



Isolation and Identification of Fungi Associated with Habanero Pepper (*Capsicum chinense* Jacq) Using Basic Molecular Techniques

*IKECHI-NWOGU, GC; ODOGWU, AB; UKOMADU, J; USIAKPONEBRO, EP

Department of Plant Science and Biotechnology, Faculty of Science, University of Port Harcourt. Choba, P. M. B. 5323, Port Harcourt, Nigeria

* Corresponding Author Email: chinyerum.ikechi-nwogu@uniport.edu.ng, Tel.: +2348032325098, +2347039338412
Other authors Email blessing.odogwu@uniport.edu.ng, Tel.: +2348036772711, josephine.ukomadu@uniport.edu.ng, Tel.: +2348147651600, ebrus_luv@yahoo.com, Tel.: +2347052744632

ABSTRACT: Habanero pepper (*Capsicum chinense* Jacq.) is an economically important vegetable. It is used as spices for foods and for medicinal purposes in many parts of the country. Despite the importance of the crop, it has been observed that during storage, the fruits of the Habanero pepper (HP) shelf-life is reduced by several post-harvest diseases caused by fungal pathogens. A study was conducted to identify the common fungal pathogens causing post-harvest rot of Habanero fruits at the daily market in Omoko Aluu Town, Rivers State. The DNA of the most common fungal isolate HP-02 was molecularly characterized using Internal Transcribed Spacer 1 (ITS-1) molecular marker. The HP-02 isolate DNA sequence was aligned using the Basic Local Alignment Search Tool for nucleotide (BLASTN) 2.8.0 version of the National Center for Biotechnology Information (NCBI) database. The results indicated that the HP-02 isolate sequence was 79% identical to *Mucor irregularis* and also *Rhizomucor viribilis* isolate SAPB3. These findings showed that *Mucor irregularis*, which was formerly known as *Rhizomucor variabilis* is one of the emerging causal fungal pathogens of post-harvest Habanero pepper.

DOI: <https://dx.doi.org/10.4314/jasem.v25i7.4>

Copyright: Copyright © 2021 Ikechi-Nwogu *et al.* This is an open access article distributed under the Creative Commons Attribution License (CCL), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Dates: Received: 10 May 2021; Revised: 28 June 2021; Accepted: 01 July 2021

Capsicum chinense Jacq. (Habanero pepper) is a hot variety of Chili pepper; it is a perennial flowering plant of the Solanaceae family. It is locally known as atarodo by the Yorubas, osendu by the Igbos and attaruhu by the Hausas of Nigeria. The fruits of *C. chinense* varies greatly in both color and shape, some appears red, yellow and orange are the most widespread matured colors, but very few also appear brown and purple (*Chinense* species, 2012). The fruits are about 1 to 2.5 inches long, 1 to 2 inches in diameter and they are either lantern- in shape, round or oblong (Pepper facts, 2018). This plant thrives well in hot weather conditions. Fruits mature at about 100 to 120 days after transplanting (Marie, 2017). The Habanero pepper can be properly dried using a dehydrator, or other methods before preservation. It is of great economic importance, its use on food, ornamental products, pharmaceutical, and cosmetics (Dias *et al.*, 2013, de Oliveira, *et al.*, 2017). However, Pepper is faced with fungi diseases such as: *Anthraco*se, *Cercospora* leaf spot, damping off, *Fusarium* wilt, gray mold, *Phytophthora* blight, powdery mildew, southern blight, *Verticillium* wilt, white mold (Asare-Bediako *et al.*, 2015). These fungi diseases are post-harvest disease that have caused 50% loss in plants (Agris, 2005). Given that there are

different fungi species and the morphological identification is not adequate for taxonomy, more tools for fungi identification are needed. Identification and taxonomy of fungi pathogens are increasingly dependent on modern molecular techniques based on Polymerase Chain Reaction (PCR) amplification. Sequencing of the resulting PCR products has proven to be more reliable alternative to traditional methods of identification (Hariharan and Prasannath, 2021; Horton and Bruns, 2001). Many molecular classification approaches have been carried out for fungal organisms (Alsohaili, *et al.*, 2018). In our world today, it is vital for fungi to be properly identified using molecular characterization. This study looks into the isolation and identification of fungi diseases associated with the Habanero pepper (*Capsicum chinense*).

