 2006;19(3):281-5.

p < 0.05, **p < 0.01, ***p < 0.001: significant p-value.

Adjusted for sex and age.

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- [5] Mulder EJ, Van Baal C, Gaist D, Kallela M, Kaprio J, Svensson DA, et al. Genetic and environmental influences on migraine: a twin study across six countries. Twin Res 2003;6(5):422–31.
- [6] Pietrobon D, Moskowitz MA. Pathophysiology of migraine. Annu Rev Physiol 2013;75:365–91.
- [7] Waeber C, Moskowitz MA. Migraine as an inflammatory disorder. Neurology 2005;64(10 Suppl 2):S9–S15.
- [8] Yan J, Dussor G. Ion channels and migraine. Headache 2014;54 (4):619–39.
- [9] Yilmaz IA, Ozge A, Erdal ME, Edgünlü TG, Cakmak SE, Yalin OO. Cytokine polymorphism in patients with migraine: some suggestive clues of migraine and inflammation. Pain Med 2010;11 (4):492–7.
- [10] Kaleagasi H, Özgür E, Özge C, Özge A. Bronchial hyperreactivity in migraine without aura: is it a new clue for inflammation? Headache 2011;51(3):426–31.
- [11] Raddant AC, Russo AF. Calcitonin gene-related peptide in migraine: intersection of peripheral inflammation and central modulation. Expert Rev Mol Med 2011;13:e36.
- [12] Franceschini A, Vilotti S, Ferrari MD, van den Maagdenberg AM, Nistri A, Fabbretti E. TNFα levels and macrophages expression reflect an inflammatory potential of trigeminal ganglia in a mouse model of familial hemiplegic migraine. PLoS ONE 2013:8(1):e52394.
- [13] Filipović B, Matak I, Lacković Z. Dural neurogenic inflammation induced by neuropathic pain is specific to cranial region. J Neural Transm 2014;121(5):555–63.
- [14] Woldeamanuel Y, Rapoport A, Cowan R. The place of corticosteroids in migraine attack management: a 65-year systematic review with pooled analysis and critical appraisal. Cephalalgia 2015;35(11):996–1024.
- [15] Lane Thomas E, Carson M, Bergmann C, Wyss-Coray T. Cytokines in CNS inflammation and disease. Cent Nerv Syst Dis Inflammation 2008:59–106, ISBN 2008; 978-0-387-73893-2.
- [16] Perini F, D'andrea G, Galloni E, Pignatelli F, Billo G, Alba S, et al. Plasma cytokine levels in migraineurs and controls. Headache 2005;45(7):926–31.
- [17] Brietzke E, Mansur RB, Grassi-Oliveira R, Soczynska JK, McIntyre RS. Inflammatory cytokines as an underlying mechanism of the comorbidity between bipolar disorder and migraine. Med Hypotheses 2012;78(5):601–5.
- [18] de Goeij M, van Eijk LT, Vanelderen P, Wilder-Smith OH, Vissers KC, van der Hoeven JG, et al. Systemic inflammation decreases pain threshold in humans in vivo. PLoS ONE 2013;8 (12):e84159.
- [19] Turner MD, Nedjai B, Hurst T, Pennington DJ. Cytokines and chemokines: at the crossroads of cell signalling and inflammatory disease. Biochim Biophys Acta 2014;1843(11):2563–82.
- [20] Bai YM, Chiou WF, Su TP, Li CT, Chen MH. Pro-inflammatory cytokine associated with somatic and pain symptoms in depression. J Affect Disord 2014;155:28–34.
- [21] Luchting B, Rachinger-Adam B, Heyn J, Hinske LC, Kreth S, Azad SC. Anti-inflammatory T-cell shift in neuropathic pain. J Neuroinflammation 2015;12:12.
- [22] Pedersen LM, Schistad E, Jacobsen LM, Røe C, Gjerstad J. Serum levels of the pro-inflammatory interleukins 6 (IL-6) and -8 (IL-8) in patients with lumbar radicular pain due to disc herniation: a 12-month prospective study. Brain Behav Immun 2015;46:132-6.
- [23] Naslednikova IO, Konenkov VI, Ryazantseva NV, Novitskii VV, Tkachenko SB, et al. Role of genetically determined production of immunoregulatory cytokines in immunopathogenesis of chronic viral hepatitides. Bull Exp Biol Med 2007;143(6):706–12.
- [24] Headache Classification Commitee of the International Headache Society. Classification and diagnostic criteria for headache disorders, cranial neuralgias and facial pain. 2nd edn. Cephalalgia, 2004, Suppl 1, 1–160.

