

Effect of ultraviolet-B radiation on biochemical and antioxidant defence system in *Indigofera tinctoria* L. seedlings

K.C.Ravindran¹, A.Indrajith¹, P.V.Pratheesh¹, K.Sanjiviraja¹ and V.Balakrishnan²

¹Department of Botany, Faculty of Science, Annamalai University, Annamalaiagar-608 002, Tamil Nadu, INDIA

²Department of Biotechnology, K.S.Rangasamy College of Technology, Tiruchengode-637 215, Tamil Nadu, INDIA
E-mail : (drkc_ravi@rediffmail.com (K.C.Ravindran), *Corresponding Author)

Abstract

The stratospheric ozone depletion and enhanced solar ultraviolet-B (UV-B) irradiance may have adverse impact on living organisms. The impact of UV-B radiation (UV-B, 280~320nm) on growth, biochemical and antioxidant enzyme activity was studied in *Indigofera tinctoria* (L.) seedling, commonly used as a green manure. The supplementary UV-B radiation significantly decreased the growth, development and changes in UV-B absorbing compounds such as anthocyanin and flavonoids. The antioxidant enzymes were unaffected and showed an enhanced activities of peroxidase, superoxide dismutase, polyphenoloxidase and phenylalanine ammonia-lyase except catalase in UV-B irradiated seedling. *Indigofera tinctoria* seedling tries to counteract high level of reactive oxygen species produced under UV-B stress through the increased activities of antioxidant enzyme. The results suggest that *Indigofera tinctoria* is resistant to UV-B radiation damage and the possible negative effect of additional UV-B radiation on the growth of seedling may have been effectively balanced by the UV-B radiation stress through increase in UV-absorbing compound and antioxidant enzymes.

Keywords: Antioxidant, flavonoids, stress, UV-B radiation.

1. Introduction

The stress affecting plants are numerous and often species or even variety or location specific. They include drought, high salinity, temperature extremes, water logging, mineral nutrients deficiency, metal toxicity, pollutants and ultraviolet-B (UV-B) radiation (Smirnoff, 1998). It has been established that depletion of the stratosphere ozone layer is increasing the level of ultraviolet-B radiation reaching the earth surface (Caldwell et al., 1998). Scenario-based chemistry-climate models shows that in the middle of 21st century, UV-B radiation at ground level is enhanced due to high concentration of greenhouse gases and halogenated species (Taalas et al., 2000).

A wide range of morphological, growth, biochemical, and physiological responses of plant have been reported to elevate UV-B radiation (Caldwell et al., 1998; Zhang et al., 2003; He et al., 2003). Plant have developed a complex biochemical defense system that including carotenoids and flavanoids. Flavonoid compounds, as secondary metabolites are considered to play a major role in protecting plants from UV-B damage (Liang et al., 2006). These flavonoids generally absorb the light in the region of 280~320 nm and thus are capable of acting as a UV filter, thereby protecting the photosynthetic tissues from damage (Siefermann, 1987). Flavonoids stabilize and protect the lipid phase of the thylakoid membrane, and are quenchers of the excited triplet state of chlorophyll and singlet oxygen (Agawal and Rathore, 2007). Apart from the flavonoids, carotenoids also have antioxidant properties which act as an internal filter against UV-B radiation. Plants scavenge reactive oxygen species by detoxification mechanism produced by enzymatic antioxidant such as catalase, peroxidase, superoxide dismutase and phenylalanine ammonia-lyase etc (Moran and Porath, 1980). Some plants are more tolerant to UV-B than others because they produce a variety of secondary metabolites that effectively absorb UV-B and prevent it from penetrating into the leaf mesophyll cells. The aim of this work was to investigate the effects of UV-B radiation on *Indigofera tinctoria* seedling against UV-B radiation under field condition.

2. Materials and Methods

2.1. Plant material and UV-B exposure

Seeds were surface sterilized with 0.2% CuSO₄ for 12h and grown in plastic trays containing a soil mixture of sand (70%) and compost (30%). Seeds were grown in a 16:8h light: dark photo period. After germination the selected seedlings were transferred to field conditions and subjected to UV-B radiation with Philips sun lamps (Philips TL 20 W/12, N.V. Philips Gloelampenfabrickan, Holland) installed 15 cm above the seedlings and oriented in an east-west direction. UV radiation was filtered through cellulose acetate filter paper to avoid transmission of wave length below 290 nm. Controls were exposed to normal day light. Seedlings were irradiated for 2 hours per day (11.00 to 13.00h) for 8 days.

