Reprinted from

International Journal d Health Research

Peer-reviewed Online Journal

http://www.ijhr.org



International Journal of Health Research

The International Journal of Health Research is an online international journal allowing free unlimited access to abstract and full-text of published articles. The journal is devoted to the promotion of health sciences and related disciplines (including medicine, pharmacy, nursing, biotechnology, cell and molecular biology, and related engineering fields). It seeks particularly (but not exclusively) to encourage multidisciplinary research and collaboration among scientists, the industry and the healthcare professionals. It will also provide an international forum for the communication and evaluation of data, methods and findings in health sciences and related disciplines. The journal welcomes original research papers, reviews, commentaries and case reports on current topics of special interest and relevance. All manuscripts will be subject to rapid peer review. Those of high quality (not previously published and not under consideration for publication) will be published without delay. The maximum length of manuscripts should normally be 10,000 words (20 single-spaced typewritten pages) for review, 6,000 words for research articles, 3,000 for technical notes, case reports, commentaries and short communications.

Submission of Manuscript: The *International Journal of Health Research* uses a journal management software to allow authors track the changes to their submissions. All manuscripts must be in MS Word or RTF format and in English, and should be submitted online at http://www.ijhr.org/jmanager/. Authors who do not want to submit online or cannot submit online should send their manuscript by e-mail attachment (in single file) to the editorial office below. Submission of a manuscript is an indication that the content has not been published or under consideration for publication elsewhere. Authors may submit the names of expert reviewers or those they do not want to review their papers.

Enquiries:

The Editorial Office International Journal of Health Research Dean's Office, College of Medicine Madonna University, Elele Campus, Rivers State *E-mail:* editor@ijhr.org or editor_ijhr@yahoo.com



Academic Publishers

International Journal of Health Research, September 2008; 1(3): 129-138

© Poracom Academic Publishers. All rights reserved.

Available at http://www.ijhr.org

Original Research Article

Open Access Online Journal

Mathematical Analysis of Corneal Oxygenation

Received: 09-Jun-08

Revision received: 13-Jun-08

Accepted for publication: 08-Jul-08

Abstract

Purpose: To develop a quasi steady state model for the time course concentration profile describing the oxygen diffusion and consumption in a multilayered corneal tissue and investigate the effect of various model parameters on the oxygen concentration for open and closed eyes.

Method: A simple mathematical model for the oxygen transport in multilayered corneal tissue was developed using Fick's law of diffusion and Michaelis-Menten kinetics of metabolism. A Crank-Nicoloson finite difference scheme of the equation describing the oxygen diffusion and consumption was written, in which spatial diffusive terms were approximated by central differences while the temporal terms were approximated by average of forward and backward time differences. A system of linear equations obtained from the Crank-Nicholoson finite differences schemes was solved by the Thomos Alaorithm.

Result: The model predict that oxygen tension without contact lens for an open and closed eye increases along the distance from the aqueous side in each of the layers and the partial pressure gradient in the stroma is higher than that in the epithelium and endothelium layers. It is also observed that the oxygen tension with contact lens in the steady and transient stares, in case of low oxygen permeability of lens decreases along the distance from the aqueous side to the stroma, whereas, at higher oxygen permeability of the lens it increases along the distance for open and closed eyes.

Conclusion: Oxygen tension as observed in the cornea of an open eye with or without contact lens is higher than that in closed eye. Also at a high oxygen permeability of contact lens enhance the oxygen tension significantly than that of low oxygen permeability.

Keywords: Oxygen transport, finite difference, metabolism, oxygen consumption, pressure gradient.

Ram Avtar Deepti Tandon

Department of Mathematics, Harcourt Butler Technological Institute, Kanpur 208002, India.

