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ALUMINUM TOXICITY IN ACID SULFATE SOIL ALLEVIATED WITH BIOGENIC LIMING COMPOSITES OF BLOOD COCKLE SHELL AND PALM KERNEL SHELL

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ABSTRACT

Low crop yield from acid sulfate soil was caused by high concentrations of aluminum and iron in soil. Aluminum ion which cause root injury at root region inhibits the uptake of the nutrients by the root and lead to nutrient deficiency. Ground magnesium limestones can be applied to overcome this problem but incurred more production cost. Biogenic liming agent from blood cockle shell and/or palm kernel shell are more cost effective and provide the essential element i.e. calcium. This study evaluate phase-association of Al in acid sulfate soil remediated using biogenic composites liming material using selective sequential extraction analysis. Biogenic liming composites were characterized for pH, proximate analysis and elemental composition. High ratio of BCS in composite retained Al in the residual form about 91.33%. Amelioration of soil with 100% PKS did not give significant reduction of the exchangeable Al.

Keywords: soil remediation; phytotoxicity; abiotic stress; calcination; alkalinity.

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1. INTRODUCTION

Low yield production of crops is one of the main issues in the agricultural sector. A major factor which contributed towards this issue is related with abiotic stress. Abiotic stress can be initiated by the non-living agents such as soil pH conditions, imbalance of plant nutrients, air pollutants or water stress. Nutrient imbalance in plant is influenced by the relationships with soil pH conditions. Fig. 1 illustrates nutrient availability as a function of the soil pH values and some plant growth best at soil pH 5.5 to 6.5 [1]. The availability of toxic elements such as Al increased when soil pH value decreases. Such scenario may occur when the sulfide-rich soils are oxidized to create sulfuric acid in the soils. In Malaysia, coastal plain area such in Kedah-Perlis zones are facing with this condition related with acid sulfate soil.

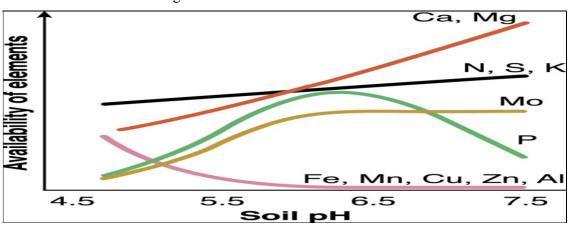


Fig.1. Effect of pH on nutrient availability [2]

Growth and yield of paddy at the granary area of Kedah-Perlis coastal plains are affected due to presence of acid sulfate soils [3]. Major area affected by acid sulfate soil is Merbok in the Sungai Petani district. Acid sulfate soil is a problematic soil, with low pH value (< 4) and high concentration of Al and Fe in soil. Aluminum toxicity severely affects the granary area in Merbok especially during the dry first cultivation season. Amelioration of Merbok soil has been carried out using lime [3], basalt [4] or calcium silicate [5]. Soil pH increased with amendment application and reduced Al toxicity.

The high availability of Al in the soil may cause root injury which inhibit the uptake of nutrient (i.e. phosphorus) by root. This condition leads to phosphorus deficiency of paddy. Deficiency of phosphorus can be traced by narrow, short and very erect dark green leaves [6]. Aluminum is strongly bound to insoluble inorganic matter and plant roots are not able to absorb this metal at

pH 5.5 or higher. However, Al in soil solution increase at pH < 5.5 [7] and is bioavailable that may be harmful to growth of plant [3].

Alleviation of aluminum phytotoxicity can be conducted by application of commercial liming materials (i.e. ground magnesium limestone [8, 3], basalt [4] or calcium silicate [5]. Commercial liming materials ameliorate the soil by increasing the pH and immobilize the heavy metals. Unfortunately, production of these commercial liming materials come from non-renewable source i.e. limestone mining quarry. Consequently, this may lead to depletion of the limestone, and may cause pollution to the air and water resources. As an alternative, waste BCS and palm oil milling biomass (i.e., PKS) can be manipulated as biogenic liming agents. BCS and PKS were selected as it is a highly abundant waste material. Addition of BCS and/or PKS may ameliorate soil acidity by providing calcium, magnesium or other nutrient element. Soil pH may be increased by addition of the liming materials and immobilized Al³⁺ as gibbsite (Al (OH) ₃). To the best of our knowledge, there has been no report on the phase-association of Al in the ameliorated acid sulfate soil.

