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# Albendazole potentiates the neurotoxic effect of ivermectin in rat

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## ABSTRACT

The study was carried out to investigate whether an interaction between albendazole and ivermectin could lead to enhanced central nervous system (CNS) toxicity. Ivermectin (0.4mg/kg body weight (b.w)) and/or albendazole (15mg/kg b.w) were daily and orally administered to albino rats for 14 days. Activities of acid phosphatase (ACP), alkaline phosphatase (ALP), catalase (CAT), Na<sup>+</sup>-K<sup>+</sup> ATPase, Ca<sup>2+</sup>-Mg<sup>2+</sup> ATPase and malondialdehyde (MDA) level which are considered essential to correct functioning of the brain were determined. The co-administration of the two drugs significantly caused reduction (p<0.05) in the activities of brain ACP, ALP, Na<sup>+</sup>-K<sup>+</sup> ATPase and Ca<sup>2+</sup>-Mg<sup>2+</sup> ATPase with corresponding increase in the serum. Separate administration of ivermectin or albendazole did not show any significant changes (p>0.05) in the activity of these enzymes. These suggest that in the presence of albendazole, ivermectin is able to reach the CNS and impair its function through neurochemical changes. Also, co-administration of ivermectin and albendazole led to a significant increase (p<0.05) in brain CAT activity as well as serum and brain MDA concentrations. This may be an indication that the drugs have other mechanisms of action, such as increasing oxidative damage in the CNS. Overall, these findings suggest that both drugs exert additive effect when co-administered. © 2012 International Formulae Group. All rights reserved.

Keywords: Combination therapy, enhanced CNS toxicity, ATPases.

#### **INTRODUCTION**

Brain is a vital organ of the body whose function controls the activities of the cellular system. It consists of three parts viz. forebrain, midbrain, and hindbrain. It is involved in sensory perception, motor control, memory association, thought and personality, regulating temperatures, water metabolism, blood sugar level, productive cycle, hunger, thirst, anger and pleasure (Cicerone et al, 2005), and several forms of reflex actions. Due to the complexity of its functions, the brain receives some 15% of the total cardiac output and accounts for about 20% of the total oxygen consumption of the body at rest (Alavijeh et al, 2005). The high oxygen utilization by the brain during metabolism increases its susceptibility to oxygen free radicals which is the major mode of action of neurotoxicant. However, the brain regulates transport of materials in and out through the action of blood-brain barrier which is composed of specialized endothelial cells that prevent various substances from entering the brain except those that are either small or lipophilic (Rubin and Staddon, 1999) thereby reducing accumulation of toxic materials in the brain.

Ivermectin and albendazole are synthetic drugs used for the treatment of

© 2012 International Formulae Group. All rights reserved. DOI : http://dx.doi.org/10.4314/ijbcs.v6i1.28 Lymphatic filariasis (Elephantiasis) and Onchocerciasis (river blindness) respectively. Both diseases are major public health problems in several tropical countries (Ottesen and Ramachandran, 1995). An estimated 120 million people in more than 80 endemic countries are infected with Lymphatic filariasis (Molyneux et al., 2003) and 50 million individuals remain at risk with 17.7 million infected with Onchocerca volvulus predominantly in 22 sub-Saharan African countries. The strategy that has been shown to be most effective is the treatment of affected populations with the drug ivermectin - a white or slightly yellow crystalline powder (Chabala et al., 1980). However, report of resistance to ivermectin in nematodes is becoming increasingly common (Richard, 1994: Sangster, 1996). In view of this and the coendemic nature of the two diseases, clinical trials of ivermectin and albendazole combination commenced in some African countries like Ghana (Awadzi et al., 2003). The advantageous pharmacologic action of this combination therapy is that while ivermectin is only microfilaricidal (Goa et al., 1991) albendazole is toxic to all intra-uterine stages (Awadzi et al., 1995). Although, this regimen has been tried in many countries with reports of high efficacy in the treatment of onchocerciasis and lymphatic filariasis (Ismail et al., 1998; Awadzi et al., 2003), their effect on various metabolic processes, tissues and cellular systems of the mass human populations receiving these drugs is yet to be clarified. This study is thus aimed at elucidating the toxic effect of the combination therapy on the cellular integrity of the brain of wistar rats.