METHODS AND METHODS

Study Area: The study was conducted in Mycology/Pathology laboratory of Plant Science and Biotechnology and Regional Centre for Biotechnology and Bio-fuel Research Laboratory where DNA extraction was carried out, University of Port Harcourt. Amplification and sequencing of the PCR products were done at the Habanero pepper with

* Corresponding Author Email: chinyerum.ikechi-nwogu@uniport.edu.ng, Tel.: +2348032325098, +2347039338412

disease symptoms were purchased Omoko Aluu Town, Port Harcourt, Rivers State, Nigeria International Institute for Tropical Agriculture (IITA) Ibadan both in Nigeria.

Isolation of Fungi from Capsicum chinense Jacq. Using Blotter Method: According to the rules of the international seed health testing association (ISTA, 2016), a standard blotter method was used to isolate fungi pathogens associated with Habanero pepper. The filter paper, distilled water (placed in a conical flask) and Petri dishes (wrapped in a foil paper) used for the work were first autoclaved at 121°C for 15mins. The

Petri-dishes were lined with 3 layers of sterilized 9cm Whatman's filter paper, the filter paper was soaked with little water and then the petri dishes were covered immediately. The pepper was surface sterilized in a beaker using 70% ethanol for 2-3minutes, discard the ethanol and rinsed with sterile distilled water twice. 4-5 pepper fruits were plated per Petri-dish (Plate 1a), the plates were then wrapped with masking tape and then labelled. Plated pepper was incubated at 25 + 2°C in the laboratory for 7 days. Unidentified fungus (Plate 1b), was sub-cultured on Potato Dextrose Agar (PDA) medium under darkness at room temperature (25+ 2°C).



Plate 1A: Pepper on Blotter Paper and **Plate 1B:** Pepper on Blotter Paper after 7 Days (red arrow showing the fungus)

Preparation of Potato Dextrose Agar: To prepare potato infusion, 200g of sliced, unpeeled potatoes was boiled in 1 liter of distilled water for 30 minutes. The boiled potato was then filtered through cheesecloth, saving effluent, which is potato infusion. The filtrate was transferred into measuring flask, 20g of dextrose and 20g of agar were added and the mixture was made up to 1litre with distilled water. The medium was shaken properly to mix well and then dispensed into 500ml flat bottom flasks. The flasks were plugged with cotton wool and foil and then autoclave at 121°C for 15 minutes. The medium was allowed to cool, then 3 drops of 25% lactic acid was added to each 500ml of medium and then it was stirred. About 10 to 20ml PDA was dispensed into each sterile petri-dish and allowed to solidify (Ataga *et al.*, 2010 and BAM, 2001). It was then shake to make sure it is very solid.

Inoculation of Fungal Organism: The inoculating loop was sterilized by dipping into 70% ethanol and then flaming in a Bunsen burner, waved a little to cool off, to prevent contamination. The sterile inoculating loop was used to pick up individual colonies that were isolated from the pepper seeds, and then transferred unto the sterilized Petri-dishes containing Potato Dextrose Agar medium. Each Petri-dish was sealed with masking tape, labelled with dates and placed upside down to prevent contamination from any moisture that may be present.

Incubation of Fungal Organism: The Petri dishes were incubated at room temperature (25 + 2°C) for 7 days to obtain pure cultures of the fungal organism. A total of 5 Petri dishes were used for each fungus. The cultures were stored at 4°C prior to DNA extraction.

