

<u>https://dx.doi.org/10.4314/jpb.v16i2.15</u> Vol. 16 no. 2, pp. 195-204 (September 2019) <u>http://ajol.info/index.php/jpb</u> Journal of PHARMACY AND BIORESOURCES

Synergistic effect of novel aspirin analogues in a colorectal cancer cell line

Asma'u I.J. Bashir^{1*}, Christopher J. Perry² and Iain D. Nicholl²

¹Department of Pharmacology and Toxicology, Faculty of Pharmaceutical Sciences, Kaduna State University, Kaduna. Nigeria. ²Research Institute in Healthcare Science, University of Wolverhampton, Wolverhampton, WV1 1LY, UK.

Received 20th March 2019; Accepted 26th June 2019

Abstract

Studies show aspirin, a non-steroidal anti-inflammatory drug (NSAID) has potential to decrease incidence of, or mortality from a number of cancers including colorectal cancer (CRC). With emphasis on the treatment of CRC involving platinum compounds, oxaliplatin has been used in several combinations with other drugs. Unfortunately, these combinations do not improve overall survival and are accompanied with side effects that include gastrointestinal (GI), hematologic, neurologic toxicities, myopathy, and most recently interstitial lung disease, particularly fibrotic hypersensitivity pneumonitis, which has high mortality rates and long-term morbidity in survivors. The need of compounds that will reduce the doses of platinum compounds required for chemotherapy in order to reduce or alleviate these side effects is long overdue. This study investigates the synergistic effect if any, of novel aspirin analogues with platinum compounds cisplatin, oxaliplatin and carboplatin in order to lower doses needed of these platinum compounds and thus reduce or alleviate common debilitating side effects. MTT assay was used to assess cell viability and the CompuSyn software (Paramus, NJ, 2005) was used to calculate CI and DRI at ED₅₀, ED₇₅ and ED₉₀. Oxaliplatin was found to exhibit synergistic effects when combined with *p*-aspirin (PN549), diaspirin (PN508) and *o*-thioaspirin (PN590). Although further investigations such as *in vivo* experiments will be needed to draw any conclusions, this study is a stepping-stone for platinum compounds and aspirin drug combinations in order to decrease doses needed for treatment and thus lessen or alleviate debilitating side effects.

Keywords: Colorectal cancer; Aspirin; Aspirin analogues; Oxaliplatin; DMSO

INTRODUCTION

The discovery of a positive effect of aspirin, a very cheap drug on colorectal cancer (CRC) [1] was a stepping-stone for the study of aspirin on various cancers. Evidence later emerged that indeed the daily intake of aspirin reduced mortality [2], incidence and metastasis of CRC [3]. These discoveries geared studies carried out on the effects caused by aspirins in different cancers [4-6]. The cell line primarily used in this study is SW480 CRC cell line (Figure 1), which has been well characterized by [7] and more recently genetically characterized [8]. The cell line was isolated from the primary adenocarcinoma in the colon of a 50-year-old Caucasian male [7,8]. The cells are polygonal in shape with microvilli on their cell surfaces when viewed under an electron microscope [7] with an intermediate growth rate [8]. They are also known to be hyperdiploid and a low producer of carcinoembryonic antigen (CEA)

^{*} Corresponding author. *E-mail*: <u>asmau.junaidu@yahoo.com</u> *Tel*: +234 (0) 8100348071 ISSN 0189-8442 © 2019 Faculty of Pharmaceutical Sciences, University of Jos, Jos. Nigeria.

and are considered a good candidate for experimental studies in CRC research [7].

The SW480 cell line is DNA mismatch repair (MMR) proficient [9] and thus a good model for the study of changes involved in the progression of late CRC [10]. Semi quantitative analysis reveals the SW480 cell line express low levels to of cyclooxygenase (COX)-1 and undetectable levels of COX-2 [11,12], thus making it a good model for the study of mechanism of action of compounds that also follow COXindependent pathways. The cytotoxicity of aspirin and its analogues have been reported in various cancers [9,13,14].

The adaption of combined therapy using several active ingredients to produce a desired effect in the treatment of different diseases by both traditional and modern medical practitioners [15] dates back thousands of years when the Chinese and African herbalists used a combination of naturally occurring herbs to treat ailments [16,17]. One of the reasons behind the increased interest in the development of combination therapies can be attributed to the enhanced understanding that cancer, as a disease, involves the disruption of different molecular pathways, which are all connected and better tackled with the combined action of two or more drugs [18,19]. Another reason for the increased interest in combination therapy is the desire to achieve therapeutic effects at reduced doses, which will result in toxicity reduction and also delay or minimize the induction of drug resistance [20]. Drug combination therapy has increased in popularity for the treatment of complicated diseases such as cancer and AIDS [21].

