

Reprinted from

International Journal
of
Health Research

Peer-reviewed Online Journal

<http://www.ijhr.org>

PORACOM

Academic Publishers

International Journal of Health Research

The *International Journal of Health Research* is an online international journal allowing free unlimited access to abstract and full-text of published articles. The journal is devoted to the promotion of health sciences and related disciplines (including medicine, pharmacy, nursing, biotechnology, cell and molecular biology, and related engineering fields). It seeks particularly (but not exclusively) to encourage multidisciplinary research and collaboration among scientists, the industry and the healthcare professionals. It will also provide an international forum for the communication and evaluation of data, methods and findings in health sciences and related disciplines. The journal welcomes original research papers, reviews and case reports on current topics of special interest and relevance. All manuscripts will be subject to rapid peer review. Those of high quality (not previously published and not under consideration for publication) will be published without delay. The maximum length of manuscripts should normally be 10,000 words (20 single-spaced typewritten pages) for review, 6,000 words for research articles, 3,000 for technical notes, case reports, commentaries and short communications.

Submission of Manuscript: The *International Journal of Health Research* uses a journal management software to allow authors track the changes to their submission. All manuscripts must be in MS Word and in English and should be submitted online at <http://www.ijhr.org>. Authors who do not want to submit online or cannot submit online should send their manuscript by e-mail attachment (in single file) to the editorial office below. Submission of a manuscript is an indication that the content has not been published or under consideration for publication elsewhere. Authors may submit the names of expert reviewers or those they do not want to review their papers.

Enquiries:

The Editorial Office
International Journal of Health Research
Dean's Office, College of Medicine
Madonna University, Elele Campus, Rivers State
E-mail: editor_ijhr@yahoo.com or editor@ijhr.org

PORACOM
Academic Publishers

Review Article



Advances in Iontophoresis for Drug Delivery

Received: 06-Jun-08

Revision received: 26-Jun-08

Accepted for publication: 24-Jul-08

Abstract

Iontophoresis is an exciting technology that was initially investigated 250 years ago. Simply defined, it is the application of an electrical potential that maintains a constant electric current across the skin or barrier that enhances the delivery of ionized as well as unionized moieties. In the past few years, different types of iontophoresis such as transdermal, ophthalmic, transungual, buccal, ural and transnasal iontophoresis have been reported. Each system has its own advantages and drawbacks. The review summarizes recent findings and applications of various iontophoresis techniques.

Keywords: *Iontophoresis; transdermal; ocular; buccal; transungual; ural; drug delivery.*

MHG Dehghan

Md Ismail Mouzam *

Dept of Pharmaceutics, Y B Chavan College of Pharmacy, Dr Rafiq zakaria campus, Rauza bagh, Aurangabad-431001, Maharashtra State, India

***For Correspondence:**

Tel: +9198604408576;

E-mail: mdismail1111@rediffmail.com

Introduction

Iontophoresis is one of the most interesting and challenging endeavors facing the pharmaceutical scientist. The systemic drug delivery systems often require large dose and are associated with gastrointestinal side effects, while topical application of solutions, suspensions, and ointments show variability in absorption patterns. Iontophoresis technique is capable of expanding the range of compounds that can be delivered through ocular, transdermal, ural, transungual, buccal or by nasal route. One key advantage of iontophoresis is that it offers the possibility of externally controlled flux modulation, carefully adjusted to the needs of the patients¹⁻⁴. Various types of iontophoresis observed are discussed below.

Ocular Iontophoresis

Delivery of drug to the inner eye still presents a critical problem in ocular therapeutics. Topical administration cannot effectively reach the ocular fluids, where as systemic drug delivery has restricted access due to blood-aqueous and blood-retinal barriers. The retrobulbar and subconjunctival injections do not produce adequate drug levels, while direct intracameral or intravitreal delivery leads to intraocular complications⁵. Ocular iontophoresis could be a solution for such problems.

Ocular iontophoresis was first investigated in 1908 by the German investigator, Wirtz, who passed an electric current through electrolyte-saturated cotton sponges placed over the globe for the treatment of corneal ulcers, keratitis and episcleritis⁶. Basically, in ocular iontophoresis, a donor electrode is placed in the eye while another electrode is placed on the body surface to complete the electrical circuit. The drug to be delivered into the eye is loaded in the donor electrode. An electric field is applied across the eye to enhance the delivery of the drug into the eye⁷. It has been suggested that this technique is relatively easy, convenient for

use, safe, and provides fast and higher drug concentration in the specific ocular site, thereby providing solution to the low bioavailability of drugs. It is also well tolerated at low electric current density⁸⁻¹⁰. However, it causes minimum discomfort to patients and is not entirely harmless for ocular tissues¹¹⁻¹².

Iontophoresis has been evaluated for various drugs into the eye¹³⁻¹⁴. This technique can reproducibly deliver therapeutic concentrations of various ophthalmic drugs, such as corticoids, antibiotics, peptides, and proteins, to both segments of the eye. The drugs can be delivered either by transscleral or transcorneal iontophoresis. Transscleral iontophoresis presents more advantages when compared to transcorneal delivery, owing to scleral larger surface area, enhanced delivery of the drugs to the posterior segment and least possibilities of systemic absorption.

Transscleral Iontophoresis

In phakic animals, the lens-iris diaphragm limits penetration of a topically applied drug to the posterior tissues of the eye, such as the vitreous and retina. Transscleral iontophoresis overcomes this barrier and delivers drugs directly into the vitreous and retina through the choroids¹⁵. Transscleral iontophoresis of steroids (dexamethasone and methyl prednisolone) can be the alternative treatment for many ocular inflammations. Detailed pharmacokinetic studies have been performed on transscleral iontophoresis for various drugs¹⁶⁻²⁰. Every drug resulted with different patterns of distribution in the vitreous. Controlled distribution concentration for 1 to 6 hours into the vitreous chamber has been reported for carboplatin after transscleral iontophoresis. A previous study also revealed that the transscleral iontophoretic treatment can be used to obtain high concentrations of drugs to the posterior segments of the eye using a short transscleral iontophoresis with low current, without removing the conjunctiva²¹.

Several investigators have conducted clinical studies using transscleral iontophoresis of the anti-inflammatory corticosteroid, methylprednisolone hemisuccinate (SoluMedrol). The procedure was safe, well tolerated and easily applied for the treatment of severe ocular inflammation thereby reducing the systemic side effects of corticotherapy²².

