

Journal of African Association of Physiological Sciences

Official Publication of the African Association of Physiological Sciences http://www.jaaps.aapsnet.org

Research Article

Effects of Ascorbic Acid on Reproductive Functions of Male Wistar Rats Exposed to Nicotine

W.A. Oyeyemi¹*, T.A. Kolawole², S.T Shittu³, R. Ajah² And B.F. Oyeyemi⁴

Departmens of Physiology, ¹Igbinedion University, School of Basic Medical Sciences, Okada, Edo State, ²Madonna University, Elele, Rivers State and, ⁴ The Federal Polytechnic Ado-Ekiti, Department of Science Technology, Ado-Ekiti, Ekiti State, Nigeria

Keywords:

Nicotine, ascorbic acid, FSH, LH, Testosterone, Sperm parameters

ABSTRACT

Nicotine is a pharmacologically active and addictive alkaloid component of the cigarette smoke, and its effects on male reproductive system and fertility are well documented. Influence of ascorbic acid on reproductive functions of male wistar rats exposed to nicotine was examined in this study. Thirty-two adult male rats of 180 ± 15 g weight were used and grouped into control, nicotine, ascorbic and nicotine with ascorbic acid. The drugs were orally administered for thirty-five days. Plasma levels of FSH, LH and testosterone were significantly reduced in nicotine exposed rats when compared with the control (p<0.05), both FSH and LH plasma levels were significantly increased in rats exposed to ascorbic acid (p<0.05) relative to the control, while ascorbic acid also increased the level of these hormone in nicotine treated group (p<0.05). The cytoarchitecture of the seminiferous tubule shows high level of degeneration in the nicotine only treated group and this was reversed in the ascorbic acid treated group. There was a significant decreased in sperm motility, counts, percentage viability and morphology (p < 0.05) in nicotine treated group relative to the control, while there was a significant improvement in these sperm parameters in co-administered ascorbic acid with nicotine when compared with the rats treated with nicotine only. In conclusion, ascorbic acid supplement may suppress nicotine toxic effects on reproductive functions in male rats.

© Copyright 2014 African Association of Physiological Sciences -ISSN: 2315-9987. All rights reserved

INTRODUCTION

Nicotine constitutes approximately 0.663.0% of the dry weight of tobacco derived from *Solanaceae* plants. It is the psychoactive substance in cigarette that acts as a stimulant and account for the highly addictive to cigarette smoking (Villégier *et al.*, 2003). Averagely, cigarette yields about 1 mg of absorbed nicotine (Bose *et al.*, 2007). Nicotine is rapidly absorbed by the brain

and metabolized to cotinine by the cytochrome P450 enzymes of the liver, it has half-life of 2 hours and 20 hours for it metabolite (Reddy *et al.*, 1995; Binnie *et al.*, 2004).

Extensive studies have been reported on the male reproductive effect of cigarette smoking and its alkaloid active constituent (nicotine) in both human and experimental animals. For instance, cigarette smoking affects male fertility (Ramlau-Hansen et al., 2006; Gaur et al., 2007), it has been associated with decreased sperm count, alteration in motility of the sperms, and overall increase in the number of abnormal sperms in humans (Jorsaraei et al., 2008). Nicotine affects spermatogenesis, sperm count, motility, and the fertilizing potential of sperms (Aydos et al., 2001; Dhawan and Sharma, 2002; Oyeyipo et al., 2011). It also inhibits pulsatile luteinizing hormone (LH) secretion (Funabashi et al., 2005), decreased plasma gonadotropin level (Jana et al., 2010; Heidary et al., 2012) and decreased the plasma level of testosterone

^{*}Address for correspondence: Email <u>oyeyemiwahab@yahoo.com</u> Tel. +2347034891903

(Sarasin *et al.*, 2003) through an inhibition in the multiple steps of testosterone biosynthesis in the rats and the mouse (Yamamoto *et al.*, 1999). It has also been reported to cause testicular degeneration in rats (Azza *et al.*, 2010; Oyeyipo *et al.*, 2010).

