



Theoretical study of the structure and acidity of condensed tannin monomers

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ABSTRACT

Low level of polyphenols absorption is due among other things to the formation of irreversible complexes between non-hemic iron (Fe^{2+} , Fe^{3+}) and these polyphenols through their hydroxyl groups. Understanding of these complexes formation mechanisms led us to explore hydroxyl groups acidity of these polyphenols monomers (catechin, epicatechin, galocatechin and epigallocatechin) mainly found in food. Quantum chemistry modelling, more precisely the functional density theory (DFT) method, associated to 6-311G (d, p) base was used in gas phase. Spectroscopic, thermodynamic descriptors and geometric parameters resulting from calculations showed a slight stability of catechin compared to epicatechin and galocatechin compared to epigallocatechin. As for the acidity of these monomers hydroxyl groups, the results show that it decreases similarly as follows: $\text{O}_{4'} > \text{O}_5 > \text{O}_7 > \text{O}_5' > \text{O}_3$ for Catechin and epicatechin; $\text{O}_3' > \text{O}_5 > \text{O}_{4'} > \text{O}_7 > \text{O}_5' > \text{O}_3$ and $\text{O}_3' > \text{O}_{4'} > \text{O}_5 > \text{O}_7 > \text{O}_5' > \text{O}_3$ respectively for galocatechin and epigallocatechin.

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INTRODUCTION

Catechin, epicatechin, galocatechin and epigallocatechin are condensed tannins monomer found in abundance in human nutrition. They are found in fruits and vegetables but also in chocolate and wine. Their study got a great interest since they had not only anticancer properties but also cardiovascular effects (Rein et al., 2000; Martin and Andriantsitohaina, 2002; Raju et al., 2014). In addition, they are used in the prevention of ulcers and hormone-dependent diseases (Funatogawa et al., 2004; Ruggiero et al., 2006; Gerber and Berta, 2006; Lainé et al., 2007). Several scientific studies have shown that inorganic or non-hemic iron (Fe^{2+} , Fe^{3+})

inhibits intestinal absorption of condensed tannins from food by formation of insoluble and irreversible complexes. (Roberta et al., 2002; Seigo et al., 2002; Scalbert et al., 2002; Jeremy, 2003; Mourad et al., 2007). It's established that this phenomenon occurs between phenolic hydroxyls of tannins and inorganic iron (Tapiero H. et al., 2002; Manach C. et al., 2004). Any solution aimed at avoiding formation of these complexes in order to optimize absorption of tannins requires first, precise knowledge of mechanisms of action. However, process of complex formation between condensed tannins and inorganic iron is not yet clearly established. Knowing that hydroxyl groups of

tannins are the sites of non heminic iron binding, our objective during this work is to study preferential sites at these hydroxyl groups. This, by theoretical determination of these hydroxyl groups acidity of monomers (catechin, epicatechin, gallic acid and epigallocatechin). Theoretical calculations of quantum chemistry using functional density theory (DFT) method, associated to 6-311G (d,p) base allowed us to carry out this study.

MATERIALS AND METHODS

The Acer predator processor runs the calculations. Catechin, epicatechin, gallic acid and epigallocatechin molecules (Figure 1) were drawn by GaussView 5.0 software (Frisch et al. 2003). Calculations were made in gas phase using the functional density theory method (DFT) with functional hybrid B3lyp, associated to 6-311G (d,p) base, incorporated in the GAUSSIAN-03 program (Frisch et al., 2003). Initial geometries have been optimised. Gibbs free energies (G) are obtained from frequencies calculation.