Synergy can be defined as the greater effect for drugs in combination than in the simple additive effect produced by each drug individually. For example, 1+1 should be >2 for a synergistic effect while an antagonistic effect is less than an additive effect [15]. The term 'combination index' (CI) is used to quantitatively depict synergism (CI<1), additive effect (CI=1) or antagonism (CI>1) [22].

With emphasis on the treatment of involving platinum compounds, CRC oxaliplatin has been used in several combinations with other drugs. As far back as 1992, a study was published on the effects that resulted from the combination of oxaliplatin, folinic acid (leucovorin) and 5-FU with an objective response from 58% of the patients [23], later known as FOLFOX, which resulted in a high response rate in CRC patients [24,25]. Bevacizumab, a VEGF antagonist is also used in combination with FOLFOX in the treatment of metastatic CRC (mCRC) and has produced promising results [26]. EGFR antagonists, cetuximab or panitumumab have also produced improved response rates when used in combination with FOLFOX in patients harbouring the KRAS gene mutation [27]. Unfortunately, these combinations do not improve overall survival [26,28] and are accompanied with side effects gastrointestinal include that (GI). hematologic, neurologic toxicities and most recently interstitial lung disease, specifically fibrotic hypersensitivity pneumonitis, which has high mortality rates [29] and long-term morbidity in survivors [30]. Treatment with oxaliplatin also associated with is upregulation of myopathy-associated genes Foxo3, MAFbx and Bnip3 [31].

Oxaliplatin, as part of a neoadjuvant chemotherapy regimen is administered to CRC patients in order to shrink tumours surgery/main before treatment [24,32]. However, resistance to oxaliplatin is an encroaching menace especially in cancers that harbour a mutation of the TP53 gene of the tumour suppressor protein p53 [33]. P53 as a transcription factor acts as guard to the cell in response to stress signals and is responsible for the regulation of the cell cycle [34] and several metabolic enzymes, which are in turn responsible for drug metabolism [32,33]. For example, mutations found in the *TP53* gene affect cytochrome P450, an enzyme involved in the metabolism of drugs such as oxaliplatin [32,35], thereby leading to resistance.

Other platinum compounds, namely, cisplatin and carboplatin are also used in cancer chemotherapy and present with a range of long-term side effects such as GI toxicity, hepatotoxicity and late-term ototoxicity.

This study aims to establish whether aspirin analogues when used in combination with platinum compounds will have an effect in either a synergistic, additive or antagonistic manner.

Terminologies and plots used in the interpretation of drug combination results.

An important plot found in both the CalcuSyn and CompuSyn software is the Fa-dosereduction index (Fa-DRI) plot. As alluded before, one of the main reason for synergistic studies is to develop drug combinations with desired effects at reduced doses in order to decrease toxicity effects [21]. The dosereduction-index, DRI>1 indicate favourable while dose-reduction DRI<1 indicate unfavourable dose-reduction. For example, (Table 1), Fa=0.5 means at 50% inhibition of cell proliferation. For a 50% inhibition of cell growth, 3.7 μ M of drug 'A' and 4.2 μ M of 'B' are required individually. However, when combined, CompuSyn software calculates the DRI value of A and B as 3.2163 and 3.5924 respectively. Thus, 3.2163-fold less 'A' plus 3.5924-fold less 'B' is required to achieve the 50% inhibition same at the chosen combination ratio (i.e., $3.7260 \ \mu\text{M} \div 3.2163 =$ 1.2 μ M of drug 'A' plus 4.1938 μ M \div 3.5924 = $1.2 \mu M$ of drug 'B'). CI values are interpreted as descriptive words or by symbols (Table 2).

This paper aims to investigate the cytotoxic effects of novel aspirin analogues, which include the *meta-* and *para-* isomers of aspirin and thioaspirin. It is also aimed to study if their cytotoxic effect against SW480

CRC cell line will result in a synergistic effect when in combination with DNA-damaging platinum compounds known to be used as chemotherapeutic agents by use of the CalcuSyn® and CompuSyn® software.

EXPERIMENTAL

Tissue culture. The SW480 CRC cell line (ECACC, Salisbury, UK) was cultured in Leibovitz's L-15 medium (11415-049, gibco® life technologiesTM, ThermoFisher Scientific) supplemented with 10% (v/v) FBS and 1% (v/v) penicillin-streptomycin. The cells were cultured at 37°C in a humidified incubator and passaged at approximately 80% confluency.

Combination assay using MTT and CompuSyn software. For the combination assay, SW480 cells were plated in 96-well plates at a density of 500 cells/well/100µl of medium and allowed to set overnight at 37°C and then treated the next day with serial dilutions of cisplatin, oxaliplatin, carboplatin analogues 72 and aspirin for h [36,37,38,39,40].