Transcorneal Iontophoresis

The transcorneal iontophoresis has demonstrated good results for various investigations. Its application in the treatment of corneal ulcers offers a potentially effective method of management. Gentamicin, tobramycin and ciprofloxacin iontophoresis have resulted in significantly fewer bacterial colonies in the cornea compared with frequent eye-drops instillation²³⁻²⁶. Using this method, it has been revealed that after a short iontophoretic treatment of 1 mA for only 1 min with transcorneal iontophoresis of dexamethasone, a 30 fold higher concentrations in the cornea (1363.7F 436.3 Ag/g) is achieved when compared with the common treatment of frequent drop instillation every 5 min for 1 hour²⁷. Berdugo et al. have used transcorneal iontophoresis (300 μ A for 5 min) to enhance *in vivo* delivery of an AS-ODN against VEGFR-2 using rat cornea. The AS-ODNs penetrated into all corneal layers²⁸.

In earlier studies⁵, the β -blocking agents, timolol maleate and betaxolol hydrochloride, showed significantly increase in permeation through transcorneal iontophoresis. However, iontophoresis does not always show good result for penetration, as iontophoresis of vancomycin, a complex glycopolypeptide antibiotic, has resulted in poor corneal penetration compared with the other antibiotics, due to its high molecular weight (1448 Dalton) that highly influence the effectiveness of the iontophoretic drug delivery²⁹.

Voigt et al³⁰ used the combined transcorneoscleral iontophoresis to enhance

intraocular penetration of rat antinitric oxide synthase II oligonucleotides (anti-NOSII AS-ODNs) in the rat model of endotoxin-induced uveitis (EIU). The anti-NOSII AS-ODNs were detected intact in all the corneal layers, iris/ciliary body, peripheral retinal layers as well as conjunctiva and sclera 1 hour post-iontophoresis³⁰.

Preliminary toxicity to the eye after iontophoresis indicated a reversible inflammation with a current intensity of 5.1 mA/cm² for 2 min. Further, iontophoresis effect on the eye surface was evaluated by histopathology of the corneal tissue 5 min and 8 h after iontophoretic current of 0.5 and 1 mA for 1 and 2 min (2.5 and 5.1 mA/cm²). Minor reversible epithelial defects and stromal edema were found 5 min after the iontophoretic treatment, which disappeared or diminished 8 hours afterwards²⁷.

Iontophoretic Devices

Different eye-cup shapes exist³¹⁻³⁵, including an annular shape silicone probe for transscleral iontophoresis (called FEyegate, Optis, France) with a 13 mm opening to avoid contact with the cornea, used by Behar-Cohen³⁶, Hayden³⁷ and Voigt³⁸. The two approaches for drug retaining in the iontophoretic device are filling an eye cup with the drug solution, while a metal electrode extended from the current supply submerges into the solution. The eye cup has two ports: one delivers the drug solution and the other holds the metal electrode and aspirates air bubbles that can disrupt the current supply, thus creates a slightly negative pressure that maintains the applicator in place. The ground electrode is attached usually to the ear of the animal, as close as possible to the former electrode, to obtain minimal resistance. Another approach is using a drug saturated gel as the delivery probe. This method was first used by were Jones and Maurice³⁹, who delivered fluorescein into the anterior chamber of the eye using a fluorescein-saturated agar-gel. The gel was placed in a plastic tube and was

partly extruded from the tube to make a direct contact with the eye.

OcuPhor™ hydrogel drug delivery applicators have been studied in 24 male and female subjects⁴⁰. The applicators were well-tolerated and no clinically significant changes in symptomatology or in ophthalmic assessments were seen following exposure to 0–3.0 mA for 20 min or 1.5 mA for 40 min. At 4.0 mA 2 of 4 subjects reported a burning sensation⁴⁰.

Transdermal Iontophoresis

Transdermal iontophoresis is the application of an electrical potential that maintains a constant electric current across the skin and enhances the delivery of ionized as well as unionized moieties⁴¹. It offers various advantages such as easier termination of therapy, better control of drug delivery, improving delivery of polar drugs as well as high molecular weight substances, benefits of bypassing hepatic metabolism and reducing considerably the inter and intra-individual variability⁴²⁻⁴³ and ability to be used for systemic delivery or local delivery of drugs.

Transungual Iontophoresis

Nowadays, persons suffering from nail diseases are growing. Fungal infections in the nail can lead to severe health problems if left untreated in immuno-suppressed individuals⁴⁴. Many nail diseases are notoriously difficult to cure owing to the nail barrier and the deep-seated target site underneath the nail plate. Long treatments are usually needed and relapses are common⁴⁵. Oral drug delivery is somewhat successful in treating the nail disorders, but side effects may be severe due to considerable high doses required. Topical monotherapy is considered less efficient in treating nail disorders, such as onychomycosis, due to poor trans-nail bioavailability of drugs⁴⁶. There are two main factors that could limit the accumulation and activity of drugs in the nail on topical application. First, the

physicochemical properties of the drug need to be favorable for absorption through nail matrix. The nail matrix is relatively more permeable to polar compounds than nonpolar compounds⁴⁷⁻⁴⁸. Second, binding of the drug to keratin reduces the availability of the free drug⁴⁹⁻⁵⁰. Water solubility being one of the criteria for drug permeation across the nail, antifungal drugs that are poorly water-soluble do not achieve significant penetration across nail plate. Provided a suitable iontophoretic device could be designed and the electrical protocols are optimized, the transport of not only the ionic drugs but also uncharged drugs could be enhanced across the nail stratum.

The transport of glucose and griseofulvin across the human nail have been studied and the results clearly indicate that the nail plate exhibits iontophoretic permselectivity similar to human skin⁵¹. At pH > 5, the anodal iontophoretic transport is high due to the net negative charge on the nail plate which attracts the cations. The decrease in the anodal iontophoretic transport of glucose at pH < 5 that is often observed is most likely due to the reversal of net charge present on the nail plate at lower pH. In another study, Murthy et al⁵¹ reported that salicylic acid delivery across human nail, where the increase in the current density directly increases the number of ions moving across the barrier. Other factors which affects the delivery are the pH and drug concentration⁵².

Griseofulvin is a sparingly water-soluble antifungal drug with log P of 2.0⁵³. Therefore its ability to permeate into hydrophilic keratinous nail plate is limited. The transport of griseofulvin could be enhanced ~8 fold by iontophoresis. The iontophoresis enhanced transport of prednisolone sodium phosphate across the thumb nail has been reported⁵⁴⁻⁵⁵.

Buccal Iontophoresis

Buccal administration of drugs is advantageous for those drugs that encounter degradation in the gastrointestinal tract or

severe hepatic first-pass metabolism and require the administration of large doses to reach effective therapeutic levels in the target site⁵⁶⁻⁵⁷. Side effects are minimized. Among the epithelial tissues, the buccal mucosa offers good performance for local/systemic pharmacological actions because of its permeability. Since a major limitation in the development of a buccal drug delivery device could be the low permeability of the buccal mucosa, because of relatively small surface area available for absorption and poor retention of the drug and/or drug formulation at the site of absorption. The drug passively crosses the membrane whereas the application of electric fields promotes drug diffusion⁵⁸. The application of a current density of 1 mA/cm² or more determines a good improvement⁵⁹.

