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Synthesis, characterization and cytotoxic activity of 5aminotetrazole Schiff bases

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Abstract

The incidence and prevalence of cancer has been on the rise in recent years and this has been linked to environmental factors, adoption of westernized lifestyle and aging populations. This situation is further complicated by the inadequacies of currently used anticancer agents such as toxicity and resistance which has prompted the search for new and more effective drugs. In this study, Schiff bases of 5-aminotetrazole were prepared by condensing 5-aminotetrazole with various aromatic aldehydes. Preliminary confirmation of compound formation was done using thin layer chromatography while FT-IR, UV-Visible, proton and carbon-13 NMR spectroscopy were used for structural characterization of the compounds. The cytotoxicity of the compounds was evaluated using the brine shrimp lethality assay. The compounds were synthesized in good yield ranging from 80 to 88 % and found to be soluble in polar solvents. Results of the spectroscopic analyses were indicative of the formation of the standard drug used, with the *o*-vanillin Schiff base (OVASB) showing the highest cytotoxicity (LC₅₀ 0.23 μ g/mL). The results indicate that the compounds have potential for further development in the search for safe and potent anticancer agents.

Keywords: Cancer; Schiff base; 5-Aminotetrazole; Aromatic aldehydes; Brine shrimp assay

INTRODUCTION

Cancer or malignant neoplasm is a group of diseases in which a group of cells display uncontrolled growth, invade adjacent cells and can even metastasize to distant sites [1]. It is a serious public health problem throughout the world and is the second leading cause of human death globally after cardiovascular diseases [2]. In the year 2018, cancer was reported to be responsible for about 9.6 million deaths, with approximately 70% of deaths due to the disease occurring in low- and middle-income countries [3]. In Nigeria in the same year, available statistics show that there were about 115,950 new cases of cancer with 70,327 deaths [4]. Currently, the main treatment options for cancer include radiotherapy, surgery, immunotherapy and chemotherapy [5]. Some of the classes of chemotherapeutic agents that are used in

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anticancer therapy include: alkylating agents (e.g. cyclophosphamide), antimetabolites (e.g. methotrexate), natural products (e.g. vincristine), hormonal agents (e.g. diethylstilbestrol), signal transduction inhibitors (e.g. lapatinib), anti-angiogenesis agents (e.g. sorafenib) and enzymes like Lasparaginase [6]. However, existing chemotherapeutic agents are limited by a number of factors such as severe and debilitating side effects and the development of resistance, usually after repeated use [7]. It is therefore important to develop new, safe and more effective drugs for treating cancer.

Schiff bases are aldehyde or ketone derivatives in which the carbonyl group is replaced by an imine or azomethine group [8]. They are widely used in industry and have also been found to exhibit a broad range of biological activities such as antifungal [9], antiinflammatory [10], Antileishmanial [11], antimicrobial [12–14], anti- cancer [15–17]. The tetrazole nucleus is well recognized in medicinal chemistry as a privileged scaffold due to its unique structural features and electronic properties [18]. As a result, this nucleus can be found in a large number of approved and marketed drugs such as antiviral, anti-inflammatory and anticonvulsants [19]. This nucleus has also been found to have anticancer activity [20, 21]. Furthermore, it has been shown that derivatization of certain molecules via Schiff base formation has the potential to boost their anticancer activity [22, 23] and it was therefore hypothesized that synthesis of Schiff bases of the tetrazole pharmacophore may produce a new class of molecules with enhanced cytotoxic activity and this formed the basis for this study. The therefore aimed present study is at synthesizing a number of 5-amino tetrazole Schiff bases and carrying out preliminary evaluation of their anticancer potential using the Brine shrimp lethality assay.

EXPERIMENTAL METHODS

Instruments. Melting points (°C) were Gallenkamp melting point recorded on apparatus and are uncorrected. Ultraviolet (UV) Spectra were recorded on Jenway 7315 Spectrometer, Fourier Transform Infra-Red (FTIR) Spectra were recorded on a Bruker Invenio-R spectrometer, Nuclear Magnetic Resonance (NMR) spectra were recorded on a Bruker Avance 400 spectrometer at frequencies of 400 MHz and 100 MHz for ¹H, ¹³C respectively.