- [25] Sanadgol N, Ramroodi N, Ahmadi GA, Komijani M, Moghtaderi A, Bouzari M, et al. Prevalence of cytomegalovirus infection and its role in total immunoglobulin pattern in Iranian patients with different subtypes of multiple sclerosis. New Microbiol 2011;34(3):263–74.
- [26] Heidary M, Rakhshi N, Pahlevan Kakhki M, Behmanesh M, Sanati MH, Sanadgol N, et al. The analysis of correlation between IL-1B gene expression and genotyping in multiple sclerosis patients. J Neurol Sci 2014;343(1–2):41–5.
- [27] Thomas B, Strouse MD. The relationship between cytokines and pain/depression: a review and current status. Curr Pain Headache Rep 2007;11(2):98–103.
- [28] Belfer I, Dai F. Phenotyping and genotyping neuropathic pain. Curr Pain Headache Rep 2010;14(3):203–12.
- [29] Migraine SatheSNeurogenetic Disorders. Curr Pain Headache Rep 2013;17(9):360.
- [30] Kors E, Haan J, Ferrari M. Migraine genetics. Curr Pain Headache Rep 2003;7(3):212–7.
- [31] Schürks M, Kurth K, Buring JE, Zeel RYL. A candidate gene association study of 77 polymorphisms in migraine. J Pain 2009;10(7):759–66.
- [32] Boækowski L, Sobaniec W, Elazowska-Rutkowska B. Proinflammatory plasma cytokines in children with migraine. Pediatr Neurol 2009;41(1):17–21.
- [33] Martelletti P, Stirparo G, Morrone S, Rinaldi C, Giacovacco M. ICAM-1, soluble ICAM-1 and interleukin-4 by nitricoxide expression in migraine patients. J Mol Med 1997;75(6):448-53.
- [34] Martelletti P, Zicari A, Realacci M, Fiore G, De Filippis S, Stirparo G, et al. Expression of NOS-2, COX-2 and Th1/Th2 cytokines in migraine. J Headache Pain 2001;2(Suppl 1):s51–6.
- [35] Sarchielli P, Alberti A, Baldi A, Coppola F, Rossi C, Pierguidi L. Proinflammatory cytokines, adhesion molecules and lymphocyte integrin expression in the internal jugular blood of migraine patients without aura assesse edictally. Headache 2006;46 (2):200-7.
- [36] Maier E, Duschl A, Horejs-Hoeck J. STAT6-dependent and independent mechanisms in Th2 polarization. Eur J Immunol 2012;42(11):2827–33.
- [37] Munno I, Marinaro M, Bassi A, Cassiano MA, Causarano V, Centoze V. Immunological aspects in migraine: increase of IL-10 plasma levels during attack. Headache 2001;41(8):764–7.
- [38] Leszek B, Joanna O, Wojciech S, Beata I, Wojciech K, Krzysztof S. Anti-inflammatory plasma cytokines in children and adolescents with migraine headaches. Pharmacol Rep 2010;62 (2):287–91.
- [39] Brenner AV, Butler MA, Wang SS, Ruder AM, Rothman N, Schulte PA, et al. Single-nucleotide polymorphisms in selected cytokine genes and risk of adult glioma. Carcinogenesis 2007;28 (12):2543-7.
- [40] Ioanna M, Till SH, Moore DJ, Wilson AG. Lack of association or interactions between the IL-4, IL-4Rα and IL-13 genes, and rheumatoid arthritis. Arthritis Res Ther 2008;10(4):R80.
- [41] Bartova J, Linhartova PB, Podzimek S, Janatova T, Svobodova K, Fassmann A. The effect of IL-4 gene polymorphisms on cytokine production in patients with chronic periodontitis and in healthy controls. Mediators Inflamm 2014, ID 185757, 11.
- [42] Micheal S, Minhas K, Ishaque M, Ahmed F, Ahmed A. IL-4 gene polymorphisms and their association with atopic asthma and allergic rhinitis in Pakistani patients. J Investig Allergol Clin Immunol 2013;23(2):107–11.
- [43] Shang H, Cao XL, Wan YJ, Meng J, Guo LH. IL-4 gene polymorphism may contribute to an increased risk of atopic dermatitis in children. Dis Markers 2016;2016:1021942.
- [44] Yu Lu, Zhitong Wu, Peng Qiliu, Ma Liping, Zhang Xiaolian, Zhao Jiangyang, et al. Role of *IL-4* gene polymorphisms in HBV-related hepatocellular carcinoma in a Chinese population. PLoS ONE 2014;9(10):e110061.