Jette Jacobsen's work showed that the iontophoretic approach was feasible to enhance and control the rate of buccal drug delivery of atenolol hydrochloride, and iontophoretic enhancement ratios valued above 100 were obtained. However, an 8 hour treatment period employing iontophoresis resulted in disordering of the outer epithelial cell layers⁶⁰. No sign of flogosis was found in any of mucosal specimens treated. The cells appeared vacuolated due to the presence of intracytoplasmic material, which is likely ascribable to drug accumulated. No severe cytopathic effects were found in any mucosal specimen treated with the lowest applied current density.

Transnasal Iontophoresis

The delivery of the drugs to the brain is a challenge, as systemically administered drugs fails to pass through the blood brain barrier to enter the brain. A great deal of efforts has been invested in developing the ways to open or defeat the blood brain barrier in order to deliver drugs from blood to the brain. Direct nose to brain delivery of small molecules, peptides and proteins are well known phenomenon. Transnasal

iontophoresis is another technique in the field of drug delivery to the brain.

Lerner et al⁶¹ studied the transnasal iontophoresis of octreotide in rabbits. He placed electrodes containing a drug reservoir into the deep nasal cavity with a return electrode placed at the back of head. The current strength, 3 mA, was applied for 60 min. The experiments resulted to elevated levels of octreotide in brain, with varying results due to electrode and tissue damage during insertion of electrode.

Ural Iontophoresis

Iontophoresis improves the tissue penetration of locally applied drugs. Thus, high local drug tissue levels can be achieved without general side effects. Canine⁶² studied iontophoresis using a specialized urethral catheter delivery system, equipped with an iontophoresis electrode and showed that it could safely deliver lidocaine to the prostate without a significant increase in serum levels. Possible applications in urology in addition to local anesthesia such as delivery of antibiotics for prostatitis, chemotherapy for prostate and bladder cancer, gene therapy, and prostate enzymatic ablation for benign prostate hyperplasia were also suggested. This exciting technology could certainly play a significant role in future⁶².

Patient Related Consideration

Neonates

Continuous monitoring of preterm infants is a critical care issue. However, 'minimal handling' is also recommended⁶³ since each medical intervention on a premature neonate increases the risk of infection, and amplifies thermal and energetic losses. In addition, conventional monitoring by blood sampling (i.e. plasma or serum) constitutes a painful and difficult procedure; not only is it difficult to find a vein from which to sample, but severe bruising and/or scarring of the

immature skin can also result⁶⁴. Furthermore, preterm neonates have a very limited blood volume.

Indeed, premature neonatal skin provides a unique portal for noninvasive transdermal monitoring by iontophoresis because the underdevelopment of the stratum corneum permits significantly increased drug permeability. To date, various applications have been envisaged for both drug monitoring (e.g. clonidine and theophylline⁶⁵) and diagnostic purposes (e.g. phenylalanine⁶⁶, lactate⁶⁷). Additional efforts to improve the extraction efficiency have been undertaken⁶⁸⁻⁶⁹.

Paediatrics

This iontophoresis system seems to provide a well tolerated method for providing dermal anesthesia with lidocaine in children that is not associated with the systemic delivery of this drug⁷⁰. Several studies have reported findings on the ultrastructure of stratum corneum after iontophoresis. Low current densities did not affect the structure of stratum corneum sheets; however, increased current densities, resulted in a number of changes to the lipid organization, suggesting that the electric field can perturb the intercellular lamellar ordering in the stratum corneum⁷¹. Fatouros et al. reported that 9 hour application of a 0.5 mA/cm² current *in vitro* and 3 hour 0.25 mA/cm² *in vivo* did not affect the skin architecture dramatically and that as far as structural changes in stratum corneum are concerned iontophoresis is a safe method⁷². It has been demonstrated that iontophoresis only leads to very mild skin erythema and edema⁷³.

Clinical Applications

Anaesthetics delivery

Delivery of anaesthetic agent during dermal surgery is the widest application of iontophoresis. The topical delivery of lidocaine for providing local anesthesia prior

to tooth extraction or root canal surgery by iontophoresis was reported by Gangarosa⁷⁴.

Pain management

Opioid analgesics have low molecular weights (300-500Da), usually positively charged and often requires low dose, usually in nanogram, to induce pharmacological effect. The physicochemical and pharmacological properties make these molecules suitable candidates for iontophoretic delivery⁷⁵. Examples of such drugs investigated are fentanyl and sufentanil. NSAID may cause serious adverse effects on the gastrointestinal tract, leading to ulceration and bleeding. Therefore local administration could be an desirable option. Diclofenac sodium⁷⁶⁻⁸¹, piroxicam⁸²⁻⁸³ have been investigated.

Glucose monitoring and insulin delivery

Electro-osmotic flow generated by application of low level current has been used for extraction of glucose through the skin. As the direction of glucose flow is in the opposite direction (in outward direction in skin) to conventional iontophoresis, it is called reverse iontophoresis. This property, in combination with *in situ* glucose sensors, has been used in GlucoWatch Biographer (Cygnus Inc., Redwood City, CA, USA)⁸⁴. This device allows noninvasive extraction of glucose across the skin, allowing a diabetic's glycemia to be evaluated every 10 min over several hours. Initial clinical trials using iontophoresis of soluble insulin were unsuccessful⁸⁵. Transdermal delivery of insulin by iontophoresis has been accomplished in laboratory animals. In a study of diabetic rats, iontophoretic delivery of bovine insulin affected glucose levels. By contrast, iontophoretic delivery of a monomeric human insulin analogue produced a significant fall in plasma glucose in the rats⁸⁶⁻⁸⁷.

Skin cancer

The treatment of skin cancers by radiotherapy is usually associated with many complications. Iontophoresis could be a solution for such complications. Chang et al⁸⁸ investigated the iontophoresis of cisplatin in the therapy of basal and squamous cell carcinomas in the skin and concluded that small lesions would respond best by iontophoresis. Vinblastine subcutaneous administration leads to necrosis and phlebitis, hence not recommended. Also, intralesional administration causes pain and reduces patient compliance. Smith et al⁸⁹ investigated the iontophoresis of vinblastine sulfate to treat cutaneous lesions associated with Kaposi sarcoma. All the patients showed significant clearing of their lesions.

Antiemetic drug delivery

Jadoul et al⁹⁰ conducted iontophoresis study using hairless rat skin to improve the domperidone delivery. A total of 6 hour iontophoresis resulted in 15 fold improvement in drug delivery.

