Removal of Cadmium by Silver Nanoparticles Incorporated in Electrospun Natural Rubber Latex/Polyvinyl Alcohol Matrix*

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Abstract

In this study, the efficiency of silver nanoparticles (AgNP) incorporated into electrospun natural rubber latex/polyvinyl alcohol (NRL/PVA) nanofibre matrix to remove Cd²⁺ from aqueous solution was examined. Electrospun AgNP-NRL/PVA nanofibre composites were produced using silver nitrate (AgNO₃) concentrations of 0.01 M and 0.015 M. Maximum Cd²⁺ was adsorbed at pH 7 for both nanofibre composites but at different reaction times of 20 min for 0.01 M AgNP and 40 min for 0.015 M AgNP. The maximum adsorption for 0.01 M AgNP was 14.9674 mg/g and 30.1129 mg/g for 0.015 M AgNP. Adsorption data were tested with Langmuir, Freundlich and Dubinin-Radushkevich (D-R) isotherms as well as pseudo-first order (PFO), pseudo-second order (PSO), Elovich and intra-particle diffusion kinetic models. Cadmium adsorption by both nanofibre composites fitted best to Freundlich isotherm. Adsorption data for 0.01 M AgNP fitted best to PFO kinetic model, whilst that of 0.015 M AgNP fitted best to Elovich kinetic model, with 0.015 M AgNP having a lower desorption constant and larger boundary layer ($\beta = 0.1793 \text{ g/mg}$; C = 7.5959 mg/g) than 0.01 M AgNP ($\beta = 1.0351 \text{ g/mg}$; C = 4.5942 mg/g). The nanofibre, 0.015 M AgNP, fitting best to Freundlich isotherm and Elovich kinetic model, showed that Cd2+ was chemisorbed. In general, 0.015 M AgNP was more efficient in adsorbing Cd2+ than 0.01 M AgNP. After the adsorption process, silver (Ag) concentration in the treated water (0.01 M AgNP = 0.0425 mg/L, 0.015 M AgNP = 0.0510 mg/L) was below US EPA and WHO guidelines of 0.10 mg/L for both nanofibre composites, rendering the treated water good enough for human use. Therefore, this work has shown the potential of AgNP incorporated in an electrospun NRL/PVA nanofibre matrix to remove contaminants (Cd²⁺) from contaminated water.

Keywords: Silver Nanoparticles, Cadmium Adsorption, Natural Rubber Latex, Polyvinyl Alcohol, Electrospun Nanofibre

1 Introduction

Cadmium (Cd²⁺) is a heavy metal with no known biological function to the human body but has detrimental health effects when cadmiumcontaminated water is consumed (Mahmood *et al.*, 2019). Due to Cd²⁺'s bioaccumulative nature, chronic exposure to low concentrations is considered harmful. Ingesting acute doses of Cd²⁺ can cause gastrointestinal tract erosion, hepatic dysfunction and kidney damage (Tchounwou *et al.*, 2012). Long-term exposure may lead to Itai-Itai disease, which is characterised by softening of the bones and severe bone pain (Aoshima, 2016).

Cadmium is released into water bodies through corrosion of galvanised water supply pipes, desorption from some plastic water supply pipes, municipal landfill leachates, runoff from the application of Cd-laden fertilisers and pesticides, and effluents from mining and smelting sites (Elkhatib et al., 2016; Salehi et al., 2017; Mahmood et al., 2019). Although humans are mostly exposed to Cd²⁺ through food consumption (Anon., 2011), Cd²⁺ in runoff and irrigation waters and aquatic bodies settle in soils and sediments (Aoshima, 2016; Elkhatib et al., 2016; Balali-Mood et al., 2021). Cadmium is assimilated by plants such as leafy vegetables, seeds, grains and potatoes, which is subsequently consumed by humans (Aoshima, 2012; Tchounwou et al., 2012; Aoshima, 2016).

This shows that Cd^{2+} ingested by humans can be limited mainly by reducing Cd-laden effluent release and Cd^{2+} concentration in irrigation water.

Treating contaminated water containing low concentrations of Cd^{2+} is challenging, particularly in countries with lax regulations on Cd^{2+} discharge. Cadmium can be removed from wastewater using techniques such as ion exchange and tight membrane filtration. However, these methods require large quantities of chemicals for resin regeneration, and since the membranes are prone to fouling, it leads to high operational costs (Colantonio and Kim, 2016). It is, therefore, not cost-effective to use these advanced methods to treat Cd^{2+} at low concentrations (Colantonio and Kim, 2016).