*For Correspondence:

E-mail: deepti_hbti@yahoo.co.in

Introduction

The structural integrity of the cornea is maintained by an active fluid transport system, which depends on metabolism. The normal metabolic processes are essential for cell growth and replacement and, in the case of the corneal epithelium and endothelium, for the maintenance of the ionic pump mechanism, which is responsible for maintaining the state of hydration. An increase in water corneal content results in the cornea becoming thicker and cloudy or opaque. The water content of the cornea is determined by the metabolic activity of the epithelium and endothelium. A critical level of oxygen tension in the cornea is required to maintain normal corneal metabolism, which is essential for the growth and development of living and reproducing cells and for the active transport mechanisms of the epithelium and endothelium. Any interference in the metabolic activity of the cornea will cause tissue changes. If there is not enough oxygen available to convert the glucose, primarily sourced from the aqueous humor, by means of glycolysis, into sufficient energy and allow the waste product, latic acid, to diffuse guickly out of the tissue, then less energy is available for cellular activity. This results in too much latic acid being produced, which builds up in the stroma and so is implicated in the cause of corneal edema by causing an osmotic imbalance¹. Oxygen tension above a critical level may alter the metabolism and corneal swelling may occur. The oxygen required for essential metabolism of the cornea is primarily derived from the atmosphere via the tears and diffusion across the corneal anterior surface ¹. The aqueous humor in the anterior chamber also provides the oxygen to the cornea. All the oxygen supplied to the cornea by the aqueous humor is consumed by the endothelial cells and the keratiocytes in the stroma. This leaves the epithelium dependent solely on oxygen supplied to the anterior cornea surface from the air or palpebral conjunctiva. Each layer of the

cornea consumes oxygen at its own rate. It has been shown ¹ that the endothelium, epithelium, and stroma use 21%, 40% and 39%, respectively, of the total oxygen consumption of the cornea.

A contact lens effectively occludes the cornea from its surrounding environment of oxygen, tears, and ocular secretions. It impedes the movement of oxygen from the atmosphere. But the oxygen tension at the anterior cornea surface must remain above a critical level, otherwise epithelial metabolism will be altered resulting in corneal swelling. The contact lens may reduce the oxygen tension below the critical level and if this occurs, corneal edema, formation of vertical striae, and epithelial cell loss may result². Thus, adequate oxygen tension and oxygen flux is required in the cornea to maintain normal metabolic processes in both open and closed eyes. Regulation of oxygen tension in the cornea without contact lens and that under a contact lens and oxygen flux into the cornea must be studied in order to investigate the factors influencing the oxygen -level (oxygen tension) in different layers of the cornea.

In addition to numerous exp studies 3,4,5,6,7 , the steady state experimental oxygen tension has been calculated using a simplified oxygen consumption rate expression^{4,5,8}. Takahashi et al.⁷, Fatt and Bieber⁴, and Fatt ⁵ estimated the oxygen tension in the cornea of an open or closed eye by assuming a constant oxygen consumption rate. Later on, the oxygen consumption was taken as a function of oxygen tension in several studies^{7,8}. In 1976, Lin⁹ developed a steady state mathematical model for the oxygen tension distribution in the cornea. The model takes into account molecular diffusion and nonlinear oxygen consumption rate equation of the Michaelis-Menten type for the metabolic process. The oxygen tension was estimated for open and closed eyes with or without contact lens. In 1977, Barr et al.³ constructed a mathematical model based on the

experimental studies. They estimated the steady state oxygen tension by using constant oxygen consumption rate.

The present work is concerned with the development of transient state mathematical model for the oxygen tension distribution in cornea. the Oxygen diffusion and consumption are assumed to occur in different layers of the cornea: the endothelium layer, stroma layer, and epithelium layer. The oxygen consumption rates in different layers are assumed to follow the Michaelis-Menten kinetics and the numerical solution to the model was estimated using the Crank-Nicholson finite difference implicit iterative scheme. Computational results of the model have been presented in this paper.

Formulation of mathematical model

Transversely, the cornea consists of three distinct cell layers, important to the physiology of the cornea: the outer epithelium, central stroma, and inner endothelium (Figure 1). The epithelium lies the outer aspect of the cornea and the stroma, comprising 90% of the total corneal thickness, is an extracellular matrix. In contrast to the epithelium, the endothelium is a thin monolayer of cells covering the posterior surface of the cornea. For the purpose of modeling the transient oxygen transport phenomenon, the cornea (which can be regarded as hemispherical shell) is treated as a one dimensional tissue in the posterior to anterior direction. The oxygen transport in different layers of the cornea occurs, by diffusion, and the oxygen consumption due to the metabolic reactions occurring in the corneal layers, follows the Michaelis-Menten kinetics^{7,8,10}.