Extraction of Fungal DNA: The genomic DNA of pure cultures of Habanero pepper was extracted using protocol of Quick- DNA™ Fungal/Bacterial Miniprep Kit (Zymo Research Group, California, USA) as pronounced by the manufacturer, with modifications. DNA quantity and concentration were measured using the Nanodrop 2000c spectrophotometer (Thermo fisher Scientific Inc. Wilmington, Delaware, USA). The DNA purity was measured as a ratio of absorbance at 280nm to that of 260nm. The quality of the gDNA HP-7A isolate was further quantified using the Agarose gel electrophoresis performed according to the modified method of Saghai-Marroof *et al.*, (1984). The DNA sample HP-7A isolate was shipped for amplification and sequencing to the International Institute of Tropical Agriculture (IITA) Bioscience Center, Ibadan, Nigeria for. The primers used to amplify fragments of the nuclear ribosomal DNA (rDNA) of the HP-7A isolate were the Internal Transcribed Spacer 4 (ITS4) with the sequence TCCTCCGCTTATTGACATGS and ITS5 with the sequence GGA ACTAAAAGTCGTAACAAGG. The

amplicons were sequenced using the capillary electrophoresis sequencer. The DNA sequence file was saved in the Bioedit file with an extension .ab1. The sequence was analyzed using the Molecular Evolutionary Genetics Analysis (MEGA) version

7.0.26 software, and aligned using the Basic Local Alignment Search Tool for nucleotide (BLASTN) of the National Center for Biotechnology Information (NCBI) database.



Plate 2a: Preparing cultures for crushing and



Plate 2b: Crushed cultures

RESULTS AND DISCUSSION

Isolation of Fungi Associated with *Capsicum chinense* Jacq: The result of the fungal isolation is presented in plate 3.1. one unidentified fungal organisms HP- 02 was isolated and found to be associated with Habanero pepper (*Capsicum chinense* Jacq.) (Plate 2).



Plate 3: Pure Culture of Fungus Isolated from Pepper on Potato Dextrose Agar

DNA Extraction and Concentration Determination: The genomic DNA of the isolate HP-02 of Habanero pepper (*Capsicum chinense* Jacq.) was successfully extracted and showed good quality. The Nanodrop result showed that the concentrations of the DNA of the isolates were 43.9ng/ul, 20.2ng/ul and 44.3ng/ul respectively. While the absorption peak of the 260nm/280nm readings were 1.59, 1.81 and 1.58 respectively and the 260nm/230nm readings were 0.33, 0.31 and 0.33 respectively. However, to reduce the cost of sequencing, the isolate HP-02 with the highest DNA concentration was selected.

Polymerase Chain Reaction (PCR) and Gel

Electrophoresis: The result of the amplified DNA or PCR band of the isolate HP-02 is presented in Fig. 1. The amplified DNA showed bands on gel when observed under UV light. From the result, the ladder used indicated that the HP-02 isolate sequence had over 569 base pairs.

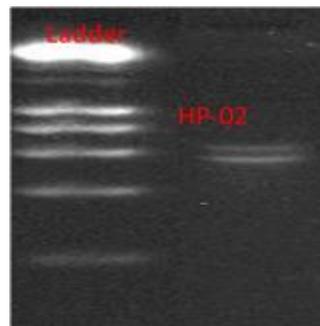


Fig 1: PCR products generated from Fungal DNA isolates

Sequence Alignment: The alignment results are presented in figure 2-3. Figure 3.4 displayed the alignment scores presented as purple lines. The scores of the alignments of all aligned sequences were below 200.

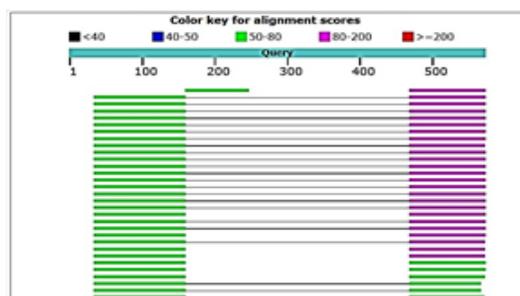


Fig 2: Alignment Scores of all Aligned Sequences

Figures 3 indicated that the HP-02 isolate sequence was 79.09% identical to *Mucor irregularis* LHL1-2 (red arrow) and 79.09% identical to *Rhizomucor*

variabilis SAPB3 (blue arrow). These findings showed that isolate HP-02 is a *Mucor* spp.