In recent years, nanoparticles have proven superior to their bulk counterparts in removing heavy metals from water (Shao-Feng *et al.*, 2005; Kango and Kumar, 2016; Tavker *et al.*, 2021). For instance, silver nanoparticles (AgNP) have the potential to adsorb Cd^{2+} efficiently from water (Al-Qahtani, 2017). However, a pressing challenge is that nanoparticles mostly remain in treated water after the removal of contaminants. Due to the toxicological nature of nanoparticles, having these nanoparticles in treated water is not advisable (Sahu and Hayes, 2017; Pietroiusti *et al.*, 2018). Furthermore, removing AgNP from the treated water will require additional investment, rendering the whole treatment process expensive (Roksana, 2016). Also, ingesting excess silver (Ag) (that is, AgNP in the treated water) can lead to argyria, which is the discolouration of the dermis and some organs (Han et al., 2011). In view of this, it is necessary to apply AgNP in water treatment when the nanoparticles are incorporated into a matrix. However, some matrices used are mostly micro or nanoparticles, which would need micro or ultrafiltration for final separation (Elkhatib et al., 2016; Tauanov et al., 2019). Given this, it would be plausible to incorporate AgNP into a matrix such as nanofibres since this material is large enough to be separated from treated water using conventional filtration.

The potential of AgNP in electrospun nanofibre matrices like natural rubber latex/polyvinyl alcohol (NRL/PVA) to adsorb metals such as Cd^{2+} has not been explored. Therefore, this study aims to investigate the efficiency of AgNP in NRL/PVA nanofibre (AgNP-NRL/PVA nanofibre composites) to sorb or remove Cd^{2+} from an aqueous solution. In the sorption experiments, the main parameters considered were pH, contact time and initial concentration of Cd^{2+} .

2 Resources and Methods Used

2.1 Materials

Electrospun NRL/PVA nanofibres prepared in previous studies were utilised (Osei et al., 2022). AgNP-NRL/PVA nanofibre composites were prepared by adding 0.01 M and 0.015 M silver nitrate (AgNO₃) solutions to two separate NRL/PVA solutions. These solutions were electrospun to form two nanofibre sheets (nanofibre composites). The nanofibre composites were named 0.01 M AgNP and 0.015 M AgNP, according to the AgNO₃ concentration used in their syntheses. Nanoparticles in the 0.01 M AgNP were about 2.35 AgNP per 10 000 nm² area with an average size of 8.15 ± 0.09 nm. The 0.015 M AgNP were about 23.66 AgNP per 10 000 nm² area with an average size of 2.82 ± 0.04 nm. Nanoparticles of the 0.01 M AgNP were mostly located near or on the surface of the fibres. On the other hand, nanoparticles of the 0.015 M AgNP were mainly located within the fibre walls. Hydrochloric acid (HCl; 25.0%), nitric acid (HNO; 68.0%) and sodium hydroxide (NaOH; > 97%) were purchased from Merck, Brenntag Ghana Ltd. and Paskem Fine Chem Pvt. Ltd., respectively. Cadmium solutions used for adsorption studies were prepared from 1 000 mg/L Cd²⁺ standard solution purchased from pg Instruments.

2.2 Methods Used

2.2.1 Adsorption Study

At an ambient temperature of 27 °C, 5×5 cm² of 0.01 M and 0.015 M AgNP were placed in separate 50 mL Cd²⁺ solutions to determine how efficiently the nanofibre composites would adsorb Cd²⁺. The solutions were shaken at 200 rpm on an orbital shaker. Contact time, pH and initial concentration were varied between $0 - 120 \min 4 - 7$ and $0.01 - 120 \min 4 - 7$ 1.00 ppm, respectively. After adsorption experiments, Cd²⁺ concentrations in solution were determined an atomic absorption using spectrophotometer (Shimadzu AA-7000). Removal efficiency and adsorption capacity of 0.01 M and 0.015 M AgNP were determined using Equations (1) and (2), respectively.

$$\operatorname{RE}(\%) = \frac{C_{o} - C_{e}}{C_{o}} \times 100 \quad (1)$$
$$q_{e}(\operatorname{mg/g}) = \frac{(C_{o} - C_{e}) \cdot V}{m} \quad (2)$$

where $C_o (mg/L) =$ initial adsorbate concentration in solution, $C_e (mg/L) =$ adsorbate concentration at equilibrium, V (L) = volume of adsorbate solution, and m (g) = mass of the AgNP in nanofibre composite used. Per calculations, 5×5 cm² (of 549.1504 cm², the total area electrospun for each nanofibre composite) of 0.01 M and 0.015 M AgNP nanofibre composites contain ~0.3929 mg and ~0.5893 mg of Ag, respectively.

