

Reprinted from

**International Journal  
of  
Health Research**

**Peer-reviewed Online Journal**

<http://www.ijhr.org>

---

---

**PORACOM**

Academic Publishers

---

# International Journal of Health Research

---

The *International Journal of Health Research* is a peer-reviewed online international journal allowing free unlimited access to abstract and full-text of published articles. The journal is devoted to the promotion of health sciences and related disciplines (including medicine, pharmacy, nursing, biotechnology, cell and molecular biology, and related engineering fields). It seeks particularly (but not exclusively) to encourage multidisciplinary research and collaboration among scientists, the industry and the healthcare professionals. It will also provide an international forum for the communication and evaluation of data, methods and findings in health sciences and related disciplines. The journal welcomes original research papers, reviews and case reports on current topics of special interest and relevance. All manuscripts will be subject to rapid peer review. Those of high quality (not previously published and not under consideration for publication) will be published without delay. The maximum length of manuscripts should normally be 10,000 words (20 single-spaced typewritten pages) for review, 6,000 words for research articles, 3,000 for technical notes, commentaries and short communications.

**Submission of Manuscript:** The *International Journal of Health Research* uses a journal management software to allow authors track the changes to their submission. All manuscripts must be in MS Word and in English and should be submitted online at <http://www.ijhr.org>. Authors who do not want to submit online or cannot submit online should send their manuscript by e-mail attachment (in single file) to the editorial office below. Submission of a manuscript is an indication that the content has not been published or under consideration for publication elsewhere. Authors may submit the names of expert reviewers or those they do not want to review their papers.

## *Enquiries:*

The Editorial Office  
International Journal of Health Research  
236-202, St David Court, Cockeysville,  
MD 21030, USA  
*E-mail:* [editor@ijhr.org](mailto:editor@ijhr.org)  
*Tel:* +1-614-535-7928

**PORACOM**  
Academic Publishers

## Original Research Article

*Open Access*  
Online Journal

# Comparative Determination of Chlorpromazine Hydrochloride Content in Multi-sourced Chlorpromazine Tablets in Nigeria

Received: 8-Feb-08

Revision received: 14-Feb-08

Accepted for publication: 5-Mar-08

## Abstract

**Purpose:** Although different brands or unbranded chlorpromazine tablets from multiple sources are available in Nigeria today, they must all contain the same active principle and satisfy the standards of quality, efficacy and safety. This study is designed to check for possible faking and adulteration of chlorpromazine tablets and also establish the possibility of inter-brand substitution based on drug content (chemical equivalence).

**Method:** The determination of the chlorpromazine hydrochloride content was carried using non-aqueous titrimetric and spectrophotometric methods. Glacial acetic acid and acetone were used as the non-aqueous solvent and equivalence points were determined using visual indicators and potentiometer.

**Results:** The results obtained showed that all the brands analyzed met the specification of the British Pharmacopoeia and so contain acceptable amounts of chlorpromazine drug content thus enabling possible brand substitution.

Statistical comparison showed no significant difference between the results obtained by determining end-point using visual indicators and the potentiometer.

**Conclusion:** The use of the visual indicator method is recommended for fast and accurate routine laboratory analytical work especially in developing nations.

**Keywords:** Chlorpromazine, non-aqueous titration, spectrophotometric determination, potentiometry

**Henry A Okeri**

**Peter O Alonge**

**Emadoye Etareri**

Department of Pharmaceutical Chemistry, Faculty of Pharmacy, University of Benin, Benin City, Nigeria.