Dissolution of platinum compounds, cisplatin, oxaliplatin and carboplatin with DMSO was avoided because DMSO reacts with platinum complexes thereby decreasing their cytotoxic effect [41]. It has been found out that the nucleophilic sulphur in DMSO reacts with platinum complexes thereby displacing their ligands, which result in a structure change, and thus making the compounds unstable in DMSO [42,43]. Due to these facts, cisplatin dissolved using PBS while both was oxaliplatin and carboplatin were dissolved using water [41]. Cell viability was measured by 3-(4,5-Dimethyl-2-thiazolyl)-2,5-diphenyl-2H-tetrazolium bromide (MTT) reduction assay [44] with modifications [45]. Combination drug treatments were performed by pairing cisplatin, oxaliplatin or carboplatin with each of the aspirin analogues in a constant ratio design based on their IC₅₀. A mixture of the two drugs was made at 2-fold and then serially diluted to 1-fold, 0.5-fold, 0.25-fold and 0.125-fold. After 24 h of seeding. the culture medium was then replaced with medium containing combinations of aspirin analogues and platinum compounds for 72 h. The medium was then aspirated from each well and cells once with fresh washed medium to completely remove drugs and replaced with 300 µl of MTT reagent (0.5 mg/ml). Cells were incubated at 37°C for 3 h after which time the medium was aspirated and replaced with 200 µl dimethyl sulfoxide (DMSO). The plates were further incubated at 37°C for 30 min at which time the conversion of MTT into formazan crystals by living cells was measured by recording changes in the absorbance at 540 nm in a microplate reader (Microplate Reader Thermo Multiskan Ascent 96 & 384). Plates were protected from light throughout this procedure. All assays were performed in duplicates, three independent times (n=3).

Doses and fraction of cells affected (Fa) of individual and combinations of compounds were fed into the CompuSyn software (Paramus, NJ., 2005), which calculated combination index (CI) and dose reduction index (DRI) at ED₅₀, ED₇₅ and ED₉₀. This produces multiple drug dose-effect calculations using the Median Effect methods described by Chou and Tatalay [22]. The CI is the quantitative measure of the degree of drug interaction in terms of additive effect (CI = 1), synergism (CI < 1), or antagonism (CI > 1) while the DRI is the measure of favourable dose reduction when two drugs are used in combination.

RESULTS

The standard curve for the MTT assay was determined to make sure that the concentration of formazan is directly proportional to the absorbance at 540 nm. Linear regression analysis shows "r-square" value as 0.99, which indicates the assay to be a good fit and thus rendering it ideal for purpose (Figure 2).

Platinum compounds in combination with aspirin analogues were used to treat SW480 CRC cells for 72 h. The cytotoxic effect and Fa were then calculated and fed into the CompuSyn software where the CI was determined at ED₅₀, ED₇₅ and ED₉₀. Against the colorectal cell line, SW480, synergistic effect was observed with cisplatin and *m*-aspirin [1:400] at ED₉₀; cisplatin and *p*aspirin [1:250] at ED₅₀, ED₇₅ and ED₉₀; cisplatin and diaspirin [1:17.5] at ED₅₀, ED₇₅ Oxaliplatin, ED90. the platinum and compound of interest showed synergistic effects with *p*-aspirin [1:100] and diaspirin [1:15] at ED₅₀ and ED₇₅. Carboplatin only had a favourable synergistic effect with *p*-aspirin [1:50] at ED₇₅ and ED₉₀ (Table 3).

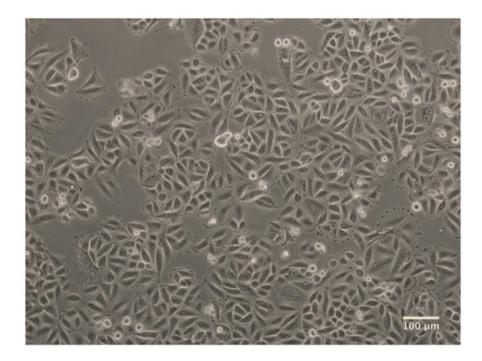


Figure 1. Morphology of SW480 CRC cell line (100X magnification).

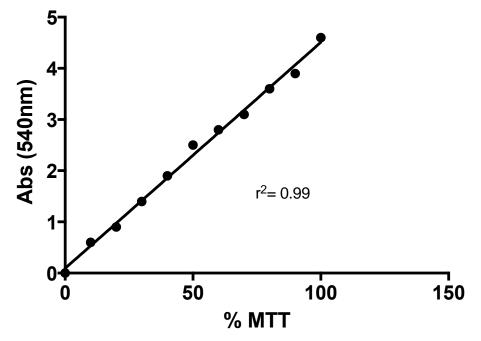


Figure 2. Standard curve for MTT solution.