Ascorbic acid is a six-carbon keto-lactone, synthesized from glucose via several intermediates (Davis et al., 1991). It is an anti-oxidant vitamin (Padayatty et al., 2003), with three biological actions of particular relevance to reproduction, each dependent on its role as a reducing agent: it is required for the biosynthesis of collagen, steroid and peptide hormones, it also prevent or reduce the oxidation of biomolecules that cause great damage to cell structures such as DNA and proteins (Luck et al., 1995). It has also been reported to prevent passive smokers and sperm from oxidative damage (Dietrich et al., 2003), improves the quality of sperm in smokers, reduces sperm agglutination and increased fertility in men (Dawson et al., 1992; Sonmez et al., 2005). Also, ascorbic acid has been reported to improve sperm parameters and chromatin quality in mice (Mangoli et al., 2012). Its low level has been associated with increased numbers of abnormal sperm, low counts reduced motility and agglutination of sperm (Wilson, 1954).

In the absence of study on the effects of ascorbic acid on the pituitary-testicular axis in male rats exposed to nicotine despite the widely reports on ascorbic acid and male reproductive functions in smokers, this study therefore aimed to evaluate the role of ascorbic acid in pituitary-testicular axis, sperm quality and testicular histology in male rats exposed to nicotine.

Materials and Methods

Nicotine Preparation

Nicotine hydrogen tartrate with product number 26140 (95% nicotine) used for this study was a product of BDH chemical Ltd Poole England. It stock solution was prepared at concentration of 1mg/ml and was stored in foil-wrapped glass bottle at 4° C for not more than 7 days (Oyeyipo *et al.*, 2010).

Experimental Animals

Thirty-two (32) male albino wistar rats weighing between 170g to 200g were used. They were kept in clean environment with standard laboratory temperature (25° C) and 12 hours light and dark cycle in accordance with U.S National Institute of Health (NIH) on the care and use of laboratory animals. The rats were fed with standard laboratory chow and had free access to water *ad libitum*. After one week of acclimatization, the animals were randomly divided into four groups with eight rats in each group. The grouping was as follows;

- 1. Control group ó the rats in this group were given normal saline.
- 2. Nicotine group ó the animals were given 1.0mg/kg of nicotine.
- 3. Ascorbic acid group ó this group was treated with 100mg/kg of ascorbic acid.
- 4. Nicotine and ascorbic acid group ó the animals in this group were co-treated with both 1.0mg/kg of nicotine and 100mg/kg of ascorbic acid.

The treatments in all the groups were done orally using oral cannula for 35 days consecutively.

Experimental Procedure

After 35 days of treatment, blood samples were collected from thiopental anesthetized animals through cardiac puncture, centrifuged at 4000 rpm for 15 minutes and the plasma samples were used for follicle stimulating hormone (FSH), leutenizing hormone (LH) and testosterone assays.

The animals were then sacrificed and the caudal epididymides were immediately dissected from the testes for sperm motility, counts, viability and morphology assessments respectively. The testes were fixed in 10% formalin for testicular histology.

Hormones Assay

The plasma levels of FSH, LH and testosterone were measured by enzyme immunoassays method. The hormonal kits used for the assay was a product of Monobind Inc Lake forest, CA 92630, USA.

Testicular Histology

The testes of all the rats were fixed in 10% formalin, dehydrated stepwise in graded ethanol, cleared in xylene and then embedded in paraffin wax. A section of 5 μ m thick paraffin section of each testicular tissue was stained with hematoxylin and eosin, followed by examination under a light microscope at 200 magnification and micrographs taken (Bancroft and Stevens, 2002).

Sperm Motility

One drop of caudal epididymis sperm was dropped into a slide and diluted with few drop of normal saline at room temperature. The slide was examined under compound microscope. The number of motile and nonmotile sperm was counted in ten random fields. The number of motile sperm was then expressed as a percentage of the total number of sperm (Cheesbrough, 2006).

Sperm Counts

Sperm count was performed as described in Cheesbrough laboratory manual (Cheesbrough, 2006) with modifications. The caudal epididymis was carefully separated from the testis and homogenized in 2 ml of normal saline and the suspension was obtained. The suspension was diluted with sodium bicarbonateformalin in ratio 1 to 20. The improved Neubauer hemocytometer chamber was filled with well diluted sperm then the sperm were counted in 2 sq mm of Neubauer hemocytometer chamber. The sperm counts were calculated in 1 ml of fluid multiplied the number counted by 100,000.