Chemicals/ reagents. The reagents: 5aminotetrazole, 3-hydroxybenzaldehyde, *o*vanillin and benzaldehyde were obtained from Sigma Aldrich, Germany. The solvents (methanol, ethyl acetate, hexane, dichloromethane, absolute ethanol, glacial acetic acid, dimethyl sulfoxide) were all analytical grade, obtained from the same source and used without further purification. Thin layer chromatography (TLC) was done using aluminum backed, Silica gel 60A F₂₅₄ TLC plates, Fluorochem, United Kingdom.

Synthesis of the Schiff bases

Synthesis of 5-Aminotetrazole/o-vanillin Schiff Base. A 0.56-gram portion of 5aminotetrazole was weighed and dissolved in 60 mL of absolute ethanol in a refluxing flask. Then 1 gram of o-vanillin was dissolved in 40 mL of absolute ethanol and slowly added to the 5-aminotetrazole solution with continuous stirring. Glacial acetic acid (4 mL) was added to the resultant mixture and refluxed for 8 hours. Thereafter, the reaction mixture was cooled to room temperature, filtered, the precipitate dried and recrystallized from ethanol.

Synthesis of 5-aminotetrazole/3-hydroxy benzaldehyde Schiff base. A 0.697-gram sample of 5-aminotetrazole was weighed and dissolved in 60 mL of absolute ethanol in a refluxing flask. Then 1 gram of 3hydroxybenzaldehyde was dissolved in 40 mL of absolute ethanol and slowly added to the 5aminotetrazole solution with continuous stirring. Glacial acetic acid (4 mL) was added to the resultant mixture and refluxed for 8 hours. Thereafter, the reaction mixture was cooled to room temperature, filtered, the precipitate dried and recrystallized from ethanol.

Synthesis of 5-aminotetrazole/benzaldehyde Schiff base. A 0.697-gram sample of 5aminotetrazole was weighed and dissolved in 60 mL of absolute ethanol in a refluxing flask. Then 1 mL of benzaldehyde was dissolved in 40 mL of absolute ethanol and slowly added to the 5-aminotetrazole solution with continuous stirring. Glacial acetic acid (4 mL) was added to the resultant mixture and refluxed for 8 hours. Thereafter, the reaction mixture was cooled to room temperature, filtered, the precipitate dried and recrystallized from ethanol.

Solubility test. Qualitative solubility tests were carried out on the synthesized Schiff bases in a range of solvents with different polarities such as hexane, dichloromethane, ethyl acetate and methanol.

Spectroscopic characterization. The compounds were characterized using Ultraviolet (UV), Fourier transform infra-red (FTIR), Proton and Carbon-13 NMR spectroscopy.

Brine shrimp lethality assay. Brine shrimp cytotoxicity assay is widely used in the bioassay for the bioactive compounds. The assay was carried out according to the principle and protocol previously described [24]. The cytotoxicity screening of the extract was carried against a simple zoological organism, brine shrimp nauplii. *Artemia salina* Leach (brine shrimp) eggs were placed in a small tank containing 3.8% non-iodized NaCl solution (sea water) for two days to hatch the shrimp eggs and to be matured as nauplii. The compound (4.0 mg) was dissolved in DMSO (dimethyl sulfoxide). Then 100 µL of solution was taken into a test tube containing 5 mL of sea water and 10 shrimp nauplii. Thus, final concentration of the first test tube solution was 400 µg/mL. Then a series of solutions of varying concentrations was prepared from the stock solution by serial dilution. Thus, the concentrations of the obtained solution in each test tube were 400, 200, 100, 50, 25, 12.5, 6.25, 3.12 and 1.56 µg/mL for 9 dilutions. 1.0 mg of cyclophosphamide (served as the positive control) was dissolved in distilled water to get an initial concentration of 100µg/mL from which serial dilutions was made using sea water prepared to get 100, 50, 25, 12.5, 6.25, 3.12, 1.56, 0.781 and 0.391 µg/mL. The positive control solutions contained 10 living brine shrimp nauplii in 5 ml sea water. For negative control, 100 µl of DMSO was added to each of the pre-marked test tubes containing 5 ml of sea water and 10 shrimp nauplii. The vials were maintained under illumination. Survivors were counted after 24 hours and the percentage mortality at each vial and control was determined using the equation: % mortality = (no. of dead nauplii/ initial no. of live nauplii) x 100. The concentrationmortality data was analyzed statistically by using probit analysis for the determination of LC₅₀ values and linear regression for the fractions and LC_{50} values less than 100 ppm (or $100 \,\mu g/mL$) was considered significant and toxic.