Table 1. Example of a DRI Data for Drug Combination involving drugs A and B.

Fa	Dose A	Dose B	DRI A	DRI B
0.5	3.7260	4.1938	3.2163	3.5924

Table 2. Description and Interpretation of CI values [Adapted from (Bijnsdorp et al., 2011, Chou, 2006)].

Combination Graded		Interpretation	Simplified CI values and			
Index (CI)	Symbols		their interpretation			
< 0.1	+++++	very strong synergism	Synergism (+)			
0.1-0.3	++++	strong synergism				
0.3-0.7	+++	synergism				
0.7-0.85	++	moderate synergism				
0.85-0.9	+	slight synergism				
0.9-1.1	*	nearly additive	Additive (*)			
1.1-1.2	_	slight antagonism	Antagonism (-)			
1.2-1.45		moderate antagonism				
1.45-3.3		antagonism				
3.3-10		strong antagonism				
>10		very strong antagonism				

 Table 3. Summary of drug combination CI values in SW480 CRC cell line.

	Cisplatin (CP)		Oxaliplatin (OX)		Carboplatin (CB)				
Compounds	CI values at								
	ED ₅₀	ED ₇₅	ED ₉₀	ED_{50}	ED ₇₅	ED ₉₀	ED ₅₀	ED ₇₅	ED ₉₀
aspirin (Kilari et al., 2019)	+	+	+	+	+	+	-	-	-
<i>m</i> -aspirin (PN548)	*	*	+	*	*	-	*	-	-
<i>p</i> -aspirin (PN549)	+	+	+	+	+	*	*	+	+
diaspirin (PN508)	+	+	+	+	+	*	-	-	+
o-thioaspirin (PN590)	*	*	*	*	*	+	-	-	*
<i>m</i> -thioaspirin (PN591)	-	-	-	*	*	-	-	-	-
p-thioaspirin (PN592)	-	-	*	*	-	-	*	-	-

Drug combinations with CI values at effective dose for 50% kill of the cell population (ED_{50}), 75% of the population (ED_{75}) and 90% of the population (ED_{90})

DISCUSSION

The mutational status of TP53 gene in SW480 CRC [9,43] and p53 dependence for sensitivity to oxaliplatin indicate that cancers made up of such cell lines will present reduced sensitivity to platinum compounds and thus cytotoxicity reduced [32,40,46,47] in such cancers. Likewise, in CRC cell lines, p53 is essential for chemotherapy-induced cytotoxicity exhibited by oxaliplatin [32]. In addition, MMR proficient CRC cells are known to survive high doses of cisplatin or oxaliplatin [48] leading to treatment failure and a high incidence of debilitating side effects [29,31]. Sensitivity to these aspirin analogues clearly differs from cell type to cell type due to differences in specific targets by

different compounds [9]. Din et al. [9] compared sensitivity to aspirin in CRC cell lines to non-CRC cell lines and found out that the dose of aspirin used was inversely proportional to cell viability in CRC cells while non-CRC cells did not show this relationship. Thus, a compound that will increase sensitivity of SW480 CRC cells to oxaliplatin will lead to reduced dose needed and eventually less serious side effects. As previously reported, aspirin was found to have synergistic effects when combined with cisplatin and oxaliplatin, but not with carboplatin [49]. The combination of cisplatin and PN548 (meta-aspirin) [1:400] in SW480 CRC cells (Table 3) had synergistic effect at ED₉₀. In addition, the ED₅₀ for cisplatin in

this combination reduced by about 10-fold, which is favourable in cancer therapy [20]. The ED₅₀ of cisplatin also reduced by almost 10-fold when in combination with PN549 (para-aspirin) [1:250] accompanied by a synergistic effect at ED₅₀, ED₇₅ and ED₉₀ (Table 3). Cisplatin in combination with PN508 (diaspirin) [1:18] (Table 3) had synergistic effects in SW480 CRC cells with about a 2-fold decrease in ED₅₀ for cisplatin. In addition, the synergistic effects of these combinations increased at higher dose, which advantageous in chemotherapy [20]. is Cisplatin in combination with PN590 (orthothioaspirin) [1:20] had an additive effect in SW480 CRC cells but both of its isomers, PN591 (meta-thioaspirin) and PN592 (parathioaspirin) (Table 3) had an antagonistic effect when used in combination with all three platinum compounds. This antagonism between the isomers of thioaspirin and the platinum compounds could be because PN591 and PN592 have lower cytotoxic effects as compared to PN590 against this CRC cell line [50].

