Dinorditerpenoids From Ricinodendron Heudelotii* (Euphorbiaceae)

Jean MOMENI^a; Thomas M. AKAM^b; Samuel F. KIMBU^{a+}; Wolfgang KREISER^c and Joseph D. CONNOLLY^d

- a) Department of Organic Chemistry, Faculty of science, University of Yaounde I, P.O. Box 812 Yaounde, Cameroon.
- b) Department of Chemistry, Faculty of Science, University of Buea, P.O. Box 63 Buea, S. W. Province, Cameroon.
- c) Naturstoff Chemie, Universitat Dortmund, 44221 Dortmund, Germany.
- d) Department of Chemistry, University of Glasgow, Glasgow G12 8QQ, Scotland.

ABSTRACT

A new dinorditerpenoid, heudelotenol 1 along with twelve known compounds namely octancosan-1-ol, E-ferulic acid n-octacosylate, stigmasterol, campesterol, â-sitosterol, â-sitosterol glycoside, taraxerone, friedelin, aleuritolic acid, heudelotinone, 1,2-dihydroheudelotinol and labda-8(17),13-dien-3â,15-diol were isolated from *Ricinodendron heudelotii*. The structure of heudelotenol 1 was established by spectral analysis and chemical transformations.

Key words: Ricinodendrn heudolotii; Euphorbiaceae; dinorditerpenoids

RÉSUMÉ

Heudelotenol 1, un nouveau dinorditerpène et douze autres composés connus : octancosan-1ol, E-ferulate de n-octacosyle, stigmasterol, campesterol, â-sitosterol, glucoside de â-sitosterol, taraxerone, friedeline, acide aleuritolique, heudelotinone, 1,2-dihydroheudelotinol et labda-8(17),13-diène-3â,15-diol ont été isolés du *Ricinodendron heudelotii*. La structure de heudelotenol 1 a été établie à partir des données spectroscopiques et des transformations chimiques.

Mots clés: Ricinodendrn heudolotii; Euphorbiacée; dinorditerpènes

^{*} Author to whom correspondence should be addressed

^{*} Part 2 in the series Studies on Ricinodendron heudelotii

INTRODUCTION

Earlier studies on this plant (KIMBU et al, 1991) resulted in the isolation of two new dinorditerpenoids heudelotinone 3 and 1,2-dihydroheudelotinol 4 and three known compounds E-ferulic acid octacosylate, 3-methylorselinate and lupeol. In continuation of our research work, an extract of this plant collected from Muyuka, South West Province, Cameroon has yielded the known compounds mentioned above together with a new dinorditerpenoid. Only one of the dinorditerpenoids heudelotinone 3 which was identified earlier in the plant collected from Yaounde was found in the extract of the plant collected from Muyuka (Momeni et al, 2005). This paper describes the results of our investigation.

EXPERIMENTAL

All melting points were determined on a Büchi melting point apparatus model 510 and are uncorrected. UV spectra were measured in methanol solutions using the Anthorpc Version 4.1 H spectrophotometer. Mass spectra were recorded using MAT 312 or MAT 8200 instruments attached to a MSCAN computer. Nuclear magnetic resonance (NMR) spectra were recorded on Bruker AMX 400 instrument (¹H, 400 MHZ, ¹³C 75.5 MHZ) using tetramethylsilane as internal reference in CDCl₃ or CD₃OD or C₅D₅N where appropriate. Chemical shifts are given in δ values.

2-D NMR, ¹H-¹H COSY, HMBC, HMQC and NOESY experiments were performed with usual pulse sequences and data processing was obtained with adequate software.

PLANT MATERIAL

The stem bark of *Ricinodendron heudelotii* was collected in Muyuka, South West Province, in August 1997. Herbarium specimens documenting the collection are deposited at the National Herbarium, Yaounde, Cameroon.

EXTRACTION AND ISOLATION OF COMPOUNDS

Dried powdered stem bark of R. heudelotii (7.7 kg) was extracted with a mixture methylene chloride/methanol (1:1) at room temperature. The methylene chloride/methanol extract was filtered and evaporated under reduced pressure in a rotatory evaporator to obtain 175 g of a crude extract.

Dried leaves of *R. heudelotii* (290 g) were extracted with a mixture of methylene chloride/methanol (1:1) at room temperature. The methylene chloride/

methanol extract was filtered and evaporated under reduced pressure in a rotatory evaporator to yield 30 g of a crude extract. Dried powdered roots of *R. heudelotii* (2.0 kg) were extracted with a mixture of methylene chloride/methanol (1:1) at room temperature. The methylene chloride/methanol extract was filtered and evaporated under reduced pressure in a rotatory evaporator to obtain 20 g of a crude extract. Comparison of this extract by thin layer chromatography (TLC) with one obtained from the stem bark, showed that they were identical and therefore combined.