The objectives of this study are (1) to evaluate the potential of using composites of BCS and PKS as liming material and (2) to investigate Al phase-association in the remediated acid sulfate soil using selective sequential extraction analysis.

2. EXPERIMENTAL

2.1. Feedstock Collection and Preparation

Blood cockle shell was collected at Sabak Bernam, Selangor. Cleaning of as-received blood cockle shell was carried out by boiling with deionized water for 2 hours. Then, the shell was oven-dried for 72 hours and pulverized to 75µm in particle diameter. Palm kernel shell from palm oil milling factory in Jengka, Pahang was sun-dried and oven-dried for 48 and 24 hours respectively. Palm kernel shell was pulverized to 75µm in particle diameter. Pulverized BCS and PKS were kept in polyethylene zip lock bags.

2.2. Combustion of Biogenic Composites Liming Material

Biogenic composites material of BCS and PKS were initially blended and homogenized at various weight ratios (i.e., 1:0, 1:1 and 0:1). About 10 grams of each composite were transferred into uncovered crucible, and were combusted at 900°C for 1 hour under ambient

condition. Composites ash were later cooled in the desiccator and kept in polyethylene zip lock bags.

2.3. Characterization of Biogenic Composites Liming Material

Two grams of samples (raw or ash) were mixed with 20 mL of 10 mM CaCl₂ and the pH values were determined using a calibrated pH meter. Proximate analysis was conducted with thermogravimetric analyzer (TGA/DSC 1 Star System, Mettler Toledo, USA) using 20 mg composites samples at 900°C peak temperature, 10°C/min of heating rate and the air flow rate was 50mL/min. Ultimate analysis was carried out using CHNS analyzer (Flash 2000 Organic Elemental Analysis, Thermo Scientific, USA). Elemental composition of biogenic composites liming material was determined using X-ray fluorescence (XRF) spectroscopy (Epsilon3-XL PANalytical, Netherlands).

2.4. Soil Collection

Sampling of acid sulfate soil was conducted at paddy field at Kampung Padang Tok Sura, Merbok, Kedah, Malaysia (5.711128N, 100.384416E). Soil samples were collected at 30cm interval, and to the depth of 0-15 cm (topsoil) using a soil auger. Soil sampling was conducted before paddy cultivation for first season (April-August 2016) and the condition was dry during this season.

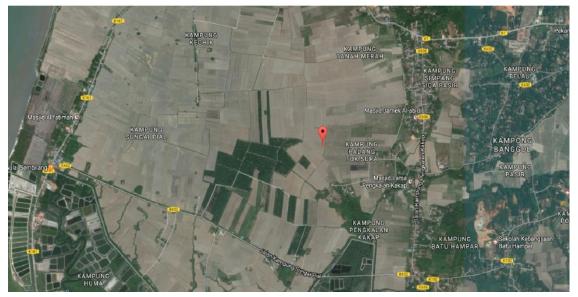


Fig.2. Soil sampling location at Kampung Padang Tok Sura, Merbok, Kedah, Malaysia (5.711128N, 100.384416E)

2.5. Soil Analyses

Samples of soil were air-dried, ground and passed through a 2 mm sieve prior to chemical analyses. Soil pH was determined on soil slurry at soil-solution (10 mM calcium chloride) ratio of 1:2.5 using a calibrated glass electrode pH meter. Soil particle distribution was determined using the hydrometer method. Cation exchange capacity (CEC) was determined using 1M NH₄OAc at pH 7 [9]. Soil organic matter was determined using loss-on-ignition method. Exchangeable Al was determined by extracting 5 gram of soil with 50 mL of 1 M KCl. The mixture shaken for 30 minutes and ICP-OES was used to analyze extracted Al.

2.6. Soil Incubation Study

Soil samples for incubation study were air-dried for 5 to 7 days, ground and sieved through 2 mm sieve (mesh no.10). Then, the soil samples were homogenized with biogenic composites liming materials with application rates of 2 g liming materials to 1 kg soil (4 Mg/ha) (Table 1). Initial and final pH was recorded during soil incubation period on day 0 and day 21 respectively. Soil incubation was carried out for 3 consecutive weeks (21 days).