Governing equations: According to Fick's law of diffusion and Michaelis-Menten equation^{7,10,11} for oxygen consumption, differential equation of oxygen tension in each layer of the cornea can be represented by:



Figure 1: Layers of the Cornea

$$k_i \frac{\partial p_i}{\partial t} = D_i k_i \frac{\partial^2 p_i}{\partial x^2} - \frac{V_i p_i}{p_i + k_{m_i}}$$
(1)

Equation (2) does not have oxygen consumption term since no oxygen is consumed in the contact lens¹².

$$k_4 \frac{\partial p_4}{\partial t} = D_4 k_4 \frac{\partial^2 p_4}{\partial x^2}$$
(2)

The parameters with subscripts i=1,2,3, respectively, represent the properties of the endothelium, stroma and epithelium. The above equation represents the oxygen tension in the cornea for an open or closed eye without contact lens. D_i is the diffusion coefficient of oxygen in tissue, k_i the

solubility of oxygen in tissue, V_i , the maximum oxygen consumption rate, and k_m , the Michaelis-Menten constant.

Initial conditions: The steady state solutions to equation (1) and (2) (subject to appropriate boundary and interface conditions in different layers) are considered to be the initial conditions for the transient problem.

Boundary and interface conditions: The physiologically relevant and mathematically consistent boundary and interface conditions are presented below:

$$p_1(0,t) = p_a \tag{3}$$

$$p_{2}(x_{1},t) = p_{1}(x_{1},t) D_{2}k_{2}(\frac{\partial p_{2}}{\partial x})_{x=x_{1}} = D_{1}k_{1}(\frac{\partial p_{1}}{\partial x})_{x=x_{1}} (4)$$

$$p_{3}(x_{2},t) = p_{2}(x_{2},t) D_{3}k_{3}(\frac{\partial p_{3}}{\partial x})_{x=x_{2}} = D_{2}k_{2}(\frac{\partial p_{2}}{\partial x})_{x=x_{2}}$$
(5)

$$p_3(x_3,t) = p_b \tag{6}$$

For an open or closed eye with contact lens, boundary condition (6) is replaced by:

$$p_{4}(x_{3},t) = p_{3}(x_{3},t) D_{4}k_{4}(\frac{\partial p_{4}}{\partial x})_{x=x_{3}} = D_{3}k_{3}(\frac{\partial p_{3}}{\partial x})_{x=x_{3}}$$
(7)
$$p_{4}(x_{4},t) = p_{b}$$
(8)

In the above equations, x is the distance measured from the interface between the aqueous humor and the endothelium. At x=0, the partial pressure (tension) of oxygen becomes the aqueous humor oxygen tension. Equation (4) and (5) indicate that there must be, at all times, the continuity of oxygen tension and flux at the interface between any two layers. Equation (6) indicates that at the anterior surface of epithelial layer, oxygen tension is constant and is equal to the oxygen tension at the posterior endothelial surface of the cornea. That the flux into the cornea (epithelium) must be equal to the flux of oxygen that is leaving the contact lens when the anterior surface of the cornea is covered with a contact lens is depicted in equation (7). Equation (8) indicates that the oxygen tension at the surface of contact lens must be equal to that at the posterior endothelial surface.

Numerical solution to the model

The analytical solution to the nonlinear equations (1)-(2), subject to the boundary and interface conditions (3)-(8) in different corneal layers, seems to be a difficult task. Hence, we solved nonlinear partial differential equations numerically to find the oxygen tension distribution in different corneal layers. First, we solved the steady state nonlinear ordinary differential equation for each layer by using the Runge-Kutta Nystrom's method¹¹ to obtain a steady-state solution which was then used as initial condition for the solution of nonlinear partial differential equation. The Crank-Nicholson implicit iterative scheme¹² was used to find the approximate solution of partial differential equation for each layer. By this method, each partial differential equation of the mathematical model was replaced by a system of simultaneous algebraic equations relating values p(m,n+1) at space x_m for all points at a certain time t=n+1.

