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A Multi-drug resistance pattern of a *Leclercia adecarboxylata* strain isolated from a urinary tract infection of a patient at China-Guinea friendship hospital of Kipé/Conakry

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ABSTRACT

Leclercia adecarboxylata (LAD) is a member of Enterobacteriaceae family that is usually reported as an opportunistic human pathogen. A few reports have described resistant strains in the literature. The aim of this paper was to describe the antimicrobial resistance pattern of a LAD strain isolated from a urinary tract infection in a 39-year-old immunocompetent man. The bacterial identification and antibiotic sensitivity tests were performed on Vitek 2 Compact 15. The results revealed the presence of LAD with a particular multidrug resistance pattern. It was sensitive only to imipenem (=1 μ g/ml), and totally resistant to association of trimethoprim/sulfamethoxazole (\geq 320 μ g/ml), ticarcillin (\geq 128 μ g/ml), nitrofurantoin (=128 μ g/ml), cefalothin (\geq 64 μ g/ml), cefoxitin (\geq 64 μ g/ml), cefotaxime (\geq 64 μ g/ml), ceftazidime (\geq 64 μ g/ml), amikacin (\geq 64 μ g/ml), approximation (\geq 16 μ g/ml), nalidxic acid (\geq 32 μ g/ml), and a combination of amoxicillin/clavulanic acid (\geq 32 μ g/ml), gentamicin (\geq 16 μ g/ml), tobramycin (\geq 16 μ g/ml), ofloxacin (\geq 8 μ g/ml), and ciprofloxacin (\geq 4 μ g/ml). It showed the intermediate sensitivity to the association of piperacillin/tazobactam (=64 μ g/ml), and ertapenem (=4 μ g/ml). The findings showed that this isolate of LAD had a multidrug resistance pattern to almost all the antibiotics tested (except imipenem). This suggests that LAD could be considered as an emergent bacterial pathogen capable of causing infections in human and carrying multidrug resistance pattern to numerous antibiotic families in Guinea. © 2018 International Formulae Group. All rights reserved.

Keywords: Leclercia adecarboxylata, multi-drug, resistance, Kipé/Conakry

INTRODUCTION

Leclercia adecarboxylata (LAD) is an opportunistic human pathogen which was designated initially as "Enteric group 41" or Escherichia adecarboxylata in 1962 by Leclerc (Kashani et al., 2014; Grantham et al., 2015). Advances of diagnostic technology particularly

in the molecular biology fields allowed to separate *Escherichia adecarboxylata* from the "*Enterobacter agglomerans*" complex and to name it as *LAD* (De Mauri et al., 2013; Kashani et al., 2014; Hurley et al., 2015).

LAD is a motile Gram negative rod belonging to the *Enterobacteriacea* family.

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The species are facultative anaerobic bacteria, oxydase negative, peritrichously flagellated bacilli (Stock et al., 2004). This organism shares many biomedical features with *Escherichia coli*.

Leclerc et al. first reported the isolation of *LAD* from drinking water in 1962 (Grantham et al., 2015). Further reports showed that it is widely distributed in nature (food, water), a part of normal flora in the gut of animals and in the stools of humans (Keren et al., 2014; Tam and Nay, 2012; Stock et al., 2004). It has also been reported in various clinical samples including blood, urine, sputum, wounds pus, synovial fluid, peritoneal fluid, gallbladder tissue, and cardiac valve tissue (Forrester et al., 2012; Tam and Nayaka, 2012; Dalamaga et al., 2015; Jean et al., 2016). *LAD* is capable to cause infection in immunocompetent human (Hess et al., 2008).

The aim of this study was to report the first isolation of *LAD* in Guinea from a urinary tract infection in humans and determine the multi-drug resistance pattern of this emerging bacterial pathogen.

MATERIALS AND METHODS

A 39 year-old-male patient admitted for hypertension in the Neurology department of China-Guinea Friendship hospital of Kipé (Conakry/Guinea) on October 13th 2016. The patient was suffering from urinary tract infection. Some biological parameters of the patient were investigated. The HIV immunoserological test performed using both HIV Combo (Alere Medical Co., Ltd, Mtsudo-shi, Japan) and Enzyme Linked Immunosorbent Assay (ELISA) on miniVidas (bioMérieux, Marcy l'Etoile, France).