Treatments	Application Rate		
Soil 100% (control)	No amendment		
Raw 0:1(BCS:PKS)	4 Mg/ha		
Raw 1:1(BCS:PKS)	4 Mg/ha		
Raw 1:0(BCS:PKS)	4 Mg/ha		
GML	4 Mg/ha		
CaCO ₃	4 Mg/ha		
900°C 0:1(BCS:PKS)	4 Mg/ha		
900°C 1:1(BCS:PKS)	4 Mg/ha		
900°C 1:0(BCS:PKS)	4 Mg/ha		

Table 1. Soil incubation study treatments and application rate

2.7. Selective Sequential Extraction (SSE) Analysis

Analysis of selective sequential extraction for soil samples were carried out after soil incubation period was completed. Five major fractions of soil Al: exchangeable, bound to carbonate, bound to manganese oxide, bound to organic matter and residual were determined [10-12].

2.7.1. Fraction 1: Exchangeable

One gram of soil samples were extracted at room temperature for 1 hour with 8 mL of magnesium chloride hexahydrate (1M MgCl₂.6H₂O, pH 7.0) with continuous agitation (end-to-end shaker, 35 rpm). Once the extraction was completed, the soil samples were centrifuged at 4000 rpm for 10 minutes. Supernatant from the samples were filtered by using 0.45µm syringe filter and deionized water were added about 8mL into the filtered supernatant.

2.7.2. Fraction 2: Bound to Carbonate

The residue from Fraction 1 was leached at room temperature with 8mL of 1M NaOAc (adjusted to pH 5.0 with 100% glacial acetic acid, HoAc). Continuous agitation (end-to-end shaker, 35 rpm) was maintained and the time necessary for complete extraction was determined to be 3 hours. Once the extraction was completed, the soil samples were centrifuged at 4000 rpm for 10 minutes. Supernatant from the samples were filtered by using 0.45µm syringe filter and deionized water were added about 8ml into the filtered supernatant.

2.7.3. Fraction 3: Bound to Iron and Manganese Oxide

Residue from Fraction 2 was extracted with 20mL of 0.04M NH₂OH.HCl in 25% (v/v) HoAc. This fraction experiment was performed at 95°C with occasional agitation for 6 hours. Once the extraction was completed, the soil samples were centrifuged at 4000 rpm for 10 minutes. Supernatant from the samples were filtered by using 0.45 μ m syringe filter.

2.7.4. Fraction 4: Bound to Organic Matter

Three milliliter 0.02M HNO₃ and 5 mL 30% H_2O_2 (pH adjusted with HNO₃ to pH 2) was added to the residue from Fraction 3 and the mixture was heated to 85 ± 2 °C for 2 hours with occasional agitation. After 2 hours, 3 mL 30% H_2O_2 (pH adjusted with HNO₃ to pH 2) were added and continue heating for another 2 hours at 85 ± 2 °C with occasional agitation. Once the samples were cooled down, add 5mL 3.2M NH₄OAc in 20% (v/v) HNO₃ (dilute to 20mL) and mixed with continuous agitation for 30 minutes. Once the extraction was completed, the soil samples were centrifuged at 4000 rpm for 10 minutes. Supernatant from the samples were filtered by using 0.45µm syringe filter.

2.7.5. Fraction 5: Residual

The residue from Fraction was digested with 9mL concentrated HCl and 3mL concentrated HNO₃ in a microwave digester according to EPA method 3051 and diluted 50:1 with $18m\Omega$ DI

water. Final dilution was filtered through a 0.45µm syringe filter and stored at 4°C. Once the entire sequential extraction was completed, the samples were analyzed using inductively couple plasma- optical emission spectrometer (ICP-OES) according to EPA method 6010B.

2.8. Statistical Analyses

All measurement in the characterization of composites and SSE analyses were conducted in duplicates, mean values and standard deviation were computed using Microsoft Excel for the data obtained from analysis.

3. RESULTS AND DISCUSSION

Table 2 shows the pH values of the biogenic liming material (raw and calcined). Increasing the content of BCS in the raw composites material causes the increasing pH values from 5.84 to 10.02. Calcination of biogenic liming material at 900°C also increased the pH with 900°C 1:1 has the highest pH compared to others composites.