The plant extracts obtained above were separated into different fractions A, B, C, D, E by flash chromatography. These fractions were then chromatographed over silica gel columns and fractions which could not be separated on columns of silica gel were further purified by preparative TLC over Merck kieselgel GF 254. Spots on analytic TLC plates were observed under uv light and by spraying with concentrated sulphuric acid. Eluents for column chromatography were hexane and increasing percentages of ethyl acetate in hexane. TLC plates were developed using appropriate concentrations of ethyl acetate in hexane. Thus the chromatographic separation of the mixture of compounds in fraction C afforded octacosan-1-ol (90 mg), Eferulic acid octacosylate (450 mg), a mixture of sterols, stigmasterol, campesterol, and β-sitosterol (400 mg). Fraction D afforded heudelotenol (120 mg); fraction E heudelotinone (12 mg); fraction F aleuritolic acid (90 mg) and fraction G β-sitosterol glycoside (400 mg).

Chromatographic separation of the crude extract from the leaves from Muyuka over silica gel and elution with hexane followed by hexane-ethyl acetate afforded friedelin (10 mg) and taraxerone (10 mg). The known compounds isolated were identified by comparison (mp, ¹H, ¹³C NMR) with published data.

Heudelotenol 1

White crystalline solid (120 mg); mp: 207-209 °C; $C_{19}H_{22}O_4$; IE m/z (rel. int. %) 314 [M]+ (98), 271 (49); 258 (100); 244 (58); 243 (58); 231 (58); 230 (88); 215 (54); 202 (50); 115 (56); 83 (39); 77 (35); 51 (29). IR (KBr, v_{max}) cm⁻¹: 3400 (OH); 1712 (α, β- unsaturated C=O); 1691 (simple C=O); 1655, 1601, 1522 (aromatic C=C), 1500; ¹H NMR (400 MHz, CDCl₃) and ¹³C NMR (75.5 MHz, CDCl₃) see Tables 1 and 2

Table 1: ¹H NMR (400 MHz, CDCl₃) Assignments or compounds (1) and (2)

			-			
Compound	δ_{H}	J (Hz)		Compound 2	$\delta_{\rm H}$	J (Hz)
1						
O-H	7.12; s	-		O-CO-C <u>H</u> 3	2,36; s	-
H-1ax	2.07; dt	13.7; 9.9)	H-lax	2.14	₄ 13.9; 9.5
H-1eq	2.46; ddd	13.7;	9.3;	H-leq	2.50	13.7; 9.3
H–2ax	2.76; ddd	1.5 18.8; 8.0	9.8;	H–2ax	2.80; dd	18.8; 9.8
H-2eq	2.76; ddd	18.8;	9.8;	H–2eq	2.80; dd	18.8; 9.8
H-11	6.81; s	, <u> </u>		H-11	6.77; s	-
H-14	7.94; s	-		H-14	7.91; s	-
H-15	2.23; s	-		H-15	2.22; s	-
H-18	1.52; s	-		H-18	1.40; s	-
H-19	1.55; s	-		H-19	1.40; s	-
H-20	1.28; s	-		H-20	0.82; s	-
O-C <u>H</u> 3	3.90; s	-		O-C <u>H</u> ₃	3.89; s	-

Table 2: ¹³C NMR (75.5 MHz, CDCl₃) Assignments for compounds 1, 2 and 5)

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Attribution	Compound 1	Compound 2	Compound 5
1	32.7 t	32.9 t	27.05 t
3	33.0 t	37.8 t	33.08 t
3	214.3 s	195.4 s	214.50 s
4	48.5 s	48.4 s	48.7 s
5	142.3 s	142.2 s	140.0 s
6	151.0 s	149.1 s	140.1 s
7	179.5 s	176.8 s	182.9 s
8	120.5 s	122.3 s	107.2 s
9	137.1 s	137.1 s	135.2 s
10	39.1 s	41.0 s	40.6 s
11	106.1 d	105.7 d	131.2 s
12	162.2 s	162.1 s	153.8 s
13	127.0 s	129.2 s	111.5 s
14	128.8 d	150.9 d	154.8 s
15	26.2 q	26.2 q	34.4 t
16	_	-	83.5 d
17	-	-	22.0 q
18	24.4 q	25.7 q	24.4 q
19	20.9 q	21.0 q	21.1 q
20	15.8 q	15.6 q	20.1 q
O- <u>C</u> H ₃	55.6 q	55.6 q	-
$CH_3 C = O$	-	168.6 s	-

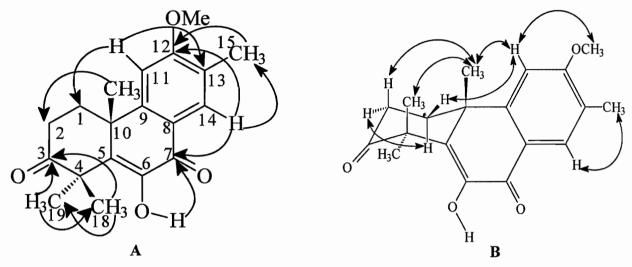


Fig. 1: Pertinent correlations observed with heudelotenol (1) (A); HMBC correlations (Htoc) (B) NOESY correlations.