The urine cultures were done on agar plates of different media: Nutrient agar

(Liofilchem, Italy), blood agar (Liofilchem, Italy), and CLED (bioMérieux, Narcy, l'Etoile, France). After 24 hours incubation cultures yielded pure and large colonies of gram negative bacilii. Gram stained smears were observed on photonic binocular microscopy (Microscope XS-213, China). The bacterial identification and antibiotic susceptibility tests (antibiogram) as well as the determination of minimal inhibition concentrations (MIC) were processed on Vitek 2 Compact 15 system (bioMérieux, Narcy, 1'Etoile, France). The Vitek 2 GN card (bioMérieux, Narcy, l'Etoile, France) was used for bacterial identification and the Vitek 2 N 233 card (BioMérieux, Narcy, l'Etoile, France) was used for antibiogram and determination of MIC, according to manufacturer's instructions.

RESULTS

Our results showed that the patient was HIV negative. The bacterial isolate was identified by Vitek 2 Compact as *Leclercia adecarboxylata*. Culture on nutrient agar plate is shown in Figure 1.

The results of antibiotic sensitivity tests of *LAD* strain are shown in Table 1.

This isolate was resistant to nearly all antibiotic families tested. In fact, it was resistant all tested quinolones, aminoglycosides, folate pathway inhibitors (sulfamides) and nitrofurantoin. For betalactams, this strain was resistant to all cephalosporins tested and nearly to all penicillins, except the association piperacillin/Tazobactam, to which it showed intermediate sensitivity. It showed total sensitivity to imipenem (carbapenem) and the intermediate sensitivity to ertapenem.

Table 1: Antibiotic sensitivity of *Leclercia adecarboxylata* isolate with the minimal inhibition concentrations (MIC).

Antibiotics	Sensitivity	MIC	Antibiotics	Sensitivity	MIC
		(µg/ml)			(μg/ml)
Ampicillin	R	≥ 32	Imipenem	S	1
Amoxicillin/Cla-vulanic acid	R	≥ 32	Amikacin	R	≥64
Ticarcillin	R	≥128	Gentamicin	R	≥16
Piperacillin/Tazo-bactam	I	64	Tobramycin	R	≥16
Cefalotin	R	≥ 64	Nalidixic acid	R	≥ 32
Cefoxitin	R	≥ 64	Ciprofloxacin	R	≥ 4
Cefotaxime	R	≥ 64	Ofloxacin	R	≥ 8
Ceftazidime	R	≥ 64	Nitrofurantoin	R	≥ 128
Ertapenem	I	4	Trimethoprim/Sulfamethoxazole	R	≥ 320

MIC: minimal inhibition concentration; I: intermediate, R: resistant; S: sensitive;



Figure 1: Colonies of *Leclercia decrboxylta* isolated on nutrient plate agar medium (cultures were incubated for 24 hours at 37 °C).

DISCUSSION

Numerous reports have revealed that LAD is widely distributed in nature (food, water), and is a part of normal flora in the gut of animals and in the stools of humans (Keren et al., 2014; Tam et Nay, 2012; Stock et al., 2004). It has also been reported in the various clinical samples including blood, urine, sputum, wounds pus, synovial fluid, peritoneal fluid, gallbladder tissue, sublingual splinter, and cardiac valve tissue (Forrester et al., 2012; Tam and Nayak, 2012; Dalamaga et al., 2015, Jean et al., 2016). Some authors have reported that the epidemiological significance of Leclercia. adecarboxylata is not clear (De Mauri et al., 2013). The paucity of reports of human infection in the past may reflect misdiagnosis, as the organism shares many biochemical features with Escherichia coli, rather than a true incidence of human infection. Some reports showed that LAD has been isolated from patients with mixed microbial infection (Saccani et al., 2017), which raises questions concerning the organism's role in some of these infections. Moreover, some investigators reported that LAD has been isolated from polymicrobial cultures from immunocompetent patients and as a pure culture from immunocompromised patients, which suggests the dependence of this bacterium on co-flora to cause a disease (Tam and Nayak, 2012).

In the present study, we isolated *LAD* in a mono-microbial urinary tract infection in immunocompetent man instead polymicrobial infection. This is different from others observations (Saccani et al., 2017). Our data suggest that this isolate could be considered as an emergent pathogen. It is therefore of interest in that it might be considered to our knowledge as the first report in the literature describing the isolation of this organism in a pure culture of a urinary tract infection of an immunocompetent man. Numerous authors (Tam et Nayak, 2012; Hess et al., 2008; Grantham et al., 2015) have reported isolation of LAD in blood cultures of immunocompetent patients (Mazzariol et al., 2003), and in an abscess (Hess et al., 2008).