### For Correspondence:

Tel: +234-802-311-2394

E-mail: [hokeri1@yahoo.com](mailto:hokeri1@yahoo.com)

## Introduction

Chlorpromazine (10-[3-dimethylaminopropyl] phenothiazine) belongs to the primary chemical group of antipsychotic agents known as the phenothiazines. It has an aliphatic side chain and is referred to as an atypical phenothiazine with a low to moderate-potency antipsychotic action. Apart from its antipsychotic activity where it is used as a tranquilizer and maintenance therapy to prevent acute relapse in chronic schizophrenic patients, chlorpromazine is also used for the treatment of vomiting and vertigo because of its sedative and extrapyramidal effects.<sup>1, 2</sup> Although chlorpromazine has been primarily replaced by newer generation of antipsychotic agents which have improved action and side effect profile, it is still used in the management of acute and chronic psychosis including schizophrenia and the manic phase of manic depression as well as amphetamine-induced psychosis, other serious psychiatric illnesses marked with agitation, and impaired reasoning.<sup>(2)</sup>

Chlorpromazine is highly lipophilic, membrane-bound, and protein-bound (especially albumin protein) and accumulate in the brain, lungs, and other tissues with a high blood supply.<sup>3</sup> The peak plasma concentration of chlorpromazine and other phenothiazines are attained within 2 – 4 hours and intramuscular administration helps to avoid much of the first-pass metabolism in the liver leading to increased bioavailability.<sup>4</sup> Chlorpromazine is mainly metabolized by oxidative processes mediated largely by the hepatic cytochrome-P450 microsomal oxidase and by conjugation processes.<sup>5</sup> Like all phenothiazine derivatives, chlorpromazine discolorizes under the influence of light and oxygen. Also it is readily oxidized in alkaline or neutral medium to the N-oxide derivative.<sup>6</sup>

Various analytical methods have been

used for the analysis of the compound in both pharmaceutical dosage forms and biological fluids. Some of the analytical methods that have been employed include thin-layer chromatography<sup>7</sup>, gas liquid chromatography<sup>8</sup>, gas chromatography/mass spectrometry<sup>9</sup>, high performance liquid chromatography<sup>10-12</sup>, spectroelectrochemical method<sup>13</sup>, spectrophotometry<sup>14</sup>, radioimmunoassay<sup>15, 16</sup> fluorimetry<sup>17</sup>, voltammetry<sup>18</sup>, chemiluminescence method<sup>19</sup>, electron spin resonance spectroscopy<sup>20</sup>, nuclear magnetic resonance (nmr) spectroscopy<sup>21</sup> and flow-injection potentiometric method.<sup>22</sup>

Presently, there is an increase in the number of generic drug products from multiple sources and this has placed the pharmacists and other health practitioners in a position of having to select one among several seemingly equivalent products. Most of the methods listed above employ expensive equipment not often available in poor countries. Non-aqueous titration is an approved assay method for chlorpromazine. The method was discontinued for the assay of the tablets because of interferences in the assay results due to formation of oxidation products in the presence of acetic acid. By overcoming this problem, non-aqueous titration is a reliable and cheap method for the assay of chlorpromazine tablets. Therefore the objective of this study is to develop a modified non-aqueous titrimetric method for the analysis of chlorpromazine tablets.

## Materials and Methods

### Sample selection and storage

Six different brands of chlorpromazine tablets labeled as DAL and CPE (with a label claim of 50mg of chlorpromazine HCl), as well as CPM, CPK, CPH and CPZ (with a label claim of 100mg of chlorpromazine HCl) were randomly purchased from pharmacies in Benin City, Nigeria. All the tablets were stored in air-tight, amber-coloured

containers kept in a dry cool place prior to the assay. Sampling and assay of chlorpromazine HCl in all the brands were done one and half years before the expiry dates.

### **Materials**

Glacial acetic acid, acetic anhydride, acetone, perchloric acid (70%) and hydrochloric acid all of Sigma-Aldrich (Germany); mercuric acetate, methyl orange powder and crystal violet crystals all of BDH (Poole, England), ascorbic acid was obtained from Merck (Germany), pH meter (Jenway, England, model 3020) and a Spectronic 21D UV/Visible spectrophotometer (Milton Roy Ltd., USA).

### **Preparation of solutions**

Perchloric acid (0.1M) that was standardized with potassium hydrogen phthalate, 2M and 1M HCl, crystal violet solution (0.2% w/v in acetic acid), mercuric acetate solution (5%w/v in acetic acid) and a saturated methyl orange solution in acetone were all prepared prior to the assay.