2.2.2 Adsorption Isotherm

Data from the adsorption experiments were fitted to Langmuir, Freundlich and the Dubinin-Radushkevich (D-R) isotherms. This was done to understand the relationship between the adsorbent and the adsorbate in the solution (Sawyer et al., 2003; Tran et al., 2017). In the Langmuir isotherm, presented in Equation (3), all active sites of the adsorbent are assumed to have the same affinity for the adsorbate. The model is associated with a separation factor (R_L) which determines whether the adsorption is favourable or not (Equation 4) (Langmuir, 1918; Tran et al., 2017).

$$q_e = \frac{q_{max} + (K_L \cdot C_e)}{1 + (K_L \cdot C_e)}$$
(3)

$$R_{\rm L} = \frac{1}{1 + (K_{\rm L} \cdot C_{\rm o})} \tag{4}$$

where q_{max} (mg/g) = the adsorbent's maximum capacity, and K_L (L/mg) = Langmuir constant. The adsorption is favourable if $0 < R_L < 1$ and unfavourable if $R_L > 1$. The adsorption may also be linear if $R_L = 1$ or irreversible if $R_L = 0$.

On the other hand, Freundlich isotherm, proposed by Freundlich (1906), assumes that the active sites of an adsorbent have different affinities for different adsorbates (Sawyer *et al.*, 2003). This is presented in Equation (5).

$$q_e = K_F \cdot C_e^{\frac{1}{n}}$$
(5)

where $K_F (mg/g)/(mg/L)^{1/n}$ = Freundlich constant, and n = change in adsorption intensity with changes in the adsorption density. Depending on the value of 1/n, the adsorption may be favourable (1/n< 1) or unfavourable (1/n > 1)

The D-R isotherm is mainly applied to porous adsorbents. Since the matrix was porous, the D-R isotherm was employed to obtain additional information on the adsorption process. The D-R isotherm, as proposed by Dubinin and Radushkevich (1947), is presented in Equation (6), and its adsorption energy, E (kJ/mol), is also shown in Equation (7).

$$q_e = q_{DR} \cdot e^{(-K_{DR} \cdot \varepsilon^2)}$$
(6)

$$E = \frac{1}{\sqrt{2 \cdot K_{DR}}}$$
(7)

where q_{DR} (mg/g) = adsorption capacity, K_{DR} (mol²/kJ²) = constant related to sorption energy and ϵ (kJ/mol) = the Polanyi potential. Polanyi potential is computed as shown in Equation (8).

$$\varepsilon = \mathrm{RT} \cdot \ln(1 + \frac{\mathrm{c}^o}{\mathrm{c}_{\mathrm{e}}}) \tag{8}$$

where R (8.314 \times 10⁻³ kJ/mol·K) = universal gas constant, T (K) = temperature for adsorption and C^o (mg/g) = concentration of the adsorbate in standard state (1 mol/dm³) (Zhou, 2020). At standard state, 1 mol/dm³ = 112 411 mg/L for Cd²⁺.

2.2.3 Adsorption Kinetics

Adsorption data were fitted to the pseudo-first order (PFO), pseudo-second order (PSO), Elovich and intra-particle diffusion kinetic models to elucidate the adsorption mechanisms and the rate of reaction. The PFO model is expressed as shown in Equation (9) (Lagergren, 1898).

$$q_t = q_e \cdot (1 - e^{-k_1 \cdot t}) \tag{9}$$

where $q_t (mg/g) =$ amount of adsorbate adsorbed at time t (min), and $k_1 (1/min) =$ PFO equation's rate constant. Mostly, the PFO model can accurately describe the first 20 to 30 min of contact time and not beyond (Ho and McKay, 1998). The PSO kinetic model, which is also widely applied to adsorption data, is presented in Equation (10) (Blanchard *et al.*, 1984).

$$q_{t} = \frac{k_{2}(q_{e})^{2}t}{1+(k_{2}q_{e}t)}$$
(10)

where $k_2 (g/(mg \times min)) = PSO$ equation's rate constant.

The Elovich kinetic model, proposed by Roginsky and Zeldovich (1934), is mainly applied to chemical adsorption data (Tran *et al.*, 2017). As expressed in Equation (11), the Elovich kinetic model provides insight into the desorption constant of the adsorption (β ; g/mg) and the initial rate constant (α ; mg/(g × min)). The model is as follows:

$$q_t = \frac{1}{\beta} ln(1 + \alpha\beta t) \tag{11}$$

Finally, the intra-particle diffusion model, proposed by Weber and Morris (1963), can predict the reaction pathway and adsorption mechanism (Tran *et al.*, 2017). The model is presented in Equation (12).

$$q_t = k_p \sqrt{t} + C \tag{12}$$

where $k_p (mg/(g \times min^{1/2})) =$ diffusion rate constant, and C (mg/g) = constant associated to the boundary layer's thickness.