(i) Numerical model

Consider a uniform grid for the flow in the direction $n-1 \rightarrow n \rightarrow n+1$. In the Crank-Nicholson scheme, the space derivative, $\frac{\partial^2 p_i}{\partial x^2}$, is replaced by the average of its finite difference approximations on the backward time line (nth step) and the forward time line $(n+1^{st} step)$. Thus, $D_i(\frac{\partial^2 p_i}{\partial x^2})_{(m=1)} = D_i[[\frac{p_i(m-1,n)+p_i(m-1,n+1)}{2}]$

$$D_{i}\left(\frac{\sigma \ P_{i}}{\partial x^{2}}\right)_{(m,n+\frac{1}{2})} = D_{i}\left[\left[\frac{P_{i}(m-1,n) + P_{i}(m-1,n+1)}{2(\Delta x_{i})^{2}}\right]$$
(9)
$$-\left[\frac{P_{i}(m,n) + P_{i}(m,n+1)}{(\Delta x_{i})^{2}}\right] + \left[\frac{P_{i}(m+1,n) + P_{i}(m+1,n+1)}{2(\Delta x_{i})^{2}}\right]$$

The time derivative in this iterative scheme is approximated by a central difference:

$$\left(\frac{\partial p_i}{\partial t}\right)_{(m,n+\frac{1}{2})} = \left[\frac{p_i(m,n+1) - p_i(m,n)}{\Delta t}\right]$$
(10)

Equation (1) governing the oxygen diffusion and consumption in the endothelium layer can be replaced by the following finite difference analogue:

$$- D_{1}r_{1}p_{1}(m-1,n+1) + (2D_{1}r_{1}+1)p_{1}(m,n+1) - D_{1}r_{1} p_{1}(m+1,n+1) = D_{1}r_{1}p_{1}(m-1,n) + (1-2D_{1}r_{1})p_{1}(m,n)$$

$$+ D_{1}r_{1}p_{1}(m+1,n) - (\frac{V_{1}p_{1}(m,n)}{p_{1}(m,n)+k_{m}})\frac{1}{k_{1}}\Delta t$$

$$(11)$$

The finite difference approximation of equation (1) governing oxygen diffusion and consumption in the stromal layer is as follows:

$$\begin{aligned} &-D_2 r_2 p_2 (m+1,n+1) + (2D_2 r_2 + 1) p_2 (m+2,n+1) \\ &-D_2 r_2 p_2 (m+3,n+1) = D_2 r_2 * p_2 (m+1,n) + (1-2D_2 r_2) \\ &p_2 (m+2,n) + D_2 r_2 p_2 (m+3,n) - (\frac{V_2 p_2 (m+2,n)}{p_2 (m+2,n) + k_{m^2}}) \frac{1}{k_2} \Delta t \end{aligned}$$

The equation (1) defining the oxygen diffusion and consumption in the epithelium layer can be replaced by the finite difference analogue as follows:

$$\begin{aligned} &-D_{3}r_{3}p_{3}(m+3,n+1)+(2D_{3}r_{3}+1)p_{3}(m+4,n+1)\\ &-D_{3}r_{3}p_{3}(m+5,n+1)=D_{3}r_{3}*p_{3}(m+3,n)+(1-2D_{3}r_{3}) \\ &p_{3}(m+4,n)+D_{3}r_{3}p_{3}(m+5,n)-(\frac{V_{3}p_{3}(m+4,n)}{p_{3}(m+4,n)+k_{m3}})\frac{1}{k_{3}}\Delta t\end{aligned}$$

The finite difference approximation of equation (2) representing the oxygen diffusion and consumption is given by:

$$-D_4 k_4 r_4 p_4 (m+5,n+1) + (2D_4 k_4 r_4 + 1) p_4 (m+6,n+1) -D_4 r_4 p_4 (m+7,n+1) = D_4 * k_4 r_4 p_4 (m+5,n) + (1-2D_4 k_4 r_4) p_4 (m+6,n) + D_4 k_4 r_4 p_4 (m+7,n)$$
(14)

where
$$r_1 = \frac{\Delta t}{2[(\Delta x_1)]^2}$$
, $r_2 = \frac{\Delta t}{2[(\Delta x_2)]^2}$,
 $r_3 = \frac{\Delta t}{2[(\Delta x_3)]^2}$, $r_4 = \frac{\Delta t}{2[(\Delta x_4)]^2}$.