RESULTS

Some physico-chemical properties of the Schiff bases including their melting points, rf values and yield obtained after synthesis and purification are displayed in Table 1 while Table 2 shows the result of the solubility assessment of the compounds in various solvents. Table 3 shows the results of the brine shrimp lethality assay displayed as median lethal concentration (LC₅₀) of the compounds. The spectroscopic data for the compounds are also displayed below.



Scheme 1: Synthesis of 5-Aminotetrazole/o-Vanillin Schiff Base



Scheme 2: Synthesis of 5-Aminotetrazole//3-hydroxybenzaldehyde Schiff base



Scheme 3: Synthesis of 5-Aminotetrazole//benzaldehyde Schiff base

Table 1: Physico-chemical properties of the Schiff bases					
cular Formula Me	elting Point (° C)	% Yield	Rf value		
N ₅ O ₂ 180	0 - 182	88 %	0.76		
N ₅ O 171	1 – 173	80 %	0.74		
N ₅ 202	2 - 203	82 %	0.75		
	$\begin{array}{c c} \text{scular Formula} & \text{Me}\\ \text{Scular Formula} & \text{Me}\\ \text{N}_5\text{O}_2 & 18\\ \text{N}_5\text{O} & 17\\ \text{N}_5 & 20\\ \end{array}$		$\begin{tabular}{ c c c c c c c c c c c c c c c c c c c$		

OVASB = 5-aminotetrazole / o-vanillin Schiff Base, HBSB = 5-aminotetrazole / 3-hydroxybenzaldehyde Schiff Base, BZSB = 5-aminotetrazole / benzaldehyde Schiff Base. TLC Solvent system: Petroleum Ether: Ethanol (1:1).

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Solvent	OVASB	HBSB	BZSB
Methanol	Soluble	Soluble	Soluble
Dichloromethane	Insoluble	Insoluble	Insoluble
Ethyl acetate	Insoluble	Insoluble	Insoluble
Hexane	Insoluble	Insoluble	Insoluble

Spectroscopic data for the Schiff bases

2-methoxy-6-{[(1H-tetrazol-5-yl)imino]methyl}phenol (OVASB)

UV-Vis (MeOH) λ_{max} : 245 nm. FTIR (neat) ν (cm⁻¹): 3384 (O–H str.), 3190 (N-H str.), 2937 (C-H str.-saturated), 1666 (C=N str.), 1587 (C=C str.), 1375 (N=N str.), 1263 (C-N str.). ¹H NMR (400 MHz, MeOD) : δ (ppm) 9.78 (1H, d), 7.46 (1H, t), 6.97 (1H, d), 3.95 (3H, s).

¹³C NMR (100 MHz, MeOD): δ (ppm) 193.04, 192.99, 154.73, 149.73, 130.73, 127.94, 116.37, 111.45, 56.48.