2.2.4 Metal Concentration in Solution

Before and after adsorption studies, Cd^{2+} concentration was determined using the Flame AAS (FAAS; Shimadzu AA 7000). Five per cent HNO₃ (7.54 M) was added to the Cd^{2+} solution prior to FAAS analysis, according to APHA 3111B standard method. This method was also applied in analysing Ag concentration after treating the contaminated water with the nanofibre composites.

2.2.5 Morphology and Elemental Composition of the Nanofibre Composite After Adsorption

Morphology of the nanofibre composites was examined using a scanning electron microscope (SEM; Zeiss EVO MA 15) before and after adsorption. The SEM was equipped with an energy dispersive x-ray spectrometer (EDS; Bruker) which was used to determine if the nanocomposite had truly adsorbed Cd²⁺ after the adsorption process.

3 Results and Discussion

3.1 Effect of pH

A solution's pH can significantly affect the surface chemistry of adsorbate and adsorbent, hence,

affecting adsorption. Fig. 1 shows how pH affects Cd²⁺ adsorption by the AgNP-NRL/PVA nanofibre composites. The initial concentration was maintained at 1.00 ppm; contact time was maintained at 20 min and 40 min for 0.01 M and 0.015 M AgNP, respectively, whilst pH was varied between 4 - 9. From Fig. 1, it can be seen that the amount of Cd²⁺ removed by 0.01 M AgNP gradually increased from 4.41 mg/g, representing 3.33%, at pH 4 to 39.82 mg/g, representing 30.04%, at pH 7. After pH 7, the amount of Cd²⁺ removed reduced from 39.82 mg/g to below detection at pH 9. For 0.015 M AgNP, a similar trend was recorded: the amount of Cd²⁺ removed increased from 2.94 mg/g, representing 3.33%, at pH 4 to 29.49 mg/g, representing 33.67%, at pH 7. After this, there was a gradual decrease in Cd²⁺ adsorbed from 29.49 mg/g (33.67%) at pH 7 to 3.83 mg/g, representing 4.32%, at pH 9.

The observed trend was similar to that of other researchers, such as Jiang *et al.* (2006), where maximum amount of Cd^{2+} removed from an aqueous solution occurred at pH 7. On the other hand, in a study by Al-Qahtani (2017) where AgNP synthesised using *Benjamina* leaf extracts was used to adsorb Cd^{2+} from solution, the maximum uptake of Cd^{2+} occurred at pH 6, with a decrease in uptake recorded at pH 7. Observations in this study contradicts that of Al-Qahtani (2017), with maximum uptake occurring at pH 7, whilst a sharp decrease in uptake was observed at pH 8.

It is worth mentioning that according to the Pourbaix diagrams of Takeno (2005), at pH 7 and Eh of 0.22 V (SHE), Cd exists as Cd^{2+} . This explains why relatively lower adsorption of Cd^{2+} was recorded at lower pH, such as 4 and 5. At lower pH, cation adsorption is limited because H⁺ is more readily and directly adsorbed onto surfaces than other cations (Drever, 1997).

On the other hand, at around pH 7, H^+ in the solution becomes limited, enabling Cd^{2+} to bond freely to the adsorbent. Also, according to Henglein (1998), Cd^{2+} adsorption could be ascribed to the cathodic charging of AgNP, which causes Cd^{2+} to reduce on the AgNP surface (Equation 13).

 $Ag_x^{(y+2)-} + Cd^{2+} \rightarrow (Ag_xCd)^{y-}$ (13)

where x = number of Ag clusters and y = moles of Ag⁺ in the AgNP synthesis reaction (yAg⁺). The AgNP can serve as a cathode for deposition due to its higher standard electrode potential than Cd (Ag⁺/Ag^o (0.80 V); Cd²⁺/Cd^o (-0.40 V)).

However, OH^- ions may become abundant when an alkali region is formed, that is, pH beyond 7. Hence Cd^{2+} may readily form $Cd(OH)^+_{(aq)}$ and $Cd(OH)_{2 (aq)}$,

therefore limiting Cd^{2+} adsorption (Equations (14) and (15)). This probably caused the decrease in per cent of Cd^{2+} adsorbed at pH beyond 7.

$$Cd^{2+}_{(aq)} + OH^{-}_{(aq)} \rightarrow Cd(OH)^{+}_{(aq)}$$
(14)

$$Cd(OH)^{+}_{(aq)} + OH^{-}_{(aq)} \rightarrow Cd(OH)_{2(aq)}$$
(15)



Fig. 1 Effect of pH on Cd²⁺ Adsorption by AgNP-NRL/PVA Nanofibre Composites

3.2 Effect of Contact Time

How long an adsorbate solution remains in contact with an adsorbent determines the optimum adsorption efficiency. At a pH of 7 and an initial concentration of 1.00 ppm, the effect of contact time on Cd^{2+} adsorption was studied, ranging from 0 – 120 min. Fig. 2 presents the effect of contact time on Cd^{2+} adsorption by NRL/PVA only and AgNP-NRL/PVA nanofibre composites.