and Δx_1 , Δx_2 , Δx_3 , Δx_4 are the step sizes of mesh in the endothelium, stroma, epithelium layers, and contact lens, respectively. The boundary and continuity conditions at different interfaces are given by:

$$p_1(m-1, n+1) = p_a \tag{15}$$

$$p_2(m, n+1) = p_1(m, n+1)$$
(16)

$$p_3(m+1,n+1) = p_2(m+1,n+1)$$
 (17)

$$p_3(m+2, n+1) = p_b \tag{18}$$

If the cornea is covered by the permeable contact lens, then condition (18) is replaced by:

$$p_4(m+2, n+1) = p_3(m+2, n+1)$$
 (19)

$$p_4(m+3, n+1) = p_b$$
 (20)

(ii) Numerical flux continuity condition

The finite difference analogue of the flux continuity condition at any interface is derived by following Fatt et al.¹³. Let D_i represent the diffusion coefficient of oxygen in the i^{th} layer and D_{i+1} represents the diffusion coefficient of oxygen in the $(i+1)^{th}$ layer.

$$Q_{i} = D_{i}k_{i}[\frac{p_{i}(m,n) - p_{i}(m-1,n)}{\Delta x_{i}}]$$
(21)

$$Q_{i+1} = D_{i+1}k_{i+1}\left[\frac{p_i(m+1,n) - p_i(m,n)}{\Delta x_i}\right]$$
(22)

$$\frac{Q_{i+1} - Q_i}{\Delta x_i} = \frac{\partial p_i}{\partial t}$$
(23)

If this equation is averaged with its counterpart at the forward time line, we obtain the finite difference approximation of the flux continuity condition at the interface in the following form:

$$D_{i}k_{i}r_{i}p_{i}(m-1,n+1) + [(D_{i+1}k_{i+1} + D_{i}k_{i})r_{i} - 1]$$

$$p_{i}(m,n+1) + D_{i+1}k_{i+1}r_{i}p_{i}(m,n+1) = D_{i}k_{i}r_{i}$$

$$p_{i}(m-1,n) + [(D_{i+1}k_{i+1} + D_{i}k_{i})r_{i} - 1]$$

$$p_{i}(m,n) + D_{i+1}k_{i+1}r_{i}p_{i}(m+1,n)$$
(24)

At the endothelium-stroma interface, the finite difference approximation of the flux continuity condition is given by:

$$D_{1}k_{1}r_{1}p_{1}(m-1,n+1) + [(D_{1}k_{1}+D_{2}k_{2})r_{1}-1]$$

$$p_{1}(m,n+1) + D_{2}k_{2}r_{1}p_{1}(m,n+1) = D_{1}k_{1}r_{1} \quad (25)$$

$$p_{1}(m-1,n) + [(D_{1}k_{1}+D_{2}k_{2})r_{1}-1]p_{1}(m,n)$$

$$+ D_{2}k_{2}r_{1}p_{1}(m+1,n)$$

Similarly, the finite difference approximation of the flux continuity at the interface between thehe stroma and epithelium layer is:

$$D_{2}k_{2}r_{2}p_{2}(m,n+1) + [(D_{2}k_{2} + D_{3}k_{3})r_{2} - 1]p_{2}(m+1,n+1) + D_{3}k_{3}r_{2}p_{2}(m+2,n+1) = D_{2}k_{2}r_{2}p_{2}(m,n) + [(D_{2}k_{2} + D_{3}k_{3})r_{2} - 1]p_{2}(m+1,n) + D_{3}k_{3}r_{2}p_{2}(m+2,n)$$
(26)

Also, the finite difference analogue of the oxygen flux continuity condition at the interface of the epithelium layer and the anterior surface of lens is:

$$D_{3}k_{3}r_{3}p_{3}(m+1,n+1) + [(D_{3}k_{3} + D_{4}k_{4})r_{3} - 1]$$

$$p_{3}(m+2,n+1) + D_{4}k_{4}r_{3}p_{3}(m+3,n+1) = D_{3}k_{3}r_{3}p_{3}(m+1,n)$$

$$+ [(D_{3}k_{3} + D_{4}k_{4})r_{3} - 1]p_{3}(m+2,n) + D_{4}k_{4}r_{3}p_{3}(m+3,n)$$
(27)