# MATERIALS AND METHODS Materials

## Experimental animals

A total of 20 male albino rats (*Rattus* novergicus) of average weight 157.50 g  $\pm 2.50$  g were obtained from the Animal Holding Unit of the Department of Biochemistry, University of Ilorin, Ilorin, Nigeria. The rats

were maintained on normal rat chow and water *ad libitum* and were housed in wooden cages. Animal husbandry and experimentation were consistent with the Guiding Principles in the Use of Animals in Toxicology (Derelanko, 2000).

## Drugs and reagents

Ivermectin and albendazole were products of Merck and Co. England and Glaxo Smithkline Beecham, Netherlands respectively. All reagents used were of analytical grade and were prepared in all-glass distilled water. The reagents were stored in clean, air tight reagent bottles.

## Methods

## Animal grouping and drug administration

The rats were randomly divided into four groups of five rats each. Group I (control) received an appropriate volume of distilled water. Group II (Iver) received a human therapeutic dose of 0.4 mg/kg body weight (b.w.) ivermectin; Group III (Alb) received a human therapeutic dose of 15 mg/kg body weight albendazole while group IV (Iver + Alb) received a dose of 0.4 mg/kg body weight ivermectin co-administered with 15 mg/kg body weight albendazole. The administration was done orally on a daily basis for fourteen (14) days at 24 h interval (at 9.00a.m). This duration was based on the clinical trial conducted by Ismail et al. (1998) to evaluate the potency of the combination therapy. The animals were sacrificed 24 h after the fourteenth day administration.

# Preparation of serum and brain homogenate

At the end of the experimental period, approximately 5 ml of venous blood was collected from each of the experimental animals according to the method of Narayanan et al. (1984) The serum was prepared by centrifuging the blood samples at 3000 rpm for 5 min (Ogbu and Okechukwu, 2001) and serum collected with a Pasteur Pipette. Immediately after the collection of blood, the brain was removed and transferred into a well labelled container of ice-cold 0.25 M sucrose solution (1:5 w/v) and homogenized. The homogenates were kept frozen overnight to ensure maximum release of the enzymes before being used for the various biochemical assays (Adebayo et al., 2003).

# Assay of enzyme activities and lipid peroxidation

Acid phosphatase activity was assayed using the method of Armstrong (1964). The method of Bessey et al. (1946) as modified by Wright et al. (1974) was employed for the assay of alkaline phosphatase activity. ATPase activities were assayed using the method of Ronner et al. (1977) and catalase by the method of Sinha (1971). Determination of malondialdehyde level was done by the method of Varshney and Kale (1990). Protein content of the homogenate and serum were determined using the Biuret method (Gornall et al., 1949).

## Statistical analysis

All data are presented as mean  $\pm$  standard deviation. Statistical analyses were carried out using Duncan Multiple Range test (Montgomery, 1976). In all cases, probability level of 95% was taken as significant.

### RESULTS

Figure 1 shows the effect of administration of ivermectin and /or albendazole for 14 days on rat brain-body weight ratios. Combined administration of both drugs significantly reduced (p<0.05) brain-body weight ratios of rats. There was no significant change in this parameter in rats administered separately with ivermectin or albendazole when compared with controls.