Sperm Viability

The caudal epididymis sperm was dropped on the slide and mixed with a drop of 0.5% eosin solution. After 2 minutes, the slide was examined under compound microscope with 40X objective lens to count the percentage of viable (unstained) and non-viable sperm (stain red) (Cheesbrough, 2006).

Sperm Morphology

A drop of sperm suspension was smeared on a glass slide, fixed with 95% ethanol for 10 minutes and was allowed to air-dry. The smear was washed with sodium bicarbonate formalin solution to remove any mucus and then rinsed with several changed of water. The smear was covered with diluted (1 in 20) carbon fuchsin and allowed to stain for 3 minutes. The stained was washed off with water and counter stained with covered smear with diluted (1 in 20) Loefflerøs methylene blue for 2 minutes. The normal sperm which were stained (nucleus of hand ódark blue; cytoplasm of head ó pale blue; middle piece and tail ó pink red) were counted and expressed in percentage (Cheesbrough, 2006).

Statistical analysis

The results were expressed as mean and standard error of mean (Mean \pm SEM). Statistical significance between the groups was assessed by one way analysis of variance (ANOVA) followed by least significant difference (LSD) test. p<0.05 was considered significant. Statistical Package for Social sciences 16 was used for the analysis.

Results

FSH, LH and Testosterone

Table 1 showed that FSH and LH were significantly reduced in groups treated with nicotine and nicotine with ascorbic acid relative to the control group (p<0.05), while these levels were significantly

FSH, LH and Testosterone

Table 1 showed that FSH and LH were significantly reduced in groups treated with nicotine and nicotine with ascorbic acid relative to the control group (p<0.05), while these levels were significantly increased in the ascorbic acid treated group when compared with the control (p<0.05). Also, the levels of FSH and LH were significantly increased (p<0.05) in nicotine and ascorbic acid group relative to nicotine group. Testosterone level was significantly reduced in nicotine, ascorbic acid, and nicotine with ascorbic acid groups when compared with the control group (p<0.05) (table 1).

Testicular Histology

The transverse section of the testes of control and ascorbic acid groups showed normal testicular architecture, the outer capsule of fibroblastic connecting tissues bound the seminiferous tubules with narrow lumen and sperm cells were normal (figure 1 A & C). There was disorganization in both the outer capsule of fibroblastic layers of the seminiferous tubule and seminiferous tubule with reduced sperm cells in the testes of rats treated with nicotine (figure 1 B). There was reorganization in the seminiferous tubule and the outer capsule of fibroblastic connecting tissues surrounding the tubule was normal in the testis of rats that were co-treated with nicotine and ascorbic acid (figure1 D).

Sperm Motility, Counts, Viability and Morphology

Table 2 showed the results of the effect of ascorbic acid on sperm motility, counts, viability and morphology of the male rats exposed to nicotine. The results revealed that the percentage forward progressive sperm motility was significantly reduced in nicotine and nicotine with ascorbic acid groups relative to the control group (p<0.05), while the progressive sperm motility was significantly improved (p<0.05) when ascorbic acid was co-administrated with nicotine relative to nicotine group. Sperm counts was significantly decreased in nicotine and nicotine with ascorbic acid treated rats relative to control group (p<0.05). The sperm count was significantly increased by ascorbic acid (p<0.05) when co-administrated with nicotine when compared with nicotine group. Percentage sperm viability in rats exposed to nicotine and nicotine with ascorbic acid showed significant decrease when compared with rats in control group (p<0.05). Also, the percentage normal

Oyeyemi et al

Groups	FSH (miu/ml)	LH (miu/ml)	Testosterone (ng/ml)	
Control	9.11 ± 0.74	13.01 ± 0.98	12.94 ± 0.050	
Nicotine	0.67 ± 0.02^{a}	$2.60\pm0.18^{\rm a}$	12.60 ± 0.021^{a}	
Ascorbic acid	10.88 ± 0.68^{a}	$21.56\pm0.83^{\rm a}$	$12.79 \pm 0.025^{\mathrm{a}}$	
Nicotine+Ascorbic	$3.44 \pm 0.26^{a,b}$	$5.35 \pm 0.26^{a,b}$	$12.65 \pm 0.021^{a,b}$	
acid				

