Lead Acetate-induced Changes in Haematological Indices and Bone Marrow of Adult Wistar Rats: Protective Role of α-Tocopherol (Vitamin E)

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ABSTRACT: Lead exposure has been associated with several adverse health effects involving multiple body systems and organ failure. Hence, the objective of this paper was to the effects of α-Tocopherol (Vitamin E) on haematological indices and bone marrow of lead acetate (LA)-exposed adult Wistar rats. Twenty (20) adult Wistar rats (n=5) were randomly assigned as follows: Control group (A) received 1ml of distilled water; Group B received 100 mg/kg body weight (BW) of LA; Group C received 50 mg/kg BW of α-Tocopherol and 100 mg/kg BW of LA; Group D received 50 mg/kg BW α-Tocopherol only. All administrations, via an oral gavage, lasted for twenty-eight days. Following the sacrifice of experimental rats, blood samples were collected in Ethylenediaminetetraacetic acid bottles for haematological analysis and the femur of rats were fixed in 10% neutral buffered formalin for histological evaluation. Results showed that the haematological indices in group B rats were significantly different (P<0.05) when compared to control. Also, the histological findings in group B rats revealed mild hypoplasia of the haematopoietic systems and organ failure. Hence, the objective of this paper was to the effects of α-Tocopherol on the haematological system, and further studies are needed to corroborate these findings and investigate the mechanisms of action.

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Lead is a poisonous ubiquitous metal that exists in the environment and is one of the most widely studied toxic substances (Enogieru and Iyoha, 2023). It enters the body system either via the respiratory structures or gastrointestinal tracts and is taken up in large amounts by the liver which acts as a repository before being passed to the kidney where a small fraction is evacuated via urine (Wani et al., 2015). Lead may also concentrate within the bone marrow, brain, and testis, with its accumulation resulting in serious damage in the respective tissues (Sanders et al., 2009). Lead toxicity has also been shown to affect the erythrocyte membrane, causing a significant reduction in erythrocyte mobility and an alteration of other haematological parameters (Suwalsky et al., 2003). Lead-induced oxidative stress has been attributed as one of the leading etiologies of lead poisoning (Enogieru and Momodu, 2022; Patra et al., 2011). Thus, low to high levels of lead exposure could result in different responses to oxidative stress within various target sites such as the blood vessels, liver, brain, testes, and lungs (Mandal et al., 2022). In rats, the beneficial role of antioxidants has been previously reported against metal toxicity (Enogieru and Egbon, 2022; Enogieru and Momodu, 2021; Enogieru and Omoruyi, 2022). For instance, the use of ascorbic acid as a potent antioxidant in rabbits (Rabbani et al., 2003), rats (Nandi et al., 2005), and other rodents (Rana et al., 2008). Similarly, α-Tocopherol, also known as Vitamin E is a dietary essential that acts as a first line
of defense against pro-oxidants (Rizvi et al., 2014). It is a fat-soluble vitamin with powerful anti-oxidative properties that protect the cell against the oxidizing activity of free radicals and lipid peroxidation, thus preventing oxidative stress (Rizvi et al., 2014).

α-Tocopherol has also been found to be beneficial in improving immune function, lowering the risk of vascular thromboembolism, and providing support for patients with coronary heart disease and stroke (Vardi et al., 2013). Taking into consideration the above effects of α-Tocopherol, hence, the objective of this paper was to investigate the effects of α-Tocopherol (Vitamin E) on haematological indices and bone marrow of lead acetate (LA) - exposed adult Wistar rats.

MATERIALS AND METHODS

Chemicals and Reagents: Lead acetate and α-Tocopherol were procured from Loba Chemie, India. Other reagents were all of the analytical grades.

Animal Care and Management: Twenty (20) adult Wistar rats, weighing between 180-200g, were purchased from the animal holdings of the Department of Anatomy, University of Benin, Benin City. Care and management of animals were done following the guidelines for the care and use of laboratory animals (National Research Council of the National Academics, 2011). The animals were allowed to acclimatize for a period of two weeks before the commencement of the experiment.