Antiviral agents

Azidothymidine, an antiviral agent, has been investigated by different groups⁹¹⁻⁹². This antiviral agent undergoes first pass hepatic metabolism after oral administration, resulting in bioavailability of approximately 60-70%. In addition, drugs having short plasma half-life of approximately 1 hour, often requires maintenance of blood levels. Oh el al⁹¹ investigated iontophoretic delivery of azidothymidine and reports an approximately 1.5-fold increase in the cumulative amount of azidothymidine delivered iontophoretically from solution over 24 hour.

Cardiovascular agents

Various studies have been conducted on cardiovascular drugs including antihypertensive drugs (calcium channel blockers and

β -adrenoreceptor blockers)⁹³⁻¹⁰⁵. The iontophoretic delivery of metoprolol using rabbit model has been investigated. Arterial pressure was induced by intravenous administration of methoxamine hydrochloride at the rate of 30 mgkg⁻¹min⁻¹ for 2 hours. High frequency pulsed iontophoresis (50 kHz, 30% duty cycle) at a current density of 0.08 mA cm⁻² was begun 15 min after onset of the IV infusion. The metoprolol iontophoresis significantly decreased the systolic blood pressure from 126±9 to 86±1 mmHg and diastolic pressure from 99±7 to 72±10 mmHg¹⁰⁶.

Dermatologic applications

Treatment of hyperhidrosis

The most successful application of iontophoresis is for the treatment of hyperhidrosis. The basis for such treatment and its practical aspects have been well described¹⁰⁷⁻¹⁰⁹. Yamashita et al¹¹⁰ studied the efficacy of iontophoretic delivery of calcium for treating hydrofluoric acid-induced burns. They observed that burn areas were significantly reduced by iontophoresis more than any other mode of calcium administration, and iontophoresis was more efficacious than topical or injection therapy for experimental hydrofluoric acid burns.

Fungal infections

There are reports of the successful treatment of dermatophytosis with the use of copper sulfate iontophoresis¹¹¹ and of sporotrichosis with potassium iodide iontophoresis¹¹².

Ischemic leg ulcers

Iontophoresis has been used for the treatment of patients with ischemic leg ulcers. The effect of histamine iontophoresis on ulcers was studied by Abramson et al¹¹³, complete healing was reported in four of the five patients.

Cosmetics

Most recently, a new generation of iontophoretic patches, containing a fully integrated power source, has become available for home use. The patches are enabled through the invention of proprietary thin and flexible, safe and non-toxic, fully disposable electrical power cells and microelectronics incorporated into a simple cosmetic patch. The developer, has revealed two types of iontophoretic patches. Type one boosts the topical delivery of lotions, gels, serum preparations and other cosmetic formulations. The other type provides immediate effects of wrinkle reduction and skin smoothing. The patches can be designed to suit and target any area of the body¹¹⁴.

Anti-wrinkle effects

Human clinical studies on dozens of subjects have shown that a single 20-minute treatment using the patch results in a visible reduction of the number and depth of wrinkles under the eye and at the crow feet's area.

Treatment of pigmentation disorders

Vitamin C is known to both inhibit melanin formation and reduce oxidized melanin. However, vitamin C does not easily penetrate the skin. In 2003, Huh et al¹¹⁵ reported that iontophoresis treatment using an active form of vitamin C (namely magnesium ascorbyl phosphate or MAP) at 3.6% for 12 weeks resulted in significant reduction of pigmentation.

Treatment of scars

In 2002, Schmidt et al¹¹⁶ reported on treatment of post-acne scars using iontophoresis with 0.025% tretinoin gel. At the end of treatment, in 94% of patients a significant decrease in the scar depth was observed clinically. In conclusion, tretinoin iontophoresis was found to be effective,

noninvasive treatment of atrophic acne scars without causing disturbing side effects.

Peyronie's disease treatment

In the practice of urology, the primary use for iontophoresis has been for the treatment of Peyronie's disease (PD). PD is characterized by the clinical symptoms of diseases is initial penile pain followed by development of plaque, penile deviation, plaque calcification, penile deformation and erectile dysfunction. The majority of patients initially prefer conservative treatment. Indication for conservative therapy seems to be the early painful and progressive stage of the disease¹¹⁷.

The surgical correction is the treatment of choice for major penile angulation and deformation. A variety of medical regimens have been used to resolve pain, plaques and minor deviation. Oral vitamin E, potassium para-aminobenzoate and tamoxifen, and intralesional injection of steroids, orgotein, collagenase, verapamil and interferon- α have shown differing grades of efficacy to reduce the symptoms of PD. However, except for vitamin E, they have considerable side effects, such as gastrointestinal symptoms, which often leads to premature termination of para-aminobenzoate therapy. Local injections are extremely painful and require local anesthesia.

Iontophoretic therapy for PD have been tried in three patients. The treatment consisted of three, 20-minute sessions weekly for two weeks with 1% cortisone cream applied directly over the penile plaque. A positive electrode was placed above the application site of cortisone cream and a negative electrode was placed at a neutral site (eg. thighs, abdominal wall). A current of 3 ± 5 mA was applied, depending upon the patient's tolerance. These three patients were successfully treated with iontophoresis in spite of having mature PD¹¹⁸. It was demonstrated that a substance, such as verapamil, could be detected in PD plaques

after iontophoresis¹¹⁹. In the controlled study, a response was observed by reduction of plaque size in 100%, on curvature in 57%, and on pain in 76%. Improvement in sexual function was observed in 51%¹²⁰.

Riedl et al¹²¹ used the self adhesive iontophoretic patch in the treatment of PD and reported that iontophoresis of dexamethasone, lidocaine and verapamil is effective for peyronie's disease and especially beneficial for painful lesions of less than 12 months in duration and for deviations less than 60 degrees. In their opinion, this procedure should be regarded as first line noninvasive therapy for PD¹²¹. Thus, iontophoresis presents a potential area of development for drug delivery in patients with PD and other penile disorders.

Conclusion

The iontophoretic field of drug delivery technology holds the promise of delivering the technology breakthrough and is moving very fast from concept to reality. Some pharmaceutical companies are also set to launch major initiatives in this direction. This technology will attract attraction in the consumer market.