Figure 2 shows the activity of ALP in brain of rats following the administration of ivermectin, albendazole and their combination for 14 days. The serum ALP activity was significantly increased (P<0.05) in all treatment groups when compared with the controls. There existed no significant

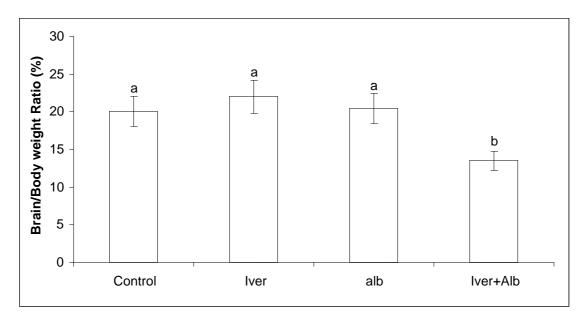
difference (P>0.05) in the brain ALP activity of rats administered separately with ivermectin or albendazole when compared with the control. However, about two folds reduction in activity was observed following co-administration of both drugs for 14 days when compared with control.

Figure 3 presents the effects of both separate and combined administration of ivermectin and albendazole for 14 days on the activity of acid phosphatase (ACP) of rat brain. The serum acid phosphatase activity increased significantly (P<0.05) following combined administration of ivermectin and albendazole while there was no significant differences (P>0.05) when each drug was seperately administered to rats compared with control. There was no significant difference (P>0.05) in the brain ACP activity of rats administered ivermectin and albendazole seperately while in rats administered combined invermectin and albendazole, there was a significant increase (P<0.05) in activity when compared with control.

Combined administration of ivermectin and albendazole for 14 days caused significant (P<0.05) drop in brain Na<sup>+</sup>-K<sup>+</sup> and Ca<sup>2+</sup>-Mg<sup>2+</sup> ATPase activities (Figures 4 and 5). However, no significant changes (p>0.05) were observed following separate administration for the same period.

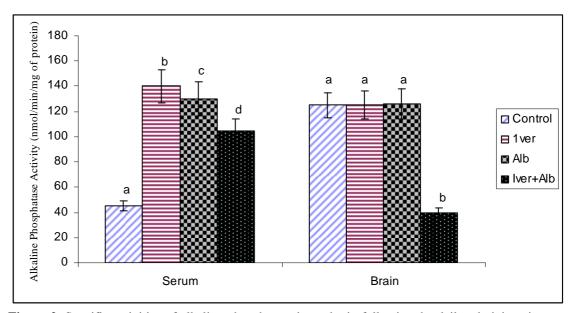
Separate administration of ivermectin and albendazole did not affect (P>0.05) brain catalase activity (Figure 6). However their co–administration led to a significant increase (P<0.05) in the enzyme activity of brain when compared with control.

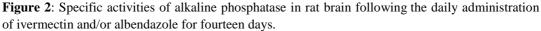
Neither of the drugs caused any significant change (P>0.05) in the concentration of MDA in the serum and brain when compared with controls following 14 days administration (Figure 7). However, co–administration of ivermectin with albendazole for 14 days caused a significant elevation (P<0.05) in both serum and brain malondialdelhyde levels.



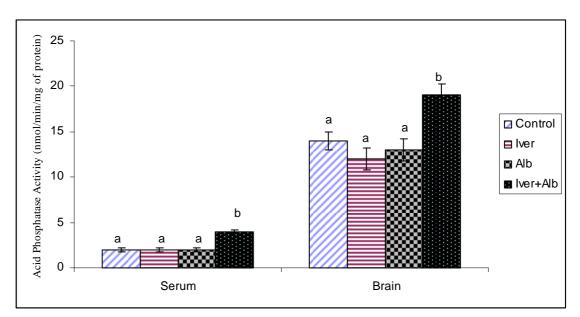
**Figure 1:** Brain-body weight ratio of rats following the daily administration of ivermectin and/or albendazole for fourteen days.

Values are means (n = 5)  $\pm$  S.D (bars with different superscripts are significantly different at P<0.05). Iver = Ivermectin; Alb = Albendazole.