2.2. Growth parameters and photosynthetic pigments

Total length of the seedlings and fresh weight were measured immediately after removing the seedling from the experimental field. Leaf area was calculated by using the Licor 3100 leaf area meter (LICOR, model LI- 3100, Lincoln, USA). For leaf thickness, leaf bits of 0.5 X 1.5 cm were cut and cross sections of leaves were prepared by using a rotary microtome. Chlorophyll content was estimated by the procedure of Moran and Porath (1980) using the formulae suggested by Inskeep and Bloom (1985). Fresh leaf disc of 100mg were cut and placed in a test tube containing 10 ml of N, N' – dimethyl formamide (DMF) and stored for 24h at 4°C. The coloured supernatant was used for chlorophyll estimation by reading the absorbance at 647 and 666 nm in a spectrophotometer with DMF as blank for chlorophyll.

Anthocyanin content was extracted in acidified methanol (1:99 HCl:methanol) in 100 mg of leaf material. Extract was kept at 0°C for 24 hours. After 24h, the content was made up to 10ml and the absorbance was read at 530 nm, as described by Mancinelli et al. (1975) Flavonoids were extracted and quantified by the Mirecki and Teramara (1985) method. 100 mg of leaves were placed in 80 per cent acidified methanol (methanol:water:HCl 80:20:1) for 12 hours in dark at 4°C to extract flavonoids and the absorbance was read at 315 nm.

2.3. Measurement of the activities of catalase, peroxidase and superoxide dismutase

Catalase (CAT, EC 1. 11.1.6) activity was measured by the method of Machly and Chance (1959). One gram of leaf tissue was homogenized in 10ml of 0.1 mM sodium phosphate buffer, pH 7 and centrifuged at 4°C for 10 minutes at 10,000g. An aliquot of 1ml of supernatant of the enzyme extract was added to the reaction mixture containing 1ml of 0.01 M H₂O₂, 3ml of 0.1M sodium phosphate buffer, pH 6.8. The reaction was stopped after an incubation of 5 minutes at 20°C by addition of 10ml of 1 per cent H₂SO₄. The acidified medium without or with the enzyme extract was titrated against 0.005N KMnO₄.

Peroxidase (POX, EC 1.11.17) activity was assayed by the method of Kumar and Khan (1971) and Superoxide dismutase (SOD, EC 1.15.1.1) activity by Beauchamp and Fridouich (1971). 1g of leaves were homogenized with 20ml of ice cold extraction medium containing 2M MgCl₂, 1mM EDTA, 10mM β-mercaptoethanol, 7%t PVP and 10 mM sodium metabisulphate. The homogenate was strained through two layers of cheesecloth and centrifuged at 10,000 g for 5 minutes and the supernatant was made up to 20ml with the same buffer and it was used as the source of enzyme.

2.4. Polyphenoloxidase (EC 1.10.3.1)

Polyphenoloxidase activity was assayed by the method of Kumar and Khan (1982). Assay mixture of polyphenoloxidase contained 2ml of phosphate buffer (pH 6.0) 1ml of 0.1M catechol 3.6 and 0.5 ml of enzyme extract. This was incubated for 5 minutes at 25°C, and then the reaction was stopped by adding 1ml of 2.5 N sulphuric acid. The absorbance of the purpurogallin formed was recorded at 495nm. The enzyme activity was expressed in units. One unit is defined as the amount of purpurogallin formed which raised the absorbance by 0.1 per minute under the assay condition.

2.5. Phenylalanine ammonia-lyase (EC 4.3.1.5)

Phenylalanine ammonia-lyase was determined by following the method of Bruseke (1980). 500 milligram fresh leaves were homogenized in 5 ml at cold 25 mM mercaptoethanol and centrifuged at 12,000 g for 20 minutes. The supernatant was used for assay. An aliquot of 0.2 ml of enzyme extract was added with 0.5 ml borate buffer and 1.3 ml distilled water. The reaction was initiated by adding 1ml of 0.1 m phenylalanine solution and incubated for 30–60 minute at 32°C. After incubation the reaction was terminated by adding 0.5 ml 1M trichloroacetic acid and the absorbance at 290 nm was measured against blank. The reaction rate was expressed as micro mole transcinamic acid formed per mg protein per minute.

2.6. Statistical Analysis

Each value is the mean of five replicate experiments (± SE). The analysis was carried out using statistical package SPSS. All data were subjected to one-way analysis of variance (ANOVA) and the significance of difference between control and each treatment was analyzed using student's t test. Levels of significance used were P<0.05.

3. Results and Discussion

3.1. Growth Parameters

Ambient UV radiation exerted a clear effect on the growth parameters, biochemical and antioxidant enzymes of *Indigofera tinctoria* seedling. In our study, foliar symptoms such as glazing, wrinkling, chlorotic lesions and necrosis were observed. Similar observations were also noticed by Feng et al (2007). This foliar damage might be due to reduction in photosynthetic pigments (Teramura, 1983) and cell and tissue damage in the upper epidermis (Teramura, 1980). According to Teramura (1980) interveinal wrinkling and leaf chlorosis were observed in UV-B treated plants. These observations conclude that interveinal wrinkling is the result of UV-B effect on cell division and development.