References

1. Banga AK, Chien YW. Iontophoretic delivery of drugs: fundamentals, developments and biomedical applications. *J Control Release*. 1988; Apr 7(1): 1–24.
2. Singh P, Maibach HI. Iontophoresis in drug delivery: Basic principles and applications. *Crit Rev Ther Drug Carrier Syst*. 1994;11(2-3):161-213
3. Nair V, Pillai O, Poduri R, Panchagnula R. Transdermal iontophoresis Part I: Basic principles and considerations. *Meth. Find. Exp Clin Pharmacol*. 1999 Mar; 21(2):139–151.
4. Kalia YN, Naik A, Garisson J, Guy RH. Iontophoretic drug delivery. *Adv Drug Deliv Rev*, 2004, Mar 27; 56(5):619–658.
5. Monti D, Saccomani L, Chetoni P, Burgalassi S, Saettone MF. Effect of iontophoresis on transcorneal permeation 'in vitro' of two b-blocking agents, and on corneal hydration. *Int J Pharm*, 2003 Jan 16;250(2):423-429.
6. Wirtz R. Die ionentherapie in der augenheilkunde. *Klinische Monatsblätter für Augenheilkunde*, 1908; 46:543–579.
7. Sarah Molokhia A, Eun-kee Jeong, William Higuchi I, Kevin Li S. Examination of barriers and barrier alteration in transscleral iontophoresis. *J Pharm Sci*, 2008 Feb; 97(2):831-844.
8. Parkinson TM, Ferguson E, Febraro S, Bakhtyari A, King M, Mundasad M. Tolerance of ocular iontophoresis in healthy volunteers. *J Ocul Pharmacol Ther*, 2003 Apr;19(2):145–151.
9. Hughes L, Maurice DM. A fresh look at iontophoresis. *Arch Ophthalmol*, 1984; 102:1825–1849.
10. Kalia YN, Naik A, Garrison J, Guy RH. Iontophoretic drug delivery. *Adv Drug Deliv Rev*, 2004; March 27; 56(5):619– 658.
11. Rootman DS, Jantzen JA, Gonzalez JR, Fischer MJ, Beurman R, Hill JM. Pharmacokinetics and safety of transcorneal iontophoresis of tobramycin in the rabbit. *Invest Ophthalmol Vis Sci*, 1988 Sep; 29(9):1397-1401.
12. Grossman R, Lee DA. Transscleral and transcorneal iontophoresis of ketoconazole in the rabbit eye. *Ophthalmol*, 1989 May; 96(5):724-729.
13. Halhal M, Renard G, Courtois Y, BenEzra D, Behar Cohen F. Iontophoresis: from the lab to the bed side. *Exp Eye Res*, 2004; 78:751–757.
14. Myles ME, Neumann DM, Hill JM. Recent progress in ocular drug delivery for posterior segment disease:emphasis on transscleral iontophoresis. *Adv Drug Deliv Rev*, 2005 Dec 13;57(14):2063–2079.
15. Eljarrat-Binstock E, Domb AJ. Iontophoresis: A non-invasive ocular drug delivery. *J Control Release*, 2006 Feb 21; 110(3):479–489.
16. Grossman R, Chu DF, Lee DA. Regional ocular gentamicin levels after transcorneal and transscleral iontophoresis. *Invest Ophthalmol. Vis Sci*, 1990 May; 31(5):909–916.
17. Chapon P, Voigt M, Gautier S, Baher-cohen FF. Intraocular tissues pharmacokinetics of ganciclovir transscleral Coulomb controlled iontophoresis in rabbits. *Invest Ophthalmol Vis Sci*, 1999; 40:S189.
18. Lam TT, Fu J, Chu R, Stojack K, Siew E, Tso MO. Intravitreal delivery of ganciclovir in rabbits by transscleral iontophoresis. *J Ocul Pharmacol*, 1994 Fall; 10(3):571–75.
19. Kralinger MT, Voigt M, Kieselbach GF, Hamasaki D, Hayden BC, Parel JM. Ocular delivery of acetylsalicylic acid by repetitive Coulomb-controlled

- iontophoresis, *Ophthalmic Res*, 2003 Mar-Apr;35(2):102–110.
20. Sarraf D, Equi RA, Holland GN, Yoshizumi MO, Lee DA. Transscleral iontophoresis of foscarnet. *Am J Ophthalmol*, 1993 Jun 15; 115(6):748–754.
 21. Esther eljarrat-binstock, Frederik Raisku, Joseph Frucht-Pery, Abrahamj Domb. Hydrogel probe for iontophoresis drug delivery to the eye. *J Biomater Sci Polymer Edn*, 2004; 15(4):397–413.
 22. Chauvaud D, Behar-Cohen FF, Parel JM, Renard G. Transscleral Iontophoresis of corticosteroids:Phase II clinical trial. *Invest Ophthalmol Vis Sci*, 2000; 41(4):S79 Abstract nr 414.
 23. Frucht-Pery J, Raiskup F, Mechoulam H, Shapiro M, Eljarrat-Binstock E, Domb A. Iontophoretic treatment of experimental pseudomonas keratitis in rabbit eye using gentamicin loaded hydrogel. *Cornea*, 2006 Dec;25(10):1182–1186.
 24. Hobden JA, Ocallaghan RJ, Hill JM, Reidy JJ, Rootman DS, Hilary TW. Tobramycin iontophoresis into corneas infected with drug-resistant pseudomonas-aeruginosa, *Curr Eye Res*, 1989 Nov;8(11):1163–1169.
 25. Hobden JA, Reidy JJ, Ocallaghan RJ, Insler MS, Hill JM. Ciprofloxacin iontophoresis for amino glycoside-resistant pseudomonas keratitis. *Invest Ophthalmol Vis Sci*, 1990; 31:1940–1944.
 26. Rootman DS, Hobden JA, Jantzen JA, Gonzalez JR, O'Callaghan RJ, Hill JM. Iontophoresis of tobramycin for the treatment of experimental pseudomonas keratitis in the rabbit. *Arch Ophthalmol*, 1988; 106:262–265.
 27. Esther Eljarrat-Binstock, Frederik Raiskup, Joseph Frucht-Pery, Abraham Domb J. Transcorneal and transscleral iontophoresis of dexamethasone phosphate using drug loaded hydrogel. *J Control Release*, 2005 Sept 2; 106(3):386–390.
 28. Berdugo M, Valamanesh F, Andrieu C, Klein C, Benezra D, Courtois Y, Behar-Cohen F. Delivery of antisense oligonucleotide to the cornea by iontophoresis, *Antisense Nucleic Acid Drug Dev*, 2003 Apr;13(2):107–114.
 29. Choi TB, Lee DA. Transscleral and transcorneal iontophoresis of vancomycin in rabbit eyes. *J Ocul Pharmacol*, 1988 Summer; 4(2):153–164.
 30. Voigt M, De Kozak Y, Halhal M, Courtois Y, Behar Cohen F. Down-regulation of NOSII gene expression by iontophoresis of anti-sense oligonucleotide in endotoxin induced uveitis. *Biochem Biophys Res Commun*, 2002 Jul 12; 295(2):336–341.
 31. Asahara T, Shinomiya K, Naito T, Shiota H. Induction of gene into the rabbit eye by iontophoresis: Preliminary report. *Jpn J Ophthalmol*. 2001 Jan-Feb; 45(1): 45:31–39.
 32. Behar-Cohen FF, Parel JM, Pouliquen Y, Thillaye-Goldenberg B, Goureau O, Heydolph S, Courtois Y, De Kozak Y. Iontophoresis of dexamethasone in the treatment of endotoxin-induced-uveitis in rats. *Exp Eye Res*, 1995 Oct; 65(4):533-545.
 33. Sarraf D, Lee DA. The role of iontophoresis in ocular drug-delivery. *J Ocul Pharmacol*, 1994 Spring;10(1):69–81.
 34. Lam TT, Edward DP, Zhu XA, Tso MO. Transscleral iontophoresis of dexamethasone. *Arch Ophthalmol*, 1989 Sep; 107(9):1368–1371.
 35. Barza M. Transscleral iontophoresis of cefazolin, cicarcillin, and gentamicin in the rabbit. *Ophthalmology*, 1986 Jan; 93(1):133–139.
 36. Behar-Cohen FF, El Aouni A, Gautier S, David G, Davis J, Chapon P, Parel JM. Transcleral Coulomb controlled iontophoresis of methyl prednisolone into the rabbit eye: Influence of duration of treatment, current intensity and drug concentration on ocular tissue and fluid levels. *Exp Eye Res*, 2002 Jan; 74(1):51–59.
 37. Hayden BC, Jockovich ME, Murray TJ, Voigt M, Milne P, Kralinger M, Feuer WJ, Hernandez E, Parel JM. Pharmacokinetics of systemic versus focal carboplatin chemotherapy in the rabbit eye: possible implication in the treatment of retinoblastoma. *Invest Ophthalmol Vis Sci*, 2004 Oct;45(10):3644–3649.
 38. Voigt M, Kralinger M, Kieselbach G, Chapon P, Anagnoste S, Hayden B, Parel JM. Ocular aspirin distribution: a comparison of intravenous, topical, and coulomb-controlled iontophoresis administration. *Invest Ophthalmol Vis Sci*, 2002 Oct;43(10):3299–3306.
 39. Jones RF, Maurice DM. New methods of measuring the rate of aqueous flow in man with fluorescein. *Exp Eye Res*, 1966 Jul;5(3):208–220.
 40. Thomas Parkinson M, Elizabeth Ferguson, Salvatore Febbraro, Arash Bakhtyari, Martin King, Mundasad M. Tolerance of ocular iontophoresis in healthy volunteers. *J Ocul Pharmacol Ther*, 2003 Apr; 19(2):145-151.
 41. Yiping Wang, Rashmi Thakur, Qiuxi Fan, Bozena Michniak. Transdermal iontophoresis: Combination strategies to improve transdermal iontophoretic drug delivery. *Eur J Pharm Biopharm*, 2005 Jul; 60(2):179–191.
 42. Williams AC, Barry BW. Skin absorption enhancers. *Crit Rev Ther Drug Carrier Syst*, 1992;9(3-4):305–353.
 43. Williams AC, Barry BW. Terpenes and the lipid-protein partitioning theory of skin penetration enhancement. *Pharm Res*, 1991 Jan;8(1):17–24.