In this study, combinations of oxaliplatin with PN548 [1:80] (Table 3) had antagonistic effects at ED₇₅ and ED₉₀. Even though there was a synergistic effect at ED₅₀ and below, this is unfavourable in cancer therapy because maximum cytotoxicity against the cancer cells is key to effective therapy [20]. Oxaliplatin combination with PN548 [1:80] (Table 3) however resulted in a slight synergistic effect at ED₅₀ and ED₇₅ increasing to moderate synergism at ED₉₀ with a 4-fold decrease in oxaliplatin ED₅₀. The ED₅₀ for the platinum compound, oxaliplatin had a 7-fold decrease when combined with PN549 and PN590 with a synergistic effect (Table 3). Although there was a synergistic effect when oxaliplatin was combined with PN508 [1:14] at ED_{50} , the effect regressed to antagonism with an increase in dose (ED₇₅ and ED₉₀). With the prominent side effect of oxaliplatin being peripheral neuropathy [27], a reduction in its

 ED_{50} as a result of the combinations with aspirin [49], PN549 and PN590 may reduce or totally alleviate this side effect because a reduction in drug concentration will result in toxicity reduction and also delay or minimize the induction of drug resistance [20]. Although, there was what seemed to be a synergistic effect between carboplatin with PN508 at ED₅₀, the doses of carboplatin at ED₅₀ increased rather than decreased when used in combination with the diaspirin. This increase in ED₅₀ of carboplatin when in combination defeats one of the main reasons for combination therapy, which is to achieve a decrease in effective dose in order to reduce or alleviate side effects. Carboplatin, in combination with the thioaspirins, PN590, PN591 and PN592 also had strong synergistic effects (Table 3) against the colorectal cancer cell line. PN549 [1:50] (Table 3) was the only aspirin analogue that showed synergistic effects against SW480 CRC cell line when in combination with carboplatin. There was a 3fold decrease in carboplatin ED₅₀.

Combinations of aspirin and its analogues with platinum compounds have also been investigated using OE33 oesophageal cancer cell line [49]. However, these combinations should further be investigated in other cancer cell lines.

In the treatment of CRC, oxaliplatin has been used in combination with a variety of cytotoxic agents producing different levels of desired effects [27]. Due to oxaliplatin not effect on the having any drug enzyme, biotransformation P450. its combination with commonly used drugs such as aspirins does not raise any concerns [51].

In conclusion, oxaliplatin was found to exhibit synergistic effects when combined with para-aspirin (PN549), diaspirin (PN508) and ortho-thioaspirin (PN590). Some of these aspirin analogues thus show promising results when combined with platinum compounds for the treatment of cancer and should be further investigated *in vivo*.

REFERENCES

- 1. Kune, G. A.; Kune, S. and Watson, L. F. (1988); Colorectal cancer risk, chronic illnesses, operations, and medications: Case control results from the melbourne colorectal cancer study; *Cancer Res.* 48, 4399-4404.
- 2. Rothwell, P. M.; Fowkes, F. G.; Belch, J. F.; Ogawa, H.; Warlow, C. P. and Meade, T. W. (2011); Effect of daily aspirin on long-term risk of death due to cancer: Analysis of individual patient data from randomised trials; *Lancet.* 377, 31-41.
- 3. Algra, A. M. and Rothwell, P. M. (2012); Effects of regular aspirin on long-term cancer incidence and metastasis: A systematic comparison of evidence from observational studies versus randomised trials; *Lancet Oncol.* 13, 518-527.
- 4. Drew, D. A.; Cao, Y. and Chan, A. T. (2016); Aspirin and colorectal cancer: The promise of precision chemoprevention; *Nat Rev Cancer*. 16, 173-186.
- 5. Nan, H.; Hutter, C. M.; Lin, Y.; Jacobs, E. J.; Ulrich, C. M.; White, E.; Baron, J. A.; Berndt, S. I.; Brenner, H.; Butterbach, K.; Caan, B. J.; Campbell, P. T.; Carlson, C. S.; Casey, G.; Chang-Claude, J.; Chanock, S. J.; Cotterchio, M.; Duggan, D.; Figueiredo, J. C.; Fuchs, C. S.; Giovannucci, E. L.; Gong, J.; Haile, R. W.; Harrison, T. A.; Hayes, R. B.; Hoffmeister, M.; Hopper, J. L.; Hudson, T. J.; Jenkins, M. A.; Jiao, S.; Lindor, N. M.; Lemire, M.; Le Marchand, L.; Newcomb, P. A.; Ogino, S.; Pflugeisen, B. M.; Potter, J. D.; Qu, C.; Rosse, S. A.; Rudolph, A.; Schoen, R. E.; Schumacher, F. R.; Seminara, D.; Slattery, M .L.; Thibodeau, S. N.; Thomas, F.; Thornquist, M.; Warnick, G. S.; Zanke, B. W.; Gauderman, W. J.; Peters, U.; Hsu, L.; Chan, A.T.; Ccfr and Gecco (2015); Association of aspirin and nsaid use with risk of colorectal cancer according to genetic variants; JAMA. 313, 1133-1142.
- 6. Thorat, M. A. and Cuzick, J. (2015); Prophylactic use of aspirin: Systematic review of harms and approaches to mitigation in the general population; *Eur J Epidemiol.* 30, 5-18.
- Leibovitz, A.; Stinson, J. C.; McCombs, W. B., 3rd; McCoy, C. E.; Mazur, K. C. and Mabry, N. D. (1976); Classification of human colorectal adenocarcinoma cell lines; *Cancer Res.* 36, 4562-4569.
- Ahmed, D.; Eide, P. W.; Eilertsen, I. A.; Danielsen, S. A.; Eknaes, M.; Hektoen, M.; Lind, G. E. and Lothe, R. A. (2013); Epigenetic and genetic features of 24 colon cancer cell lines; *Oncogenesis*. 2, e71.