Experimental Design: The rats were randomly assigned into a control group (A) and three treatment groups B, C, and D containing n=5 rats/group. Control group (A) received 1ml of distilled water; Group B received 100 mg/kg body weight (BW) of LA; Group C received 50 mg/kg BW of α-Tocopherol and 100 mg/kg BW of LA; Group D received 50 mg/kg BW α-Tocopherol only. The administration was carried out daily using an oral gavage for a period of twenty (28) days.

Sacrifice of Rats: At the end of the experimental period, the animals were sacrificed under chloroform anaesthesia, and blood samples were collected, by Cardiac puncture, in Ethylenediaminetetraacetic acid (EDTA) bottles for determination of hematological indices. The femur of the experimental rats was dissected out and fixed in 10 % Neutral buffered formalin for tissue processing.

Hematological Analysis: Blood analysis, using an automatic hematological assay analyzer, was used to analyze the blood samples at the laboratory of Accident and Emergency (A & E), University of Benin Teaching Hospital, Benin City, as previously reported (Kanu et al., 2016). Hematological parameters analyzed of interest include: Red Blood Cell (RBC) count, White Blood Cell (WBC) counts, differential white blood cell counts including Lymphocyte, Monocyte, and Granulocyte Counts, Hemoglobin concentration (HbC), Erythrocyte indices analyzed were Mean Corpuscular Volume (MCV), Mean Corpuscular Hemoglobin Concentration (MCHC), Mean Corpuscular Hemoglobin (MCH) and Packed cell volume (PCV).

Histology of the Bone marrow: The bones were fixed in 10% neutral buffered formalin for 48 hours. The bones were decalcified in 10% nitric acid for 7 days. They were then processed and stained in Hematoxylin and Eosin as previously reported (Drury and Wallington, 1980).

Statistical Analysis: All values are presented as mean ± standard error of the mean for all groups. The significance of the difference in the means of all parameters was determined using a one-way analysis of variance (95% confidence interval). Least Square difference, post-hoc tests were carried out for all groups with control and comparison of all pairs of groups respectively. All statistical analysis was carried out using the Statistical Package for Social Sciences (SPSS) (version 16) manufactured by the International Business Machine Corporation (IBM) in Armonk, New York.

RESULTS AND DISCUSSION

Effects on Hematological Indices: Results showed that the RBC, Hb concentration, PVC, MCV, MCH, and MCHC of rats in group B was significantly (P<0.05) decreased when compared with the control. These parameters were not significantly (P>0.05) different in rats treated with α-Tocopherol only when compared with control. However, there was a significant increase in these parameters in group C rats (α-Tocopherol + lead acetate) when compared with group B rats treated with lead acetate only (Table 1).

Table 1: Effects of α-tocopherol and lead acetate administration on packed cell volume, red blood cell count, hemoglobin concentration, and erythrocyte indices