In the present study after eight days of UV-B treatment the shoot length, fresh weight, dry weight and leaf area were reduced to 23.44%, 37.3%, 28.9% and 28.7% respectively. However, leaf thickness increased with increasing time after UV-B radiation treatment (Table 1). The highest increase of 16 per cent was observed after 8 days in treated seedling. Krause et al (2007) observed that UV-B radiation reduced the height of spring wheat. However, field study indicates that plant growth is less sensitive to UVB radiation. Teramura and Murali (1986) observed that the reduction on plant growth under UV-B radiation in soybean is between 26 and 38 per cent in green house and between 11 to 22 per cent in field study. Increase in shoot length is due to IAA, which absorbs in the UV-B range and is readily destroyed by UV-B *in vitro* and *in vivo* (Tevini et al., 1990).

Table 1. Effect of UV-B radiation on shoot length, fresh weight, dry weight, leaf area and leaf thickness in *Indigofera tinctoria* L. Seedlings.

Day(s)	Treatments	Shoot length (cm)	Fresh weight (g plant ⁻¹)	Dry weight (g plant ⁻¹)	Leaf area (cm ² plant ⁻¹)	Leaf thickness (mm)
2	Control	3.6 ± 0.18	0.42* ± 0.021	0.25 ± 0.01	0.32 ± 0.02	0.07 ± 0.03
	UV-B	3.9 ± 0.19	0.45 ± 0.023	0.26* ± 0.01	0.31 ± 0.02	0.08 ± 0.03
4	Control	5.8* ± 0.29	0.50 ± 0.03	0.29 ± 0.02	0.35* ± 0.02	0.08 ± 0.03
	UV-B	5.1 ± 0.25	0.47 ± 0.03	0.28 ± 0.01	0.33 ± 0.02	0.10* ± 0.04
6	Control	7.5* ± 0.38	0.55* ± 0.03	0.33* ± 0.02	0.40* ± 0.03	0.85 ± 0.045
	UV-B	5.8* ± 0.29	0.49 ± 0.03	0.31* ± 0.02	0.35* ± 0.02	1.09 ± 0.07
8	Control	8.9* ± 0.44	0.64* ± 0.04	0.38* ± 0.03	0.46* ± 0.03	0.92* ± 0.06
	UV-B	6.1* ± 0.31	0.52* ± 0.03	0.32* ± 0.02	0.37* ± 0.03	1.18* ± 0.08

Results are means ± S.E. of 5 replicates. Significant level (p) for Student 't' test is shown of * p<0.05

UV-B radiation significantly depressed the biomass accumulation of many species such as rice (Pal et al., 1998), *Vigna mungo* (Hofmann, 2000) and *Trifolium repens* (Pal et al., 1999). The decrease in dry weight observed in the present study may be caused by reduced photosynthesis rates and enzyme activities. The changes in the leaf area measurements observed under supplemental UV-B radiation were similar to those seen for fresh and dry weight. In musfard, leaf area reduction was observed under UV-B treatment (Santos et al., 2004). Similar trend in leaf area reduction have also been observed in potato (Gao et al., 2003), cotton plants (Krizek, 1997) and spring wheat (Vu et al., 1981). Reduction in leaf area could be an adaptive mechanism to minimize the exposure to UV-B radiation. There are also report correlating the tolerance of the species to their ability to increase leaf thickness in response to UV-B exposure (Vu et al., 1981; Campbell, 1975).

3.2. Biochemical Constituents

The supplemental UV-B radiation significantly reduced the chlorophyll content (20.1%) and carotenoid (30.3%) (Table 2) throughout the study period except during the initial period. The chloroplast was the first organelle to show injury response when irradiated with UV-B radiation. In a field experiments with *Vigna radiata*, Pal et al. (1999) observed an initial increase and subsequent decrease in chlorophyll content, which was also reflected in the present study. In our study, an initial increase in chlorophyll was observed and this might be due to accumulation of UV-B absorbing pigments. The carotenoid content gradually declined due to UV-B radiation treatment. Previous work demonstrated that carotenoid content decreased under UV-B radiation (El-Mansy and Salisbury, 1971). The reduction in carotenoid content may result either from inhibition of synthesis or from breakdown of the pigments (Stapleton and Walbot, 1992). Since the carotenoids are involved in the light harvesting and protection of chlorophyll from photo-oxidative destruction, any reduction in carotenoid could have serious consequences of chlorophyll pigments.