There are few published data in the literature dealing with antimicrobial susceptibility patterns of LAD. In all cases, only a small number of antibiotics (up to a maximum of 13) were tested. In addition, only a few data are available on the natural antimicrobial susceptibilities of LAD. It is generally sensitive to most antibiotics and only a few reports revealed some resistant isolates (Dalamaga et al., 2009; Tam and Nayak, 2012; Shin et al., 2012; Eiland et al., 2013; Anuradha, 2014; Allawh and Camp, 2015). In the present study, data on the antibiotic sensitivity tests showed that our LAD isolate was sensitive only to imipenem. It was totally resistant to ampicillin. the association clavulanic acid. amoxicillin/ ticarcillin, cefalotin, cefoxitin, cefotaxime, ceftazidime, amikacin, gentamicin, tobramycin, nalidixic acid, ciprofloxacin, ofloxacin, nitrofurantoin trimethoprim/sulfamethoxazole. and Finally, it showed the intermediate sensitivity association ertapenem and the piperacillin/tazobactam (Table 1).

The antimicrobial sensitivity tests revealed that this strain was resistant to nearly all antibiotic families tested. In fact, it was resistant to all tested quinolones, aminoglycosides, folate pathway inhibitors (sulfamides) and nitrofurantoin. For betalactams, this strain was resistant to all cephalosporins tested and to all penicillins, except the association piperacillin/tazobactam, to which it showed intermediate sensitivity. The only one total sensitivity was shown to imipenem (carbapenem) while intermediate sensitivity was shown to ertapenem. These data are very different from those of numerous authors Kashani et al., 2014; Hurley et al., 2015; Saccani et al. 2017). Indeed, in Germany, Stock and his colleagues conducted the first large study on natural antimicrobial susceptibility patterns and biochemical profiles of 101 LAD strains (Stock et al., 2004). Their data revealed that most of the strains tested were sensitive to all tested antimicrobial agents, apart from benzylpenicillin. Anuradha (2014) in his reports on LAD strains isolated in two case

reports showed that his strains were sensitive to nearly all antibiotics tested (Anuradha, 2014). Also, Kashani et al. (2014) reports on antimicrobial susceptibility tests of LAD strains showed that these were susceptible to all the antibiotics tested (ampicillin, cefazolin, ciprofloxacin, gentamicin, imipenem, levofloxacin, tobramycin, trimethoprim /sulfamethoxazole. More recently, Saccani et al. (2017)showed that Leclercia adecarboxylata has a broad sensitivity to the majority antibiotics (beta-lactams, quinolones, azithromycin, aminoglycosides, and tetracyclines). Mazzariol et al. (2003) have previously reported that their strain was sensitive to numerous antibiotics including amoxicillin/clavulanic acid. cefoxitin, gentamicin, ciprofloxacin (Mazzariol et al., 2003). Their isolate was susceptible to imipenem but resistant to cefotaxime and ceftazidime. This data is then very similar to ours. Shin et al. (2012) showed their LAD isolate was resistant only to carbapenems and but quinolones, it was sensitive aminoglycosides (amikacin, gentamicin, tobramycin), most beta-lactactams, including broad spectrum cephalosporins (cefotaxime cefixime) and and trimethoprime/sulfamethoxazole.

Considering these and other authors published data, there is evidence that Leclercia adecarboxylata strains are usually sensitive to numerous antimicrobial agents. Hurley et al. (2015) have reported a pan-drug sensitive isolate of LAD, except for intermediate sensitivity for piperacilline/tazobactam.. contrast, our data showed that the isolate of Leclercia adecarboxylata described in the present study was resistant to nearly all the antibiotics tested (15/16= 93,75%) except to imipenem. To our knowledge, this is the first report in the literature that described a Leclercia adecarboxylata strain isolated in pure monomicrobial culture from urinary tract infection in immunocompetent patient, and which was resistant to 93,75% of antibiotics tested belonging to different families. In fact, this multidrug-resistance pattern suggest, an obvious therapeutic difficulty for treatment of

LAD infections. Then, *LAD* seems becoming a microbial pathogen like other classical microbes in the world (Makanéra et al., 2003; Okorundu et al., 2013; Koanga Mogtomo et al., 2016; Abba et al., 2017; Makanéra et al., 2017; Tchapdie Ngassam et al., 2017).

Our findings revealed that *LAD* could be considered as an emerging bacterial threaten pathogen to human health, because of its multidrug-resistance pattern which includes numerous antibiotic families.

COMPETING INTERESTS

The authors declare that they have no competing interests.

AUTHORS' CONTRIBUTIONS

TD and AOB: collected urine samples; AM, MC, MC, DC, MAD: contributed in the laboratory analyses; AM and MLK contributed in the collection of bibliography and manuscript drafting. AM: the main investigator did the conception of the protocol and manuscript writing.

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