<table>
<thead>
<tr>
<th></th>
<th>A (Control)</th>
<th>B (Lead acetate only)</th>
<th>C (α-Tocopherol + Lead Acetate)</th>
<th>D (α-Tocopherol)</th>
</tr>
</thead>
<tbody>
<tr>
<td>RBC Count (x 10^6/μl)</td>
<td>7.70 ± 0.11</td>
<td>6.03 ± 0.36 *</td>
<td>6.90 ± 0.51 *</td>
<td>7.54 ± 0.26</td>
</tr>
<tr>
<td>Hb (g/dl)</td>
<td>13.27 ± 0.07</td>
<td>10.43 ± 0.52 *</td>
<td>13.23 ± 0.18 *</td>
<td>12.33 ± 0.24</td>
</tr>
<tr>
<td>PCV (%)</td>
<td>42.20 ± 0.20</td>
<td>30.58 ± 1.42 *</td>
<td>40.80 ± 0.76 *</td>
<td>38.43 ± 0.70</td>
</tr>
<tr>
<td>MCV (fl/cell)</td>
<td>54.77 ± 0.53</td>
<td>36.77 ± 3.68 *</td>
<td>51.03 ± 1.13 *</td>
<td>52.20 ± 1.13</td>
</tr>
<tr>
<td>MCH (pg/dl)</td>
<td>17.50 ± 0.20</td>
<td>15.13 ± 0.07 *</td>
<td>16.97 ± 0.69 *</td>
<td>16.47 ± 0.03</td>
</tr>
<tr>
<td>MCHC (g/dl)</td>
<td>31.40 ± 0.00</td>
<td>26.73 ± 0.75 *</td>
<td>32.43 ± 0.32 *</td>
<td>32.07 ± 0.03</td>
</tr>
</tbody>
</table>

Data is represented as Mean ± SEM, * and # represent P<0.05 when compared with control and Lead acetate groups respectively.
It was also observed that the Lymphocyte, Granulocyte, Monocyte counts, and the WBC of rats in group B were significantly \((P<0.05)\) increased when compared with the control. These parameters were not significantly \((P>0.05)\) different in rats treated with \(\alpha\)-Tocopherol only when compared with control. However, there was a significant decrease in these parameters in group C rats \((\alpha\)-Tocopherol + lead acetate) when compared with group B rats treated with lead acetate only (Table 2). In addition, there were no significant \((P>0.05)\) differences in the Platelet count of rats across experimental groups.

### Table 2: Effect of \(\alpha\)-tocopherol and lead acetate administration leucocyte count and leucocyte indices.

<table>
<thead>
<tr>
<th></th>
<th>A (Control)</th>
<th>B (Lead acetate only)</th>
<th>C ((\alpha)-Tocopherol + Lead Acetate)</th>
<th>D ((\alpha)-Tocopherol)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WBC Count (x 10^3/\mu l)</td>
<td>9.13 ± 0.47</td>
<td>14.57 ± 0.52 *</td>
<td>10.17 ± 0.62 *</td>
<td>9.87 ± 0.20</td>
</tr>
<tr>
<td>Lymphocyte Count (x 10^3/\mu l)</td>
<td>4.53 ± 0.44</td>
<td>8.70 ± 0.66 *</td>
<td>6.17 ± 0.15 *</td>
<td>5.83 ± 0.59</td>
</tr>
<tr>
<td>Granulocyte Count (x 10^3/\mu l)</td>
<td>4.10 ± 0.10</td>
<td>5.13 ± 0.24 *</td>
<td>4.07 ± 0.44 *</td>
<td>3.90 ± 0.17</td>
</tr>
<tr>
<td>Monocyte Count (x 10^3/\mu l)</td>
<td>1.02 ± 0.06</td>
<td>1.34 ± 0.19 *</td>
<td>1.18 ± 0.20 *</td>
<td>1.22 ± 0.18</td>
</tr>
<tr>
<td>Platelet Count (x 10^3/\mu l)</td>
<td>612.00 ± 21.00</td>
<td>550.00 ± 18.00</td>
<td>608.00 ± 29.46</td>
<td>528.33 ± 52.454</td>
</tr>
</tbody>
</table>

Data is represented as Mean ± SEM; * and # represent \(P<0.05\) when compared with control and Lead acetate groups respectively.

**Effects on Bone Marrow Histology:** The photomicrographs of control group A showed normal meshwork of bone trabeculae (BT) with the intertrabecular spaces containing hemopoietic cells with adequate cellularity (HC). Group B rats treated with lead acetate showed mild hypoplasia of haematopoietic cells (HP) and severe vacuolation (V). Group C rats treated with \(\alpha\)-tocopherol and lead acetate showed relatively normal features of the bone marrow with few vacuolations. Group D rats treated with \(\alpha\)-tocopherol showed normal meshwork of bone trabeculae with the intertrabecular spaces containing hemopoietic cells with adequate cellularity (Figure 1).