Table: 2. Effect of UV-B radiation on Chlorophyll, Carotenoid, Anthocyanin and Flavonoid in *Indigofera tinctoria* L. Seedlings

Day (s)	Treatments	Chlorophyll (mg g ⁻¹ fr. wt.)	Carotenoid (mg g ⁻¹ fr.wt.)	Anthocyanin (A ₅₃₀ g fr.wt.)	Flavonoid (A ₃₁₅ g fr. wt.)
2	Control	1.82 ± 0.99	0.12 ± 0.06	0.03 ± 0.01	0.100 ± 0.05
	UV-B	1.83 ± 0.99	0.13* ± 0.07	0.05 ± 0.02	0.200* ± 0.10
4	Control	1.91* ± 0.09	0.135* ± 0.07	0.037 ± 0.02	0.115* ± 0.05
	UV-B	1.84* ± 0.10	0.132 ± 0.08	0.070* ± 0.03	0.280* ± 0.14
6	Control	2.12* ± 0.12	0.148* ± 0.08	0.044* ± 0.02	0.123* ± 0.06
	UV-B	1.95* ± 0.10	0.138* ± 0.09	0.097* ± 0.05	0.356* ± 0.17
8	Control	2.40* ± 0.13	0.162* ± 0.09	0.054 ± 0.02	0.133* ± 0.077
	UV-B	2.03* ± 0.11	0.145* ± 0.09	0.132* ± 0.06	0.448* ± 0.02

Results are means ± S.E. of 5 replicates. Significant level (p) for Student 't' test is shown of *p<0.05

3.3. UV- Absorbing pigments

Increased anthocyanin and flavonoid content was observed in UV-B treated seedling. The highest accumulation of anthocyanin (84%) and flavonoids (91%) were noticed in UV-B treated seedling after eight days of treatment when compared to control seedlings (Table 2). Anthocyanin concentration was significantly increased in UV-B radiation treatment when compared to control seedlings. Several studies have shown that increased anthocyanin was mainly due to UV-B irradiation effect (Ntefido and Manetas, 1996; Ravindran et al., 2001). Ravindran et al., (2001) demonstrated over 171% increase in anthocyanin content in a halophyte species, *Suaeda maritima* under field study. Ambasht and Agarwal (1998) observed a more than 275% increase in the anthocyanin content in maize. Anthocyanin has very weak absorption in the UV-B regions and regarded as UV screens only at very high concentration. Olssen et al., (1999) suggested that UV-B induced accumulation of anthocyanin which protects the photosynthetic apparatus from the damaging effect of UV-B radiation. Similar to anthocyanin, flavonoid concentration was also increased in UV-B treated seedlings after eight days of treatment. In general, the flavonoid accumulation is linearly depends on UV-B influence (Vu et al., 1981; Baumbusch et al., 1998). Feng et al (2007) concluded that flavonoid concentration can reduce the UV-B penetration and protect the photosynthetic apparatus to some extent depending on a threshold level, which may vary in different species. Flavonoid accumulation is also considered a defence mechanism against UV-B radiation and protects the mesophyll tissue through epidermal screening. UV-B screening by epidermal flavonoids is often proposed as an adaptive mechanism to prevent this radiation from reaching the mesophyll.

Table: 3. Effect of UV-B radiation on Catalase, Peroxidase, Polyphenol oxidase, superoxide dismutase and phenylalanine ammonia-lyase in *Indigofera tinctoria* L. Seedlings

Day (s)	Treatments	Catalase (μ mol H ₂ O ₂ decomposed min ⁻¹ g ⁻¹ fr.wt.)	Peroxidase (μ mol pupurogallin formed min ⁻¹ g ⁻¹ fr.wt.)	Polyphenol oxidase (μ mol pupurogallin min ⁻¹ g ⁻¹ fr.wt.)	Superoxide dismutase (μ mol annamic acid min ⁻¹ g ⁻¹ fr.wt.)	Phenylalanine ammonia-lyase (units h ⁻¹ mg ⁻¹ protein fr.wt.)
2	Control	7.8 ± 0.39	22.6 ± 1.13	19 ± 0.95	12.7 ± 0.64	8.4 ± 0.42
	UV-B	6.9 ± 0.34	28.9 ± 1.45	22 ± 1.10	14 ± 0.70	11.2 ± 0.56
4	Control	9.3* ± 0.46	26.4* ± 1.32	22.7* ± 1.14	14.9 ± 0.75	9.9 ± 0.49
	UV-B	8.1* ± 0.40	36.4* ± 1.82	32.5* ± 1.63	18.2* ± 0.91	14.8* ± 0.74
6	Control	10.3 ± 0.52	27.4* ± 1.37	23.5* ± 1.18	15.6* ± 0.78	10.2* ± 0.51
	UV-B	8.4* ± 0.42	40.9* ± 2.05	33.5* ± 1.68	21.5* ± 1.08	16.4* ± 0.82
8	Control	11.6* ± 0.58	28.5 ± 1.43	25.1* ± 1.26	16.5* ± 0.83	10.9* ± 0.55
	UV-B	8.7* ± 0.44	47* ± 2.35	38* ± 1.90	24.5* ± 1.23	19.1* ± 0.96

Results are means ± S.E. of 5 replicates. Significant level (p) for Student 't' test is shown of *p<0.05