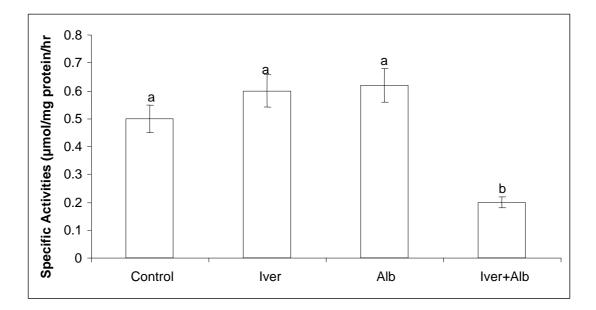


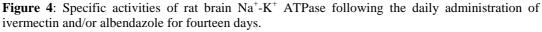
Values are means  $(n=5) \pm S.D$  (bars with different superscripts are significantly different at P<0.05). Iver = Ivermectin; Alb = Albendazole.



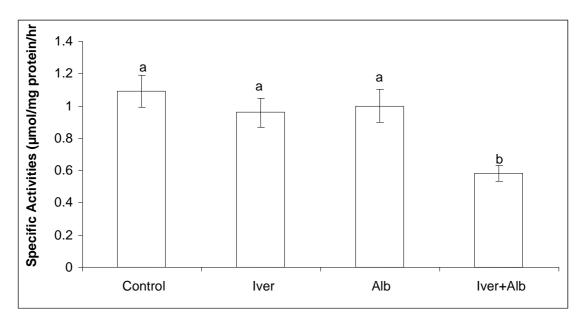
**Figure 3**: Specific activitivies of acid phosphatase in rat brain following the daily administration of ivermectin and/or albendazole for fourteen days.

Values are means  $(n=5) \pm S.D$  (bars with different superscripts are significantly different at P<0.05). Iver = Ivermectin; Alb = Albendazole.





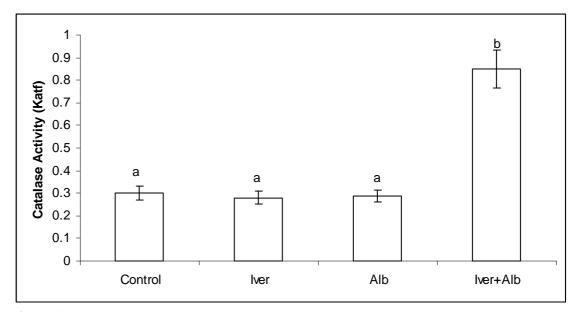
Values are means  $(n=5) \pm S.D$  (bars with different superscripts are significantly different at P<0.05). Iver = Ivermectin; Alb = Albendazole.

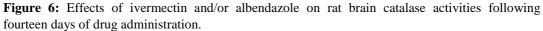


**Figure 5:** Specific activities of rat brain  $Ca^{2+}-Mg^{2+}$  ATPase following the daily administration of ivermectin and/or albendazole for fourteen days.

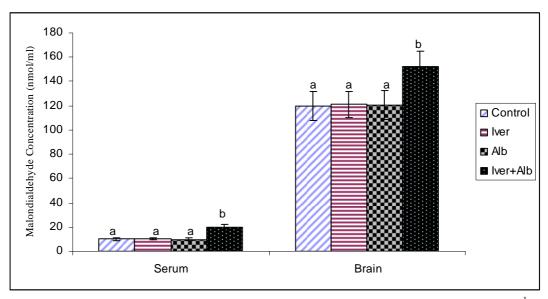
Values are means (n=5)  $\pm$  S.D (bars with different superscripts are significantly different at P<0.05).

Iver = Ivermectin; Alb = Albendazole.





Values are means  $(n=5) \pm S.D$  (bars with different superscripts are significantly different at P<0.05). Iver= Ivermectin; Alb= Albendazole.



**Figure 7**: Effects of ivermectin and/or albendazole on malondialdehyde (MDA) concentration<sup>1</sup> in rat brain followingr fourteen days of drug administration.