Description	Max Score	Total Score	Query Cover	E value	Per. Ident	Accession
<input type="checkbox"/> <i>Mucor irregularis</i> strain LHL1-2 small subunit ribosomal RNA gene, partial sequence, internal transcribed spacer 1 and 5.8S ribosomal RNA gene, complete sequence, and internal transcribed spacer 2, part	82.4	82.4	18%	1e-11	79.09%	M550071.1
<input type="checkbox"/> <i>Mucor irregularis</i> strain MF22455 small subunit ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and	82.4	146	40%	1e-11	79.09%	M691149.1
<input type="checkbox"/> <i>Rhizomucor variabilis</i> isolate F1 small subunit ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and	82.4	146	40%	1e-11	79.09%	M271193.1
<input type="checkbox"/> <i>Rhizomucor variabilis</i> strain SAPB3 18S ribosomal RNA gene, partial sequence	82.4	146	40%	1e-11	79.09%	K034395.1
<input type="checkbox"/> <i>Rhizomucor</i> sp. BAR-408 18S ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and 28S ribosomal R	82.4	146	40%	1e-11	79.09%	KJ521493.1
<input type="checkbox"/> <i>Ganoderma</i> sp. BAR-494 18S ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and 28S ribosomal R	82.4	223	40%	1e-11	79.09%	KR249038.1
<input type="checkbox"/> <i>Ganoderma</i> sp. BAR-492 18S ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and 28S ribosomal R	82.4	223	40%	1e-11	79.09%	KR549038.1
<input type="checkbox"/> <i>Ganoderma</i> sp. HN 18S ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and 28S ribosomal RNA gene	82.4	146	40%	1e-11	79.09%	KM930391.1
<input type="checkbox"/> <i>Mucor irregularis</i> strain A21 18S ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and 28S ribosomal r	82.4	146	40%	1e-11	78.70%	KM871199.1
<input type="checkbox"/> <i>Rhizomucor variabilis</i> strain D11 18S ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and 28S ribosom	82.4	142	40%	1e-11	79.09%	KJ582095.1
<input type="checkbox"/> <i>Rhizomucor variabilis</i> strain V1 18S ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and 28S ribosom	82.4	146	40%	1e-11	79.09%	KJ582095.1
<input type="checkbox"/> <i>Ganoderma lucidum</i> strain MAJ V 18S ribosomal RNA gene, partial sequence, internal transcribed spacer 1, 5.8S ribosomal RNA gene, and internal transcribed spacer 2, complete sequence, and 28S ribos	82.4	146	40%	1e-11	79.09%	KM275027.1

Fig 3: The Sequence Alignments of HP-02 isolate sequence with NCBI database sequences

Several organisms have been associated with the post-harvest fruits of the Habanero pepper. Among the casual pathogens, *lasiodiplodia*, *Aspergillus flavus*, *Penicillium corylophilum*, *Asperillus fumigates*, *Aspergillus niger*, *Rhizopus stolonifer* and *Verticillium spp.* have all caused various diseases such as, damping off, wilting, powdery-mildew, soft rot and general spoilage in pepper and other plant species (Lema *et al.*, 2018). In this study, *Mucor irregularis* was identified as a seed-borne pathogen associated with Habanero pepper. From the DNA alignment results, it was also noticed that the HP-02 isolate sequence was 79% identical to the partial sequence of the small subunit of ribosomal RNA gene of *Rhizomucor variabilis* isolate SAPB3 and this may be as a result of the ITS region sequenced. From PCR and gel electrophoresis result it was found that an organism of the *Rhizomucor* spp. was noticed to have mixed and was mimicking the *Mucor irregularis* spp. *Mucor irregularis* is an emerging fungal pathogen that can cause cutaneous infection and could cause death. However, little is known about its mechanism of pathogenesis, but for the purpose of this study, it brought to light that it can affect the fruit of pepper, thus damaging it. *Mucor irregularis* is a *Mucor* spp. belonging to the family Mucoraceae. It belongs to the fungi causing the group of infections referred to as zygomycosis. Zygomycosis infections includes, rhinocerebral infections as well as

septic arthritis, dialysis associated peritonitis, renal and pulmonary infections. This microbe is highly relevant to the safety and care plans of modern human population. (Doctor Fungus, 2019)