Heudelotenol Acetate 2

Compound 1 (25 mg) in pyridine (5 ml) and acetic anhydride were allowed to stand at room temperature for 24 h. Methanol (25 ml) was added to the reaction mixture and the solution obtained passed over activated alumina. The resulting solution was evaporated to dryness under reduced pressure in rotatory evaporator. The acetate (20 mg) was obtained by crystallisation in a mixture of hexane/ethyl acetate (3:2).

RESULTS AND DISCUSSION

Compound 1 was isolated as a white crystalline solid, mp. 207-209°C, from the methylene chloride/ methanol extract of the dried powdered stem bark/roots as described in section 3. Its molecular formula C₁₀H₂₂O₄ was deduced from microanalysis EIMS and NMR data. It showed IR absorption bands for an hydroxyl group at V_{max} 3400 cm⁻¹; an α, β- unsaturated carbonyl at 1712 and a simple carbonyl function at 1691 cm⁻¹. The ¹H NMR spectrum of compound 1, revealed the presence of three tertiary methyl groups [$\delta_{\rm H}$ 1.52 (3H, s); 1.55 (3H, s) and 1.28 (3H, s)], two aromatic protons [$\delta_{\rm H}$ 7.94 (1H, s) and 6.81 (1H, s)], one aromatic methyl group $[\delta_{\rm H}$ 2.23 (3H, s)] and one methoxy group $[\delta_{\rm H}$ 3.90 (3H, s)]. The ¹³C NMR and DEPT spectra of compound 1 showed two carbonyl carbons at δ_c 214.3 and 179.5; eight sp² carbons at $\delta_{\rm C}$ 106.1, 120.0, 127.0, 128.8, 137.1, 142.3, 151.0 and 162.2. Other signals observed include two methylenes, two methines and two quaternary carbons (Table 2).

Thus compound 1 is a tricyclic dinorterpenoid including an aromatic ring, two carbonyl groups and an additional tetrasubstituted double bond. The appearance of only two aromatic protons signals in the ¹H NMR spectrum suggest that the benzene ring is tetrasubstituted bearing a methyl at H 2.23 and one methoxy group at $\delta_{_{\rm H}}3.90$. The location of the different groups on the tricyclic skeleton which is similar to that of teuvincenone A 5 (CARRIERAS et al, 1990.) was accomplished by HMBC and NOE correlations. Differences between the two compounds were the absence of two hydroxyl groups on aromatic ring, C-16 and C-17. The 'H NMR spectrum showed the presence of two adjacent groups [$\delta_{\rm H}$ 2.01 (dt 13.7, 9.9 Hz); 2.46 (ddd 13.7, 9.3; 1.5 Hz); 2.76 (ddd 18.8, 9.0, 8.0 Hz); 2.76 (ddd 18.8, 9.8, 2.2 Hz)] (Table 1) one of which is a to the carbonyl group. Analysis of the HMBC spectrum (summarized in Fig. 1A) showed how various parts were fixed together and pertinent correlations were observed on the spectrum between protons H-14 and C-7, C-12, C-15; H-11 and C-9, C-13, C-1; H-20 and C-2, C-5 and between H-19 and C-18, C-3. These long range couplings in the HMBC spectrum enabled the assignment of the structure of the new compound as 1 and this was further confirmed by correlations observed in the NOE's (summarized in Fig. 1B) and the fragmentations observed in the mass spectrum (Fig.

Acetylation of heudelotenol 1 with acetic anhydride in pyridine gave a white crystalline compound

Fig. 2: Principal fragmentations of heudelotenol acetate (2)

which gave M⁺ m/z 356 corresponding to the molecular formula C₂₁H₂₄O₅. Conspicuously absent in the IR spectrum of compound 2 was the absorption band at 3400 cm⁻¹ for the hydroxyl group. Comparison of the ¹H NMR spectroscopic properties of the acetylated product (2) and (1) (Table 1) showed much similarity the main difference being

the absence of the signal at $\delta_{\rm H}$ 7.10 for the hydroxyl group and the presence of the additional signal at $\delta_{\rm H}$ 2.36 for the acetyl group. Thus acetylation transformed the additional group giving compound 2.

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