(iii) Procedure of resolution of the algebraic system

By applying the iterative (implicit) described above, finite difference scheme, each partial differential equation describing oxygen diffusion and consumption can be reduced to the tridiagonal system of N simultaneous algebraic equations. The Gauss elimination method was used to solve the tridiagonal system of 30 algebraic equations. The variable values p_i (m, n+1) were determined backward substitution at the by the successive time intervals, $t^{1} = 0$, $t^{2} = \Delta t$, $t^{2} = \Delta t$ $2\Delta t$ seconds. All the numerical calculations have been performed in the 20 cells without the contact lens and 22 cells with the contact lens. In the finite difference (implicit) iterative scheme, 6,10,4 grid points were considered in the endothelium layer, stromal layer, and epithelium layer of the cornea, respectively, at the successive time intervals. The width of each layer was set to be 0.002 cm, 0.045 cm, 0.01 cm and 0.02 cm, respectively.

The computational matrix which describes the oxygen tension in different layers of the cornea, as a function of time, is defined as follows:



Results and Discussion

The computational results of the model for the oxygen tension distribution in different layers of the corneal tissue have been obtained by using typical values of model parameters for a representative eye⁹ given in Table 1. The steady and unsteady states tension distributions oxygen in the endothelium, stroma, and epithelium layers of the cornea without contact lens under open eye conditions are depicted in Figure 2. It is evident from the curves in the Figure that the oxygen tension increases along the distance from the aqueous side in each of

Avtar & Tandon

the layers and that the partial pressure gradient of oxygen in the stroma layer is higher than that in the epithelium and endothelium layers. The oxygen tension becomes constant at the anterior surface of the epithelium because the cornea can not receive much oxygen from the aqueous humor unless the oxygen tension in the epithelium layer becomes 155 mmHg.

 Table 1: Computed oxygen tension distribution in different corneal tissue layers

Symbol	Explanation	Numerical Value			
		Endothelium	Stroma	Epithelium	Lens
х	Thickness (cm)	0.01	0.45	0.04	0.02
Vi	Oxygen consumption rate (ml(O ₂) ml - sec)	11x10 ⁻⁴	1.5x10 ⁻⁷	2.1x10 ⁻⁴	0
k_{m_i}	Michaelis- Menten constant (mmHg)	4.834	4.834	4.834	4.834
P _a	Aqueous humor oxygen tension	55 mm Hg (open and closed eye)	-	-	-
Pb	Posterior endothelial oxygen tension	-	-	155 mm Hg (open eye) 55mm Hg (closed eye)	-

Besides, the oxygen tension in the transient state is higher than that in the steady-state in different lavers of the cornea and it increases with time. Under closed eye condition, the oxygen tension profiles in the steady and transient states in the endothelium, stroma and epithelium layers of the cornea are depicted in Figure 3. In this case, the oxygen tension decreases along the distance from the aqueous side in the endothelium layer, whereas, the tension increases along the depth in the stroma and epithelium. The pressure gradient at the stroma-epithelium interface is higher than that at the endothelium-stroma interface. Oxygen tension becomes constant at the anterior surface of the epithelium.

The curves shown in Figure 4 represent the steady and transient states oxygen tension profiles in the endothelium, stroma and epithelium layers of the cornea in open eye with contact lens for high and low oxygen permeabilities of the lens. It is observed from figure that the tension in the steady and

transient states, in case of low oxygen permeability of the lens, decreases along the distance from the aqueous side to the stroma, whereas. at high oxygen permeability of the lens, it increases along the distance. Thus the tension increases with an increase in the oxygen permeability of the contact lens in steady and transient states. It is evident from the curves that the oxygen tension increases along the depth in the stroma and epithelium towards the anterior surface of the cornea.

An increase in the oxygen permeability of the contact lens enhances the oxygen tension in different layers of the cornea. The tension achieves the prescribed value at the anterior surface of the contact lens. It increases with time

in different layers of the cornea at high and low oxygen permeabilities of the contact lens.

The curves in Figure 5 represents the steady and transient states oxygen tension profiles in different layers of the cornea in a closed eye with contact lens. Evidently, the oxygen tension decreases along the depth from the aqueous side in the endothelium and stroma. This decrease in the stroma is linear. However, the tension increases along a distance in the epithelium-contact lens layer from the stroma-epithelium interface and it attains a constant value at the anterior surface of the contact lens.