3-{[(1H-tetrazol-5-yl)imino]methyl}phenol (HBSB)

 $\begin{array}{l} \text{UV-Vis} \ (\text{MeOH}) \ \lambda_{\text{max}}: 245 \ \text{nm}. \\ \text{FTIR} \ (\text{neat}) \ \nu \ (\text{cm}^{-1}): \ 3339 \ (\text{O}-\text{H} \ \text{str.}), \ 3190 \ (\text{N}-\text{H} \ \text{str.}), \ 2959 \ (\text{C}-\text{H} \ \text{str.-saturated.}), \ 1666 \ (\text{C}=\text{N} \ \text{str.}), \ 1580 \ (\text{C}=\text{C} \ \text{str.}), \ 1356 \ (\text{N}=\text{N} \ \text{str.}), \ 1282 \ (\text{C}-\text{N} \ \text{str.}). \\ \ ^{1}\text{H} \ \text{NMR} \ (400 \ \text{MHz}, \ \text{MeOD}): \ \delta \ (\text{ppm}) \ 9.91 \ (1\text{H}, \ \text{s}), \ 7.40 \ (1\text{H}, \ \text{m}), \ 7.31 \ (1\text{H}, \ \text{d}), \ 7.11 \ (1\text{H}, \ \text{d}). \end{array}$

¹³C NMR (100 MHz, MeOD): δ (ppm) 194.41, 194.35, 159.45, 139.47, 131.30, 123.03, 122.84, 115.86.

1-phenyl-N-(1H-tetrazol-5-yl)methanimine (BZSB)

UV-Vis (MeOH) λ_{max}: 240 nm.

FTIR (neat) ν (cm⁻¹): 3190 (N-H str.), 3071 (C–H str.-unsaturated), 2929 (C-H str.-saturated.), 1640 (C=N str.), 1602 (C=C str.), 1319 (N=N str.), 1203 (C-N str.).

¹H NMR (400 MHz, MeOD) : δ (ppm) 8.04 (1H, d), 7.61 (2H, t), 7.48 (2H, t), 6.33 (1H, s).

¹³C NMR (100 MHz, MeOD): δ (ppm) 170.11, 158.24, 134.09, 131.96, 130.73, 130.22, 129.49, 129.17.

Table 3: Results of brine shrimp lethality assay		
Compound	LC ₅₀ (µg/mL)	
OVASB	0.23 ± 0.15	
HBSB	0.59 ± 0.16	
BZSB	0.33 ± 0.21	
Cyclophosphamide (Standard)	0.09 ± 0.02	

The LC_{50} values are expressed as mean \pm SEM, n = 10 in each group and were determined using Finney Probit analysis via SPSS 17.0 software.

DISCUSSION

The Schiff bases were obtained in good yield ranging from 80 to 88 % (Table 1) and this yield is higher than that obtained in a previous study (17 - 87%) where antimicrobial Schiff bases of 5-amino tetrazole were prepared [14]. The Schiff bases were found to be poorly soluble in non-polar solvents but soluble in polar solvents such as methanol (Table 2) and this is a useful property with respect to their distribution in vivo since the physiological milieu is mostly made up of water. However, for a compound to have good bioavailability, it must possess reasonable between hydrophilicity balance and lipophilicity and this may imply that the compounds may need to be further optimized to improve their lipohilicity.

The spectroscopic data prove that the compounds were successfully synthesized and the results are in agreement with published

data for the component molecules [25,26]. Also, occurrence of the key transformation which involved reaction between the aldehyde and amine functional groups to form the imine was backed up by the spectroscopic data. For instance, the o-vanillin Schiff base of 5aminotetrazole (OVASB) showed maximum UV absorption at a wavelength of 245 nm. Ordinarily, the electronic spectrum of unsubstituted tetrazole is characterized by an absorption band with a maximum in the vacuum UV region (λ_{max} below 200 nm). However, conjugation of the tetrazole nucleus with the benzene ring of o-vanillin led to a considerable red shift of the maximum into the normal UV range. In the FTIR spectrum of the compound, the Carbon-Nitrogen stretching band of the newly formed imine bond can be seen at 1666 cm⁻¹. It was also observed that the strong carbonyl stretching band of the aldehyde usually seen around 1700 cm⁻¹