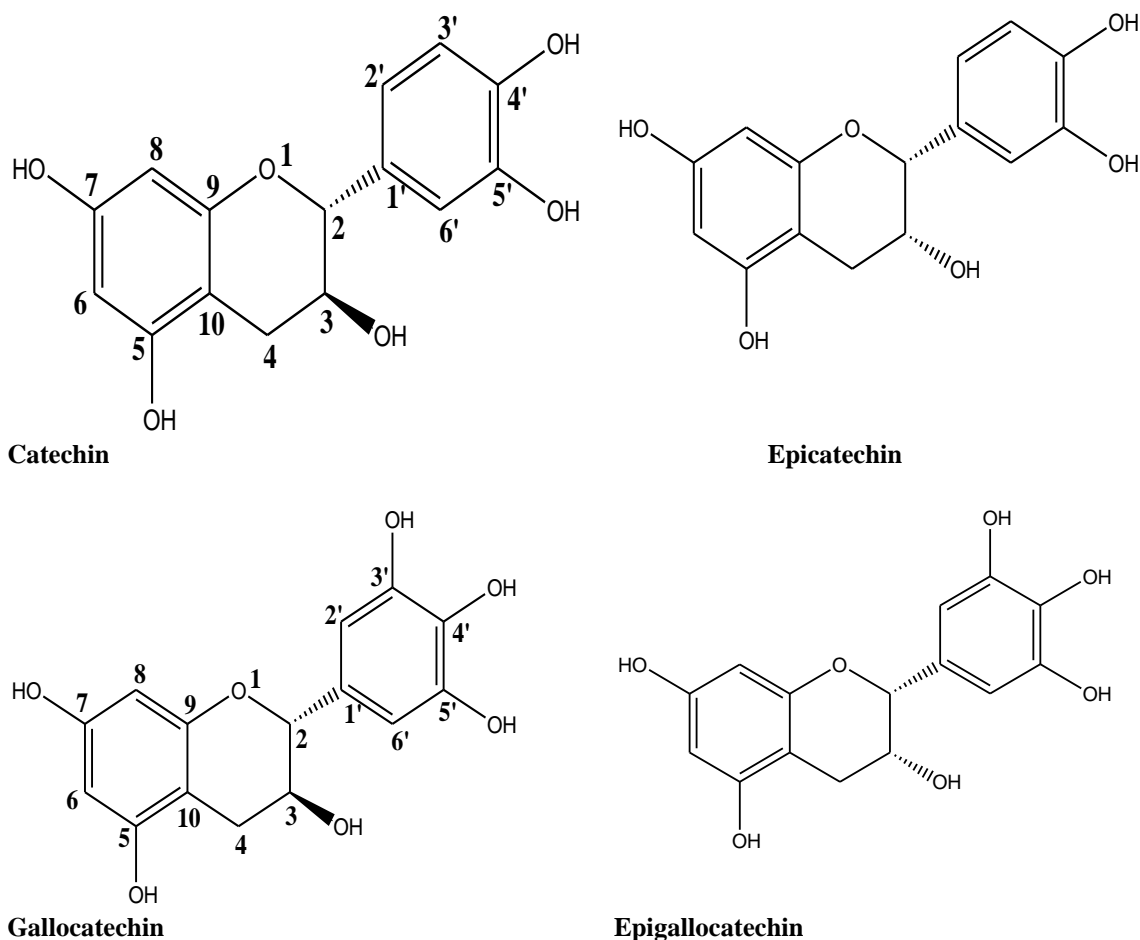


Figure 1: structures of catechin, epicatechin, gallic acid and epigallocatechin.

RESULTS

Geometric parameters

Figure 1 shows the structures of catechin, epicatechin, gallic acid and epigallocatechin. Bond lengths and measured dihedral angles are contained in Table 1. C-O bond lengths of catechin derivatives varied very little compared to those of epicatechin. Dihedral angle $O_1-C_2-C_1-C_6'$ is smaller in epicatechin compared to catechin. It's also smaller in epigallocatechin compared to gallic acid. Same remarks are made at the dihedral angle $O_1-C_2-C_1-C_2$.

Theoretical frequencies

Catechin and epicatechin are composed of 35 atoms, they have ($3N - 6 = 99$) normal modes of vibration. Gallic acid and epigallocatechin had 36 atoms which gave 102 normal modes of vibration. We are particularly interested in vO-H elongations because oxygen atoms are preferred sites for complexation with iron. Values are contained in Tables 2 and 3. In general, vO-H elongations range from 3786 cm^{-1} to 3850 cm^{-1} .

Stability

Gibbs energy (G), at computational level B3LYP / 6-311G (d, p), helps to discuss

relative stability of catechin and its derivatives. Table 4 summarises its values; ΔG is difference between Gibbs energy of catechin and epicatechin on one hand and on other hand between gallic acid and epigallocatechin. The data in Table 4 show relative stability of catechin and gallic acid compared to epicatechin and epigallocatechin, respectively.

Acidity

Catechin, epicatechin, gallic acid and epigallocatechin have hydrogenated sites (hydroxyl groups) which may be deprotonated. Choice of the site of deprotonation depends of strength of acid. ΔG variation of associated reaction ($AH \rightarrow A^- + H^+$) helps to evaluate acidity of oxygen atoms of catechin and its derivatives (Remko, 2003):

$$\Delta G = \Delta H - T\Delta S$$

$$\Delta G = G(A^-) + G(H^+) - G(AH)$$

$$G(H^+) = 2,5RT - TS^\circ(H^+) = 1,48 - 7,76 = -628\text{ Kcal. mol}^{-1}\text{ in gas phase}$$

With: **AH**: acid molecule; **A⁻**: deprotonated molecule; **G**: Gibbs energy or free enthalpy reaction; **ΔG**: variation of free enthalpy reaction; **T**: temperature; **R**: perfects gas constant; **S^o**: standard entropy reaction; **ΔS**: variation of entropy reaction.

