# THE NUTRIENT AND PHYTOCHEMICAL COMPONENTS OF Monodara myristica SEED

Obiloma Appolonia A.<sup>1</sup>, Madu Wenceslaus C.<sup>2</sup>, Osuji Godson O.<sup>3</sup> and \*Ogbonna Patrick C.<sup>1</sup>,

<sup>1</sup>Department of Nutrition and Dietetics, Imo State Polytechnic, Umuagwo, Imo State, Nigeria.

<sup>2</sup> Science Laboratory Technology, Imo State Polytechnic, Umuagwo, Imo State, Nigeria.

<sup>3</sup> Cooperative Agricultural Research Center, Prairie View A&M University, Prairie View, Texas, USA.

# \*Corresponding author: patogbonna22@yahoo.com

## **Abstract**

**Background:** Monodora myristica is one of the common herbal spices used in the ethnomedicine of the Igbo people of Southeastern Nigeria alternative medicine. It is claimed to possess valued therapeutic and nutritive properties.

**Objective**: The aim of this study was to determine the nutrient and phytochemical components of *M. myristica* seed, sold in Owerri, Imo State, Nigeria.

**Methods**: The seed was processed into spice powder before subjection to proximate, minerals, phytochemicals and terpenes evaluations using standard methods.

**Results:**The proximate composition found: Protein 3.85±0.10%, moisture 9.02±0.37%, fat 9.74±0.02%, ash 11.75±0.21%, crude fiber5.85±0.41% and carbohydrate64.65±1.31%. Vitamins present were Vitamin A 1.27±0.03%, Vitamin .B 0.63±0.02% and Vitamin C 0.34±0.02%. The spice contained the mineral elements potassium, sodium and magnesium in high amounts, and calcium, iron, manganese, copper and zinc in low amounts. Tannins, alkaloids, flavonoids, saponins, phytates, oxalates and phenols were the phytochemicals detected. Delta 3 carene was the major terpene detected at 0.4231% followed by rho cymene (0.2506%), alpha pinene (0.1567%), beta myrcene (0.1102%). Majority of the terpenes were present in trace amounts. The spice powder is vitamins and minerals rich.

**Conclusion:** The medicinal functions of the plant can be attributed to its diverse phytochemical potentials.

**Keywords:** *Monodora myristica*, proximate, minerals, phytochemicals, terpenes.

#### Introduction

Monodora myristicais a perennial tree of the family Annonecea native that thrives in evergreen forest. It is known colloquial as 'Ehuru' (Igbo), 'Ariwo' (Yoruba), 'Awerewa' (Hausa) and African nutmeg in Nigeria generally [1]. M. myristica is an important economic tree used as timber because it is hard and easy to work on [2]. The seeds which are embedded in a white sweetsmelling pulp of the sub-spherical fruit are the most economically important [3]. Studies have shown that the seeds are used as condiment for their aromatic flavour and medicinal potency. The kernel is obtained by cracking the hard nut after roasting and ground into powder for use as spice in traditional cuisines in Nigeria [4]. Ethnomedicinally, M. myristica is used to treat arthritis, rheumatism, haemorrhoids, cutaneous and subcutaneous infections [5]. The aromatic seeds are used widely as tonic, antiemetic, aperients, stimulant, and stomachic [6].

Spices contain nutrients required for growth and normal body functions and as such add to the bulk of nutrients in a diet to alleviate the effects of their deficiencies [7]. Spices also contain phytochemical such as alkaloids, glycosides, flavonoids, tannins, saponin and terpenes which are responsible for their medicinal properties [8-10]. *M. myristica* considered as a cheap substitute to nutmeg for the preparation of spicy meals contains several nutritional and non-nutritional compounds [11]. Plants produce Phytochemicals which have a wide array of functions both for the benefit of animals and humans [12]. Phytochemicals are secondary metabolites found in plants that exert physiological and metabolic effects on humans, which might be beneficial for the prevention

and management of several diseases [13-15]. Beneficial effect of phytochemicals against cancers, coronary heart disease, diabetes, high blood pressure, inflammation, microbial and parasitic infections, psychotic diseases, spasmodic conditions, ulcers have been reported [15,16]. Terpenes represent the largest class of phytochemicals in several plants with diverse bioactive properties [17]. Spices are increasingly finding useful applications in medicine aside their common culinary usage [8]. For the full nutritional and health benefits of any spice to be harnessed, a compositional study needs be carried out. Therefore, the aim of this study was to evaluate the nutrient and phytochemical contents of *M. myristica* spice powder sold in Owerri, Imo State, Nigeria.