### **Assay methods**

#### **Non-aqueous titration using acetic acid:**

Twenty tablets of each brand were accurately weighed and reduced to fine powder. From the powdered drug, an amount equivalent to 0.35g of chlorpromazine HCl were taken and transferred into a 250ml conical flask containing 50ml of glacial acetic acid. The mixture was shaken and 10ml of 5% w/v mercuric acetate, 2g of ascorbic acid and 2 drops of 0.2% w/v crystal violet indicator were added. The solution was stirred for 15 min with a magnetic stirrer and then titrated with 0.1M acetous perchloric acid to a blue end-point. For the potentiometric method, the crystal violet indicator was omitted and the end-point was determined potentiometrically.<sup>23</sup> All the determinations were done in triplicates.

#### **Non-aqueous titration using acetone:**

A modification of the International Pharmacopoeia method<sup>24</sup> was used. Twenty tablets of each brand were accurately weighed and reduced to fine powder. From the powdered drug, an amount equivalent to 0.35g of chlorpromazine HCl was transferred into a 250ml conical flask containing 100ml of acetone. The mixture was shaken and 10ml of 5% w/v mercuric acetate and 3ml of methyl orange/acetone solution were added. The solution was stirred for 15 min with a magnetic stirrer and then titrated with 0.1M acetous perchloric acid to a blue end-point. For the potentiometric method, the methyl orange indicator was omitted and the end-point was determined potentiometrically. All the determinations were done in triplicates.<sup>24</sup>

#### **Spectrophotometric determination:**

Twenty tablets of each brand were accurately weighed and reduced to fine powder. From the powdered drug, an amount equivalent to 0.05g of chlorpromazine HCl was taken and transferred into a 500ml volumetric flask containing 5ml of 2M HCl and 200ml of distilled water. The mixture was shaken for 15 min with a magnetic stirrer and then diluted to the 500ml with distilled water. The mixture (50ml) was centrifuged at 1000 rpm for 15 min and 5ml of a clear supernatant was pipetted into a 100ml volumetric flask. Hydrochloric acid (10ml of 1M) was added to this solution and made up to the 100ml with distilled water. The absorbance of the resulting solution was then taken at 254nm against a reagent blank of 10ml 1M hydrochloric acid solution diluted to 100ml with distilled water. The content of chlorpromazine HCl was calculated and all the determinations were done in triplicates.<sup>25</sup>

### **Results and Discussion**

The results of the non-aqueous titration using acetic acid with mercuric acetate, acetone and spectrophotometry are given

**Table:** Assay of different brands of chlorpromazine HCl using non-aqueous titration and spectrophotometry

Brand	Label claim/ Manufacturer	Non-aqueous Titration using acetic acid		Non-aqueous Titration using acetone		Spectrophotometric determination (%)
		Visual Indicators (%)	Potentiometer (%)	Visual Indicators (%)	Potentiometer (%)	
DAL	50mg, Ernest Chemist ltd, Ghana	97.87 ± 0.23	98.12 ± 0.16	98.52 ± 0.31	99.07 ± 0.23	97.74 ± 0.34
CPE	50mg, England	99.83 ± 0.37	100.28 ± 0.29	100.67 ± 0.27	99.85 ± 0.18	98.33 ± 0.26
CPM	100mg, Kinapharma ltd, Ghana	94.97 ± 0.22	95.31 ± 0.17	95.33 ± 0.15	95.08 ± 0.20	94.71 ± 0.29
CPK	100mg, XL-Laboratory PVT ltd, India	98.75 ± 0.31	97.82 ± 0.24	99.45 ± 0.36	98.84 ± 0.28	97.17 ± 0.30
CPH	100mg, Medopharm malur-India	96.53 ± 0.21	95.88 ± 0.19	96.37 ± 0.23	96.05 ± 0.19	95.29 ± 0.25
CPZ	100mg, Hans- E-Jembcke, Germany	99.67 ± 0.27	100.28 ± 0.15	99.48 ± 0.25	99.71 ± 0.34	98.89 ± 0.21

in the Table. Some of the official monographs ((International Pharmacopeia <sup>24</sup>, British Pharmacopoeia <sup>25</sup> and United States Pharmacopoeia <sup>26</sup>) specified perchloric acid titration for the assay of chlorpromazine HCl. Results of the assay using acetic acid with mercuric acetate was not significantly different from those of the spectrophotometric assay method (2-tailed  $p < 0.05$ ; 95% confidence interval). All the brands passed the different assay tests since they were all within the acceptable range of 92.5 – 107.5%. <sup>25</sup>