By comparing the removal efficiency of NRL/PVA only, which does not contain AgNP, to that of 0.01 M and 0.015 M AgNP, it can be seen that NRL/PVA adsorbed a relatively smaller amount of Cd²⁺. This signifies that the AgNP in AgNP-NRL/PVA nanofibre composites improved Cd2+ removal from the aqueous solution. For 0.01 M AgNP, maximum amount of Cd²⁺ was removed at 20 min, representing 14.9674 mg/g (11.90%), which later decreased with an increase in uptake at 120 min. On the other hand, the maximum uptake of Cd²⁺ for 0.015 M AgNP occurred at 40 min, representing 30.1129 mg/g (35.92%), with a slight drop in uptake at 60 min. The maximum uptake recorded at 40 min for 0.015 M AgNP was similar to that recorded by Al-Qahtani (2017). However, Jiang et al. (2006) observed the maximum uptake of Cd^{2+} by tourmaline at 60 min.

From Fig. 2, it can be seen that the adsorption of Cd^{2+} by 0.015 M AgNP was relatively better than 0.01 M AgNP. This could be attributed to the fact that there are more AgNP in 0.015 M than in 0.01 M AgNP; hence the availability of relatively more active sites in 0.015 M AgNP for Cd^{2+} adsorption.

Also, from Equation (13), it can be seen that the higher the moles of Ag^+ (yAg^+), the higher the charge of the reduced complex formed ($(Ag_xCd)^{y_-}$). It is also worth mentioning that the higher the magnitude of the charge of the reduced complex, the stronger its attraction will be. Hence, better adsorption was observed in 0.015 M AgNP than in 0.01 M AgNP. The relatively smaller amount of active sites in 0.01 M AgNP probably resulted in the desorption and readsorption of Cd²⁺ after 20 min and at 120 min, respectively.

3.3 Effect of Initial Concentration

How concentration of Cd²⁺ in an aqueous solution affects its adsorption by AgNP-NRL/PVA nanofibre composites was studied. Whilst pH was held constant at 7, time at 20 min for 0.01 M AgNP and 40 min for 0.015 M AgNP, the initial concentration of Cd²⁺ was varied between 0.00 – 1.00 ppm. Results on how initial Cd²⁺ concentration affects adsorption by the nanofibre composites are presented in Fig. 3. It can be seen that maximum uptake occurred at 0.05 ppm (93.40%), with a minor drop in uptake at 0.10 ppm (93.30%) for Cd^{2+} adsorption by 0.01 M AgNP. One of the highest percentages of Cd²⁺ removed by 0.015 M AgNP occurred at 0.10 ppm (93.80%), similar to Cd²⁺ adsorption by 0.01 M AgNP. A significant drop in Cd²⁺ adsorbed by both nanofibre composites was recorded at 0.20 ppm (0.01 M AgNP = 27.00%; 0.015 M AgNP = 28.50%).



Fig. 2 Effect of Contact Time on Cd²⁺ Adsorption by NRL/PVA and AgNP-NRL/PVA Nanofibre Composites

This was followed by a gradual decrease in the percentage of Cd^{2+} removed until 1.00 ppm (0.01 M AgNP = 19.68%; 0.015 M AgNP = 19.68%). The gradual decrease in Cd^{2+} adsorbed by both nanofibre composites was due to the relatively smaller number of active sites available for adsorption as the initial concentration increased.



Fig. 3 Effect of Initial Concentration on Cd²⁺ Adsorption by AgNP-NRL/PVA Nanofibre Composites

3.4 Adsorption Isotherm

Fig. 4 presents the Langmuir, Freundlich and D-R isotherms for Cd²⁺ adsorption by 0.01 M and 0.015 M AgNP. The experimental data were fitted to the aforementioned models to understand the adsorption behaviour between Cd²⁺ on the surfaces of the nanofibre composites. Table 1 shows the adsorption isotherm model parameters as well as their respective coefficient of determination (R²) and mean square error (MSE) for Cd²⁺ adsorption by 0.01 M and 0.015 M AgNP. The adsorption data for both nanofibre composites were a weak fit to Langmuir isotherm. However, for both nanofibre composites, the experimental data were a good fit to D-R isotherm but the best fit to the Freundlich isotherm with 0.01 M AgNP having an R² and MSE values of 0.7413 and 10.5678 $(mg/g)^2$, respectively. The nanofibre composite, 0.015 M AgNP, had R^2 and MSE values of 0.8066 and 3.6705 $(mg/g)^2$, respectively.