3.4. Antioxidant enzymes

Most physiological stresses including UV-B enhancement disturb plant metabolism and cause oxidative injury by enhancing the production of reactive oxygen species. The metabolism of reactive oxygen species depend on low molecular anti-oxidant systems as well as enzymes such as super oxide dismutase, peroxidase, polyphenoloxidase and phenylalanine ammonia-lyase. In the present study the catalase activity was inhibited under supplemental UV-B radiation treatment. The highest decrease of catalase activity (22.7%) was observed after eight days in treated seedlings. In contrast to catalase activity, peroxidase, polyphenoloxidase, superoxide dismutase and phenylalanine ammonia-lyase activities were increased (36.5%, 39.9%, 41.5% and 40.7% respectively) under supplemental UV-B radiation (Table 3). This reduction trend in catalase activity under UV-B radiation was also observed

previously by Baumbush et al (1998) and Yang et al. (2007). Catalase is the most efficient antioxidant enzyme which protects plants by scavenging free radicals and H₂O₂ (Gao et al., 2008). Decreased catalase activity in the *vtc1* mutants of *Arabidopsis thaliana* during the course of the UV-B exposure experiment was observed and it could be due to destruction of the peroxisome via rampant lipid peroxidation (Yannarelli et al., 2006).

Along with catalase activity, peroxidase activity was also an important component of antioxidant defence system for scavenging H₂O₂. In the present study, the peroxidase activity increased with increasing treatment period of supplemental UV-B radiation. Sheen and Calvert (1969) demonstrated that phenol-oxidizing peroxidase unlikely contribute to UV tolerance as a result of their oxygen radical scavenging activity.

Polyphenoloxidase is also responsible for the oxidation of phenolic compound as reported by Jansen et al. (2001). Several studies have shown increased polyphenoloxidase due to UV-B irradiation (Balakrishnan et al., 2005; Santos et al., 1999). Enhanced UV-B radiation activates antioxidant enzymes and induces polyamines and causes leaf damage exemplified by an increase in polyphenoloxidase activity and a decrease in chlorophyll concentration. Superoxide dismutase showed a linear increase in activity throughout the study period. UV-B caused 325% increase in superoxide dismutase activity when compared to control (Krizek et al., 1993). Similar to superoxide dismutase, phenylalanine ammonia-lyase activity also increased under supplemental UV-B radiation treatment. Studies in cucumber (Beggs et al., 1985) have shown that exposure of seedlings to supplemental UV-B radiation caused 78% increase in the activity of phenylalanine ammonia-lyase (Braun and Tevini, 1993). Phenylalanine ammonia-lyase is an important enzyme in regulating flavonoid biosynthesis and transcriptionally induced by UV-radiation. Thus, it is concluded from the present study that the increase in phenylalanine ammonia-lyase activity stimulates the synthesis of flavonoid and anthocyanin.

4. Conclusion

Our results show that after eight days of treatment, *Indigofera tinctoria* exhibits a different sensibility to supplemental UV-B radiation. UV-B adversely affected the growth parameters and photosynthetic pigments. However, accumulation of UV-B absorbing compounds and antioxidant enzymes content indicates that supplemental UV-B radiation induces a photosynthetic protection mechanism in a short time growth period. Finally, it suggests that antioxidant enzymes and UV-B absorbing compounds provide protection during oxidative injury. The balance between the antioxidant enzymes and UV-absorbing compounds contribute the growth and development of *Indigofera tinctoria* seedling under the oxidative stress condition.