Values are means  $(n=5) \pm S.D$  (bars with different superscripts are significantly different at P<0.05). Key: Iver= Ivermectin; Alb= Albendazole

### DISCUSSION

Organ/body weight ratios are normally investigated to determine change in size of the organ relative to the body (Bailey et al., 2004). High Organ to body weight ratio has been associated to inflammation while otherwise is necrosis. Xenobiotics (including drugs) have been reported to induce inflammatory responses (Omiecinski, 2011) disrupting respiration and affecting energy production. This may result in dependence of the brain cells on anaerobic respiration to generate energy. The eventual accumulation of the product of this process- lactic acid in brain cells may lead to cell damage as a result of lactic acidosis with consequent Efflux mechanism at the inflammation. blood-brain barrier is a limiting factor in the penetration of drugs. Drugs including ivermectin and albendazole are actively transported out of brain, thus limiting the concentration of these compounds in the brain (Taylor and Yan, 2002). The significant decrease in the brain/body weight ratio of rats administered combined ivermectin and

albendazole, suggests appreciable accumulation of ivermectin in the brain thus causing cell death. Ivermectin is known to be neurotoxic because of its action on GABA receptors (Leo et al., 1996). Both ivermectin and albendazole have been reported as cosubstrates of cytochrome P<sub>450</sub> (CY3A4) (Edwards, 2003). However it has also been proposed that albendazole may be substrate to *p*-glycoprotein (Redondo et al., 1999). Therefore, since *p*-glycoprotein may have higher affinity for albendazole, it may in the presence of ivermectin mediate the efflux of albendazole ivermectin allowing to accumulate appreciably in the brain. The significant increase in serum ALP activity

The significant increase in serum ALP activity of all the treatment groups following 14 days administration (P<0.05) when compared with the control is suggestive of a possible damage to tissue cell plasma membrane by the administration of the drugs either singly or in combination, thus leading to leakage of membrane components into the extracellular fluid (Akanji et al., 1993). This is further supported by significant reduction in brain ALP activity following the co-administration of ivermectin with albendazole for 14 days. This effect may be attributed to the role of pglycoprotein of blood brain barrier (BBB) and the fact that albendazole has been proposed to be a substrate to p-glycoprotein (Redondo et al., 1999). It may be possible that pglycoprotein has higher affinity for albendazole and in the presence of ivermectin mediated the efflux of albendazole thereby allowing ivermectin accumulate to appreciably in the brain to exert its neurotoxic effects on the brain cells, one of which is membrane derangement and hence the observed loss of ALP activity.

Acid phosphatase is an enzyme of the lysosomal membrane (Zhang et al., 2009). Acid phosphatase was chosen for assay on the basis of its specificity for lysosomal membrane. Its activity before and after exposure to insult with chemical compounds (in this case ivermectin and/or albendazole) gives a picture of the sequence of cell damage if any. Alteration in acid phosphatase activity will thus explain the suspected effects of these drugs on lysosomes (Zhang et al., 2009). The increase in ACP activity observed following combined administration of the drug could be a consequence of de novo synthesis induction by accumulation of ivermectin in the brain. Another possible explanation is the release of the enzyme from the brain lysosomal membrane, an indication of tissue damage.