 Table 1: Effect of ascorbic acid on Follicle stimulating hormone (FSH), Luteinizing hormone (LH) and testosterone of male rats exposed to nicotine

Data are expressed in Mean \pm SEM of 8 rats, ^{a, b} are Mean significant difference relative to control and nicotine groups respectively at p<0.05

Table 2: Effect of ascorbic acid on sperm motility, counts, viability and morphology of male rats exposed to nicotine

Groups	Sperm motility	Sperm counts	Sperm viability	Sperm morphology
	(%)	(million/ml)	(%)	(%)
Control	68.4 ± 1.33	47.4 ± 2.79	76.6 ± 2.00	79.0 ± 2.86
Nicotine	32.2 ± 2.60^{a}	23.0 ± 2.56^{a}	46.6 ± 1.92^{a}	55.2 ± 2.15^{a}
Ascorbic acid	72.6 ± 3.57	44.8 ± 2.48	79.6 ± 2.81	80.0 ± 1.00
Nicotine + Ascorbic	$55.2 \pm 2.06^{a,b}$	$33.6 \pm 1.54^{a,b}$	$66.2 \pm 2.65^{a,b}$	$71.8 \pm 1.36^{a,b}$
acid				

Data are expressed in Mean \pm SEM of 8 rats, ^{a, b} are Mean significant difference relative to control and nicotine groups respectively at p<0.05

sperm morphology was significantly reduced both in nicotine and nicotine with ascorbic acid treated animals (p<0.05). The percentage sperm viability and morphology were significantly improved in nicotine and ascorbic acid group (p<0.05) when compared with nicotine group.

Discussion

Pituitary-testicular axis is immensely important in regulation of male reproductive functions. Testosterone which is regulated by LH from anterior pituitary gland plays a vital role in final maturation of spermatozoon and while FSH is needed for the maintenances of the gametogenic function of the testis (Barrett *et al.*, 2011). The results of this study showed significant reduction in FSH, LH and testosterone in rats that were exposed to nicotine, although, Zavos and Zarmakoupis-Zavos (1999) reported high plasma FSH level in male smokers. The possibility of the low levels of plasma FSH and LH concentration following nicotine exposure

has been reported to be probably due to elevation of glucocorticoid and corticosterone secretion from adrenal gland (Jana *et al.*, 2010), glucocorticoid may suppress the sensitivity of the gonadotroph cells to gonadotropin-releasing hormone and, therefore, may prevent gonadotropin secretion (Kamel and Kubajak, 1987). Inhibition of FSH and LH by nicotine may be as a result of its negative effect on central nervous system that can inhibit the neural stimulus essential for the release of pituitary gonadotrophins (Reddy *et al.*, 1995), which lead to a lack of pituitary gonadotrophins essential for initiating and completing spermatogenesis and steroidogenesis in the testis (Aydos *et al.*, 2001).

Fávaro *et al.* (2006) and Ahmadnia *et al.* (2007) reported many alterations attributed to the direct cytotoxic effects of nicotine leading to decrease testosterone synthesis.

Ascorbic Acid, Reproductive Functions and Nicotine

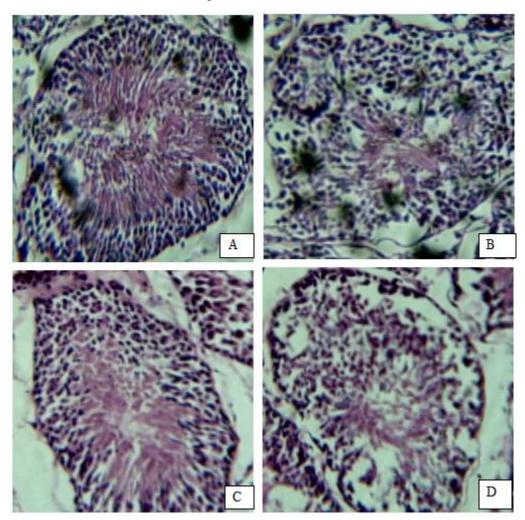


Fig.1:

Histology micrograph of transverse testes section of rats (H&E, 200X): (A) Control group shows normal testis structure with seminiferous tubule; (B) Nicotine treated group showed disarray architectural of seminiferous tubule; (C) Ascorbic acid treated group with normal testis structure and seminiferous tubule; (D) Nicotine with Ascorbic acid treated group shows reorganization of seminiferous tubule

The results of this study revealed that ascorbic acid significantly increased the plasma level of FSH, LH and testosterone both in ascorbic acid group and when supplemented in nicotine treated group. The significant increase in testosterone level observed in this study may be as a result of ascorbic acid roles in the synthesis of testosterone (Sonmez et al., 2005). The elevation in FSH and LH by ascorbic acid supplement in rats exposed to nicotine may be related to the fact that ascorbic acid can be a vitaminergic transmitter that activates the release of LH and FSH from the anterior pituitary gland (Karanth et al., 2001). The ameliorating action of ascorbic acid in elevating FSH, LH and testosterone in nicotine treated rats may also be attributed to its anti-oxidant property (Padayatty et al., 2003) which may prevent the depressive effects of

nicotine on the hypothalamus that secrete gonadotrophin that control pituitary gonadotrophins. These results were in agreement with Fernandes *et al.* (2011); Obianime and Roberts (2009), who earlier reported that ascorbic acid supplement revert FSH, LH and testosterone secretion in hyperglycemic and cadmium induced toxicity in male rats.

The structural architecture of the testis was intact with a normal spermatogenic process in the control rats of this study, while there was a destruction of both fibrous connective tissue surrounding the seminiferous tubule and spermatogenesis process in nicotine exposed rats. This may account for the reduction in sperm motility, counts and viability and plasma testosterone observed in this study. The disarray in testicular histology observed was gradually reorganized in an

Oyeyemi et al

ascorbic acid supplemented group. This further provides evidence to buttress both the detrimental effects of nicotine and ameliorative effects of ascorbic acid on sperm parameters.

From this study, results indicated that nicotine induces significant reduction in sperm motility, counts, viability and increased the percentage of abnormal sperm, an observation which was in accordance with earlier reports on the detrimental effect of nicotine on sperm parameters (Oyeyipo *et al.*, 2011; Jana *et al.*, 2009). It has been proven that nicotine increased the production of reactive oxygen species (ROS) by increased generations of testicular H_2O_2 and hydroxyl radicals in experimental rats (Bandopadhyay *et al.*, 2008).

All these observations were significantly improved in ascorbic acid supplemented rats following exposure to nicotine. Low ascorbic acid levels have been reported to be associated with low sperm counts, increased number of abnormal sperm, reduced motility and agglutination (Wilson, 1954). Also, it has been reported that dietary supplement of ascorbic acid improved sperm quality (Lucket al., 1995). The beneficial effects of ascorbic acid seen in this study may result from its antioxidant activity in mopping nicotine induced free radicals (Dietrich et al., 2003). The elevation of FSH and testosterone secretion in rats that were exposed to both ascorbic acid and nicotine in this study may be used to support the improved level of sperm motility, count, viability and normal morphology that were observed in this group, since both hormones are essential in early and final maturation of sperm cells (Barrett et al., 2011).

We therefore suggested that the nicotine induced degenerative changes in the testicular structure, reduction sperm parameters and inhibition of testosterone production acts primarily at the level of hypothalamic-pituitary axis to inhibit the release of gonadotropins and ascorbic acid supplementation could ameliorate the toxic effects of nicotine on male reproductive functions in rats.

References

- Ahmadnia, H., Ghanbari, M., Moradi, M.R. & Khaje-Dalouee, M. (2007). Effect of Cigarette Smoke on Spermatogenesis in Rats. *Urology*. 4: 3-11.
- Aydos, K., Guven, M.C., Can, B. & Ergun, A. (2001). Nicotine toxicity to the ultra-structure of the testis in rats. *BJU Int.* 88: 6226626.
- Azza, M.G., Alia, M.I., Aziza, M.A. & Sherin, R. (2010). Morphometrical, Histopathological, and Cytogenetical ameliorating Effects of Green tea

Extract on Nicotine Toxicity of the Testis of Rats. *Journal of American Science*. 6 (11): 401-411.