	рН		
Composite Ratio (BCS:PKS)	PKS) Raw 90		
0:1	5.84	9.24	
1:1	6.40	13.23	
1:0	10.02	13.12	

Table 2. pH of biogenic liming material (raw and calcined)

Proximate and ultimate analysis of biogenic liming material is illustrated in Table 3 and 4 respectively. Pure BCS, (i.e., 1:0) generated more ash as compared to pure PKS (i.e., 0:1). Lignocellulosic component in PKS contains high percentage of carbon as compared to other composites material.

1:0

0.00

Moisture	Proxi Volatile	imate Analysis			
Moisture	Volatile	T * 1			
		Fixed	Ash Contents (%)		
Content (%	5) Matter (%	%) Carbon (%)			
4.83	21.89	47.11	26.17		
2.28	18.69	36.77	42.26		
0.50	0.00	31.67	68.33		
ble 4. Ultimate and	alysis of bioger	nic liming material			
Ultimate Analysis					
Nitrogen (%)	Carbon (%)	Hydrogen (%)	Sulfur (%)		
0.02	46.79	5.76	0.00		
0.00	11.60	0.01	0.00		
	2.28 0.50 ble 4. Ultimate and Nitrogen (%) 0.02	2.28 18.69 0.50 0.00 ble 4. Ultimate analysis of biogen Ultimate Nitrogen (%) 0.02 46.79	2.28 18.69 36.77 0.50 0.00 31.67 ble 4. Ultimate analysis of biogenic liming material Ultimate Analysis Nitrogen (%) 0.02 46.79		

Table 3. Proximate analysis of biogenic liming material

Table 5 illustrates elemental composition of biogenic liming materials from X-ray fluorescence (XRF) spectrophotometer. Calcined BCS (i.e., 900°C 1:0) contained the most calcium oxide (CaO) content at 61.4%. Silicon oxide (SiO₂) in of calcined PKS (i.e., 900°C 0:1) was highest as compared to other materials at 55.2%. Calcium oxide content increased as the BCS content in the composite increased. Calcination of biogenic liming composites at 900°C produced more calcium oxide as compared to raw composites due to liberation of carbon dioxide from calcium carbonate in the BCS.

11.63

0.13

0.00

BCS: PKS	Elemental Composition (%)					
Composites	CaO	K ₂ O	MgO	Al ₂ O ₃	Fe ₂ O ₃	SiO ₂
Raw 0:1	0.52	1.54	3.85	3.76	6.70	7.26
Raw 1:1	37.7	0.68	4.37	7.62	1.21	4.52
Raw 1:0	55.7	0.14	4.08	6.45	0.07	0.95
900°C 0:1	1.80	5.42	4.35	15.87	20.07	55.2
900°C 1:1	56.7	0.60	5.56	8.65	1.23	6.20
900°C 1:0	61.4	0.17	3.52	7.84	0.06	0.78

 Table 5. Biogenic composites liming materials elemental composition

Effect of commercial and raw biogenic composites liming materials on soil pH at the application rate of 2 g/kg soil is illustrated in Fig. 3. Raw 1:0 composite liming materials increased the initial soil pH (i.e., 3.14) to maximum of pH 6.48 on the 9th day of incubation. The trend of soil pH incubated with GML and calcium carbonate are similar for the first 9 days of incubation. Presence of CaO (55.7%) in the raw BCS helped increased soil pH and reduced soil acidity. Reduction of pH value after 9 days of incubation may due to following dissolution reaction:

(i) $CaCO_3+H_2O+CO_2 \rightarrow Ca^{2+}+2HCO_3$

(ii) MgCa (CO₃)₂ + 2H₂O+2CO₂ \longrightarrow Mg²⁺ + Ca²⁺ + 4HCO⁻₃[13]

Increased of H^+ in the soil samples in form of carbonic will reduced the pH value as showed in the Fig. 3.