Conclusion: This study revealed that *Mucor irregularis* is one of the causal fungal pathogens of post-harvest rot of Habanero pepper. Sequence alignment result from the NCBI database showed that the HP-02 isolate sequence was 79.09% identical to *Mucor irregularis* LHL1-2. This research therefore paves way for further research using molecular tools in the identification of pathogenic organisms associated with diverse crops.

REFERENCES

- Agrios, GN (2005). Plant Pathology. Fifth (5th) Edition. pp. 27-557
- Alshohaili, SA; Bayan, MB (2018). Morphological and Molecular Identification of Fungi Isolated from Different Environmental Sources in the Northern Eastern Desert of Jordan. *Jordan Journal of Biological Sciences*. 11:329-337
- Asare-Badiako, E; Addo-Quaye, A; Boakye, B; Sarbah, JM, Asante, P; Dorm, E (2015). Incidence and Severity of Viral and Fungi Diseases of Chili

- Pepper (*Capsicum frutescens*) in Some District in Ghana. *International Journal of Plant and Soil Science*. 7(3):147-159
- Ataga, AE; Elenwo, EN; Nwachukwu, EO (2010). Laboratory Exercises and Series in Mycology. ACOTEC Technology, P.H., Nigeria. pp. 13-25, 97-183
- Chinense Species". *Capsicum Species*. The Chili Man. Archived from the original on 23 February 2012/ Retrieved 8/06/2011
- De Oliveira¹, CVS; Matos, KS; De Albuquerque, DMC; Hanada, RE; Da Silva, GF (2017) Identification of *Colletotrichum* isolates from *Capsicum chinense* in Amazon. *Genetics and Molecular Research*. 16 (2)
- Dias, GB; Gomes, VM; Pereira, UZ (2013). Isolation, characterization and antifungal activity of proteinase inhibitors from *Capsicum chinense* Jacq. Seeds. *Protein Journal*.32: 15-26
- Doctor Fungus (2019) "The *Mucor* Species" .<http://drfungus.org/knowledge-base/> Retrieved 10/10/2019
- Hariharan, G; Prasannath, K (2021). Recent Advances in Molecular Diagnostics of Fungal Plant Horton, TR; Burns TD (2001). The Molecular Revolution in Ectomycorrhizal Ecology: Peeking into the Black-Box. *Molecular Ecology*. 10:1855-1871.
- ISTA (International Seed Testing Association) (2016). International Rules for Seed Testing. Rules Amendments. *Seed Science Technology*. 29: 1-127.
- Lema, AA; Mudansiru, a; Alenxander, BA; Sakinatu, MJ (2018). Evaluation of Fungal Species Isolated from Three Different Varieties of Pepper (*Capsicum chinense*, *C. frutescens* and *C. annum L.*) in Dutsin-ma, Katsina State. *Annals of Biological Sciences*. 6(1):13-17.
- Marie, D (2017). Nutrition information. <https://www.livestrong.com/article/349143-habanero-pepper-nutrition-information/> Retrieved 10-09-2019. Pathogens. Mii Review: *Frontiers in Cellular and Infection Microbiology*. 1-14
- Pepper Facts (2018). <http://www.ballintemple.com/archive/organic/pepperfacts.html>/Retrieved 09-9-2019.
- Saghai-Marroof, M. A., Soliman, K. M, Jorgensen, R. A and Allard, R. W. (1984) Ribosomal DNA spacer-length polymorphism in barley: mendelian inheritance, chromosomal location and population dynamics. *Proceedings of National Academy of Sciences of the United States of America*. 81(24):801-8018.