# **Materials and Methods Sample Collection**

Monodara myristica seeds were purchased from Relief Market in Owerri Municipal, Imo state, Nigeria. The samples were identified by plant taxonomists at Imo state Polytechnic Umuagwo.

# **Sample Preparation**

The *M. Myristica* seeds were dried at room temperature and in a cabinet oven at temperature of 60°C for 30 minutes. The kernel was removed before grinding with Marlex electroline grinder to obtain a smooth powder. The preparation process is shown in Figure 1.



**Figure 1:** Flow chart for processing of *M. myristica* spice powder

## Chemical analysis

The moisture, ash, fat, protein, crude fiber and carbohydrate content of *M. myristica* were determined using standard AOAC methods [18]. Vitamins A, B<sub>1</sub> and C were determined by Pearson [19] method. The atomic absorption spectrophotometer (AAS) method of AOAC [18] was used to determine mineral (iron, calcium, zinc, copper, manganese, sodium, magnesium and potassium) and heavy metal content of the spices. Terpene content was determined using FET headspace GC-FID according to method of Hilliard *et al.* [20]. The analysis was carried out by ANRESCO Laboratories Inc., San Francisco, CA, USA.

subjected to statistical analysis using SPSS v20 to obtain mean and standard deviation.

#### Results

The mean was calculated from duplicate determination. Proximate composition of M. myristica seed presented in Table 1, revealed protein content of  $3.85\pm0.10\%$ , moisture  $9.02\pm0.37$ , fat  $9.74\pm0.02$ , ash  $11.75\pm0.21$ , crude fibre 5.85.41 and carbohydrate  $3.85\pm0.10\%$ .

# **Statistical Analysis**

The data obtained from different analyses were

Table 1: Proximate composition of M. Myristica spice powder

Property	Percentage (%)	
Protein	$3.85 \pm 0.10$	
Moisture	$9.02 {\pm} 0.37$	
Fat	$9.74 \pm 0.02$	
Ash	$11.75 \pm 0.21$	
Crude fiber	$5.85 \pm 0.41$	
Carbohydrate	64.65±1.31	

Table 2 shows the mineral contents (Potassium, sodium magnesium, calcium, iron, manganese, copper and zinc) evaluated.

**Table 2:** Mineral composition of *M. Myristica* spice powder

Property	Percentage (%)	
Mg	$0.266 \pm 0.02$	
K	$1.124\pm0.01$	
Na	$0.191 \pm 0.01$	
Ca	$0.003 \pm 0.01$	
Mn	$0.001 \pm 0.006$	
Fe	$0.009 \pm 0.003$	
Cu	$0.002 \pm 0.001$	
Zn	$0.001 \pm 0.001$	

Table 3 shows the vitamin composition *M. myristica* spice powder. Vitamins A, B<sub>2</sub> and C were analyzed, with concentrations  $1.27\pm0.03\%$ ,  $0.63\pm0.02\%$  and  $0.34\pm0.02\%$  respectively.

**Table 3:** Vitamin composition of *M. myristica* spice powder

Property	Percentage (%)	
Vitamin-A	$1.27 \pm 0.03$	
Vitamin-B <sub>2</sub>	$0.63\pm0.02$	
Vitamin-C	$0.34 \pm 0.02$	

Table 4 revealed the result for phytochemicals analyzed; tannins, alkaloids, flavonoids, saponins, phytates, oxalates and phenols.