The adsorption data fitting best to the Freundlich isotherm means the adsorbates formed a multi-layer on the surface of the adsorbent. Since 1/n values for 0.01 M (0.2245) and 0.015 M AgNP (0.2315) were greater than 0 but less than 1, it showed that the adsorption was favourable. As expressed in Equation (13), it is possible that as Cd^{2+} bonded with Ag to form an alloy, (Ag_xCd)^{y-}, and the negatively charged alloy promoted further adsorption of Cd²⁺ at the surface, hence the multilayered adsorption. Also, since in Freundlich isotherm, it is assumed that active sites available for adsorption have different affinities for the adsorbate, AgNP within the fibre may have contributed to the varied affinities across the surface of the nanofibre composites, resulting in Cd²⁺ multilayered adsorption. Though in general, 0.015 M AgNP adsorbed Cd²⁺ better than 0.01 M AgNP, the higher q_{DR} value of 0.01 M AgNP (47.7059 mg/g) than 0.015 M AgNP (35.4335 mg/g) means 0.01 M AgNP could have adsorbed more Cd²⁺ if the experiment's time frame had been

increased. The parameter E, which represents the adsorption energy, and may suggest whether the adsorption was physical or chemical was 20.8791 kJ/mol for 0.01 M AgNP and 20.3996 kJ/mol for 0.015 M AgNP. It is worth mentioning that the E values of both nanofibre composites were beyond the range for physical (< 8 kJ/mol) and chemical adsorption (8 – 16 kJ/mol) (Das *et al.*, 2013; Maneechakr and Mongkollertlop, 2020). Therefore, further studies may be required to understand the adsorption behaviour better.





Fig. 4 (a) Langmuir (b) Freundlich and (c) Dubinin-Radushkevich Isotherm for the Adsorption of Cd²⁺ by 0.01 M and 0.015 M AgNP

Table 1 Parameters of Adsorption Isotherms for the Adsorption of Cd²⁺ by 0.01 M and 0.015 M AgNP

Adsorbent	Langmuir parameters				MSE	Freundlich parameters			MSE
	q _{max} (mg/g)	K _L (L/mg)	- R L	R ²	[(mg/g) ²]	1/n	$\frac{K_F}{[(mg/g)/(mg/L)^1}$	\mathbf{R}^2	[(mg/g) ²]
0.01 M AgNP	28.9897	3.0981	0.2440- 0.9700	0.4483	22.5407	0.2245	19.2994	0.7413	10.5678
0.015 M AgNP	16.1689	5.9596	0.1437- 0.9438	0.5187	9.1348	0.2315	13.6678	0.8066	3.6705
A Jacob ou 4	D-R parameters					D ²			
Adsorbent	$q_{DR}(mg/g)$ K_{L}		DR (mol ² /kJ ²)		E (kJ/mol)		K-	$MSE[(mg/g)^{-}]$	
0.01 M AgNP	47.7059		0.0011		20.8791		0.7238	11.	2840
0.015 M AgNP	35.4335		0.0012		20.3996		0.7904	3.9789	

3.5 Adsorption Kinetics

The adsorption data were fitted to the PFO, PSO, Elovich and intra-particle diffusion models to understand the reaction rate and adsorption mechanism. Fig. 5 shows the plot for the four adsorption kinetic models, and Table 2 shows the parameters of the kinetic models. The experimental data for Cd²⁺ adsorption by 0.01 M AgNP was a weak fit to the PFO ($R^2 = 0.5384$; MSE = 7.8203 (mg/g)²), PSO ($R^2 = 0.5349$; MSE = 7.8790 (mg/g)²) and Elovich ($R^2 = 0.5338$; MSE = 7.8978 (mg/g)²) kinetic models and a poor fit to the intra-particle diffusion model ($R^2 = 0.3333$; MSE = 11.2950 (mg/g)²). Although Cd²⁺ adsorption by 0.01 M AgNP fitted best to the PFO kinetic model, which

represents physical adsorption, its experimental data fitted best to Freundlich isotherm, which represents chemical adsorption. The discrepancy observed was likely due to the weak fit of the experimental data to the PFO model, hence not adequately describing the type of adsorption taking place. The poor and weak fit of Cd²⁺ adsorption by 0.01 M AgNP data to all four kinetic models used in this study might have been due to the desorption of Cd²⁺ recorded after 20 min and subsequent readsorption at 120 min. This implies that the experiment should have been cut short at 20 min or extended beyond 120 min to obtain more precise information about the adsorption kinetics. Also, Tran et al. (2017) and Revellame et al. (2020) explains that stopping an experiment before reaching equilibrium or including experimental data after reaching equilibrium can result in wrong inferences.