- Bancroft, J.D. & Stevens, A. (2002). *Theory and Practice of Histological Techniques*. 4th Edition, Queen's Medical Center, Notingham, University Hospital NHS Trust.
- Bandopadhyay, G., Sinha, S., Chattopadhyay, B.D. & Chakraborty, A. (2008). Role of cucumin against nicotine induced genotoxicity on rat liver under restricted dietary protein. *Eur. J. Pharmacol.* 588: 1516157.
- Barrett, K.E., Barman, S.M., Boitano, S. & Brooks, H.L. (2011). *Ganong's review of medical physiology*. 23rd edition,Tata Mc Graw Hill Education Private Limited. p. 404.
- Binnie, V., McHugh, S., Macpherson, L., Borland, B., Moir, K. & Malik, K. (2004). The validation of selfreported smoking status by analysing cotinine levels in stimulated and un-stimulated saliva, serum and urine. *Oral Dis.* 10: 287-293.
- Bose, M., Debnath, D., Chen, Y. & Bose, H.S. (2007). Folding, activity and import of steroidogenic acute regulatory protein into mitochondria changed by nicotine exposure. *J. Mol. Endocrinol.* 39: 67679.
- Cheesbrough, M. (2006). *District laboratory practice in tropical countries*. Cambridge University press. p. 131 ó 132.
- Davis, M.B., Austin, J. & Partridge, D.A. (1991). Ascorbic acid: Its Chemistry and Biochemistry. Cambridge: Royal Society of Chemistry.
- Dawson, E.B., Harris, W.A., Teter, M.C. & Powell, L.C. (1992). Effect of ascorbic acid supplementation on the sperm quality of smokers. *Fertil Steril*. 58:103469.
- Dhawan, K. & Sharma, A. (2002). Prevention of chronic alcohol and nicotine-induced azospermia, sterility and decreased libido, by a novel trisubstituted benzoflavone moiety from *Passiflora incarnate* Linneaus in healthy male rats. *Life Sciences*. 71:3059-3069.
- Dietrich, M., Block, G., Bnowitz, N.L., Morrow, J.D., Hudes, M., Norkus, E.P. & Packer, L. (2003). Ascorbic acid supplementation decreases oxidative stress biomaker f2-isoprostanes in plasma nonsmokers exposed to environmental tobacco smoke. *Nutr Cancer.* 45 (2): 176-184.
- Favaro, W.J. & Cagnon, V.H. (2006). Morphometric and morphological features of the ventral prostate in rats submitted to chronic nicotine and alcohol treatment. *Tissue and Cell*. 38: 311623.
- Fernandes, G.S.A., Fernandez, C.D.B., Campos, K.E., Damasceno, D.C., Anselmo-Franci, J.A. &

J. Afr. Ass. Physiol. Sci. 2 (2): 2014

Oyeyemi et al

115

Kempinas, W.D.G. (2011). Vitamin C partially attenuates male reproductive deficits in hyperglycemic rats. *Repro Biol Endocr.* 9: 100.