![Fig 1: Histology of the bone marrow across experimental groups.](image)

The major important target of lead is the hematological system and induces changes in the composition of red blood cell membrane protein, and lipids and inhibits the synthesis of hemoglobin (Collin et al., 2022). This present investigation revealed that PCV, Hb concentration, MCV, MCH, MCHC, and RBC of rats treated with lead acetate were significantly decreased compared with control, in agreement with previous studies (El-Bahr et al., 2021; Suradkar et al., 2009). Reduction in Hb concentration has been attributed to the ability of lead to convert co-protoporphyrinogen III to protoporphyrin IX (Sachar et al., 2016). A reduction in RBC count, haemoglobin, and packed cell volume has been linked to decreased rate of erythrocyte production or an increased rate of erythrocyte loss (Sutton and Sellon, 2013). Reports show that lead may inhibit the body’s ability to make hemoglobin via interference with several enzymatic steps in the heme pathway. Precisely, lead reduces heme biosynthesis by impeding aminolaevulinc acid dehydratase and ferro chelatase activity (El-Shater et al., 2022). Lead affects the loss of Hb molecule stability and erythrocyte
morphology and survival (El-shater et al., 2022). In this study, treatment with α-tocopherol significantly increased PCV, Hb concentration, MCH, MCV, and MCHC when compared with rats treated with lead acetate alone. The mechanism by which α-tocopherol ameliorates lead-induced hematological changes is not clear but reports have linked this effect to its potent antioxidant properties (Antosik et al., 2018; Das et al., 2012). In agreement with previous studies, the WBC count, Lymphocyte count, and Platelet count of rats treated with lead acetate were significantly increased when compared with control (Andjelkovic et al., 2019; El-shater et al., 2022). White blood cells, also known as Leucocytes, play a vital role in protecting the body against foreign invaders. They are produced by multipotent cells in the bone marrow and circulate in the blood and the lymphatic system (El-shater et al., 2022). Elevated WBCs could be a result of inflammation induced by lead Increased Pb concentration in the blood induced an increase in leukogram including lymphocytes, neutrophils, eosinophils, and monocytes (Abubakar et al., 2019). Leukocytosis with lymphocytosis, indicative of bone marrow toxicity and implicated in the development of a lymphoproliferative neoplasm has been linked to lead toxicity (Aprioku and Obianime, 2014; El-shater et al., 2022). This study demonstrated that the values of WBC count, Lymphocyte count, and Platelet count were significantly restored to near normal following treatment with α-tocopherol. This effect against lead acetate may be due to its anti-inflammatory and immunomodulatory effects, as previously reported (Lewis et al., 2019; Nazrun et al., 2012).

The histology of the bone marrow of rats treated with lead acetate showed mild hypoplasia of haematopoietic cells with severe vacuolation. These observations are in agreement with other studies showing lead acetate-induced toxic damage to the normal functioning of bone and bone marrow (Haleagrahara et al., 2010; Othman et al., 2004). In some studies, lead toxicity was reported to cause a weak clastogenic effect on rat bone marrow cells which was attributed to excessive ROS generation or failure of the cellular antioxidant system (Haleagrahara et al., 2010; Hamadouche, 2009). However, in lead acetate-exposed rats treated with α-tocopherol, there were relatively normal features of the bone marrow with few vacuolations, thus indicating that α-tocopherol treatment may have a bone marrow growth stimulating effect.

Conclusion: Findings from this study demonstrated that α-tocopherol protected the haematological system and bone marrow of experimental rats from the toxic effects of lead acetate and highlighted its usefulness as a possible therapeutic agent against disorders linked to lead exposure. Further studies are recommended to investigate its mechanisms of action in other models of lead toxicity.

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