Property	Result (mg/Kg)	
Tannin	1.20±0.01	
Alkaloid	$13.42 \pm 0.13$	
Flavonoid	$3.10\pm0.02$	
Saponin	$2.84 \pm 0.07$	
Phytate	$15.20 \pm 0.05$	
Oxalate	$0.79 \pm 0.01$	
Phenol	$5.50\pm0.14$	

Table 5 shows the terpene profiling result of the M. myristica spice powder, camphene,

**Table 5:** Terpene profile of *M. myristica* spice powder

Analyte	0/0	
a –Pinene	0.1567	
Camphene	ND	
β-Myrcene	0.1120	
β –Pinene	0.0089	
d 3-Carene	0.4231	
Limonene	0.0798	
α-Terpinene	0.0018	
Ocimene 1	0.0299	
Ocimene 2	0.0366	
p-Cymene	0.2506	
Eucalyptol	BLOQ	
y-Terpinene	0.0020	
Terpinolene	BLOQ	
Linalool	0.0628	
Isopulegol	ND	
Menthol	ND	
(-)-Borneol	BLOQ	
Terpineol	0.0098	
Citronellol	0.0332	
Geraniol	BLOQ	
β –Caryophyllene	0.0049	
α –Humelene	0.0031	
Nerolidol 1	0.0060	
Nerolidol 2	ND	
Guaiol	ND	
Caryophyllene Ox	ND	
α -Bisabolol	ND	
Eudesmol	ND	
TOTAL	1.2212	

ND = Not Detected; BDL = Below Detection Limit (0.0015%)

## Discussion

The form and the analytical methods used in analysis can actually affect quantitative components or active ingredients in plant samples. The proximate composition of M. Myristica showed it contained protein 3.85±0.10%, moisture 9.02±0.37%, fat  $9.74\pm0.02\%$ , ash  $11.75\pm0.21\%$ , crude fibre 5.85±0.41%, and carbohydrate 64.65±1.31 %. The protein value (3.85±0.10) was significantly lower than previous findings[51] 12% and [52] 14.02±0.87%. This difference may be attributed to the extraction methods used [53]. Soil content and period of harvest of the sample may have affected the value. Although, M. myristica is only added as a seasoning as such does not contribute significantly to daily protein intake. The value of carbohydrate shows that M. myristica seed is a good source of carbohydrate. The fat (9.74±0.02%) and protein (3.85±0.10 %) contents were low compared to values of  $12.35\pm0.05\%$  and  $14.78\pm0.15\%$  reported in previous studies [21] while moisture and crude fibre contents are comparable to values of 8.78±0.02% and 4.93±0.02% in the same study. The moisture content was low which implies that the spice can be stored for a long a time without deterioration in quality or microbial spoilage. Dietary fibre helps in bowel movement to prevent constipation [22]. Fibre also have antiinflammatory function and may contribute to reduce symptoms or bring relieve from inflammatory bowel diseases [23].

The ash content 11.75±0.21% indicates high mineral presence in the spice. Potassium, sodium and magnesium, calcium, iron, manganese, copper and zinc were the minerals present in the spice powder. Minerals constitute only 4–6% of the human body but they play critical roles in metabolism and cellular function [24]. According to Sanchez-Castillo et al. [25] consumption of minerals such as sodium, potassium, magnesium, calcium, manganese, copper and zinc could potentially reduce risk of cardiovascular and other diseases. Magnesium, potassium, and sodium were the most abundant and they are among the major mineral elements. Sodium and potassium are responsible for osmotic equilibrium in the body [26]. Magnesium is an essential cofactor in many enzymatic reactions in intermediary metabolism [27].