Table 1: Bond lengths (Å) and dihedral angles (°).

Geometric parameters	Catechin	Epicatechin	Gallic acid	Epigallocatechin
O_1-C_2	1,44	1,44	1,44	1,44
O_1-C_9	1,37	1,37	1,37	1,37
O_3-C_3	1,42	1,42	1,42	1,42
O_5-C_5	1,37	1,37	1,37	1,37
O_7-C_7	1,36	1,37	1,36	1,37
O_3-C_3'	-	-	1,38	1,37
O_4-C_4'	1,38	1,38	1,37	1,37
O_5-C_5'	1,36	1,36	1,36	1,36
C_2-C_1'	1,52	1,52	1,52	1,52
$O_1-C_2-C_1-C_6'$	166,08	155,46	166,5	154,12
$O_1-C_2-C_1-C_2'$	-13,22	-24,50	-12,87	-26,09

Table 2: IR Frequencies (cm⁻¹) and Intensities of O-H Elongations of Catechin and Epicatechin.

Elongations O-H	Catechin		Epicatechin	
	Frequency	IR intensity	Frequency	IR intensity
vO ₃ -H	3808	30	3797	26
vO ₅ -H	3836	67	3835	66
vO ₇ -H	3833	65	3833	64
vO ₄ -H	3850	83	3850	84
vO ₅ -H	3789	114	3789	117

Table 3: IR Frequencies (cm⁻¹) and Intensities of O-H elongations of gallocatechin and epigallocatechin.

O-H Elongations	Galocatechin		Epigallocatechin	
	Frequency	IR intensity	Frequency	IR intensity
vO ₃ -H	3808	29	3788	20
vO ₅ -H	3836	68	3836	68
vO ₇ -H	3833	65	3833	65
vO ₃ -H	3848	66	3846	67
vO ₄ -H	3806	108	3805	108
vO ₅ -H	3787	112	3786	131

Table 4: Gibbs energies of catechin and its derivatives.

Compounds	G (a.u)	ΔG (Kcal. mol ⁻¹)
Catechin	-1031,404	0
Epicatechin	-1031,403	0,628
Galocatechin	-1106,64581	0
Epigallocatechin	-1106,64478	0,646

(a.u): atomic unit; 1 a.u = 627,51 kcal/mol.

Table 5: Acidity of Catechin and Epicatechin.

Compounds	G (a.u)	ΔG (Kj.mol ⁻¹)
Catechin	-1031,404	
Catechin (O ₃ -anion)	-1030,814	1548,8
Catechin (O ₅ -anion)	-1030,857	1435,9
Catechin (O ₇ -anion)	-1030,853	1446,3
Catechin (O ₄ -anion)	-1030,868	1407,0
Catechin(O ₅ -anion)	-1030,846	1464,8
Epicatechin	-1031,403	
Epicatechin (O ₃ -anion)	-1030,821	1527,8
Epicatechin (O ₅ -anion)	-1030,854	1442,8
Epicatechin (O ₇ -anion)	-1030,850	1451,6
Epicatechin (O ₄ -anion)	-1030,870	1399,1
Epicatechin (O ₅ -anion)	-1030,849	1454,3

(a.u): atomic unit; 1 u.a = 2625 kj/mol

Table 6: Acidity of gallocatechin and epigallocatechin.

Compounds	G (au)	ΔG (Kj/mol)
Gallocatechin	-1106,646	
Gallocatechin (O ₃ -anion)	-1106,057	1546,1
Gallocatechin(O ₅ -anion)	-1106,102	1428,0
Gallocatechin (O ₇ -anion)	-1106,098	1438,5
Gallocatechin(O ₃ -anion)	-1106,109	1409,6
Gallocatechin(O ₄ -anion)	-1106,100	1433,3
Gallocatechin(O ₅ -anion)	-1106,087	1467,4
Epigallocatechin	-1106,645	
Epigallocatechin (O ₃ -anion)	-1106,063	1527,8
Epigallocatechin(O ₅ -anion)	-1106,099	1433,3
Epigallocatechin (O ₇ -anion)	-1106,095	1443,8
Epigallocatechin(O ₃ -anion)	-1106,112	1399,1
Epigallocatechin(O ₄ -anion)	-1106,103	1422,8
Epigallocatechin(O ₅ -anion)	-1106,091	1454,3