Figure 2: Oxygen tension profiles in the (a) endothelium, stroma (b) and epithelium (b) layers under open eye condition at different times $(a_3:0 \text{ sec}, a_2:100 \text{ sec}, a_1: 200 \text{ sec})$

Figure 3: Oxygen tension profiles in the (a) endothelium, stroma (b) and epithelium (b) layers under closed eye condition at different times (b_3 :0 sec, b_2 :100 sec, b_1 : 200 sec)



Figure 4: Effect of oxygen permeability of contact lens on the oxygen tension in the endothelium, stroma, epithelium layer in open eye (c_3 , c_2 , c_1 : $D_4k_4 = 500 \times 10^{-11}$ ml(O_2) – cm_2 / ml(tissue) – mmHg – sec at t=0 sec, 100 sec, 200 sec; c6, c5, c4 : D4k4 = 13.1×10^{-11} ml(O_2) – cm_2 / ml(tissue) – mmHg – sec at t=0 sec, 100 sec, 200 sec, respectively)



Figure 5: Effect of oxygen permeability of contact lens on the oxygen tension in the endothelium stroma, epithelium layer in closed eye (c_3 , c_2 , c_1 : $D_4k_4 = 500 \times 10^{-11}$ ml(O_2) - cm_2 / ml(tissue) mmHg - sec at t=0 sec, 100 sec, 200 sec; c6, c5, c4: $D4k4 = 13.1 \times 10^{-11}$ ml(O_2) - cm_2 / ml(tissue) mmHg - sec at t=0 sec, 100 sec, 200 sec, respectively)

Avtar & Tandon

Transient oxygen tension in the cornea with contact lens is higher than steady state oxygen tension. The effect of oxygen permeability of the contact lens on the oxygen tension in different layers of the cornea has also been depicted. It is observed from the figures that the oxygen tension in the endothelium, stroma and epithelium increases with an increase in the oxygen permeability of the contact lens in the steady and transient state. This observation is supported from that fact that more oxygen enters into the cornea at higher oxygen permeability of the contact lens increasing the oxygen tension in the cornea.

Conclusion

The computational results of the model predict that the oxygen tension in the cornea of an open eye with or without contact lens is higher than that in closed eye. Also, at a high oxygen permeability of the contact lens, the transient oxygen tension increases significantly comparative that at to the low oxygen permeability. The understanding achieved through present study may be useful in the design of contact lens.

References

- 1. Morris J. The physiological causes of contact lens complications. Optometry today. 1999; 3: 28-33.
- 2. Kennth PA, Marriane D. Oxygen tension under a

contact lens. Invest Ophthalmol. Vis Sci. 1979; 18(2): 188-193.

- 3. Barr RE, Hennessey M, Murphy VG. Diffusion of oxygen at the endothelial surface of the rabbit cornea. J Physiol. 1977; 270: 1-8.
- Fatt I, Bieber MI. The steady state distribution of oxygen and carbon dioxide in the in vivo cornea. The open eye in air and the closed eye. Exptl Eye Res. 1968; 198: 103-112.
- Fatt I. The steady state distribution of oxygen and carbon dioxide in the in vivo cornea II. The open eye in nitrogen and the covered eye. Exptl Eye Res. 1968; 7: 413-430.
- Ralph FD, Fatt I. Oxygen permeability of the limiting layers of cornea. Biophys J. 1972;12:237-247.
- Takahashi GH, Fatt I, Goldstick TK. Oxygen consumption rate of tissue measurement by a micropolarographic method. J Gen Physiol. 1967; 50: 317-335.
- 8. Walse BM. Oxygen requirements of chironmid Larvae. J Exptl Biol. 1948; 25: 35-44.
- 9. Lin S.H: Oxygen tension in the vivo cornea. Bull Math Biol. 1976; 38 :269-275.
- Amberson WR. The influence of oxygen tension on the respiration of unicellular organisms. Biol Bull. 1968; 55: 79-85.
- 11. Jain MK. Numerical Solution of Differential Equations. Wiley Eastern Limited, New York, 1984.
- 12. Gupta SK. Numerical Methods for Engineers. New Age International Limited, India, 1995.
- **13.** Fatt I, Claude GJ, Thomas MD. Non-steady state diffusion in a multilayered tissue initiated by manipulation of chemical activity at the boundaries. Biophys J. 1998; 74(1): 475-486.