With the exception of calcium the other minerals present in trace amounts are among the trace element. Calcium is essential for bone formation, muscular, digestive and circulatory system function. The mineral also have regulatory effects on proteins and enzymes [28]. Zinc is an essential trace element needed for proper central nervous system function and for hormonal balance in the body [29]. Zinc is a co-factor in the function of enzymes and influence gene expression for protein synthesis [30]. It has thyroid [31], nervous system and hormonal balance functions [29], and ensures stronger immunity [32]. Special interest is been paid to spice for their minerals and vitamin content to tackle the problem of deficiency in diet [7]. Vitamin A is needed for good vision, growth and development, and immune system function Tannins (1.20±0.01 mg/kg), alkaloids (13.42±0.13 mg/kg), flavonoids (3.10±0.02 mg/kg), saponins  $(2.84\pm0.07 \text{ mg/kg})$ , phytates  $(15.20\pm0.05 \text{ mg/kg})$ , oxalates (0.79±0.01 mg/kg) and phenols (5.50±0.14 mg/kg) were the phytochemicals detected in the seeds of M. myristica. Similar phytochemicals with the exception of phytate were reported present in the seeds of M. myristica in another study in Umuahia, Abia State [8]. Phytochemicals are known to have physiological and metabolic roles in living system [35, 8]. Plant alkaloids have analgesic, antispasmodic and antibacterial effect [36]; flavonoids have antioxidant properties [38]; plant phenolics have antioxidant function [39]; saponins serve as expectorant and emulsifying agent [35]; tannins can inhibit microbial growth and metabolism [40] and steroid have cardiovascular and hormonal functions [41].

The *M. myristica* seeds contained Delta 3 carene, p-cymene, alpha pinene, beta myrcene, beta pinene, limonene, alpha terpinene, gamma terpinene, linalool, citronellol, terpineol, beta caryophylleneocimene 1, alpha humelene, nerolidol 1, ocimene 2 and and p-cymene. Nguefack [2] reported that the seeds of *M. myristica* contained alpha phellandrene which was not detected in this study and alpha pinene, myrcene, limonene and beta pinene which were detected.

Terpenes are the largest class of chemical compounds in plant [17]. They have gained much attention because of their importance in plants physiology as structural compounds and for the defensive role they play in plant ecology [42,43]. These volatile aromatic compounds are beneficially to man industrially as solvents, cleaning agents and as flavour and fragrance additive [44, 45]. In medicine, alpha-pinine regarded as one of the most potent and widely distributed terpene among plants, functions as bronchodilator, anxiolytic, anticancer, anti-inflammatory and antimicrobial agent [46]. P-cymene is reported to have anti-inflammatory and antidiabetic effect [47]. Limonene is used in traditional medicine to provide relieve from heartburn, gallstone and gastro-esophageal reflux disease [48]. Intake of 1000 mg of d-limonene has been shown to reduce symptoms of gastro-esophageal reflux disease [49]. Safety and toxicity report suggest that high doses of terpenes can cause allergic dermatitis [45], affect fertility and reproductive performance [50].

# Conclusion

From a nutritional view point seeds of *M. myristica* is rich in minerals and vitamins which have nutritional benefits. Though they are part of the major staples consumed, their nutritional quality makes them important in supplementing the vitamins and minerals present in diets to which they are added. They also contain several beneficial phytochemicals which have functional properties as antioxidants. The challenge in

the use is to produce it in commercial quantities and forms which will have easy introduction in our delicacy