- 9. Din, F. V.; Dunlop, M. G. and Stark, L. A. (2004); Evidence for colorectal cancer cell specificity of aspirin effects on nf kappa b signalling and apoptosis; *Br J Cancer.* 91, 381-388.
- Hewitt, R. E.; McMarlin, A.; Kleiner, D.; Wersto, R.; Martin, P.; Tsokos, M.; Stamp, G. W. and Stetler-Stevenson, W. G. (2000); Validation of a model of colon cancer progression; *J Pathol.* 192, 446-454.
- 11. Li, M.; Wu, X. and Xu, X. C. (2001); Induction of apoptosis in colon cancer cells by cyclooxygenase-2 inhibitor ns398 through a cytochrome c-dependent pathway; *Clin Cancer Res.* 7, 1010-1016.
- 12. Richter, M.; Weiss, M.; Weinberger, I.; Furstenberger, G. and Marian, B. (2001); Growth inhibition and induction of apoptosis in colorectal tumor cells by cyclooxygenase inhibitors; *Carcinogenesis*. 22, 17-25.
- 13. Claudius, A. K.; Kankipati, C. S.; Kilari, R. S.; Hassan, S.; Guest, K.; Russell, S. T.; Perry, C. J.; Stark, L. A. and Nicholl, I. D. (2014); Identification of aspirin analogues that repress nf-kappab signalling and demonstrate anti-proliferative activity towards colorectal cancer in vitro and in vivo; *Oncol Rep.* 32, 1670-1680.
- 14. Gurpinar, E.; Grizzle, W. E. and Piazza, G. A. (2013); Cox-independent mechanisms of cancer chemoprevention by anti-inflammatory drugs; *Front Oncol.* 3, 181.
- 15. Foucquier, J. and Guedj, M. (2015); Analysis of drug combinations: Current methodological landscape; *Pharmacol Res Perspect.* 3, e00149.
- 16. Odugbemi, T. O.; Akinsulire, O. R.; Aibinu, I. E. and Fabeku, P. O. (2007); Medicinal plants useful for malaria therapy in okeigbo, ondo state, southwest Nigeria; *African Journal of Traditional Complementary and Alternative Medicines*. 4, 191-198.
- 17. Yuan, R. and Lin, Y. (2000); Traditional chinese medicine: An approach to scientific proof and clinical validation; *Pharmacol Ther.* 86, 191-198.
- Podolsky, S. H. and Greene, J. A. (2011); Combination drugs--hype, harm, and hope; *N Engl J Med.* 365, 488-491.
- 19. Zimmermann, G. R.; Lehar, J. and Keith, C. T. (2007); Multi-target therapeutics: When the whole is greater than the sum of the parts; *Drug Discov Today*. 12, 34-42.
- 20. Chou, T. C. (2010); Drug combination studies and their synergy quantification using the chou-talalay method; *Cancer Res.* 70, 440-446.