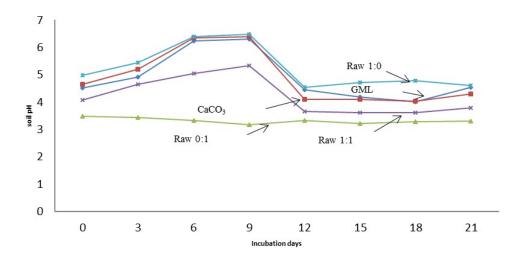


Fig.3. Effect of commercial and raw biogenic composites liming materials (2g/kg) on soil pH

Fig. 4 illustrated effect of calcined biogenic composites liming materials with application rate 2 gram/kilogram soil on soil pH. Calcined BCS (i.e., 900°C 1:0) increased soil pH to 6.67 and 6.64 on day 6 and day 9 of incubation, respectively. Content of CaO (i.e., 61.4%) in the calcined BCS (i.e., 900°C 1:0) has played a significant role in increasing the soil alkalinity. The pH value of soils ameliorated with 900°C 1:0 increased from day 0 to day 6, and decreased from day 12 onwards. The trend of soil pH for 900°C 1:0 showed similar trend as raw 1:0 samples. In contrast, soils amended with calcined composites of 50% BCS: 50% PKS (i.e., 900°C 1:1) has soil pH value with slightly changes for the entire duration of incubation period. Calcination of biogenic liming materials will make the calcium become less soluble.

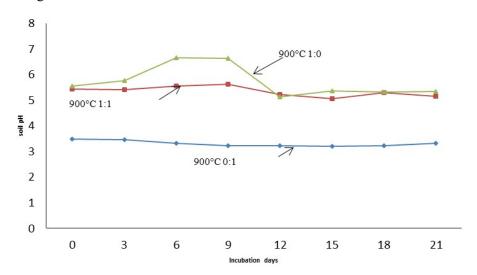
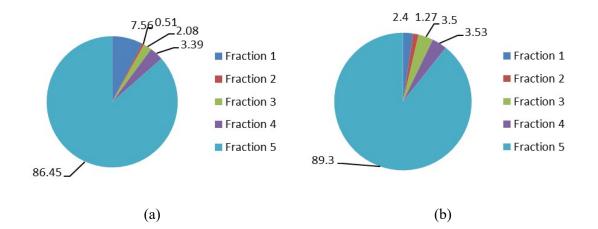


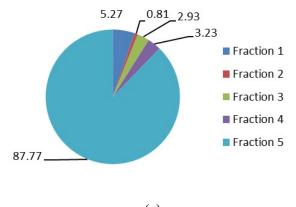
Fig.4. Effect of calcined biogenic composites liming materials (2g/kg) on soil pH

Aluminium distributions for soil incubated with commercial liming materials were analyzed using selective sequential extraction analysis and results were illustrated in Fig. 5. Soil without any treatment with commercial liming agent has highest percentage of exchangeable Al (7.56 %) as compared to other soil treatment. More Al was leached out from soil due to increasing mobility and solubility of Al. In contrast, less Al (2.4 %) was leached out from soil treated with GML, due to lower solubility and decreasing mobility of Al at higher soil pH [14].

Soil amended with GML has highest percentage of Al for Fraction 2 (Bound to carbonate) compared with another soil treatment with 1.27%. Treatment of soil using $CaCO_3$ has slightly lower percentage of Al for Fraction 2 (Bound to carbonate) with 0.81%.

Fraction 5 (Residual) showed that soil treated with GML has more residues of Al with 89.3%. Only about 10.7% of Al present in the soil reacted with previous four fractions of selective sequential extraction (Fraction 1-4). Control sample (soil 100%) has less residues of Al with 86.45%. More than 13.55% of Al present in the soil able to be extract out with previous four fractions of selective sequential extraction (Fraction 1-4).





(c)

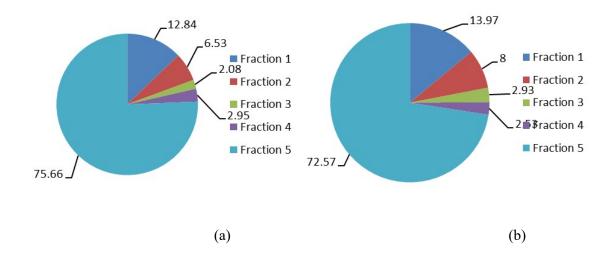
Fig.5. Aluminium distributions by Selective Sequential Extraction (SSE) analysis for soil incubated with commercial liming material (a) Soil 100% (b) GML (c) CaCO₃