 $Na^+-K^+$  ATPase is a membrane bound enzyme invoved in maintaining potential difference across the plasma membrane and also mediates transport of several other molecules into the cells. The brain  $Na^+$ - $K^+$  ATPase activity was significantly reduced in the combination group. This may be ascribed to the modulatory effect of albendazole in potentiating the neurotoxic effect of ivermectin by competitively preventing its efflux by *p*-glycoprotein while albendazole gets pumped out of the brain. The reduced activity observed could be attributed to the binding of the drug to various subunits essential for the activity of the enzyme possibly via cysteine residues, as reported in the interaction of ivermectin with cytochrome P<sub>450</sub> CY3A (Beers and Berkow, 1999; Rossi, 2000) which may reduce the enzyme activity. Ca<sup>2+</sup> plays a very important role as second messenger, effecting brain signals. Its gradient is maintained by several of its transporter across the cellular membrane. The significant reduction in activity of Ca<sup>2+</sup>-Mg<sup>2+</sup> ATPase following ivermectin, and albendazole administration may be attributed to membrane derangement or inhibition of the enzyme by the drugs. Ahern et al. (1999) reported that ivermectin increased Ca2+ release from sarcosplasmic reticulum vesicles and from endosplasmic reticulum by inhibiting Ca<sup>2+</sup> uptake by  $Ca^{2+}$  -Mg<sup>2+</sup> ATPase.

Exposure to xenobiotics results into many complex processes which can be evaluated by antioxidant enzyme activity as well as by lipid peroxidation measures (Romeo et al., 2000). Increase in catalase activity could be attributed to compromise of the tissue xenobiotic protective biotransformation mechanism which may resulted accumulation have from of ivermectin in the brain mediated by albendazole thus leading to the production of reactive oxygen species and concomittant increased catalase activity to cope with these.

Lipid peroxidation is assessed by maximal rate of malondialdehyde formation (Chield et al., 1999). The integrity of cells could be assessed by evaluating the malondialdehyde level. Usually, the level is elevated under conditions such as oxidative stress where reactive oxygen group could react with unsaturated fat and increase malondialdehyde formation. In the brain, malondialdehyde levels can be used to measure various diseased conditions. Weigand et al. (1999) investigated cerebral formation of malondialdehyde as an index of lipid peroxidation in relation to different sources of reactive oxygen species in patients undergoing carotid endarterectomy. It has been proposed to be one of the major mechanisms of secondary damage in traumatic brain injury. The brain is particularly prone to oxidative injury because the membrane lipids are essentially rich in polyunsaturated fatty acid (Halliwell and Gutteridge, 1989). Increased serum MDA levels have been found in patients with cancer (Huang et al., 1999). The increase in MDA level observed in this study suggests that the antifilarials are capable of inducing oxidative stress which is in agreement with the report of Yarsan et al.(2002) that albendazole caused increased formation of MDA in the blood of healthy mice.

## Conclusion

The current study suggested that albendazole may mediate neurotoxic effect of ivermectin during combined administration for the treatment of filarial diseases. This study however, gives way to further study for the determination of kinetic parameters as a measure of affinity of p- glycoprotein for ivermectin and albendazole.

#### REFERENCES

- Adebayo JO, Yakubu MT, Egwim EC, Owoyele BV, Enaibe BU. 2003. Effect of ethanolic extract of *Khaya senegalensis* on some biochemical parameters of rat kidney. J. Ethnopharmacol., 88: 69-72.
- Ahern GP, Junankar PR, Pace SM, Curtis S, Mould JA, Dulhunty AF. 1999. Effects of Ivermectin and midecamycin on ryanodine receptors and the Ca<sup>2+</sup> -ATPase in Sarcoplasmic reticulum of rabbit and rat skeletal muscle. *J. Physiol.*, 514: 313 – 326.
- Akanji MA, Olaore OA, Oloyede OB. 1993. Effect of chronic consumption of metabisulphite on the integrity of the rat kidney cellular system. *Toxicol.*, 81: 173-179.
- Alavijeh MS, Chishty M, Qaiser MZ, Palmer AM 2005. Drug Metabolism and Pharmacokinetics, the Blood-Brain

Barrier, and Central Nervous System Drug Discovery. *NeuroRx*, **2**(4): 554– 571.