## REFERENCES

- 1. Enabulele, S.A, Oboh, F.O.J., and Uwadiae, E.O. (2014). Antimicrobial, Nutritional and Phytochemical properties of *Monodoramyristica* seeds. *Journal of Pharmacy and Biological Sciences* 9(4): 1-6.
- Nguefack, J., Leth, V., Amvam, V., and Zollo, P.H,. (2004). Evaluation of five essential oils from aromatic plants of Cameroon for controlling food spoilage and mycotoxin producing fungi. *Int. J. Food Microbiol*.94(3):329–334.
- 3. Burubai, W., Akor, A.J, Igoni, A.H, and Puyate, Y.T. (2007). Effects of temperature and moisture content on the strength properties of African nutmeg (Monodoramyristica). International Agrophysics. 21: 217–223.
- 4. Ekeanyanwu, C.R., Ogu, I.G, and Nwachukwu, U.P. (2010). Biochemical Characteristics of the African Nutmeg, *Monodoramyristica Agricultural Journal*. 5(3): 303-308.
- 5. Burkill, H.M.(1985). The useful plants of West Tropical Africa, 2<sup>nd</sup> ed. Royal botanic gardens, kew, UK,. Available at http://plants.jstor.org/upwta/2\_580.
- Agiriga, A., and Siwela, M.,(2017).
   *Monodoramyristica* (Gaertn.) Dunal: A
   Plant with Multiple Food, Health and
   Medicinal Applications: A Review.
   *American Journal of Food Technology.* 12:
   271-284.
- 7. Borquaye, L.S., Darko, G., Laryea, M.K., Gasu, E.N., Amponsah, N.A.A., and Appiah, E.N. Nutritional and anti-nutrient profiles of some Ghanaian spices. *Cogent Food & Agriculture*. 3: 1348185
- 8. Osuagwu, G.G.E., and Onwuegbuchulam, N.P. (2015). The Phytochemical Screening and antimicrobial activity of the leaves of *Monodoramyristica*, (Gaertn) Dunal, *Acanthus montanus*(Ness) T. Anders and *Alstoniabonnei* De wild. *International Journal of Pharmacy and Pharmaceutical Research*. 2(4): 85-102.
- 9. Erhirhie, E.O., and Moke, G.E. *Xylopiaaethiopica*: A review of its ethnomedicinal, chemical and

- pharmacological properties. *Am. J. PharmTech Res.* 4(6):21-37.
- 10. Imo, C., and Uhegbu, F.O. (2015).

  Phytochemical analysis of GongronemalatifoliumBenth leaf using gas chromatographic flame ionization detector. International Journal of Chemical and Biomolecular Science. 1(2): 60-68.
- Nigeria Natural Medicine Development Agency (NNMDA) (2008). Medicinal plants of Nigeria; South – East Nigeria. Vol.1. Lisida consulting Lagos Nigeria. pp. 32
- Croteau, R., Kutchan, T.M., and Lewis, N.G., (2000). Natural products (secondary metabolites). In Buchanan B, Gruissem W, Jones R (Eds.), Biochemistry and molecular biology of plants. Rockville, MD: American Society of Plant Physiologists. 2000; 1250-1318.
- 13. Ugochukwu, N.H., and Babady, N.E.(2002).

  A n t i o x i d a n t e f f e c t s o f

  Gongronemalatifoliumin hepatocytes of rat
  models of noninsulin dependent diabetes
  mellitus. Fitoterapia, 73:612-618.
- 14. Ugochukwu, N.H., Babady, N.E., Cobourne, M., and Gasset, S.R.(2003). The effect of Gongronemalatifoliumextracts on serum lipid profile and oxidative stress in hepatocytes of diabetic rats. J. Biosci., 20(1):1-5.
- 15. Cora, J.D., and Bruce, G.J. (2000). Review Phytochemicals: nutraceuticals and human health. *Journal of the Science of Food and Agriculture*;80: 1744-1756.
- Manas, K.M., Pratyusha, B., and Debjani, N. (2015). Phytochemicals—biomolecules for prevention and treatment of human diseases a review. *International Journal of Scientific and Engineering Research*, 3(7):1-32.
- 17. Wang, G., Tang, W., and Bidigare, R.R. (2005). Terpenoids as therapeutic drugs and pharmaceutical agents. *In* Natural products: Drug discoveries and therapeutic medicine. Zhang L. and Dermain A.L. (Eds), Humana Press, 197-227.
- 18. AOAC. Official methods of *analysis 20th Ed. (2016)*. Washington D.C.: Association of Official Analytical Chemists.
- 19. Pearson, W.R. (1996). Effective protein sequence comparison. Methods in