The addition of mercuric acetate in the non-aqueous titrimetric method is based on the principle of removing the chloride counter ion so as to prevent the interference of the halide ion released by the titrant (acetous perchloric acid). The addition of mercuric acetate (which is undissociated in acetic acid) replaces the halide ion in chlorpromazine with a quantitative acetate ion which is a strong base in acetic acid. An intense red coloured oxidation product was formed when ascorbic acid was not added. This made end-point detection with visual indicators to be difficult.

The addition of ascorbic acid solution prevents the formation of the red colouration due to the reduction of ascorbic acid to dehydroascorbic acid. It has been shown that ascorbic acid and its oxidation product (dehydroascorbic acid) are neutral to acetous perchloric acid and do not interfere with the titration. <sup>23</sup> Thus, the addition of ascorbic acid allows titrations using a visual indicator and also sharpens the potentiometric end-point. When acetone was employed as the solvent in the place of glacial acetic acid, there was no colour formation which means that the oxidation of chlorpromazine does not occur in acetone. However, because of the questionability of the non-aqueous titrimetric method resulting from the catalyses of oxidation by the necessary addition of mercuric acetate, the British Pharmacopoeia (BP) later adopted the spectrophotometric method for the determination of chlorpromazine and other phenothiazines. This method we are reporting has overcome the problem for which non-aqueous titration was discontinued in the BP.

## Conclusion

A non-aqueous titrimetric method for the assay of chlorpromazine tablet, using glacial acetic acid or acetone as solvents, has been successfully developed. The method is simple, cheap and reliable, and produced assay results that are not significantly different from those of the official monographs assay methods. It can easily be used in the quality control of chlorpromazine dosage forms.