For Cd²⁺ adsorption using 0.015 M AgNP, the experimental data were a good fit to the PFO ($R^2 =$ 0.9214; MSE = 6.2582 (mg/g)²), PSO (R² = 0.9443; $MSE = 4.4371 \text{ (mg/g)}^2$), and intra-particle diffusion $(R^2 = 0.8006; MSE = 15.8823 (mg/g)^2)$ kinetic models. The best fit was for the Elovich kinetic model ($R^2 = 0.9461$; MSE = 4.2957 (mg/g)²). Since Freundlich isotherm and Elovich kinetic model best describes Cd2+ adsorption by 0.015 M AgNP, it could be that chemisorption might have occurred during the adsorption process. The Elovich model also showed that the initial rate of adsorption of Cd²⁺ by 0.015 M AgNP ($\alpha = 12.8106 \text{ mg/(g \times min)}$) was slower than that of 0.01 M AgNP ($\alpha = 679.0428$ $mg/(g \times min)$). However, the desorption constant showed that a relatively smaller amount of Cd²⁺ will desorb from 0.015 M AgNP ($\beta = 0.1793$ g/mg) than from 0.01 M AgNP ($\beta = 1.0351$ g/mg) within the experiment's time frame. Also, from the intraparticle diffusion model, 0.015 M AgNP (C = 7.5959 mg/g) had a relatively larger boundary layer for adsorption than 0.01 M AgNP (C = 4.5942 mg/g). From this, it can be inferred that the presence of relatively more AgNP in 0.015 M AgNP (due to the relatively higher yAg^+) resulted in the larger boundary layer and lower desorption constant; hence, the better adsorption observed in 0.015 M AgNP.

3.6 Elemental Composition of AgNP-NRL/PVA After Adsorption

Fig. 6 shows an SEM image of AgNP-NRL/PVA nanofibre composite and its corresponding EDS spectra before Cd^{2+} adsorption. The whitish specks in Fig. 6a are the AgNP clusters on the nanofibre composite. Its EDS spectra (Fig. 6b) confirm these white specks as AgNP clusters. Fig. 7 shows an SEM image of AgNP-NRL/PVA nanofibre composite and its corresponding EDS spectra after Cd^{2+} adsorption.



Fig. 5 (a) Pseudo-First Order (b) Pseudo-Second Order (c) Elovich and (d) Intra-Particle Diffusion Kinetic Models for the Adsorption of Cd²⁺ by 0.01 M and 0.015 M AgNP

Madal	Danamatang	Adsorbent			
Model	Farameters	0.01 M AgNP	0.015 M AgNP		
DEO	$q_e (mg/g)$	10.1604	28.5447		
IIU	k ₁ (1/ min)	0.2074	0.0831		
	R ²	0.5384	0.9214		
MSE	$E\left[(mg/g)^2\right]$	7.8203	6.2582		
DSO	$q_e (mg/g)$	10.6641	31.9701		
130	$k_2 [g/(mg \times min)]$	0.0416	0.0037		
	R ²	0.5349	0.9443		
MSE	$E\left[(mg/g)^2\right]$	7.8790	4.4371		
Flowich	$\alpha [mg/(g \times min)]$	679.0428	12.8106		
Elovicii	β (g/mg)	1.0351	0.1793		
	R ²	0.5338	0.9461		
MSI	$E\left[(mg/g)^2\right]$	7.8978	4.2957		
Intro portiolo differsion	$k_p [mg/(g \times min^{1/2})]$	0.7467	2.5093		
Intra-particle diffusion	C (mg/g)	4.5942	7.5959		
	R^2	0.3333	0.8006		
MSE	$E\left[(mg/g)^2\right]$	11.2950	15.8823		

(a)

Table 2 Parameters of Adsor	ption Kinetics for the Adsorr	otion of Cd ²⁺ by 0.01 N	A and 0.015 M AgNP

(a)







Similar to Fig. 6a, the whitish specks in Fig. 7a also represent AgNP clusters. From Fig. 7a, it can be seen that upon removal of the nanofibre composite from the aluminium foil (substrate for collecting nanofibres during electrospinning) and when in contact with water, the nanofibres collapse and clamp together, forming larger and fewer pores as seen in Fig. 7a (Osei et al., 2022).









The EDS analysis of the red circled area in Fig 7a is shown in Fig. 7b. It can be seen from the EDS spectra that Cd²⁺ is present where AgNP clusters are. This shows that the nanoparticles had indeed removed Cd²⁺ from the aqueous solution. The presence of aluminium in the spectra was likely due to fragments of the aluminium foil sticking to the nanofibre composite during its removal (Osei et al., 2022) because aluminium foil was used to support the matrix nanocomposite during electrospinning.