- *Enzymology;* 266: 227-258. Wiley online library visited 23 September, 2015.
- 20. Hilliard, C., Rigdon, A., Schroeder, W., Schroeder, C., and Flood, T. (2014). A fast, simple FET headspace GC-FID technique for determining residual solvents in C a n n a b i s c o n c e n t r a t e s , http://www.restek.com/Technical-Resources/Technical-Loibrary/Foods-Flavour.
- 21. Ehirim, F.N., and Onugha, F.C., and Agomuo, J.K. (2017). Effect of different processing methods on the chemical composition and organoleptic properties of A f r i c a n n u t m e g (Monodoramyristica). Journal of Food and Nutrition Sciences. 5(6): 232-235.
- Grabitske, H.A., and Slavin, J.L. (2016).
   Gastrointestinal effects of low-digestible carbohydrates. *Critical. Reviews in Food Science and Nutrition*. 49(4): 327-60
- 23. Ewaschuk, J.B., and Dieleman, L.A. (2006).

  Probiotics and prebiotics in chronicinflammatory bowel diseases. World

  J. Gastroenterol. 2006; 12(37): 5941-50
- 24. Ozcan, M. (2004). Mineral contents of some plants used as condiments in Turkey. *Food Chemistry*. 84:437–440.
- Sanchez-Castillo, C., Dewey, P., Aguirre, A., Lara J.J., Vaca. R., and Barra, P.L. (2008). The mineral content of Mexican fruits and vegetables. *Journal of Food Composition* and Analysis. 11: 340–356.
- 26. lewis, J.L (2008). Hormonal andmetabolic disorders electroly tebalance; overview of electrolytes. https://www.msdmanuals.com. Visited 12th September, 2018.
- 27. Jahnen-Dechent, W., and Ketteler, M. (2012). Magnesium basics. Feb;5(Suppl 1):i3-i14. doi:10.1093/ndtplus/sfr163
- 28. Clapham, D.E. (2007). CalciumsignalingCell. 131(6): 1047-1058. <a href="https://doi.org/10.1016/j.cell.2007.11">https://doi.org/10.1016/j.cell.2007.11</a>. <a href="https://doi.org/10.1016/j.cell.2007.11">028</a> visited August, 2018.
- 29. Tyszka-Czochara, M., Grzywacz, A., Gdula-Argasinka, J., Librowski, T., Wilinski, B., and Opoka W. (2014). The role of zinc in the pathogenesis and treatment of central nervous system (CNS) diseases implications of zinc homeostasis for proper CNS function. *ActaPoloniaePharmaceutica Drug Research*. 71(3): 369-377.

- 30. Muhammad, A., Dangoggo, S.M., Tsafe, A.I., Itodo, A.U., and Atiku, F.A. (2011). Proximate mineral and anti-nutritional factors of *Gardenia aqualla*(*Gautandutse*) fruit pulp. *Pakistani journal of Nutrition*. 10(6): 577-581.
- 31. Maxwell, C., and Volpe, S.L. (2007) Effect of zinc supplementation on thyroid hormone function. Annals of Nutrition and Metabolism 51:188-194. Doi: 1 0 . 1 1 5 9 / 0 0 0 1 0 3 3 2 4 https://www.karger.com/article/Abstract/10 3324
- 32. Shankar, A.H., and Prasad, A.S. (1998). Zinc and immune function: the biological basis of altered resistance to infection. *Am J ClinNutr.* 68 (suppl):447S-63S.
- 33. Tanumihardjo, S.A. (2011).Vitamin A: Biomarkers of nutrition for development. *The American Journal of Clinical Nutrition*. 94(2): 658S-65S.
- 34. Vunchi MA, Umar AN, King MA, Liman AA, Jeremiah G. and Aigbe, C.O. (2011). Proximate Vitamins and Minerals Composition of *Virtexdonianna* (black plum) Fruit Pulp. *Nigerian Journal of Basic and Applied Science*. 19(1): 97-101.
- 35. Smart, M. (1999). The vitamins: fundamental aspects in nutrition and health, 3<sup>rd</sup>ed. *The Canadian Vertinary Journal*. 40(11): 813-814.
- 36. Edeoga, H.O., and Eriata, D.O. (2001)Alkaloid,tannin and saponin contents of some Nigerian medicinal plants. *Journal of Medicinal and Aromatic Plant Science*. 23:244-249.
- 37. Ogukwe, C.E., Oguzie, E.E, Unaegbu, C., and Okolue, B.N. (2004). Phytochemical screening of the leaves of Sansevieratrifasciata. J. Chem. Soc. Nigeria. 29(1):8-10.
- 38. Okwu, D.E. (2006). Evaluation of the chemical composition of indigenous species and flavouring agents. *Global Journal of Pure and Applied Sciences*; 7(3): 455–459.
- 39. Rein, D., Paglieroni, J., Wun., Pearson, D.A., Schmhz, H.H., Gossenlin, R., and Keen, C.L. (2000). Cocoa inhibits platelet activation and function. *Am. J. Clin. Nutr.* 272: 30–35.
- 40. Ijeh, I.I., Njoku, O.U., and Ekenze, E.C.