References

- Agawal, S.B., Rathore, D., 2007. Changes in oxidative stress defense in wheat (*Triticum aestivum* L.) and mung bean (*Vigna radiata* L.) cultivars grown with and without mineral nutrients and irradiated by supplemental ultraviolet- B, *Environ. Exp. Bot.* Vol.59, No.5, pp.21-33.
- Ambasht, N.K, Agrawal, M., 1998. Physiological and biochemical responses of *orghum vulgare* plants to supplemental ultraviolet- B radiation, *Can. J. Bot.*, Vol.76, pp. 1290-1294.
- Balakrishnan, V., Venkatesan, K., Ravindran, K.C., Kulandaivelu, G., 2005. Protective mechanism in UV-B treated *Crotalaria juncea* L. seedling, *Plant Protect. Sci.*, Vol.41, No.6, pp.115-120.
- Baumbusch, L.O, Eiblmeir, M., Schnitzler, J.P, Heller, W., Sandermann, H., Polle, A., 1998. Interactive effects of ozone and UV-B radiation on antioxidants in spruce (*Picea abies*) and Pine (*Pinus sylvestris*) needles, *Physiol. Plant*, Vol.104, No.3, pp.248-254.
- Beauchamp, C.O., Fridovich, I., 1971. Superoxide dismutase: Improved assays and an assay applicable to acrylamide gel, *Anal. Biochem.*, Vol.44, No.8, pp.275-287.
- Beggs, C.J., Wellmann, E., 1985. Analysis of light controlled anthocyanin formation in coleoptiles of *Zea mays* L. the role of UV-B, blue, red and far-red light, *Photochem. Photobiol.* Vol.41, No.7, pp. 481-486.
- Braun, J., Tevini, M., 1993. Regulation of UV protective pigment synthesis in the epidermal layer of rye seedlings (*Secale cereale* L. CN. Kustro). *Photochem. Photobiol.* Vol.57, No.4, pp. 318-323.
- Bruseke, C.H., 1980. Effect of UV-B radiation on leguminous plants, *Physiol. Plant Pathol.*, Vol.16, No.1, p 409.
- Caldwell, C.R., Britz, S. J., 2006. Effect of supplemental ultraviolet radiation on the carotenoid and chlorophyll composition of green house-grown leaf lettuce (*Lactuca sativa* L.) cultivars, *Journal of Food Composition and Analysis*, Vol.19, No.8, pp. 637-644.
- Caldwell, M.M., Bjorn, L.O., Bomman, J.F., Flint, S.D., Kulandaivelu, G., Teramura, M, Tevini, M., 1998. Effect of increased solar ultraviolet radiation on terrestrial ecosystem, *Journal of Photochemistry and Photobiology*. Vol.46, No.5, pp. 40-52.
- Campbell, W.S., 1975. Ultraviolet induced ultra-structural changes in Mesophyll cells of *Glycine max*. In: Impacts of climatic changes on the Biosphere, part I. Ultraviolet radiation effects. (D.W. Nachtwey, M.M. Caldwell, R.H. Biggs, eds.). Monogr. 5 Climatic Impact Assessment Program, US Dept. Transportation Rep No. DOT-IST-75-55. Nat. Tech Info. Serv, Springfield, VA, pp. 167-176.
- El-Mansy, H.L., Salisbury, F.B., 1971. Biochemical responses of *Xanthium* leaves to ultraviolet radiation, *Radiat. Bot.* Vol.11, No.8, pp.325-328.