44. Repka MA, O'Hare J, See CH, Gutta K, Munjal M. Nail morphology studies as assessments for onychomycosis treatment modalities. *Int J Pharm*, 2002 Oct 1; 245(1-2):25–36.
45. Jinsong hao, Kevin Li S. Transungual Iontophoretic Transport of Polar Neutral and Positively Charged Model Permeants: Effects of Electrophoresis and Electroosmosis. *J Pharm Sci*, 2008; 97(2):893–905.
46. Murdan S. Drug delivery to the nail following topical application. *Int J Pharm*, 2002 Apr 2; 236(1-2):1–26.
47. Walters KA, Flynn GL, Marvel JR. Physicochemical characterization of the human nail: Solvent effects on the permeation of homologous alcohols. *J Pharm Pharmacol*, 1985 Nov; 37(11):771–775.
48. Mertin D, Lippold BC. In-vitro permeability of the human nail and of a keratin membrane from bovine hooves: Influence of the partition coefficient octanol/water and the water solubility of drugs on their permeability and maximum flux. *J Pharma Pharmacol*, 1997 Jan; 49(1):30–34.
49. Hashiguchi T, Kodama A, Ryu A, Otagiri M. Retention capacity of topical imidazole antifungal agents in the skin. *Int J Pharm* 1998 Feb 23; 161(2):195–204.
50. Narasimha Murthy S, Dora E. Wiskirchen, Christopher Paul Bowers. Iontophoretic Drug Delivery across Human Nail. *J Pharm Sci*, 2007 Feb; 96(2):305–311.
51. Diego M, Guy RH, Delgado-Charro BM. Characterization of the iontophoretic permselectivity properties of human and pig skin. *J Control Release*, 2001 Jan 29; 70(1-2): 213–7.
52. Narasimha Murthy S, Waddell DC, Shivakumar HN, Balaji A, Bowers CP. Iontophoretic permselective property of human nail. *J Dermatol Sci*, 2007 May; 46(2):150–152.
53. Balakrishnan A, Rege B, Amidon GL, Polli JE. Surfactant mediated dissolution: Contributions of solubility enhancement and relatively low micelle diffusivity. *J Pharm Sci*, 2004 Aug; 93(8):2061–75.
54. Kassar DG, Lynch AM, Steller MJ. Physical enhancement of dermatologic drug delivery: Iontophoresis and phonophoresis. *J Am Acad Dermatol*, 1996 Apr; 34(4): 657–66.
55. James MP, Graham RM, English J. Percutaneous iontophoresis of prednisolone -a pharmacokinetic study. *Clin Exp Dermatol*, 1986 Jan; 11(1): 54–61.
56. Hao J, Heng PWS. Buccal delivery systems. *Drug Dev. Ind. Pharm*, 2003 Sep; 29(8): 821–832.
57. Rossi S, Sandri G, Caramella CM. Buccal drug delivery: A challenge already won. *Drug Discov Today*, 2005 spring 2(1):59–65.
58. Libero Italo Giannola, Viviana De Caro, Giulia Giandalia, Maria Gabriella Siragusa, Claudio Tripodo, Ada Maria Florena, Giuseppina Campisi. Release of naltrexone on buccal mucosa: Permeation studies, histological aspects and matrix system design. *Eur J Pharm Biopharm*, 2007 Sep; 67(2):425–433.
59. Giannola LI, De Caro V, Giandalia G, Siragusa MG, Campisi G, Florena AM, Ciach T. Diffusion of naltrexone across reconstituted human oral epithelium and histomorphological features. *Eur J Pharm Biopharm*, 2007 Feb; 65(2):238–246.
60. Jette Jacobsen. Buccal iontophoretic delivery of atenolol. HCl employing a new in vitro three-chamber permeation cell. *J Control Release*, 2001 Jan 29; 70(1-2):83–95.
61. Eduard Lerner N, Elske Van Zanten H, Gregory Stewart R. Enhanced delivery of octreotide to the brain via transnasal iontophoretic administration. *J Drug Target*, 2004 Jun; 12(5):273–280.
62. Karen Hanson A. Identifying Iontophoresis. (New urologic drug delivery device). *Urol Nurs*, 1999; 19:46.
63. Langer VS. Minimal handling protocol for the intensive care nursery. *Neonatal Netw*, 1990 Oct; 9(3):23–27.
64. Barrett DA, Rutter N. Transdermal delivery and the premature neonate. *Crit Rev Ther Drug Carrier Syst*, 1994; 11(1):1–30.
65. Glikfeld P, Hinz RS, Guy RH. Noninvasive sampling of biological fluids by iontophoresis. *Pharm Res*, 1989 Nov; 6(11):988–990.
66. Merino V, Lopez A, Hochstrasser D, Guy RH. Noninvasive sampling of phenylalanine by reverse iontophoresis. *J Control Release*, 1999 Aug 27; 61(1-2):65–69.
67. Numajiri S, Sugibayashi K, Morimoto Y. Non-invasive sampling of lactic acid ions by iontophoresis using chloride ion in the body as an internal standard. *J Pharm Biomed Anal*, 1993 Oct; 11(10):903–909.
68. Santi P, Guy RH. Reverse iontophoresis-parameters determining electro-osmotic flow I. pH and ionic strength. *J Control Release*, 1996 Feb; 38(23):159–165.
69. Santi P, Guy RH. Reverse iontophoresis-parameters determining electro-osmotic flow II. Electrode chamber formulation. *J Control Release*, 1996 Oct; 42(1): 29–36.
70. Gregory Kearns L, PharmD, Jeanellen Heacock, SallyAnn J. Daly, Hena Singh, Sarah W. Alander, and Shankang Qu. Percutaneous Lidocaine Administration Via a New Iontophoresis System in Children: Tolerability and Absence of Systemic