- Armstrong K. 1964. Enzymes. In *Text of Clnical Chemistry*. Tietz NW (ed.). W.B Saunders Company; 704-757.
- Awadzi K, Addy ET, Opoku NO, Plenge-Borig A, Burtner DW. 2003. *O. volvulus* resistance to ivermectin: evidence for resistance and resistance monitoring in the field. Final report of the conference on the eradicatibility of onchocerciasis. *Filarial J.*, **2**: 2-17.
- Awadzi K, Addy ET, Opoku NO, Lenge-Borig A, Burtner DW. 1995. The chemotherapy of onchocerciasis: Ivermectin in combination with albendazole. *Trop. Med. Parasitol.*, **46**: 213-220.
- Bailey SA, Zidell RH, Perry RW. 2004.
  Relationships Between Organ Weight and Body/Brain Weight in the Rat: What Is the Best Analytical Endpoint? *Toxicologic Pathology*, **32**: 448–466.
- Beers MH, Berkow R. 1999. In *the Merck Manual of Diagnosis and Therapy* (17<sup>th</sup> edn), Beers MH, Berkow R (eds). White House Station: Washington; 55.
- Bessey OA, Lowry OA, Brock MD. 1946. Appearance of specific acid phosphatase isoenzymes in the synovial fluid of patients with rheumatoid arthritis. *Nature*, **216** : 274 – 277.
- Chabala JC, Mrozik H, Tolma RL, Eskola PLA, Fisher GH. 1980. Ivermectin : A new broad spectrum antiparasitic agent. *J. Med. Chem.*, **23**: 1136-1934.
- Chield R, Brown S, Dunelly A, Roper H, Saxton J. 1999. Changes in indices of antioxidant status, lipid peroxidation and inflammation in human skeletal muscle after eccentric muscle action. *Clin. Sci. Calch.*, **96**: 105 – 115.
- Cicerone K, Dahlberg C, Malec JF, Langenbahn DM, Felicetti T, Kneipp S, Ellmo W, Kalmar K, Giacino JT, Harley JP. 2005. Evidence-based cognitive

rehabilitation: Updated review of the literature from 1998 through 2002. *Archives of Physical Medicine and Rehabilitation*, **86**: 1681–1692.

- Derelanko MJ. 2000. Guiding principles in the use of animals in toxicology. In *Toxicologist's Pocket Handbook*. CRC Press: Boca Raton, London; 1-7.
- Edwards G. 2003. Ivermectin: does pglycoprotein play a role in neurotoxicity? *Filarial. J.*, **2**: S8.
- Goa KL, McTavish D, Clissold S. 1991. Microfilaricidal activity of ivermectin. *Drugs*, **42**: 640-658.
- Gornall A, Bardsmill CT, David MM. 1949. Determination of serum protein by means of biuret reaction. *J. Biol. Chem.*, **177**: 751-766.
- Halliwell B, Gutteridge JMC. 1989. Free radicals in biology and medicine. *Mol. Aspects. Med.*, **8**: 89-93.
- Huang VL, Sheu JY, Lin JH. 1999: Association between oxidative stress and changes of trace elements in patients with breast cancer. *Clin. Biochem.*, **32**: 131– 136.
- Ismail MM, Jayakoly RL, Weil GGJ. Nirmalan, N, Jaya-Single KSS, Abeyewickrema W, Rezrisheriff MH, Rajaratnam HN, Amarisekera N, Desilva DCL, Michalski MLK, Dissanaike AS. 1998. Efficacy of single dose combination of albendazole, ivermectin and diethyl carbamazine for the treatment of bancroftian filariasis. Trans. R. Soc. Trop. Med. & Hyg., 92: 94-97.
- Leo PA, Tranquilli WJ, Seward RL, Todd, KS, Dipietro JA. 1996. Clinical observations in collies given ivermectin orally. *Am. J. Vet. Res.*, **48**: 684-685.
- Molyneux DH, Bradley M, Hoerauf A, Kydem D, Taylor MJ. 2003. Mass drug treatment for lymphatic filariasis and onchocerciasis. *Trends Parasitol.*, **19**: 516-522.