- Funabashi, T., Sano, A., Matsusima, D. & Kimura, F. (2005). Nicotine inhibits pulsatile luteinizing hormone secretion in human males but not in human females, and tolerance to this nicotine effect is lost within one week of quitting smoking. *J. Clin. Endocrinol. Metab.* 90: 390863913.
- Gaur, D.S., Talekar, M. & Pathak, V.P. (2007). Effect of cigarette smoking on semen quality of infertile men. *Singapore Med J.* 48 (2): 119.
- Heidary, F., Ahmadi, R. & Lotfi, A. (2012). The Effects of Cigarette or Hookah Smoking on Serum Levels of LH, FSH and Testosterone in Male Rats. International Conference on Medical, Biological and Pharmaceutical Sciences. 17-18.
- Jana, K., Samanta, P.K. & De, D.K. (2010). Nicotine Diminishes Testicular Gametogenesis, Steroidogenesis, and Steroidogenic Acute Regulatory Protein Expression in Adult Albino Rats: Possible Influence on Pituitary Gonadotropins and Alteration of Testicular Antioxidant Status. *Toxicological Sciences*. 116(2): 6476659.
- Jorsaraei, S.G.A., Shibahara, H., Hirano, Y., Shiraishi, Y., Khalatbari, A., Pasha, Y.Y. & Suzuki, M. (2008). The in-vitro effects of nicotine, cotinine and leptin on sperm parameters analyzed by CASA system. *Iranian Journal of Reproductive Medicine*. 6 (3): 157-165.
- Kamel, F.A. & Kubajak, C.L. (1987). Modulation of gonadotropin secretion by corticosterone interaction with gonadal steroids and mechanism of action. *Endocrinol.* 121: 5616565.
- Karanth, S., Yu, W.E., Walczewska, A., Mastronardi, C.A. & McCann, M. (2001). Ascorbic acid stimulates gonadotropin release by autocrine action means of NO. *Proc Natl Acad Sci.* 98 (20): 11783-11788.
- Luck, M.R., Jeyaseelan, I. & Scholes, R. (1995). Ascorbic acid and fertility. *Biology of Reproduction*. 52: 262-266.
- Mangoli, E., Pourentezari, M., Anvari, M., Talebi, A.R. and Nahangi, H. (2012). The improvement of sperm parameters and chromatin quality by vitamin C. Researcher. 4(11): 43-49
- Obianime, A.W. & Roberts, I. (2009). Antioxidants, cadmium induced toxicity, serum biochemical and the histological abnormalities of the kidney and testes of the male wistar rats. *Nig J. Physiol Sci.* 24(2): 177-

185.

- Oyeyipo, I.P., Raji, Y., Emikpe, B.O. & Bolarinwa, A.F. (2010). Effect of oral administration of nicotine on organ weight, serum testosterone level and testicular histology in adult male rats. *Nig J Physiol Sci.* 25 (1): 81-86.
- Oyeyipo, I.P., Raji, Y., Emikpe, B.O. & Bolarinwa, A.F. (2011). Effects of nicotine on sperm characteristics and fertility Profile in Adult Male Rats: A Possible Role of Cessation. *J Reprod Infertil.* 12 (3): 201-207.
- Padayatty, S.J., Katz, A., Wang, Y., Eck, P., Kwon, O., Lee, J.H, Chen, S. & Corpe C. (2003).. Ascorbic acid as an antioxidant: evaluation of its role in disease prevention. *Journal of the American College of Nutrition.* 22 (1): 18-35.
- Ramlau-Hansen, C.H., Thulstrup, A.M. & Aggerholm, A.S. (2006). Is smoking a risk factor for decreased semen quality? A cross-sectional analysis. *Human Reproduction*. 22 (1): 1886196.
- Reddy, A., Sood, A., Rust, P.F., Busby, J.E., Varn, E. & Mathur, R.S. (1995). The effect of nicotine on invitro sperm motion characteristics. *J Assist Repord Genet.* 12: 217-223.
- Sarasin, A., Schlumpf, M., Muller, M., Fleischmann, I., Lauber, M.E. & Lichtensteiger, W. (2003). Adrenalmediated rather than direct effects of nicotine as a basis of altered sex steroid synthesis in fetal and neonatal rat. *Reprod Toxicol.* 17: 1536162.
- Sonmez, M., Turk, G. & Yuce, A. (2005). The effect of ascorbic acid supplementation on sperm quality, lipid peroxidation and testosterone levels of male Wister rats. *Theriogenology*. 63 (7): 2063-2072.
- Villégier, A.S., Blanc, G., Glowinski, J. & Tassin, J.P. (2003). Transient behavioral sensitization to nicotine becomes long-lasting with monoamine oxidases inhibitors. *Pharmacol Biochem Behav.* 76 (2): 2676 74.
- Wilson, L. (1954). Sperm agglutinations in human serum and blood. *Proc Soc Exp Biol Med.* 85: 652-655.
- Yamamoto, Y., Isoyama, E., Sofikitis, N. & Miyagawa, I. (1998). Effects of smoking on testicular function and fertilizing potential in rats. *Urol Res.* 26: 45648.
- Zavos, P.M. & Zarmakoupis-Zavos, P.N. (1999). Impact of cigarette smoking on human reproduction: its effects on male and female fecundity. *Technology*. 6: 9-16.