- 21. Chou, T. C. (2006); Theoretical basis, experimental design, and computerized simulation of synergism and antagonism in drug combination studies; *Pharmacol Rev.* 58, 621-681.
- 22. Chou, T. C. and Talalay, P. (1984); Quantitative analysis of dose-effect relationships: The combined effects of multiple drugs or enzyme inhibitors; *Adv Enzyme Regul.* 22, 27-55.
- 23. Levi, F.; Misset, J. L.; Brienza, S.; Adam, R.; Metzger, G.; Itzakhi, M.; Caussanel, J. P.; Kunstlinger, F.; Lecouturier, S.; Descorps-Declere, A. and et al. (1992); A chronopharmacologic phase ii clinical trial with 5-fluorouracil, folinic acid, and oxaliplatin using an ambulatory multichannel programmable pump. High antitumor effectiveness against metastatic colorectal cancer; *Cancer*. 69, 893-900.
- 24. Andre, T.; Boni, C.; Navarro, M.; Tabernero, J.; Hickish, T.; Topham, C.; Bonetti, A.; Clingan, P.; Bridgewater, J.; Rivera, F. and de Gramont, A. (2009); Improved overall survival with oxaliplatin, fluorouracil, and leucovorin as adjuvant treatment in stage ii or iii colon cancer in the mosaic trial; *J Clin Oncol.* 27, 3109-3116.
- 25. de Gramont, A.; Tournigand, C.; Louvet, C.; Andre, T.; Molitor, J. L.; Raymond, E.; Moreau, S.; Vignoud, J.; Le Bail, N. and Krulik, M. (1997); [oxaliplatin, folinic acid and 5-fluorouracil (folfox) in pretreated patients with metastatic advanced cancer. The gercod]; *Rev Med Interne*. 18, 769-775.
- 26. Saltz, L. B.; Clarke, S.; Diaz-Rubio, E.; Scheithauer, W.; Figer, A.; Wong, R.; Koski, S.; Lichinitser, M.; Yang, T.S.; Rivera, F.; Couture, F.; Sirzen, F. and Cassidy, J. (2008); Bevacizumab in combination with oxaliplatin-based chemotherapy as first-line therapy in metastatic colorectal cancer: A randomized phase iii study; *J Clin Oncol.* 26, 2013-2019.
- 27. Alcindor, T. and Beauger, N. (2011); Oxaliplatin: A review in the era of molecularly targeted therapy; *Curr Oncol.* 18, 18-25.
- 28. Bendell, J. C.; Hochster, H.; Hart, L. L.; Firdaus, I.; Mace, J. R.; McFarlane, J. J.; Kozloff, M.; Catenacci, D.; Hsu, J. J.; Hack, S. P.; Shames, D. S.; Phan, S. C.; Koeppen, H. and Cohn, A. L. (2017); A phase ii randomized trial (go27827) of first-line folfox plus bevacizumab with or without the met inhibitor onartuzumab in patients with metastatic colorectal cancer; *Oncologist.* 22, 264-271.
- 29. Chen, C. Y.; Chen, C. H.; Liao, W. C.; Liang, S. J. and Tu, C. Y. (2018); Fibrotic hypersentivity pneumonitis induced by oxaliplatin: A unexpected

but serious side effect in treating colorectal cancer; *Am J Resp and Crit Care Med.* 197, A6648.

- 30. Staff, N. P.; Grisold, A.; Grisold, W. and Windebank, A. J. (2017); Chemothearpy-induced peripheral neuropathy: A current review; *Ann Neurol.* 81, 772-781.
- Feather, C. E.; Lees, J. G.; Makker, P. G. S.; Goldstein, D.; Kwok, J. B.; Moalem-Taylor, G. and Polly, P. (2018); Oxaliplatin induces muscle loss and muscle-specific molecular changes in mice; *Muscle Nerve.* 57, 650-658.
- 32. Yang, C.; Zhou, Q.; Li, M.; Tong, X.; Sun, J.; Qing, Y.; Sun, L.; Yang, X.; Hu, X.; Jiang, J.; Yan, X.; He, L. and Wan, C. (2016); Upregulation of cyp2s1 by oxaliplatin is associated with p53 status in colorectal cancer cell lines; *Sci Rep.* 6, 33078.
- Lowe, S. W.; Ruley, H. E.; Jacks, T. and Housman, D. E. (1993); P53-dependent apoptosis modulates the cytotoxicity of anticancer agents; *Cell*. 74, 957-967.
- 34. Brown, C. J.; Lain, S.; Verma, C. S.; Fersht, A. R. and Lane, D. P. (2009); Awakening guardian angels: Drugging the p53 pathway; *Nat Rev Cancer.* 9, 862-873.
- 35. Lee, W.; Belkhiri, A.; Lockhart, A. C.; Merchant, N.; Glaeser, H.; Harris, E. I.; Washington, M. K.; Brunt, E. M.; Zaika, A.; Kim, R. B. and El-Rifai, W. (2008); Overexpression of oatp1b3 confers apoptotic resistance in colon cancer; *Cancer Res.* 68, 10315-10323.
- 36. Chan, M.; Gravel, M.; Bramoulle, A.; Bridon, G.; Avizonis, D.; Shore, G. C. and Roulston, A. (2014); Synergy between the nampt inhibitor gmx1777(8) and pemetrexed in non-small cell lung cancer cells is mediated by parp activation and enhanced nad consumption; *Cancer Res.* 74, 5948-5954.
- 37. Luo, H. Y.; Wei, W.; Shi, Y. X.; Chen, X. Q.; Li, Y. H.; Wang, F.; Qiu, M. Z.; Li, F. H.; Yan, S. L.; Zeng, M. S.; Huang, P. and Xu, R. H. (2010); Cetuximab enhances the effect of oxaliplatin on hypoxic gastric cancer cell lines; *Oncol Rep.* 23, 1735-1745.
- 38. Yan, K. H.; Yao, C. J.; Chang, H. Y.; Lai, G. M.; Cheng, A. L. and Chuang, S. E. (2010); The synergistic anticancer effect of troglitazone combined with aspirin causes cell cycle arrest and apoptosis in human lung cancer cells; *Mol Carcinog*. 49, 235-246.
- 39. Zhou, J.; Zhou, Y.; Yin, B.; Hao, W.; Zhao, L.; Ju, W. and Bai, C. (2010); 5-fluorouracil and oxaliplatin modify the expression profiles of micrornas in human colon cancer cells in vitro; *Oncol Rep.* 23, 121-128.