- Montgomery DC 1976. *Design and Analysis* of *Experiment*. John Wiley: New York; 48-50.
- Narayanan CR, Joshi DD, Mujumdar AM. 1984. Hypoglycemic action of Boungavillea spectabilis leaves. Curr. Sci., 53: 579-581.
- Ogbu SI, Okechukwu EI. 2001 The effect of storage temperature prior to separation on plasma and serum potassium. *J. Med. Lab. Sci.*, **10**:1-4.
- Omiecinski CJ, Vanden Heuvel JP, Perdew GH, Peters JM. 2011. Xenobiotic Metabolism, Disposition, and Regulation by Receptors: From Biochemical Phenomenon to Predictors of Major Toxicities. *Toxicological Sciences*, **120**(S1): S49-S75.
- Ottesen EA, Ramachandran CP. 1995. Lymphatic filariasis in tropical countries. *Parasitol. Today*, **11**: 129 – 131.
- Redondo PA, Alvarez AI, Garcia JL, Larrode OM, Merino G, Prieto JG. 1999.
  Presystemic metabolism of albendazole: experimental evidence of an efflux process of albendazole sulfoxide to intestinal lumen. *Drug. Metab. Dispos.*, 27: 736- 740.
- Richard LD. 1994: Ivermectin in loaloasis and concomitant O. volvulus and M. perstans infections. Am. J. Trop. Med. Hyg., 39: 480-483.
- Romeo M, Bennani N, Gassia-Barelli M, Lafaurie M, Girard JP. 2000. Cadmium and copper display different responses towards oxidative stress in the kidney of the sea bass *Dicentrarchus labrax*. Aquat. Toxicol., 48: 185-194.
- Ronner P, Gazzotti P, Carafoli E. 1977. A lipid requirement for the Ca<sup>2+</sup>, Mg<sup>2+</sup>-ATPase of erthrocyte membranes. *Arch. Biochem. Biophys.*, **179**: 578-583.
- Rossi V. 2000. Factors affecting drug response. In *The Merck Manual of Medical Information*, Berkow R, Beers MH (eds). Home ed. Whitehouse Station NJ. Merck .Sect2; 101-113.

- Rubin LL, Staddon JM. 1999. The cell biology of the blood-brain barrier. *Ann. Rev. Neurosci.*, **22**: 11-28.
- Sangster NC. 1996. Managing parasiticide resistance. *Vet. Parasitol.*, **98**: 89-109.
- Sinha KA. 1971. Colorimetric assay of catalase. *Anal. Biochem.*, **47**: 389-394.
- Taylor EM, Yan R. 2002. Impact of efflux transporters in brain on development of drugs for CNS disorders. *Clin. Pharmacokinet.*, 2:112-123.
- Varshney R, Kale RK. 1990. Effect of calmodulin antagonist on radiation induced lipid peroxidation in microsome. *Int. J. Rad. Biol.*, **58**: 733-743.
- Weigand MA, Laipple A, Plaschke K, Eckstein HH, Martin E, Bardenlever HJ. 1999. Concentration changes of

malondialdehyde across the cerebral vascular bed and shedding of L– selecting during carotid endarterectomy. *Stroke*, **30**: 306–311.

- Wright PJ, Plummer DT. 1974. The use of urinary enzyme measurement to detect renal damage caused by nephrotoxic compounds. *Enzymologia*, **42**: 317-327.
- Yarsan E, Ceik S, Eraslan G, Aycicek H. 2002. Effects of albendazole treatment on lipid peroxidation of healthy and *Toxocaris canis* infected mice. *Isr. Vet. Med. Ass. J.*, 57: 1-11.
- Zhang L, Sheng R, Qin Z. 2009. The lysosome and neurodegenerative diseases. *Acta Biochimica et Biophysica Sinica*, **41**(6): 437-445.