3.7 Silver (Ag) Released into Solution After Water Treatment

After the adsorption study, it was necessary to determine the amount of Ag released into solution. Since AgNP was synthesised to remove Cd²⁺ from contaminated water, it would not be ideal for excess Ag to be released into the water and become part of the contamination problem. The amount of Ag released into solution after Cd²⁺ adsorption by the AgNP-NRL/PVA and guidelines values for Ag in drinking water are presented in Tables 3 and 4, respectively. According to both the US Environmental Protection Agency (US EPA) and the World Health Organisation (WHO), Ag in drinking water should not exceed 0.10 mg/L (Table 4). Table 3 shows that 0.01 M and 0.015 M AgNP released 0.0425 mg/L and 0.0510 mg/L of Ag into the solution, which is less than the US EPA and WHO guidelines. This means Ag released into the water would not pose detrimental effect upon human usage.

 Table 3 Ag Concentration in Solution After Cd²⁺

 Adsorption

Fibre Matrix (AgNP)	Ag Concentration (mg/L)
0.01 M	0.0425
0.015 M	0.0510

 Table 4 Guidelines for Ag Concentration in Drinking Water

Guideline/Standa rd	Ag Concentratio n (<i>mg/L</i>)	Referenc e
US EPA SMCL*	0.1000	(Anon.,
		2013)
WHO-drinking	0.1000	(Anon.,
water		2011)

*Secondary Maximum Contaminant Level

Since the matrix for AgNP synthesis, that is, NRL/PVA, is partially soluble, the surfaces of the fibres bearing AgNP might have dissolved into the solution. Hence, accounting for the Ag concentrations recorded. It is also worth mentioning that Ag released by 0.015 M AgNP was greater than that of 0.01 M AgNP due to the higher molar ratio of Ag in 0.015 M AgNP.

4 Conclusion

The efficiency of AgNP in electrospun NRL/PVA nanofibre matrix (0.01 M and 0.015 M AgNP) to remove Cd^{2+} from contaminated water has been

investigated. Factors such as pH, contact time, and initial Cd²⁺ concentration that play a critical role in sorption processes were considered. For both adsorbents, the maximum uptake of Cd²⁺ occurred at pH 7 and around 20 min for 0.01 M AgNP (14.9674 mg/g; 11.90%) and 40 min for 0.015 M AgNP (30.1129 mg/g; 35.92%). As the initial Cd²⁺ concentration increased, the nanofibre composites' removal efficiency decreased, and the adsorption probably resulted in Cd^{2+} forming an alloy with Ag. The experimental data for Cd^{2+} adsorption by 0.01 M and 0.015 M AgNP fitted best to Freundlich isotherm, implying that the adsorption occurred in a multilavered fashion. For the adsorption kinetics of Cd²⁺, the experimental data for adsorption by 0.01 M AgNP was poor and weak fit to all kinetic models utilised in this study. This showed that the time range selected was inappropriate for Cd²⁺ adsorption by 0.01 M AgNP. Hence, the experiment must be stopped at 20 min or extended beyond 120 min to understand its kinetics better.

On the other hand, the adsorption data of Cd^{2+} by 0.015 M AgNP was a good fit to the Elovich kinetic model, showing that Cd²⁺ was chemically adsorbed. In general, 0.015 M AgNP adsorbed Cd²⁺ better than 0.01 M AgNP, and this was plausible because of its lower desorption constant ($\beta = 0$.1793 g/mg) and larger boundary layer (C = 7.5959 mg/g) than 0.01 M AgNP ($\beta = 1.0351$ g/mg; C = 4.5942 mg/g). The better efficiency of 0.015 M AgNP was mainly due to relatively higher number of AgNP present in the nanofibre for adsorption as well as higher molar ratio of Ag⁺ in 0.015 M AgNP during its synthesis. SEM and EDS analysis confirmed that the nanofibre composite had adsorbed Cd2+ from the aqueous solution. After the adsorption process, the concentration of Ag released into the treated water was below US EPA and WHO limits of 0.10 mg/L for both 0.01 M AgNP (0.0425 mg/L) and 0.015 M AgNP (0.0510 mg/L), which renders the water safe in terms of Ag contamination.

Utilising these nanofibre composites as an adsorbent for Cd^{2+} removal in contaminated water is promising since the methodology is simple, and the materials used are readily available locally. Also, since the concentration of Ag released into the treated water is not detrimental to health, no further treatment process may be required to reduce or eliminate Ag in this water. Therefore, further research would be necessary in order to enhance the efficiency of the AgNP-NRL/PVA in adsorbing Cd^{2+} from an aqueous solution since this adsorbent looks promising.

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