- 40. Toscano, F.; Parmentier, B.; Fajoui, Z. E.; Estornes, Y.; Chayvialle, J. A.; Saurin, J. C. and Abello, J. (2007); P53 dependent and independent sensitivity to oxaliplatin of colon cancer cells; *Biochem Pharmacol.* 74, 392-406.
- 41. Hall, M. D.; Telma, K. A.; Chang, K. E.; Lee, T. D.; Madigan, J. P.; Lloyd, J. R.; Goldlust, I. S.; Hoeschele, J. D. and Gottesman, M. M. (2014); Say no to dmso: Dimethylsulfoxide inactivates cisplatin, carboplatin, and other platinum complexes; *Cancer Res.* 74, 3913-3922.
- 42. Farrell, N.; Kiley, D. M.; Schmidt, W. and Hacker, M. P. (1990); Chemical-properties and antitumoractivity of complexes of platinum containing substituted sulfoxides [ptcl(r'r'so)(diamine)]no3 chirality and leaving-group ability of sulfoxide affecting biological-activity; *Inorganic Chemistry*. 29, 397-403.
- 43. Kerrison, S. J. S. and Sadler, P. J. (1985); Pt-195 nmr-studies of platinum(ii) dimethylsulfoxide complexes; *Inorganica Chimica Acta-Articles and Letters*. 104, 197-201.
- 44. Mosmann, T. (1983); Rapid colorimetric assay for cellular growth and survival: Application to proliferation and cytotoxicity assays; *J Immunol Methods*. 65, 55-63.
- 45. Carmichael, J.; DeGraff, W. G.; Gazdar, A. F.; Minna, J. D. and Mitchell, J. B. (1987); Evaluation of a tetrazolium-based semiautomated colorimetric assay: Assessment of chemosensitivity testing; *Cancer Res.* 47, 936-942.
- 46. Zhang, X.; Cheng, L.; Minn, K.; Madan, R.; Godwin, A. K.; Shridhar, V. and Chien, J. (2014);

Targeting of mutant p53-induced foxm1 with thiostrepton induces cytotoxicity and enhances carboplatin sensitivity in cancer cells; *Oncotarget*. 5, 11365-11380.

- 47. Perego, P.; Giarola, M.; Righetti, S. C.; Supino, R.; Caserini, C.; Delia, D.; Pierotti, M. A.; Miyashita, T.; Reed, J. C. and Zunino, F. (1996); Association between cisplatin resistance and mutation of p53 gene and reduced bax expression in ovarian carcinoma cell systems; *Cancer Res.* 56, 556-562.
- 48. Sergent, C.; Franco, N.; Chapusot, C.; Lizard-Nacol, S.; Isambert, N.; Correia, M. and Chauffert, B. (2002); Human colon cancer cells surviving high doses of cisplatin or oxaliplatin in vitro are not defective in DNA mismatch repair proteins; *Cancer Chemother Pharmacol.* 49, 445-452.
- 49. Kilari, R. S.; Bashir, A. I. J.; Devitt, A.; Perry, C. J.; Safrany, S. T. and Nicholl, I. D. (2019); The cytotoxicity and synergistic potential of aspirin and aspirin analogues towards oesophageal and colorectal cancer; *Curr Cin Pharmacology*. 14, 1-10.
- 50. Bashir, A. I. J.; Kankipati, C. S.; Jones, S.; Newman, R. M.; Safrany, S. T.; Perry, C. J. and Nicholl, I. D. (2019); A novel meachanism for the anticancer activity of aspirin and salicylates; *Int Journ Oncology*. 54, 1256-1270.
- 51. Masek, V.; Anzenbacherova, E.; Machova, M.; Brabec, V. and Anzenbacher, P. (2009); Interaction of antitumor platinum complexes with human liver microsomal cytochromes p450; *Anticancer Drugs*. 20, 305-311.