- Feng, H., An L., Tan, L., Hou wang, X., 2000. Effect of enhanced ultraviolet-B radiation on pollen germination and tube growth of 19 taxa in vitro, *Environ. Exp. Bot.* Vol.42, No.12, pp.45-43.
- Feng, H., Shiwen Li., Xue L., An, L., 2007. Xunling wang, The interactive effects of enhanced UV-B radiation and soil drought on spring wheat, *South African Journal of Botany*, Vol.73, No.10, pp.429-434.
- Gao, Q., Zhang, L., 2008. Ultraviolet-B- Induced oxidative stress and antioxidant defense system responses in ascorbate deficient vtc 1 mutants of *Arabidopsis thaliana*, *Journal of Plant Physiology*, Vol.165, No.12, pp.138-148.
- Gao, W., Zhen, Y., Slusser, J.R., Gordon, M., 2003. Heisler, Impact of enhanced ultraviolet- B irradiance on cotton growth, development, yield, and qualities under field conditions, *Agricultural and Forest Meteorology*, Vol.120, No.5, pp.241- 248.
- He, D.L., Wong, C.H., He, Y.H., 2003. The effect of reduction of ultraviolet -B radiance on the content of flavonoid in leaves of wheat, *Chinese Journal of Agrometeorology*, Vol.24, No.4, pp.32.
- Hofmann, R.W., Swinny, E.E., Bloor, S.J., Markham, K.R., Ryan, K.G, Campbell, B.D., Jordan, B.R., Fountain, D.W., 2000. Responses of nine *Trifolium repens* L. populations to ultraviolet-B radiation: Differential flavonol Glycoside accumulation and biomass production, *Annals of Botany*, Vol.86, No.4, pp. 527-537.
- Inskeep, W.P., Bloom, P.R., 1985. Extinction co-efficient of chlorophyll 'a' and 'b' in N N¹ – Dimethyl formamide and 80 per cent acetone, *Plant Physiology*, Vol.77, No.9, pp. 483-485.
- Jansen, M.A.K., Vanden Noort, R.E., Tan, M.Y.A., Prinsen, E., Lagrimini, L.M., Thorneley, R.N.F., 2001. Phenol oxidizing peroxidases contribute to the protection of plants from ultraviolet radiation stress, *Plant Physiol.*, Vol.126, No.2, pp. 1012-1023.
- Krause, G.H., Jahns, P., Vigor, A., Garcia, M., Aranda, J., Wellmann, E., Winter K., 2007. Photoprotection, photosynthesis and growth of tropical tree seedlings under near-ambient and solar reduced solar ultraviolet-B radiation, *Journal of Plant Physiology*, Vol.164, No.3, pp.1311-1322.
- Krizek, D.T., Hramer, G.P., Upadhyaya, A., Mirecki, R.M., 1993. UV-B response of cucumber seedling grown under metal halide and high pressure sodium/ deluxe lampe, *Physiologia Plantarum*, Vol.88, No.5, pp.350-358.
- Krizek, D.T., Kramer, T., Mirecki, R. M., 1997. Influence of UV-B radiation and putrescine on shoot and root growth of cucumber seedlings grown in nutrient solution, *Journal of Plant Nutrition*, Vol.20, No.5, pp. 613-623.
- Kumagai, T., Hidema, J., Kang, H.S., Sato, T., 2001. Effects of supplemental UVB radiation on the growth and yield of two cultivars of Japanese low land rice (*Oryza sativa* L.) under the field in a cool rice growing region of Japan, *Agriculture, Ecosystems and Environment*, Vol.83, No.5, pp.201-208.
- Kumar, K.B., Khan, P.A., 1982. Peroxidase in excised ragi (*Eleusine coracana* Cv. PR. 202.) leaves during senescence, *Indian Journal of Experimental Botany*, Vol.20, No.8, pp.412-416.
- Liang, B., Huang, X., Zhang, G., Zhou, Q., 2006. Effect of Lanthanum on plant under supplementary ultraviolet-B radiation: Effect of Lanthanum on flavonoid contents on soybean seedling exposed to supplementary ultraviolet-B radiation, *Journal of Rare Earths*, Vol.24, No.5, pp. 613- 616.
- Machly, A.C., Chance, B., 1959. The assay of catalase and peroxidase. In: methods of biochemical analysis. Glich, D. (Ed.), Interscience publishers Inc., New York, Vol.1 No.3, pp.357-425.
- Mancinelli, A.L., Yang, C.P.H., Lindquist, P., Anderson, O.R., Rabino, J., 1975. Photoregulation of anthocyanin synthesis III. The action of streptomycin on the synthesis of chlorophyll and anthocyanin, *Plant Physiology*, Vol.55, No.4, pp.251-254.
- Mirecki, R.M., Teramura, A.H., 1984. Effects of ultraviolet- B irradiance on soybean. The dependence of plant sensitivity on the photosynthetic photon flux density during and after leaf expansion. *Plant Physiology*, Vol.74, No.8, pp.475-480.
- Mishra, S., Agarwal, S.B., 2006. Interactive effects between supplemental UV-B radiation and heavy metals on growth and biochemical characteristics of *Spinacia oleracea* L., *Braz. J. Plant Physiol.* Vol.18, No.5, pp.1-8.
- Moran, R., Porath, D., 1980. Chlorophyll determination in intact tissue using N, N¹ – dimethyl formamide, *Plant Physiology*, Vol.65, No.4, pp. 478-479.
- Ntefido, M., Manetas, Y., 1996. Optical properties of hairs during the early stages of leaf development in *Plantanus orientalis* Aus. *Plant Physiol.* Vol.117, No.6, pp.173-181.
- Olsson, L.C., Veit, M., Borman, J.F., 1999. Edidermal transmittance and phenotic composition in leaves of altrazne tolerant and sensitive cultivars of *Brassica napus* grown under enhanced UV-B radiation, *Physiol. Plant*, Vol.107, No.4, pp.259-266.
- Pal, M., Jain, V., Sengupta, U.K., 1998. Influence of enhanced UV-B radiation on mustard: Cultivar response, *Ind. J. Plant Physiol.*, Vol.3, No.1, pp.188-193.
- Pal, M., Sengupta, U.K., Srivastava, A.C., Jain, V., Meena, R.C., 1999. Changes in growth and photosynthesis of mung bean induced by UV-B radiation, *Ind. J. Plant. Physiol.* Vol.4, No.5, pp. 79-84.
- Pal, M., Sengupta, U.K., Srivastava, A.C., Jain, V., Meena, R.C., 1999. Changes in growth and photosynthesis of mung bean induced by UV-B radiation, *Ind. J. Plant. Physiol.*, Vol.4, No.5, pp.79-84.
- Ravindran, K.C., Maheshkumar, N., Amirthalingam, V., Ranganathan, R., Chellappan, K.P., Kulandaivelu, G., 2001. Influence of UV-B supplemental radiation on growth and pigment content in *Suaeda maritima* L. *Biol. Plant*, Vol.44, No.5, pp.465-469.
- Santos, I., Almedia, J., Salema, R., 1999. The influence of UV-B radiation on the superoxide dismutase of maize, potato, sorghum and wheat leaves, *Can. J. Bot.*, Vol.77, No.5, pp. 70-76.
- Santos, I., Fidalgo, F., Almeida, J.M., Salema, R., 2004. Biochemical and Ultrastructural changes in leaves of potato plants grown under supplementary UV-B radiation, *Plant Science*, Vol.167, No.8, pp.925-935.