- Bioavailability. *Pediatrics*, 2003 Sep; 112(3 Pt 1): 578-582.
71. Craane-Van Hinsberg IW, Verhoef JC, Spies F, Bouwstra JA, Gooris GS, Junginger HE, Boddé HE. Electroperturbation of the human skin barrier in vitro (II): Effects on stratum corneum lipid ordering and ultrastructure. *Microsc. Res Tech*, 1997 May 1; 37(3):200-213.
 72. Fatouros DG, Groenink HW, de Graaff AM, van Aelst AC, Koerten HK, Bouwstra JA. Visualization studies of human skin in vitro/in vivo under the influence of an electrical field. *Eur J Pharm Sci*, 2006 Oct 1; 29(2):160-170.
 73. Li G.L, De Vries J.J, Van Steeg T.J, Van den Bussche H, Maas H.J, Reeuwijk H.J.E.M, Danhof M, Bouwstra J.A, Van Laar T. Transdermal iontophoretic delivery of apomorphine in patients improved by surfactant formulation pretreatment. *J Control Release*, 2005 Jan 3; 101(1-3):199-208.
 74. Gangarosa LP. Iontophoresis for surface local anesthesia. *J Am Dent Assoc*, 1974 Jan; 88(1):125-128.
 75. Kalia YN, Naik A, Garrison J, Guy RH. Iontophoretic drug delivery. *Adv Drug Deliv Rev*, 2004 Mar 27; 56(5):619-658.
 76. Fang JY, Sung KC, Lin HH, Fang CL. Transdermal iontophoretic delivery of diclofenac sodium from various polymer formulations: in vitro and in vivo studies. *Int J Pharm*, 1999 Feb 1; 178(1):83-92.
 77. Fang JY, Wang RJ, Huang YB, Wu PC, Tsai YH. Influence of electrical and chemical factors on transdermal iontophoretic delivery of three diclofenac salts. *Biol Pharm Bull*, 2001 Apr; 24(4):390-394.
 78. Hui X, Anigbogu A, Singh P, Xiong G, Poblete N, Liu P, Maibach HI. Pharmacokinetic and local tissue disposition of [(14)C]sodium diclofenac following iontophoresis and systemic administration in rabbits. *J Pharm Sci*, 2001 Sep; 90(9):1269-1276.
 79. Koizumi T, Kakemi M, Katayama K, Inada H, Sudeji K, Kawasaki M. Transfer of diclofenac sodium across excised guinea pig skin on high-frequency pulse iontophoresis: I. Equivalent circuit model. *Chem Pharm Bull (Tokyo)*, 1990 Apr; 38(4):1019-1021.
 80. Sugibayashi K, Kagino M, Numajiri S, Inoue N, Kobayashi D, Kimura M, Yamaguchi M, Morimoto Y. Synergistic effects of iontophoresis and jet injector pretreatment on the in vitro skin permeation of diclofenac and angiotensin II. *J Pharm Pharmacol*, 2000 Oct; 52(10):1179-1186.
 81. Varghese E, Khar RK. Enhanced skin permeation of diclofenac by iontophoresis: in vitro and in vivo studies. *J Control Release*, 1996 Jan; 38(1):21-27.
 82. Gay CL, Green PG, Guy RH, Francoeur ML. Iontophoretic delivery of piroxicam across the skin in vitro. *J Control Release*, 1992 Sept; 22(1):57-68.
 83. Curdy C, Kalia YN, Naik A, Guy RH. Piroxicam delivery into human stratum corneum in vivo: Iontophoresis versus passive diffusion. *J Control Release*, 2001 Sep 11; 76(1-2):73-79.
 84. Potts RO, Tamada JA, Tierney MJ. Glucose monitoring by reverse iontophoresis. *Diabetes Metab Res Rev*, 2002 Jan-Feb; 18 Suppl 1:S49-S53.
 85. Stephen RL, Petelenz TJ, Jacobsen SC. Potential novel methods for insulin administration: I. Iontophoresis. *Biomed Biochim Acta*, 1984; 43(5): 553-8.
 86. Narayanasamy Kanikkanna. Iontophoresis-Based Transdermal Delivery Systems. *Biodrugs*, 2002 16(5):339-347.
 87. William Cefalu T. Evaluation of Alternative Strategies for Optimizing Glycemia: Progress to Date. *Am J Med*, 2002 Oct 28; 113 Suppl 6A:S23-35.
 88. Chang BK, Guthrie TH, Hayakawa K, Gangarosa LP. A pilot study of iontophoretic cisplatin chemotherapy of basal and squamous cell carcinomas of the skin. *Arch Dermatol*, 1993 Apr; 129(4):425-427.
 89. Smith KJ, Konzelman JL, Lombardo FA, Skelton HG, Holland TT, Yeager J, Wagner KF, Oster CN, Chung R. Iontophoresis of vinblastine into normal skin and for treatment of Kaposi's sarcoma in human immunodeficiency virus-positive patients. The Military Medical Consortium for Applied Retroviral Research. *Arch Dermatol*, 1992 Oct; 128(10):1365-1370.
 90. Jadoul A, Preat V. Electrically enhanced transdermal delivery of domperidone. *Int J Pharm*, 1997 Aug 2; 154(2):229-234.
 91. Oh SY, Jeong SY, Park TG, Lee JH. Enhanced transdermal delivery of AZT (Zidovudine) using iontophoresis and penetration enhancer. *J Control Release*, 1998 Feb 12; 51(2-3):161-168.
 92. Wearley L, Chien YW. Enhancement of the in vitro skin permeability of azidothymidine (AZT) via iontophoresis and chemical enhancer. *Pharm Res*, 1990 Jan; 7(1):34-40.
 93. Hirvonen J, Guy RH. Iontophoretic delivery across the skin: electroosmosis and its modulation by drug substances. *Pharm Res*, 1997 Sep; 14(9):1258-1263.
 94. Chesnoy S, Durand D, Doucet J, Couarraze G. Structural parameters involved in the permeation of propranolol HCl by iontophoresis and enhancers. *J Control Release*, 1999 Mar 29; 8(2):163-175.