## References

1. Marder SR. Antipsychotic medications. In: The American Psychiatric Press Textbook of Psychopharmacology. Schatzberg AF and Nemeroff CB (eds.). American Psychiatric Press. Washington DC, 1998, pp. 309 – 321.
2. Malcolm L. Introduction to psychopharmacology. Upjohn company, Michigan, 1983, pp 51 – 61.
3. Baldessarini RJ, Tarazi FI. Drugs and the treatment of psychotic disorders; Psychosis and Mania. In: Goodman and Gilman's The Pharmacological Basis of Therapeutics. Hardman JG, Limbard LE and Gilman AG (editors). 10th International Edition. McGraw-Hill Medical Publishing Co., USA, 2001, pp 485 – 520.
4. Baldessarini RJ, Cohen BM, Teicher MH Significance of neuroleptic dose and plasma level in the pharmacological treatment of psychosis. Arch. Gen. Psychiatry, 1988; 45: 79 – 91.
5. Morselli PL. Psychiatric Drugs. In: Drug disposition during development. Morselli PL (ed.). Halsted Press, New York, 1977, pp 431 – 474.
6. Martindale: The Extra Pharmacopoeia 29<sup>th</sup> Edition. Reynolds EF (ed.). The Pharmaceutical Press, London, 1989, pp 722 – 725.
7. Chan TL, Sakalis G, Gershon S. Quantification of chlorpromazine and its metabolites in human plasma and urine by direct spectrodensitometry of thin-layer chromatogram. In: Advances in biochemical psychopharmacology. Raven Press, New York, 1974; 9: 305 – 333.
8. Mofatt AC. Clarke's isolation and identification of drugs. 2<sup>nd</sup> Edition. The Pharmaceutical Press, London, 1986, pp 460 – 461.
9. Gruenke LD, Craig JC, Klein FD, Nguyen TL, Hitzemann BA, Holaday JW, Loh HH, Braff L, Fischer A, Glick ID. Determination of chlorpromazine and its major metabolites by gas chromatography/mass spectrometry. Biomed. Mass Spectrom, 1985; 12(12): 707 – 713.
10. Onkubo T, Shimoyama R, Sugawara K. Determination of chlorpromazine in human breast milk and serum by high performance liquid chromatography. J. Chromatogr., 1993; 614(2): 328 – 332.
11. Erzen NK. Analytical procedure for the determination of chlorpromazine residue in muscle, tissue and urine of food-producing animals. Slov. Vet. Res., 2001; 38(4): 297 – 304.
12. Takahashi DM. Rapid Determination of Chlorpromazine Hydrochloride and Two Oxidation Products in Various Pharmaceutical Samples Using High-Performance Liquid Chromatography and Fluorimetric Detection. J. Pharm. Sci., 1980; 69(2): 184 – 187.
13. Daniel D, Gutz IG Spectroelectrochemical determination of chlorpromazine hydrochloride by flow-injection analysis. J. Pharm. Biomed. Anal., 2005; 37(2): 281 – 286.
14. Murty BSR, Baxter RM. Spectrophotometric determination of chlorpromazine in pharmaceutical dosage forms. J. Pharm. Sci., 1970; 59(7): 1010 – 1011.
15. Midha KK, Loo JCK, Hubbard JW, Rowe PL, McGilveray LJ. Radioimmunoassay for chlorpromazine in plasma. Clin. Chem., 1979; 25(1): 166 – 168.
16. Midha KK, Cooper JK, McGilveray, Bulterfield AG, Hubbard JW. High performance liquid chromatography assay for nanogram determination of chlorpromazine and its comparison with a radioimmunoassay. J. Pharm. Sci., 1981; 70(9): 104 – 106.
17. Kaul PN, Conway MW, Clark ML, Huffine J. Chlorpromazine metabolism 1: Quantitative fluorimetric method for 11 chlorpromazine metabolites. J. Pharm. Sci., 2006; 59(12): 1745 – 1749.
18. Dermis S, Biryol Y. Voltammetric determination of chlorpromazine hydrochloride. Analyst., 1989; 114(4): 525 – 526.
19. Shi W, Yang J, Huang Y. Ion-pair complex-based solvent extraction combined with chemiluminescence determination of chlorpromazine hydrochloride with luminol in reverse micelles. J. Pharm. Biomed. Anal., 2004; 37(2): 281 – 286.
20. Minakata K, Suzuki O, Ishikawa Y, Seno H, Asano M. Quantitative analysis of chlorpromazine by electron spin resonance (ESR) spectroscopy. Forensic Sci. Int. 1991; 50(2): 167 – 177.
21. Zarembo JE, Warren RJ, Staiger DB. Quantitative determination of chlorpromazine.HCl in tablets, spansules, injectables and bulk chemical by nuclear magnetic resonance spectroscopy. J. Assoc. Off. Anal. Chem., 1978; 61(1): 52 – 54
22. Sales MGF, Tomas JFC, Lavandeira SR. Flow injection potentiometric determination of chlorpromazine. J. Pharm. Biomed. Anal., 2006; 41(4): 1280 – 1286.
23. Soliman SA, Abdine H, Zakhari NA. Chemistry of non-aqueous titration of chlorpromazine. J. Pharm. Sci. 1975; 64(1): 129 – 132.
24. The International Pharmacopoeia. Quality specification. 3<sup>rd</sup> Edition, Vol 2. World Health Organization, Geneva, 1981, pp 74 – 75.
22. British Pharmacopoeia. Her Majesty's Stationery

*Okeri HA, Alonge PO, Etareri E. Chlorpromazine tablets assay*

Office, London. Vol III, 2003, pp 2183.

26. The United States Pharmacopoeia XX and National Formulary XV, 15<sup>th</sup> Edition. United States Pharmacopoeial Convention, Inc, Rockville, 1980, pp 142 – 143.