- Selvakuamr, V.,2008.Ultraviolet-B radiation (280-315 nm) invoked antioxidant defense systems in *Vigna unguiculata* (L.) Walp. and *Crotalaria Jancea* L, *Photosynthetica*, Vol. 46, No.5, pp.98-106.
- Sheen, S.J., Calvert, J.,1969. Studies on polyphenol content activities and isoenzymes of polyphenol oxidase and peroxidase during air-curing in three tobacco types, *Plant Physiol.* Vol.44, No.3, pp. 199-204.
- Siefermann, D., Harms, H.,1987. The last harvesting and protective function of carotenoids in photosynthetic membranes. *Physiol plant.* Vol.69,No.6, pp.51-568.
- Smirnoff, N.,1998.Plant resistance to environmental stress, *Curr. opin. Biotech.*, Vol.9,No.5,pp.214-219.
- Stapleton, A.E., Walbot, U.,1992. Anthocyanin protects DNA from UV-B damage, Maize Genetics Cooperation Newsletter, Vol.66,No.5, p.102.
- Taalas, P., Kaurola, J., Kylling, A., Shindedll, D., Sausen, R., Dameris, M., Grewe, V., Herman, J.,2000. The impact of green house gases and halogenated species on the future solar UV radiation doses, *Geophys. Res. Lett.*, Vol.27, No.8, pp.1127-1130.
- Teramura, A.H., Murali, N.S.,1986. Intraspecific differences in growth and yield of soybean exposed to ultraviolet-B radiation under green house and field conditions, *Environ. Exp. Bot.* Vol.26,No.5, pp. 89-95.
- Teramura, A.H.,1980. Effects of ultraviolet-B irradiances on soybean, Importance of photosynthetically active radiation in evaluating ultraviolet-B irradiance effects on soybean and wheat growth, *Physiol. Plant*, Vol.48 ,No.8,pp.333-339.
- Teramura, A.H.,1983. Effects of ultraviolet-B radiation on the growth and yield of crop plants, *Physiol. Plant*, Vol.58,No.5, pp.415-427.
- Tevini, M., Mark, U., Saile, M.,1990. Plant experiments in growth chambers illuminated with natural sunlight. In: Environmental research with plants in closed chambers, Air Pollution Research Report No. 26. Commission of the European Communities, Belgium: pp. 240-251.
- Vichnevetskaia, K.D., Roy, D.N.,2001. Oxidative stress and antioixdative defense with an emphasis on plants antioxidants, *Environ. Rev.*, Vol.7,No.2,pp.31-35.
- Vu, C.V., Allen, L.H., Garrard, L.A.,1981. Effects of supplemental UV-B radiation on growth and leaf photosynthetic reaction of soybean, *Physiol. Plant*, Vol.52 ,No.5,pp.353-362.
- Yang, S.H., Wang, L.J.,2007. freezing tolerance in relation to antioxidant system in winter wheat (*Triticum destivum* L.) leaves. *Environ. Exp. Bot.* Vol.60,No.4, pp. 300-307.
- Yannarelli, G.G., Noriega, G.O., Batlle, A., Tomaro, M.L.,2006. Heme- oxygenize up-regulation in ultraviolet-B irradiated soybean plants innvalves reactive oxygen species plant, *Environ. and Expt. Botany*, Vol.224 ,No.5,pp.1154-62.
- Zhang, M., An, L., Feng, H., Chen, T., Chen, K., Liu, Y., Tang, H., Wang, C.H.,2003.The cascade mechanisms of nitric oxide as second messenger of ultraviolet – B in inhibiting mesocotyl elongation, *Photochemistry and Photobiology*, Vol,77,No.8,pp. 219-225.

Biographical notes

Dr.K.C.Ravindran is working as a Reader in Botany, Faculty of Science, Annamalai University, Annamalinagar, Tamilnadu, India. He got his Doctoral degree from Madurai Kamaraj University, Palakali nagar, Madurai under the Supervision of Dr.G.Kulandaivelu. He has produced Three Ph.D.Research Scholars and presently Supervising two Ph.D. Research Scholars in the area of saline Soil Reclamation in Tsunami affected soils. His area of specialization is Light intensity in spice plants, UV-B radiation on Legumionus plants. He has produced 14 M.Phil Research Scholars in the branch of Botany.

A.Indrajith is working as a Ph.D research Scholar in Botany, Faculty of Science, Annamalai University, Annamalinagar, Tamilnadu, India.

P.V.Pratheshh was worked as a Ph.D research Scholar in Botany, Faculty of Science, Annamalai University, Annamalinagar, Tamilnadu, India.

K.Sanjiviraja is working as a Ph.D research Scholar in Botany, Faculty of Science, Annamalai University, Annamalinagar, Tamilnadu, India.

Dr.V.Balakrishnan is working as an Assistant Professor in the Department of Biotechnology, K.S.Rangasamy College of Technology,Tiruchengode-637 215,Tamilnadu,India.