together with the two signals around 2700 and 2800 cm⁻¹ of the aldehydic C-H and the two sharp spikes of the primary amine signal at about 3300cm⁻¹ had all disappeared and these observations are indicative of the formation of the new Schiff base. Some other bands for the compound that can be seen in the IR spectrum include the broad O-H stretch band at 3384 cm⁻ ¹ and the C=C stretch at 1587 cm⁻¹ and the C-H stretch aliphatic at 2937 cm⁻¹. In the ¹H-NMR, a signal at 9.78 ppm as a doublet belongs to the aromatic proton at position 4 of the aromatic ring and it has experienced a strong downfield shift from the usual values due to the deshielding effect arising from its close proximity to the strongly electronegative oxygen atom of the methyl ether substituent at position 3 of the aromatic ring while the triplet at 7.46 ppm can be attributed to the aromatic proton at position 5 of the aromatic ring. The doublet at 6.97 is attributable to the aromatic proton at position 6 while the singlet at 3.95 is attributable to the three protons of the methyl ether substituent at position 3 of the aromatic ring. The imine and phenolic protons of the compound are exchangeable and since the compounds were characterized using a protic deuterated solvent (CD₃OD), the NH and OH protons were exchanged with deuterium and the peaks disappeared completely and were not seen on the spectrum. In the ¹³C-NMR, the signal at 193.04 ppm belongs to the imine carbon (C=N) while the signal at 192.99 is due to the single Carbon atom of the tetrazole ring. The signals at 154.72 and 149.73 are due to the oxygen bearing Carbon atoms of the aromatic rings i.e. Carbon 2 and 3 respectively. The resonances for the other unsubstituted aromatic carbon atoms can be seen at 130.73, 127.94, 116.37 and 111.45 ppm while the methyl carbon atom of the methyl ether substituent at position 3 of the aromatic ring gave a signal at the most upfield position of 56.48 ppm. Similar assignments were done for the other compounds (HBSB and BZSB) which are structurally similar to OVASB with slight

differences in substitution around the aromatic ring.

The results of the Brine shrimp lethality assay shown in table 3 indicate that all the compounds possess cytotoxic activity with LC₅₀ values in the range $0.23 - 0.59 \,\mu$ g/mL. Of all the compounds, the o-vanillin Schiff base (OVASB) was found to be the most active having the lowest LC₅₀ value of 0.23 μ g/mL. In exploring the structure-activity relationships among this group of Schiff bases, it was observed that the o-vanillin Schiff base which possessed multiple substitutions on the benzene ring was the most active. It possesses a hydroxyl group at the ortho position and a methoxy group at the meta position. It is believed that the methoxy group at the meta position exerts an electron withdrawing effect thereby reducing the electron density in the aromatic system which may be beneficial for cytotoxic activity. The hydroxyl group at the ortho position is electron donating but its effect may be moderated by steric effects due to its closeness to the imine substitution on the Benzene ring. The trend mentioned above is further supported by the observation that removal of the methoxy group and moving the hydroxyl group to the meta position which enhances the electron density in the Benzene ring is not beneficial and results in a decline in cytotoxic activity as seen by the higher LC_{50} of the HBSB Schiff base. When the Benzene ring is unsubstituted as seen in BZSB, the cytotoxicity is intermediate and the conclusion that can be reached from the above substitution is that for this class of Schiff bases. introduction of electron withdrawing substituents on the aromatic ring will lead to enhanced cytotoxic activity. It was further observed that all the compounds were less cytotoxic when compared to the standard anticancer agent cyclophosphamide and this implies that there is still room for further structural modification to improve their potency and activity.

Conclusion. The synthesis and cytotoxicity evaluation of several 5-aminotetrazole Schiff bases has been reported. The compounds showed moderate cytotoxic activity with the *o*vanillin Schiff base having the highest activity. The structure-activity relationships (SAR) within the class of compounds were also examined in order to provide a guide for the systematic design of more active candidates.

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