(a.u): atomic unit; 1 u.a = 2625 kj/mol

DISCUSSION

Catéchines, epicatechin, gallo catechin and epigallocatechin have different geometric parameters. In fact, the C₇-O₇ bond is 1,36 Å in catechin and gallo catechin. It is 0,01 Å longer in epicatechin and epigallocatechin. This difference is explained by fact that the O₃-H bond is in front of plane in catechin and in gallo catechin and behind plane in epicatechin and in epigallocatechin. C_{4'}-O_{4'} bond is 1,38 Å in catechin and epicatechin. It is 0,01 Å shorter in gallo catechin and epigallocatechin. This inequality is due to presence of hydroxyl group O₃'-H in gallo catechin and in epigallocatechin. Dihedral angle O₁-C₂-C₁'-C₆' is 166,08 ° in catechin and is 10,62 smaller in epicatechin. It is 166,5 ° in gallo catechin and is 11,96 smaller in epigallocatechin. Dihedral angle O₁-C₂-C₁'-C₂' is -13,22 ° in catechin and is 11,28 smaller in epicatechin. It is -12,87 ° in gallo catechin and is 13,22 smaller in epigallocatechin. Orientation of the O₃-H bond thus influences the value of the dihedral angles (Ana et al., 2006; Hong et al., 2018).

O-H (νO-H) elongations of catechin and its derivatives appear between 3786 cm⁻¹ and 3850 cm⁻¹. The νO₄'-H elongation is observed at 3850 cm⁻¹ in catechin and epicatechin, at 3806 cm⁻¹ in gallo catechin and at 3805 cm⁻¹ in epigallocatechin. This difference is explained by the presence of the hydroxyl group O₃-H in gallo catechin and in epigallocatechin. (νO₅-H) elongation is virtually identical in the four compounds. It comes out at 3836 cm⁻¹ in catechin, gallo catechin and epigallocatechin and at 3835 cm⁻¹ in epicatechin. νO₇-H elongation appears at 3833 cm⁻¹ in the four compound. νO₃-H elongation is least intense. It appears at 3808 cm⁻¹ in catechin and gallo catechin, at 3797 cm⁻¹ in epicatechin and at 3788 cm⁻¹ in epigallocatechin (Jayshree et al., 2010).

Difference between the frequencies of catechin and epicatechin on one hand and gallo catechin and epigallocatechin on other hand is certainly due to fact that O₃-H bond is in front of or behind plane. Elongation νO₅-H is most intense. It was released at 3789 cm⁻¹ in

catechin and epicatechin, at 3787 cm⁻¹ in gallo catechin and 3786 cm⁻¹ in epigallocatechin.

With regard to stability, results in Table 4 show that catechin is more stable than epicatechin with an energy difference of 0.628 Kcal/mol; similarly, gallo catechin is more stable than epigallocatechin with an energy difference of 0.646 Kcal/mol.

Acid nature of oxygen atoms of catechin and its derivatives is related to the ΔG value. Lower it is, more oxygen in the position is acid. Under these conditions, O₄' oxygen atom is most acid site of catechin, followed by O₅ oxygen atom. Oxygen atom O₄' is also most acid site of epicatechin. Definitely, in catechin and epicatechin acidity of oxygen atoms varies in same direction. The descending order is O₄' > O₅' > O₇' > O₅' > O₃'. The oxygen atom O₃' is most acid site of gallo catechin, then oxygen O₅, after oxygen O₄'. In general, in gallo catechin, the acidity of oxygen atoms decreases in order O₃' > O₅' > O₄' > O₇' > O₅' > O₃'. Oxygen O₃' is also most acid site of epigallocatechin, followed by oxygen O₄'. Decreasing order of epigallocatechin acidity oxygen atoms is O₃' > O₄' > O₅' > O₇' > O₅' > O₃'. The oxygen atom O₃ is thus least acid site of catechin and its derivatives (Ana et al., 2006; Jayshree et al., 2010; Hong et al., 2018).

Conclusion

The acidity of catechin and epicatechin decreased in the order O₄' > O₅' > O₇' > O₅' > O₃' while that of gallo catechin and epigallocatechin decreased in the order O₃' > O₅' > O₄' > O₇' > O₅' > O₃' and O₃' > O₄' > O₅' > O₇' > O₅' > O₃'. Knowing the theoretical acidity order of the different hydroxyl groups of the monomers, we will focus our researches to modelize different models of polyphenol inorganic iron complexes likely to be formed in the human organism according to these results. In a second step, complexation sites of inorganic iron will also be modelized to possibly reduce affinity of these condensed tannins with respect to iron.

COMPETING INTERESTS

The authors declare that there are no competing interests.

AUTHORS' CONTRIBUTIONS

Bioinformatics calculations were carried out by KDY, BAA and LAB. Results interpretation was done by KDY, BAA and BRN. Finally, manuscript writing was done by KDY, BAA and MVS.

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