- (2004). Medicinal evaluation of extracts of *Xylopiaaethipica* and *Ocimumgratissimum.J.* of Med. Arm. Plant Sc. 2004; 26: 41–49.
- 41. Perrella, J, Berco., Cecutt, A., and Bhavnani, B. (2003). Potential role of the interaction between equine estrogen low density lipoprotein (LDL) and high-density lipoprotein (HDL) in the prevention of coronary heart and neurodegenerative disease in postmenopausal women. *Lipid in Health Disease*, 2: 4.
- 42. Jiang, Z., Kempinski, C., and Chappell, J. (2016). Extraction and Analysis of Terpenes/Terpenoids. *CurrProtoc Plant Biol.*, 1:345–358.
- Kempinski, C., Jiang, Z., Bell., and Stephen,
   C.J. (2015).Metabolic Engineering of
   Higher Plants and Alage for Isoprenoid
   Production. Adv Biochem Eng Biotechnol.
   148:161–199.
- 44. Breitmaier, E. (2008). Terpenes: Flavors, Fragrances, Pharmaca, Pheromones. 1st Edition, Wiley-VCH Verlag GmbH & Co. KGaA, Germany.
- 45. Sarkic, A., and Stappen, I. (2018). Essential oils and their single compounds in cosmetics-a critical review. *Cosmetics*. 5, 11.
- 46. Cushnie T, Cushnie B, Lamb A. (2014). Alkaloids: An overview of their antibacterial, antibiotic-enhancing and antivirulence activities. Int J Antimicrob Agents, 44(5):377–386.
- 47. Lotfi, P, Yaghmael P., and Ebrahim-Habibi. (2015). Cymene and metformin treatment effect on biochemical parameters of male NMRI mice fed with high fat diet. *Journal of Diabetics and Metabolic Disorder.* 14:52.
- 48. Sun, J. (2007). D-limonene: Safety and clinical applications. *Alternative Medicine Review*. 12(3): 259-264.
- 49. Qiu, S., Sun, H., Zhang, A., Xu, H., Yan, G. and Han Y. (2014). Natural alkaloids: basic aspects, biological roles, and future perspectives. Chin J Nat Med., 2014;12(6):401–406
- 50. Paumgartten, F.J.R., De-carvalho, R.R., Souza, C.A.M., Madi, K. and Chaloud I. (1998). Study of the effects of β-myrcene on rat fertility and general reproductive performance. *Brazilian Journal of Medical*

- and Biological Research. 31: 966-965.
- 51. Faleyimu, O.I and Oluwalana, S.A (2008).

  Proximate Analysis of monodoraMyristica
  (African Nutmeg) in Ogun State Nigeria.

  World Journal of Biological Research
  001:2.
- Amponsah, N.A.A and Appiah, E.N. (2017). Nutritional and anti-nutrient profiles of some Ghanian spices. Cogent Food and Agriculture 3:1348185
- 53. Obiloma, A.A., Ogbonna, P.C., Okere, T.O., Adumanya, O.C.U. and Maduforo, A.N (2017). Chemical composition of Hippocratea Welwitchi: A common Indigenous spice consumed in South-Eastern Nigeria. *Journal of Dietitians Association of Nigeria* 8:131-135.