95. Ganga S, Ramarao P, Singh J. Effect of azone on the iontophoretic transdermal delivery of metoprolol tartrate through human epidermis in vitro. *J Control Release*, 1996 Oct; 42(1):57–64.
96. Gupta SK, Kumar S, Bolton S, Behl CR, Malick AW. Optimization of iontophoretic transdermal delivery of a peptide and a non-peptide drug. *J Control Release*, 1994 Jul; 30(3):253–261.
97. Hirvonen J, Kontturi K, Murtomaki L, Paronen P, Urtti A. Transdermal iontophoresis of sotalol and salicylate: the effect of skin charge and penetration enhancers. *J Control Release*, 1993 Aug; 26(2):109–117.
98. Kanikkannan N, Singh J, Ramarao P. Transdermal iontophoretic delivery of timolol maleate in albino rabbits. *Int J Pharm*, 2000 Mar 20; 197(1-2):69–76.
99. Okabe K, Yamaguchi H, Kawai Y. New iontophoretic transdermal administration of the beta-blocker metoprolol. *J Control Release*, 1986 Aug; 4(2):79–85.
100. Stagni G, O'Donnell D, YLiu YJ, Kellogg DL, Morgan T, Shepherd AM. Intradermal microdialysis: Kinetics of iontophoretically delivered propranolol in forearm dermis. *J Control Release*, 2000 Feb 3;63(3):331–339.
101. Tashiro Y, Sami M, Shichibe S, Kato Y, Hayakawa E, Itoh K. Effect of lipophilicity on in vivo iontophoretic delivery: II. Beta-blockers. *Biol Pharm Bull*, 2001 Jun; 24(6):671–677.
102. Thysman S, Preat V, Roland M. Factors affecting iontophoretic mobility of metoprolol. *J Pharm Sci*, 1992 Jul; 81(7):670–675.
103. Wearley L, Liu JC, Chien YW. Iontophoresis-facilitated transdermal delivery of verapamil: II. Factors affecting the reversibility of skin permeability. *J Control Release*, 1989 Aug; 9(3):231–242.
104. Wearley L, Liu JC, Chien YW. Iontophoresis-facilitated transdermal delivery of verapamil: I. In vitro evaluation and mechanistic studies. *J Control Rel*, 1989 Mar; 8(3):237–250.
105. Zakzewski CA, Li JK. Pulsed mode constant current iontophoretic transdermal metoprolol tartrate delivery in established acute hypertensive rabbits. *J Control Release*, 1991 Oct; 17(2):157–162.
106. Zakzewski CA, Amory DW, Jasaitis DK, Li JK. Iontophoretically enhanced transdermal delivery of an ACE inhibitor in induced hypertensive rabbits: preliminary report. *Cardiovasc Drugs Ther*, 1992;6(6):589–595.
107. Grice K. Hyperhidrosis and its treatment by iontophoresis. *Physiotherapy*, 1980 Feb; 66(2):43–4.
108. Morgan K. The technique of treating hyperhidrosis by iontophoresis. *Physiotherapy*, 1980 Feb; 66(2):45.
109. Levit F. Treatment of hyperhidrosis by tap water iontophoresis. *Cutis*, 1980 Aug; 26(2):192–4.
110. Yamashita M, Suzuki M, Hirai H, Kajigaya H. Iontophoretic delivery of calcium for experimental hydrofluoric acid burns. *Crit Care Med*, 2001 Aug; 29(8):1575–1578.
111. Jersild O, Plesner N. Treatment of epidermophytosis in the extremities with iontophoresis of copper. *Acta Derm Venereol (Stockh)*, 1940; 21:268–79.
112. Shaffer W, Zackheim HS. Sporotrichosis. *Arch Derm Syph*, 1947; 56:244–7.
113. Abramson DI, Tuck S, Chu LS, Buso E. Physiologic and clinical basis for histamine by ion transfer. *Arch Phys Med Rehabil*, 1967 Nov; 48(11):583–91.
114. Dov Tamarkin. Using Iontophoresis to Enhance Cosmetics Delivery. *Cosmet Toilet*, 2004 March; 1119:63–74.
115. Huh CH, Seo KI, Park JY, Lim JG, Eun HC, Park KC. A randomized, double-blind, placebo-controlled trial of vitamin C iontophoresis in melasma. *Dermatology*, 2003;206(4):316–320.
116. Schmidt JB, Donath P, Hannes J, Perl S, Neumayer R, Reiner A. Tretinoin-iontophoresis in atrophic acne scars. *Int J Dermatol*, 1999 Feb; 38(2):149–153.
117. Ekkehard WH, Thorsten Diemer, Hans US, Weidner W. A Critical Analysis of Nonsurgical Treatment of Peyronie's Disease. *Eur Urol*, 2006 Jun; 49(6):987–997.
118. Lewis R. Review of intraurethral suppositories and iontophoresis therapy for erectile dysfunction. *Int J Impot Res*, 2000 Oct; 12 Suppl 4:S86–S90.
119. Levine LA, Estrada CR, Shou W, Cole A. Tunica albuginea tissue analysis after electromotive drug administration. *J Urol*, 2003 May;169(5):1775–8.
120. Di Stasi SM, Giannantoni A, Stephen RL, Capelli G, Giurioli A, Jannini EA, Vespasiani G. A prospective, randomized study using transdermal electromotive administration of verapamil and dexamethasone for Peyronie's disease. *J Urol*, 2004 Apr;171(4):1605–8.
121. Riedl RC, Plas E, Engelhardt P, Daha K, Pfluger H. Iontophoresis for treatment of peyronie's disease. *J Urol*, 2000 Jan; 163(1):95–9.