Fig. 6 illustrates Al distribution for soil incubated with raw and calcined composites ash. Soil treated with raw 0:1 composite has highest percentage of exchangeable Al (12.84%) as compared to other soil treated with raw composites. More Al was leached out from soil treated with raw 0:1 composite due to increasing mobility and solubility of Al, possibly due to complexation with dissolved organic compounds from PKS [13]. In contrast, less Al (0.71%) was leached out from soil treated with raw 1:0 composite, due to lower solubility and decreasing mobility of Al at higher soil pH [14]. Highest percentage of exchangeable aluminium (13.97%) was detected from soil treated with 900°C 0:1 composite ash as compared to control sample (i.e. 7.56%). Aluminium mobility and solubility may leach out from PKS ash. This supported that the soil samples incubated with 900°C 0:1 composite ash were leached out more than the others soil samples. On the other hand, incubated soil with 900°C 1:0 composite ash has no significant contribution toward mobility of exchangeable Al at Fraction 1 with 0% of aluminium.

Raw 0:1 composite treated soil has highest percentage of Al for Fraction 2 (Bound to carbonate) compared with another soil treatment using raw composites with 6.53%. Present of carboxyl group from lignocellulosic compounds (lignin) in PKS possibly contributed toward high percentage bound to carbonate in Fraction 2 [15]. Other composites materials i.e. raw 1:0 and raw 1:1 contained calcium carbonate that may not react much with Al, due to stable form of carbonate. Similar condition occurs for both GML and calcium carbonate. Soil treated with 900°C 1:1 composite ash has highest percentage of carbonate-bound Al (Fraction 2)

compared with another soil treatment using calcined composites ash with 10.53%. Carbonation via atmospheric CO₂ absorption or lime addition during the soil incubation results in significant carbonate abundance in soil samples. According to [14], Al present in the soil can be influenced by the addition of lime and reacted with the carbonate present. Result obtained from this fraction differ from the Fraction 2 for raw composites materials due to calcination of materials at 900°C. Calcium carbonate will be converted to calcium oxide when calcined at higher temperature i.e. 900°C. Calcium oxide in 900°C 1:0 and 900°C 1:1 composite ash contribute toward high Al percentage bound to carbonate.

Fraction 5 (Residual) showed that soil treated with raw 1:0 composite has more residues of Al with 91.33%. Only about 8.67% of Al present in the soil reacted with previous four fractions of selective sequential extraction (Fraction 1-4). Raw 0:1 composites treated soil has less residues of Al with 75.66%. More than 24.34% of Al present in the soil able to be extract out with previous four fractions of selective sequential extraction (Fraction 1-4).



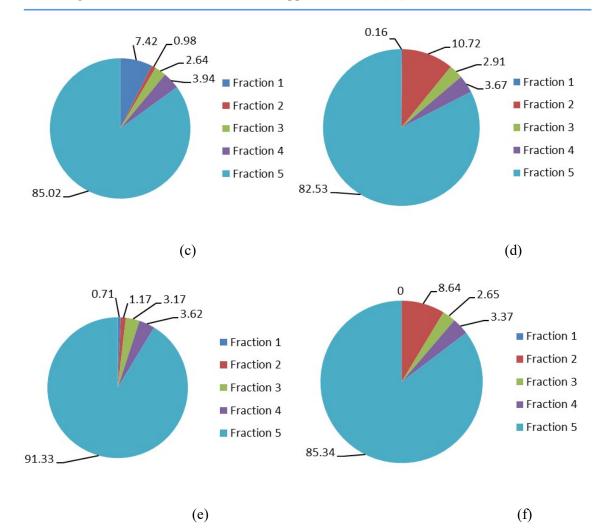


Fig.6. Aluminium distributions by Selective Sequential Extraction (SSE) analysis for raw and calcined composites ash (a) Raw 0:1 (b) 900 °C 0:1 (c) Raw 1:1 (d) 900 °C 1:1 (e) Raw 1:0 (f) 900 °C 1:0

4. CONCLUSION

Aluminum in the treated soil was immobilized by the addition of biogenic liming agent. Raw 1:0 and 900°C 1:0 composites liming material retained residual Al about 91.33% and 85.34% respectively. Raw 1:0 is recommended as biogenic liming agent to immobilized aluminum due to its capability to retain higher Al in the residual form as compared to other liming materials. Amelioration of soil with 100% palm kernel shell (PKS) did not result in significant outcome as more exchangeable aluminum leached out at Fraction 1 of selective sequential extraction (SSE).

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